

The mechanism of ions in pulmonary hypertension

Guogu Liu^{1,2}, Daiyan Fu², Heshen Tian^{1,2}  and Aiguo Dai³

¹Department of Graduate School, University of South China, Hengyang, China; ²Department of Respiratory Medicine, Hunan Provincial People's Hospital, Changsha, China; ³Department of Respiratory Diseases, Hunan University of Chinese Medicine, Changsha, China

Abstract

Pulmonary hypertension (PH) is a kind of hemodynamic and pathophysiological state, in which the pulmonary artery pressure (PAP) rises above a certain threshold. The main pathological manifestation is pulmonary vasoconstriction and remodelling progressively. More and more studies have found that ions play a major role in the pathogenesis of PH. Many vasoactive substances, inflammatory mediators, transcription-inducing factors, apoptosis mediators, redox substances and translation modifiers can control the concentration of ions inside and outside the cell by regulating the activity of ion channels, which can regulate vascular contraction, cell proliferation, migration, apoptosis, inflammation and other functions. We all know that there are no effective drugs to treat PH. Ions are involved in the occurrence and development of PH, so it is necessary to clarify the mechanism of ions in PH as a therapeutic target for PH. The main ions involved in PH are calcium ion (Ca^{2+}), potassium ion (K^+), sodium ion (Na^+) and chloride ion (Cl^-). Here, we mainly discuss the distribution of these ions and their channels in pulmonary arteries and their role in the pathogenesis of PH.

Keywords

Ca^{2+} , K^+ , Na^+ , Cl^- , PH

Date received: 2 October 2020; accepted: 23 December 2020

Pulmonary Circulation 2021; 11(1) 1–20

DOI: 10.1177/2045894020987948

Introduction

Considering that the increase of cardiac output and pulmonary artery wedge pressure will also affect the mean pulmonary artery pressure (mPAP), the sixth World Health Organization newly defined pulmonary hypertension (PH) as the $\text{mPAP} \geq 20$ mmHg detected by right cardiac catheterization and pulmonary vascular resistance ≥ 3 Wood Units (WU) with precapillary morphology.^{1,2} At present, according to pathology, hemodynamics and clinical diagnosis and treatment, PH is divided into five categories: (1) arterial PH, (2) PH caused by left heart disease, (3) PH caused by lung disease or/and hypoxia, (4) chronic thromboembolism PH, (5) PH caused by unidentified multifactorial mechanism. At the *Sixth World Health Organization Symposium*, it was proposed that long-term responses to calcium channel blockers should be included in the first group of PH because these patients had a special prognosis and treatment, and a continuous relationship between arterial, capillary and pulmonary capillary angiomas was found.¹ The main clinical manifestations of PH are progressive shortness of breath, decreased exercise endurance, chest tightness,

palpitation, and related manifestations caused by right heart failure, etc. The main mechanism of PH is pulmonary artery contraction and remodelling, which leads to the increase of pulmonary artery resistance. At present, the treatment of PH is based primarily on vascular tension and pulmonary vascular remodelling.^{2,3} From the past to the present research results, it can be known that ions play an important role in the pathogenesis of PH. The regulation of ion concentration inside and outside the cell mainly depends on the opening or closing of ion channels. The pathogenesis of PH involves a variety of substances, and finally take the ion channel as the target to achieve the corresponding function by controlling the ion concentration. It can be observed that the ions are at the center of different signal pathways. As a common and frequently occurring disease, there is no effective treatment for PH,

Corresponding author:

Aiguo Dai, Department of Respiratory Diseases, Hunan University of Chinese Medicine, Changsha, Hunan, China.

Email: daiaguo2003@163.com



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

© The Author(s) 2021
Article reuse guidelines:
sagepub.com/journals-permissions
journals.sagepub.com/home/pul



and the disability rate and fatality rate are very high. From the point of view of ions, it is necessary to clarify the distribution and mechanism of ions and their corresponding ion channels in PH, so as to develop more effective drugs for the treatment of PH.

Ca^{2+}

As one of the indispensable ions in many physiological functions of human body, Ca^{2+} participates in contraction and relaxation of muscle tissue, cell proliferation, stability of biological potential inside and outside the cell membrane, neuromuscular signal transduction, coagulation, regulation of enzyme activity, etc.⁴ Free Ca^{2+} is mainly distributed outside the cell, and a small part of it is distributed in the cell. About 90% of the intracellular calcium ions are stored in the endoplasmic reticulum (ER) and mitochondria, also known as stored calcium. Calcium channels regulate the distribution of calcium ions inside and outside the cell and coordinate various physiological functions. Ca^{2+} plays an important role in the pathogenesis of PH. Different types of calcium channels are distributed in pulmonary vessels. These calcium channels regulate pulmonary vasoconstriction and remodelling alone or interacting with each other, thus taking part in the formation of PH. The main calcium channels are store-operated calcium channels (SOCCs), receptor-operated calcium channels (ROCCs) and voltage-dependent calcium channels (VDCCs). The following describes the mechanism of Ca^{2+} release-activated Ca^{2+} channel (CRAC), inositol 1,4,5-triphosphate receptors (IP3Rs), transient receptor potential channel (TRPCs), L-type calcium channels (LTCCs) in PH.

CRAC. CRAC is the highest channel in SOCCs, which is found in the electrophysiological study of mast cells and T lymphocytes.^{5,6} CRAC regulates the store-operated calcium entry (SOCE). Stromal interaction molecule (STIM) and Orai cationic channels consist of the CRAC. STIM is an ER Ca^{2+} receptor. In mammals, STIM subtypes can be separated into STIM1 and STIM2. Orai is the storage operation channel, which is divided into three subtypes of Orai1, Orai2, Orai3.⁷ Here, we equate SOCCs with CRAC. In the inactivated state, STIM is dispersed in the ER and ORAIL is dispersed in the plasma membrane (PM). When G protein-coupled receptor (GPCR) activates phospholipase C (PLC), PLC decomposes phosphorylated lipid phosphatidylinositol 4,5-bisphosphate (PIP2) into diacylglycerol (DAG) and inositol 1,4,5-trisphosphate (IP3), IP3 acts on the corresponding target on ER, triggering the release of Ca^{2+} in ER, resulting in a decrease in the concentration of stored calcium ions. When STIM perceives Ca^{2+} depletion in ER, it aggregates between ER and PM, binds Orai and activates Orai pore-forming channels to promote Ca^{2+} into ER filling calcium pool (Fig. 1). Through the feedback mechanism, after dozens of milliseconds of Ca^{2+} , STIM and ORAI are separated, and then CRAC is inactivated.⁸⁻¹¹

SOCC exists widely in different cells, regulating vascular tension, blood pressure, immunity, nerve, muscle, secretion, gene transcription and translation, etc.^{12,13} SOCC plays an important role in cardiovascular regulation. It was found that inhibition of SOCC could inhibit the increase of intracellular Ca^{2+} concentration ($[Ca^{2+}]_i$) induced by endothelin-1 (ET-1) and angiotensin II (Ang II), reduce systemic blood pressure (including systolic and diastolic blood pressure) and inhibit the proliferation of vascular smooth muscle cells (VSMCs).¹³ Conversely, ET-1 can also regulate SOCC. It had been found that in the model of coronary artery ischemia-reperfusion injury, ET-1 induced vasoconstriction requires activation of both SOCC and LTCCs.¹⁴ In the pathogenesis of PH, it had been observed that the expression of STIM2 was up-regulated in pulmonary artery smooth muscle cells (PASM) of patients with PH. STIM2 could up-regulate $[Ca^{2+}]_i$, and promote the proliferation of PASM by up-regulating cell proliferation-related signal molecules such as CREB, STAT3, AKT, NFAT, Ki67, etc., and up-regulate the expression of Bcl-2 (an anti-apoptotic protein) to inhibit PASMcs apoptosis. Using siRNA to down-regulate the expression of STIM2 in PASMcs can reduce the expression of the above-mentioned cell proliferation signal molecules and anti-apoptosis proteins.¹⁵ SOCC not only promotes the contraction and proliferation of PASM, but also inhibits its apoptosis. SOCC causes pulmonary artery contraction and remodelling in many ways, which is an important mechanism for the formation of PH.

IP3R. IP3R is a channel made up of subunits of 4-260 kDa, which belongs to ROCC.¹⁶ IP3R is mainly distributed in ER and releases stored calcium in a short oscillatory manner, which is the main channel of calcium release in VSMCs.¹⁷ IP3R has IP3R1, IP3R2 and IP3R3 phenotypes, among which IP3R1 plays a leading role in human pulmonary arterial smooth muscle cells (HPSMCs). Under the regulation of some stimuli (hypoxia, inflammation) or substances (Ang II, norepinephrine, ET-1, etc.), IP3R expression is up-regulated and increases $[Ca^{2+}]_i$, which promotes vasoconstriction and remodelling and leads to increased vascular resistance. However, when Ca^{2+} concentration rises to a certain level, IP3R will in turn be inactivated.¹⁸⁻²⁰ IP3 binds with IP3R in PASMcs to activate the channel. PKG, PKC, CaMKII, and so on, can further promote the binding of IP3 and IP3R through phosphorylation, thus strengthening the opening of the channel. Channel activation promotes the release of stored calcium, up-regulates $[Ca^{2+}]_i$, and regulates the contraction and proliferation of PASMcs.²⁰⁻²² IP3Rs is associated with a number of different signal pathways to regulate diverse functions in different cells.²³ It is activated or inhibited by different substances in vivo, such as calcium binding protein with EF-hand structure, proto-oncogene, tumour suppressor gene, protein kinase, phosphatase, microRNAs, cellular redox products

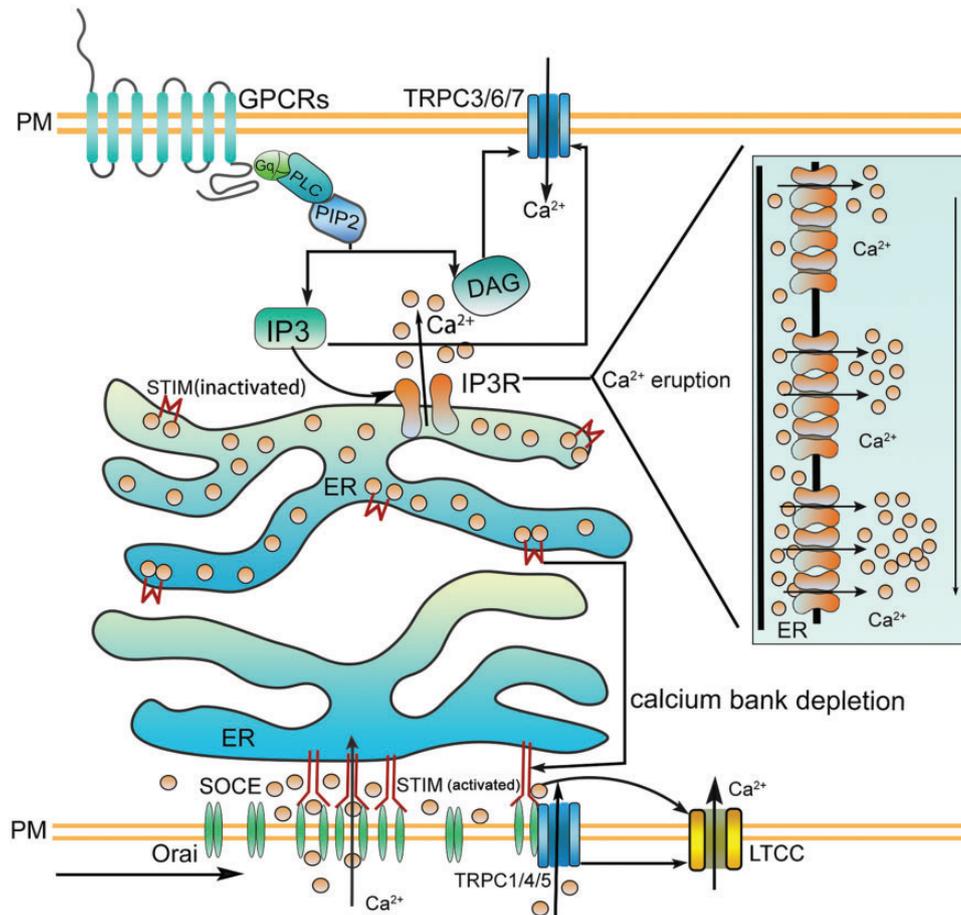


Fig. 1. Regulation of CRAC, IP3R, TRPC, LTCC channels. In the inactivated state, STIM is dispersed in the ER and ORAIL is dispersed in the plasma membrane (PM). When GPCR activates PLC, it then decompresses phosphorylated PIP2 into DAG and IP3. IP3 acts on IP3R of the endoplasmic reticulum, triggering the release of Ca²⁺ from the endoplasmic reticulum, leading to a decrease in the concentration of stored Ca²⁺. When STIM senses endoplasmic reticulum Ca²⁺ depletion, it accumulates between the endoplasmic reticulum and plasma membrane, binds ORAI, activates ORAI pore forming channels, and promotes Ca²⁺ entry. In general, DAG directly activates TRPC3/6/7, but IP3 can also directly activate TRPC channels in some specific cells. STIM and Orai combine with TRPC1/4/5 to activate TRPC. TRPC openness promotes Ca²⁺ influx and depolarization of cells, which in turn activates LTCCs hyperpolarization. Under the stimulation of IP3 and Ca²⁺, from the activation of a single IP3R channel to the cooperative opening of IP3R organized into clusters, the final Ca²⁺ eruption is produced.

CRAC: Ca²⁺ release-activated Ca²⁺ channel; DAG: diacylglycerol; ER: endoplasmic reticulum; GPCR: G protein coupled receptor; IP3: 1, 4, 5-triphosphoinositol; IP3R: inositol 1,4,5-triphosphate receptor; LTCC: L-type calcium channels; PIP2: phosphorylated phosphatidylinositol 4, 5-diphosphate; PLC: phospholipase C; PM: plasma membrane; SOCE: store-operated calcium entry; STIM: stromal interaction molecule; TRPC: transient receptor potential channel.

and so on.^{24–28} In pulmonary arteries, vasoactive substances and signal pathway molecules such as ET-1, 5-HT, NE, Ach and CaSR, CN/NFAT pathway can regulate pulmonary vascular tension by up-regulating the activity of IP3Rs, promoting the release of stored calcium and regulating pulmonary vascular tension.^{29,30} The released Ca²⁺ can further regulate the activity of IP3R and give positive feedback to the activity of IP3R, thus further promoting the increase of [Ca²⁺]_i. However, the increase of Ca²⁺ to a certain extent gives negative feedback on the activity of IP3R.^{31–33} The mutual regulation mechanism between IP3Rs and calcium ion depends in part on its spatial distribution in the cell. IP3R can be organized into clusters and open in coordination to form calcium eruptions (Fig. 1).³³ However,

inhibition occurs in some spatial distributions. It was found that the endothelial cell (EC) were flattened by a certain pressure on the artery, and the influx of Ca²⁺ mediated by IP3 decreased with the increase of pressure. Considering the change of endothelial shape, the geometry of the diffusion space between IP3R and nearby Ca²⁺ was changed, which limited the IP3-mediated Ca²⁺ signal.³⁴ However, it does not rule out that there are differences in the regulatory mechanism of calcium in different cells by IP3R. Interestingly, the same channel phenotype may have opposite function in different blood vessel cells. Moutian et al. found that the expression of IP3R1 in EC of spontaneously hypertensive rats was less than that of the control group, while the expression of IP3R2 was higher.³⁵

To further clarify the role and possible mechanism of IP3R1 in blood pressure regulation, recently, Yuan et al. knocked out the IP3R1 gene in mouse EC and found that mice were the most likely to develop hypertension. After IP3R1 gene knockout, the reactivity of EC to Ach decreased, and the expression of nitric oxide synthase (NOS) and NFAT decreased.³⁶ Because NOS is involved in the synthesis of nitric oxide (NO), it is speculated that IP3R1 can promote the synthesis of NO, and IP3R1 can maintain the vasodilation function in EC. However, different phenotypes in the same cells may have opposite functions. It was found that in PSMC, PAP increased in PH animal model after knockout of IP3R2 gene. The specific mechanism may be that IP3R2 promotes the apoptosis of PSMC and inhibits intracellular calcium ion concentration.³⁷ IP3R can also interact with other channels to regulate calcium concentration. The main function of SOCC is to sense the change of calcium concentration in calcium store. When the IP3R channel is opened, the calcium ion in ER is released, which reduces the calcium concentration in the calcium store. Calcium depletion activates SOCC to promote the influx of extracellular calcium ions, thus increasing $[Ca^{2+}]_i$. IP3R and SOCC cooperated to regulate intracellular calcium concentration.³⁸ Release of Ca^{2+} by IP3Rs can also activate channels such as VDCC and TRP to further increase Ca^{2+} influx.¹⁸ IP3Rs is regulated by a variety of substances or signal pathways, which is the principal channel for calcium storage and release, and can be used as an important therapeutic target for PH. However, it is worth noting that different or the same IP3R subtypes have different effects on blood vessels in different cells and different distribution spaces.

TRPC. TRPC was first discovered in the study of light mutants in *Drosophila melanogaster*.³⁹ TRPC is a receptor-type cationic channel with six transmembrane regions (S1–S6) and the ends of NH2 and COOH.⁴⁰ TRPC is widely distributed in mammalian tissues and cells and has a wide range of functions. In blood vessels, it can participate in the mechanical stimulation of blood vessels and GPCR-related signal pathways, EC-related vasodilation, vascular permeability, neovascularization, vasoconstriction, remodelling and so on. The above functions are mainly realized by regulating $[Ca^{2+}]_i$. It plays an important role in the formation of cardiovascular disease.^{41,42} TRPC can be separated into multiple subtypes of TRPC1–7. All subtypes (except TRPC2, which does not combine with any TRPC protein) can combine with each other to form homotetramers or heterotetramers to perform their corresponding functions.⁴³ TRPC regulates Ca^{2+} mainly through receptor-operated channel (ROC) and store-operated channel (SOC).^{44,45} ROC is mainly TRPC3/6/7, SOC is mainly TRPC1/4/5. TRPC3/6/7 activation is closely linked to GPCRs/PLC. PLC decomposes PIP2 to produce IP3 and DAG, DAG directly activates TRPC3/6/7. IP3 can also

directly activate TRPC channels in some specific cells (Fig. 1).^{46–49} SOC can direct the transfer of TRPC1 to the PM through Orail-mediated Ca^{2+} , and then the interaction between STIM1 and TRPC1 triggers the Gq/PLC-related signal pathway to regulate calcium concentration (Fig. 1).^{50,51} TRPC1, TRPC4 and/or TRPC5 can also participate in SOCC directly without the mediation of STIM1 and Orail.⁵² There is also a certain relationship between TRPC and other ion channel proteins. The regulation of TRPC on Ca^{2+} depolarizes cells and then activates LTCCs hyperpolarization (Fig. 1). TRPC can also promote RyRs-mediated Ca^{2+} release, activate BK channels, cause potassium ion outflow and regulate smooth muscle cell membrane contraction.^{41,53} It had been found that in the model of PH, activation of TRPC not only regulated the contraction and proliferation of PSMC, but also mediated the proliferation and migration of pulmonary adventitia fibroblasts. TRPC mediated the synthesis of extracellular matrix proteins by up-regulating the expression of type I collagen and fibronectin, thus participating in pulmonary adventitia remodelling.⁵⁴ TRPC has a direct or indirect effect on the regulation of calcium ions. GPCRs and various angiogenic substances (5-HT, Ang II, ET-1, etc.) can activate TRPC to regulate intracellular calcium concentration, mediate vascular contraction and remodelling. The role of TRPC in the pathogenesis of PH cannot be ignored.

LTCC. LTCC is a Ca_v1 channel, which belongs to VDCC, subtypes including $Ca_v1.1$, $Ca_v1.2$, $Ca_v1.3$ and $Ca_v1.4$. Different phenotypes have different biophysical characteristics. $Ca_v1.2$ and $Ca_v1.3$ are widely distributed and expressed in most excitable cells (smooth muscle, myocardium, etc.) with high sequence homology. The distribution of $Ca_v1.1$ and $Ca_v1.4$ is relatively limited. $Ca_v1.1$ is mainly distributed in skeletal muscle and $Ca_v1.4$ is mainly distributed in the retina. LTCC is a voltage-dependent non-specific calcium channel, which can maintain a long-term activation state and form a long-term inward current.^{39,55,56} LTCC is the primary channel involved in the influx of extracellular calcium ions into cells. When endovascular pressure increases, vascular smooth muscle membrane depolarizes, activates LTCC channels, and allows a large amount of calcium ions to flow into the cytoplasm, causing vasoconstriction. $Ca_v1.2$ is the main subtype of LTCC channels to regulate vascular smooth muscle contraction.⁵⁷ It has been observed that knockout of $Ca_v1.2$ gene can reduce the frequency of calcium discharge by about 70%.^{58,59} The calcium influx varies with different channel activity. When the channel activity is low, a small amount of isolated LTCC randomly opens to produce a small amount of Ca^{2+} influx. When associated with some substances, such as protein kinase C α , a large number of clustered LTCC channels increase activity and cooperatively open, resulting in persistent and sizable amount of Ca^{2+} influx. PKC α is essential for LTCC to produce persistent substantial amounts of

calcium current.⁶⁰⁻⁶² LTCC-mediated Ca^{2+} flow into the cytoplasm can activate RyR2 channels and produce calcium sparks through the mechanism of calcium-induced calcium release.⁶³ It has been found that the β subunit of LTCC structure plays an important role in the up-regulation of LTCC activity in VSMCs, and participates in the increase of blood pressure induced by Ang II.⁶³ LTCCs play an important role in the vasoconstriction of PH. The expression of LTCCs is up-regulated in the model of hypoxic PH, which leads to PASMC contraction. The use of inhibitors of LTCCs can significantly reduce the absolute contraction of pulmonary artery.⁶⁴ A variety of vasoconstrictor substances can regulate vasoconstriction by mediating LTCCs. LTCCs inhibitors can significantly inhibit arterial constriction induced by ET-1, U46619 and 5-HT. It is worth noting that there are species (rat and mouse) differences in vascular responses to LTCC and vasoconstrictors.⁶⁵ It is speculated that there may be differences in the expression of corresponding receptors or phenotypes of certain substances in different species. LTCC does not function independently and is associated with other ion channels to mediate vasoconstriction. LTCC channels can interact with RyR, TRPC, BK and other channels to regulate vascular tension. LTCC can couple with RyR to produce calcium sparks. The specific mechanism is that LTCC regulates $[\text{Ca}^{2+}]_i$, which affects the concentration of calcium in sarcoplasmic reticulum, and finally triggers the release of calcium sparks from RyR.⁵⁹ The increase of $[\text{Ca}^{2+}]_i$ induced by opening LTCC can activate the potassium channel, which leads to the outflow of K^+ , and then the cell membrane is hyperpolarized. Hyperpolarization of cell membrane leads to the decrease of LTCC activity, then the decrease of Ca^{2+} influx and the decrease of $[\text{Ca}^{2+}]_i$, and finally leads to contractile vasodilation. It can be used as a feedback regulation mechanism.⁶⁶ It has been found that $\text{Ca}_v1.2$, Orail and TRPC1 channels can form macromolecular complexes in close distribution areas, and functional crosstalk exists to regulate $[\text{Ca}^{2+}]_i$. After 5-HT and Thapsigargin (TG) stimulation, crosstalk between the three channels was enhanced.⁵⁷ Weigand et al. found that hypoxia-induced pulmonary vasoconstriction could be achieved by up-regulation of $[\text{Ca}^{2+}]_i$ by SOCE and VDCC. SOCE or VDCC alone could not provide sufficient calcium concentration to trigger PASMC contraction, and SOCE activation could mediate the secondary activation of VDCC.⁶⁷ On the contrary, some studies had found that after STIM1 perception calcium store was depleted and activated, the activated domain binds to the C-terminal of $\text{Ca}_v1.2$ channel, which markedly inhibits $\text{Ca}_v1.2$ channel.^{68,69} It is possible that SOCE has different regulatory effects on calcium channels of different phenotypes. LTCCs not only regulate Ca^{2+} , but also couple with potassium channels and mediate K^+ currents. It has been found that $\text{Ca}_v1.3$ channels can activate large and small conductance potassium channels activated by calcium, which can weaken K^+ currents in the absence of $\text{Ca}_v1.3$.⁵⁵

Since dihydropyridine cannot completely block this channel at present, there is no highly selective LTCC inhibitor at present.⁵⁶ The effects of different phenotypes of LTCCs are different, and the effects are also different in different species. It is necessary to further clarify the sequence differences between subtypes and strictly grasp the differences between human and animal models to achieve accurate medical care.

Ca^{2+} plays an important role in the occurrence and development of PH. Many signal molecules and vasoactive substances eventually affect the concentration of Ca^{2+} , resulting in vasoconstriction and remodelling. At the same time, Ca^{2+} can also activate other ion channels. Calcium channels are widely distributed and have different expressions and functions in different cells, tissues and species. Therefore, it is necessary to clarify the differences and develop highly selective targeted drugs to achieve personalized treatment.

K^+

K^+ is the main cation of intracellular fluid, most of which exist in cells, and play an important role in human circulation, endocrine, immune inflammation, nerve, etc. In the pathogenesis of PH, K^+ participates in the change of membrane potential, cell proliferation, apoptosis, inflammation, regulation of vascular tension, etc. K^+ plays an important role in the pathogenesis and treatment of PH. There are mainly five different functional types of potassium channels: voltage-gated potassium channels (Kv), calcium-activated potassium channels (K_{Ca}), ATP-sensitive potassium channels (K_{ATP}) and double-pore domain K^+ channels ($\text{K}_{2\text{P}}$), inward rectifier potassium channels (Kir).⁷⁰ The following mainly discusses the role of Kv, K_{Ca} , K_{ATP} and $\text{K}_{2\text{P}}$ channels in the pathogenesis of PH.

Kv. Kv is separated into Kv1-12 subfamily members, which are expressed in almost all VSMC, involved in VSMC proliferation, migration and secretion, mediating vasoconstriction and remodelling.⁷¹ Kv channel plays an important role in the pathogenesis of PH. It has been found that Kv channel function is inhibited in secondary (chronic hypoxia) or primary PH, but the inhibition degree of primary PH is higher. Inhibition of Kv channel can cause cell membrane depolarization, induce $[\text{Ca}^{2+}]_i$ increase and eventually lead to pulmonary vasoconstriction and PASMCs proliferation.^{72,73} However, later studies found that there were differences in the expression and function of different subtypes of Kv channel. Among the subtypes of Kv channel, Kv1.3 is mainly involved in cell proliferation. In the proliferation model, it was found that the expression of Kv1.3 and its subsidiary subunit $\text{Kv}\beta 2$ was up-regulated, and inhibition of Kv1.3 would inhibit cell proliferation (Fig. 2).⁷⁴⁻⁷⁶ There was a negative correlation between Kv1.5 and Kv1.3 expressions, and the up-regulation of Kv1.5 was accompanied by the down-regulation of Kv1.3 (Fig. 2).⁷⁶⁻⁷⁸ Other factors

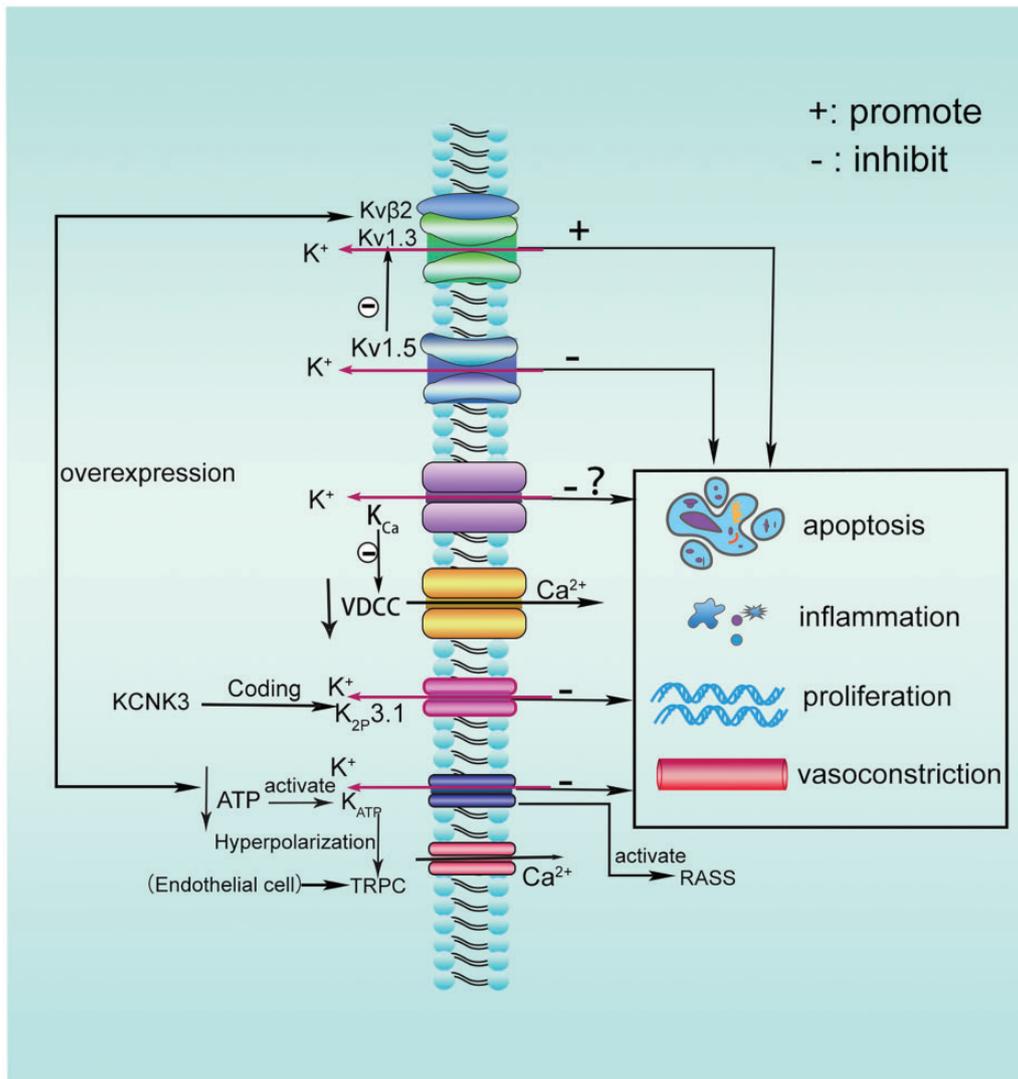


Fig. 2. Regulatory effects of overexpression of different potassium channels on vasoconstriction and remodelling. In general, activation of potassium channels inhibits inflammation, cell proliferation and vasoconstriction to be involved in the pathogenesis of PH. However, individual phenotypes showed the opposite effect. The expression of Kv1.3 and its subunit Kvβ2 is up-regulated to promote cell proliferation. Up-regulation of Kv1.5 is accompanied by down-regulation of Kv1.3, which can inhibit cell proliferation mediated by Kv1.3 channel. Activation of K_{Ca} channel can promote K⁺ outflow, make membrane hyperpolarization, inactivate VDCC, and reduce [Ca²⁺]_i, thus effectively relaxing blood vessels and inhibiting vascular remodelling. However, inhibition of KCa3.1 in some laboratories can alleviate vasoconstriction and remodelling. KCNK3 encodes K_{2P}3.1 channel, and k_{2p}3.1 overexpression can inhibit vasoconstriction and remodelling. The decrease of ATP activates K_{ATP} channel, hyperpolarizes the membrane, relaxes blood vessels, lowers systemic blood pressure and activates RASS system. In vascular endothelial cells, K_{ATP} activation can activate TRPC channel to make Ca²⁺ influx. PH: pulmonary hypertension; RASS: renin-angiotensin-aldosterone system; TRPC: transient receptor potential channel; VDCC: voltage-dependent calcium channel.

that regulate cell proliferation, such as microRNA and mitogen, also regulate cell proliferation by down-regulating the expression of Kv1.5.^{79,80} Kv channel can not only promote cell proliferation, but also inhibit apoptosis. It has been found that after inhibiting the anti-apoptotic protein Survivin, the expression of Kv1.5 and Kv2.1 was up-regulated, and the apoptosis of PASMCs was promoted in the model of PH.^{81,82} Kv channel plays a central role in cell proliferation, and its expression and activity are the key

to VSMCs dedifferentiation and transformation into proliferation. Kv1.3 is an important subtype involved in cell dedifferentiation. PAP-1(a Kv1.3 channel blocker) can down-regulate the expression of αSMA, FGFR-1, PDGFR-b and EGRF-1(cell cycle regulatory factor) to inhibit the proliferation of SMC and effectively improve the degree of vascular stenosis.^{83,84} To sum up, highly selective inhibitors can effectively inhibit pulmonary vascular remodelling and prevent the further progress of the

pulmonary artery. One of the main pathological mechanisms of PH is the continuous progression of pulmonary vascular remodelling, so targeting Kv channel will be a potential and effective treatment direction.

K_{Ca} . K_{Ca} was first found in red blood cells.⁸⁵ K_{Ca} can be divided into three subtypes: large conductance Ca^{2+} -activated potassium channels(BK), small-conductance Ca^{2+} -activated potassium channels(SK), intermediate Ca^{2+} -activated potassium channels (IK). SK ($K_{Ca}2.1$, $K_{Ca}2.2$, $K_{Ca}2.3$) and IK ($K_{Ca}3.1$) channels are not widely distributed in smooth muscle cells. SK is mainly distributed in the nervous system. At present, BK channels are widely studied in vascular diseases.^{86,87} BK is a channel of seven transmembrane domains (S0–S6), which is composed of several subunits (α , β , γ).⁸⁸ The tertiary structure of BK channel is composed of transmembrane region and cytoplasmic region. Pore domain and voltage sensor constitute transmembrane region. Calcium sensor is distributed in cytoplasmic region, which is composed of RCK1 and RCK2. RCK1 and RCK2 are potassium conductance structures.⁸⁹ When the intracellular calcium concentration increases, the calcium ion binds to the calcium sensor, and then opens the BK channel, which causes potassium ion outflow, leading to membrane hyperpolarization and vasodilation. As a negative feedback regulation of vasoconstriction and calcium influx,⁹⁰ the presence of $\beta 1$ subunit in the transmembrane region can increase channel calcium sensitivity to regulate the contraction and relaxation of smooth muscle cells. In SMC, calcium sparks released from sarcoplasmic reticulum activate BK channels, and the concentration of calcium ions can reach micromoles. The specific mechanism is that calcium ions in calcium sparks gather at the edge of α and $\beta 1$ subunits in the structure of BK channels, and the channels are metamorphosed and opened. It was found that β -1 subunit gene knockout could eliminate the outward potassium current produced by calcium spark activated BK channel.^{91,92} CAMP, PKC and H2S can also promote the opening of BK channel in rat PASM. BK is regulated by many different substances.⁹² K_{Ca} channels are voltage- and calcium-dependent, and can mediate vasoconstriction or relaxation by regulating the activity of signal molecules such as PKG, PKA, NO, CO, ROS, ET-1, Ang II and H2O2.^{66,93–96} After the activation of the channel, the outflow of K^+ was increased, which led to the hyperpolarization of the cell membrane. Hyperpolarization of cell membrane leads to the inactivation of VDCC, which in turn leads to the decrease of $[Ca^{2+}]_i$, which plays an important role in the regulation of arterial blood pressure.⁶⁶ This channel plays an important part in the regulation of pulmonary artery tension, and many substances achieve pulmonary artery relaxation by opening K_{Ca} channels.^{97,98} Through gene knockout of BK-Beta1 subunit, it was found that blood pressure in gene knockout group was significantly higher than that in control group.⁹⁹ Si et al found

that EC and SMC in blood vessels of $K_{Ca}3.1$ –/– mice showed a decreased response to Ach hyperpolarization and endothelium-derived hyperpolarizing factor (EDHF)-mediated vasodilation, resulting in an increase in arterial blood pressure.¹⁰⁰ Minami et al. found that ANG II and ET-1 can contract arterial vessels by inhibiting K_{Ca} channels.¹⁰¹ Interestingly, Peng et al. found that under normoxic conditions, ET-1 plays a dual role in PASM. Under different concentrations. When ET-1 is 5 nM, BK is activated by up-regulating $[Ca^{2+}]_i$. When the concentration of ET-1 increased to 10 nM, ET-1 inhibited BK. However, under hypoxia, ET-1 in HPSMCs markedly inhibited the opening of BK channel at 5 nM. It is suggested that there may be differences in the regulation of BK channels by upstream molecules in different concentrations and different environments.¹⁰² It is worth noting that recently, Guo et al found that the expression of $K_{Ca}3.1$ in PASM. Induced by chronic hypoxia was up-regulated than that in the control group. By injecting TRAM-34(a $K_{Ca}3.1$ highly selective inhibitor) into the abdominal cavity of male SD rats with intermittent hypoxia, it was found that it could reduce pulmonary artery contraction or remodelling and right ventricular pressure, inhibit ERK/p38MAPK signal transduction in PASM, and reduce the proliferation or differentiation of PASM. It is suggested that the function of different subtypes of K_{Ca} channel may be different.¹⁰³ From the above contradictory experimental results, it is suggested that we should pay attention to the drug concentration and action conditions of K_{Ca} channels in the treatment of PH in the future, and clarify the differences of action mechanisms of different families of channels.

K_{2P} . K_{2P} channel was found in yeast in 1995.¹⁰⁴ K_{2P} is a K^+ channel with the characteristics of four transmembrane domains and two P domains in series, so it is named K_{2P} .¹⁰⁵ K_{2P} is encoded by KCNK1-18 gene and has 15 members. According to its physiological and pharmacological characteristics, K_{2P} is separated into five families: (1) TWIK (Tandem of P domains in Weak Inward rectifier K^+ channel), (2) TREK/TRAAK (TWIK-Related K^+ channel/TWIK Related Arachidonic Acid-Stimulated K^+ channel), (3) THIK (Tandem pore domain Halothane-Inhibited K^+ channel), (4) TALK (TWIK-related Alkaline pH-activated K^+ channel) and (5) TASK (TWIK-related Acid-Sensitive K^+ channel). K_{2P} is involved in many physiological and pathological processes, including PH, endocrine, pH homeostasis, nerve excitability, heart rhythm, temperature, etc.^{70,106,107} The electrophysiological characteristics of K_{2P} channel are different from other K^+ ion channels. K_{2P} channel has no voltage sensor and is not subject to the change of membrane potential. K_{2P} channel produces a kind of instantaneous inactive current, which is called K^+ leakage current. Studies have shown that it is almost or weakly sensitive to clinical K^+ channel blockers, but can be regulated by different stimuli, such as GPCR, PKC, PH, NO, oxides,

anaesthetics, arachidonic acid, mechanical force, Ca^{2+} , etc.^{106,108,109} $\text{K}_{2\text{P}}$ channel plays an important role in the occurrence and development of PH. It has been observed that in the model of PH, $\text{K}_{2\text{P}}$ is involved in pulmonary artery relaxation and endothelial hyperpolarization to maintain arterial blood pressure.¹¹⁰ At present, the most widely studied gene in PH is *KCNK3*, which encodes $\text{K}_{2\text{P}3.1}$ channel. It was noted that the *KCNK3* gene encoding $\text{K}_{2\text{P}}$ channel was mutated and down-regulated in some non-hereditary or hereditary PH patients. In the experimental model of PH, it was found that *KCNK3* plays an important role in regulating vascular tension and can inhibit distal neovascularization and some hemodynamic changes in the early stage of PH. Up-regulation of *KCNK3* can inhibit vasoconstriction, proliferation of PSAMC/EC/fibroblasts, inflammatory response, and improve ventricular hypertrophy and dysfunction associated with PH (Fig. 2).^{111,112} It was found that the expressions of ERK1/2, AKT, Src, HIF1- α , Survivin and vWF were up-regulated in the tissues of *KCNK3* gene mutant rats, thus promoting cell proliferation or inhibiting apoptosis, while the expression of vasodilators eNOS and EDHF was down-regulated. The efficacy of sildenafil was significantly decreased in *KCNK3* gene mutant PH rats.¹¹³ In PH, RELM- β , PLC and IP3 can regulate *KCNK3* to achieve vasoconstriction, cell proliferation, inflammation and other functions.¹¹⁴ The study of *KCNK3* gene in PH shows that *KCNK3* gene can be used as an index to detect the susceptibility of PH patients and can be used as a target for gene therapy.

K_{ATP} The K_{ATP} channel is an octameric complex composed of four subunits with pore-forming inward rectifier K^+ channels (Kir6.1 or Kir6.2) and four regulatory sulfonylurea receptors (SUR1 or SUR2). The genes *ABCC8* and *KCNJ11* encode SUR-1 and Kir6.2 subunits respectively, which are situated on human chromosome 11. *ABCC9* and *KCNJ8* encode SUR-2 and Kir6.1 subunits, respectively, which are situated on human chromosome 12. *ABCC9* (SUR2) and *KCNJ8* (Kir6.1) are mainly expressed in smooth muscle and ECs, while *ABCC8* (SUR1) and *KCNJ11* (Kir6.2) are mainly expressed in pancreas and neurons. However, there were also reports of *ABCC8* mutation and loss of function in children with idiopathic pulmonary arterial hypertension (IPAH).^{115–117} As the name implies, K_{ATP} channels can be opened due to the decrease of cell ATP, which hyperpolarizes the membrane and connects metabolism with electrical excitation. In the endocrine system, insulin release can be regulated to regulate blood sugar. K_{ATP} channels are widely expressed in the vascular system and play an important role in vascular tension regulation and systemic blood pressure maintenance. K_{ATP} participates in the formation of PH and hypertension.¹¹⁸ In PSMC, Kir can control calcium influx through VDCCs, and it may regulate extracellular potassium concentration to regulate PSMC membrane potential to

mediate vasoconstriction or relaxation. It is worth noting that in vascular ECs, K_{ATP} activation can increase $[\text{Ca}^{2+}]_i$ through receptor- or store-operated channels, such as activating TRPC channels to make Ca^{2+} inflow (Fig. 2).^{119,120} Vasoconstrictor substance (ET-1, Ang II, NE, NPY1), vasodilator substance (NO, H₂S, A₂, EET), translational modifier substance (PKC, PI3-AKT, PKA), mediating immune inflammatory substance (LPS, TLR3, TNF, IFN), redox substance (ROS, H₂O₂, GSH) and transcription inducing factor (HIF-1 α , NF- κ B, FOXO1) can regulate vascular tension, cell proliferation and inflammation by mediating K_{ATP} activity.^{121,122} It was found that the expression of SUR-2 and Kir6.1 was down-regulated in hypertensive rat model.^{123–125} The use of K_{ATP} channel opener iptakalim can effectively reduce PAP, right ventricular systolic pressure and pulmonary arteriole wall remodelling in the model of hypoxic PH. K_{ATP} can decrease the expression of IL- β (IL-10) in peripheral blood and lung tissue, enhance the expression of platelet EC adhesion molecule-1 and eNOS, and finally reduce the inflammatory reaction and EC damage.^{126,127} Interestingly, long-term use of some K^+ channel openers (diazoxide, minoxidil) could cause some side effects: abnormally aggravating PAP, promoting myocardial hypertrophy, increasing vascular volume, etc. The specific mechanism was that channel openers can activate renin-angiotensin-aldosterone system to promote water and sodium retention due to significant reduction of systemic pressure (Fig. 2).^{128,129} Therefore, in the treatment of PH with K_{ATP} channel opener, we should pay attention to its side effects. We can cooperate with diuretics to improve the side effects caused by the feedback mechanism.

Na^+

Na^+ is the principal extracellular cation, which is closely related to the excitability of cells. In human body, Na^+ participates in water electrolysis, blood vessel, muscle, nerve function, etc. Ca^{2+} , K^+ , H^+ participate in Na^+ entry and exit under different chemical conditions. Na^+ is widely disseminated in different cells. In blood vessels, it is allocated in epithelial cells, SMC, EC, etc. Under certain conditions, Na^+ affects cell proliferation, migration and contraction, which mediate vasoconstriction and remodeling, resulting in increased vascular resistance. The following is mainly about the pathogenesis of epithelial Na^+ channels (ENaCs), Sodium-calcium exchanger (NCX) and Na^+ - H^+ exchanger (NHE) in PH.

ENaC. ENaC is a mechanically sensitive ion channel, which can fall into three subtypes: α , β and γ . ENaC belongs to the degenerin protein family and participates in mechanically related myogenic responses.^{130,131} Angiotensin and aldosterone can act on heart, artery and kidney by regulating ENaC activity. Short-term regulation can maintain electrolyte, volume and blood pressure stability, but promote mechanical myogenic occurrence, vascular sclerosis,

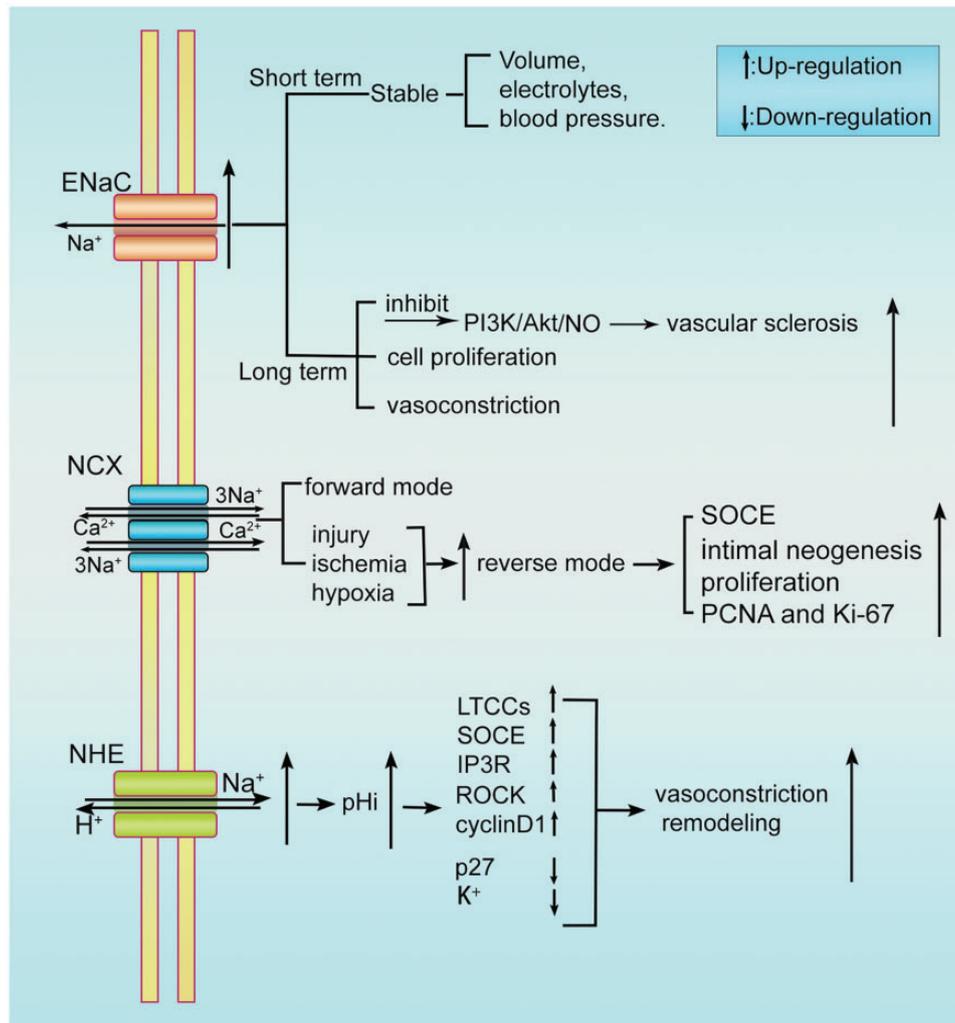


Fig. 3. Regulation of sodium channel in vasoconstriction and remodelling. When ENaC is overexpressed, electrolyte, volume and blood pressure can be retained in the short term. However, when ENaC is overexpressed for a long time, it may inhibit the release of nitric oxide by endothelial cells through PI3K/Akt signal pathway, which leads to vascular hardening. NCX is divided into forward mode and reverse mode. In cardiomyocytes, forward mode is important for diastolic calcium excretion. Under some conditions (injury, ischemia, hypoxia, etc.), activation of reverse mode can mediate SOCE, to promote calcium influx. NHE activation can increase intracellular pH and up-regulate calcium channels (LTCC, SOCE and IP3R), cyclinD1, ROCK, down-regulate p27, resulting in increased intracellular calcium concentration, cell proliferation, vasoconstriction, etc.

ENaC: Epithelial Na⁺ channels; IP3R: inositol 1,4,5-triphosphate receptor; LTCCs: L-type calcium channels; NCX: Sodium-calcium exchanger; NHE: Na⁺-H⁺ exchanger; PCNA: proliferating cell nuclear antigen; ROCK: Rho kinases; SOCE: store-operated calcium entry.

vascular remodelling, increased vascular resistance and systemic hypertension in the long term (Fig. 3).¹³² ENaC is widely distributed. ENaC is expressed in SMC, baroreceptor nerve endings and nodular ganglion (inferior ganglion of vagus nerve), cardiovascular regulatory area of brain (outside hypothalamic nucleus), kidney, taste buds and EC. ENaC regulates humoral circulation and blood pressure through nerves, heart, kidney, blood vessels, biological receptors, etc.^{39,133–137} Vascular sclerosis may occur when ENaC is over-expressed for a long time. At present, EC sclerosis is mainly considered, and the degree of sclerosis is positively correlated with ENaC density. The specific mechanism of sclerosis may be through PI3K/Akt signal

pathway to inhibit the release of NO from EC, thus regulating vasoconstriction and inflammation. High sodium and aldosterone can aggravate vascular sclerosis, and the use of aldosterone or ENaC inhibitors can alleviate the degree of atherosclerosis.^{90,130,138–141} Under the action of mechanical stimulation (pressure, traction, shear force, etc.), ENaC can promote the myogenic occurrence of blood vessels, contraction and proliferation of VSMC. Inhibition of ENaC could eliminate pressure-induced contraction of VSMC and increase of intracellular Ca²⁺ and Na⁺ in mouse kidney.¹⁴² Kim et al. found that inhibition of ENaC in the posterior cerebral artery could inhibit the increase of Ca²⁺ concentration, myogenic response and myosin light chain

phosphorylation induced by pressure.^{143,144} Recently, Choi et al. have confirmed that the expression of ENaC is up-regulated in the posterior cerebral artery of spontaneous PH rats, which significantly increases the myogenic response and myogenesis.¹⁴⁵ In the ENaC study, we know that ENaC can be regulated by angiotensin, aldosterone, blood pressure, shear force, baroreceptor and so on to harden blood vessels. When vascular sclerosis occurs, vascular pressure, shear force of blood flow, expression of substances regulating vascular tension and renal sodium excretion change, and baroreceptor is stimulated, which finally increases the expression of ENaC.^{146–148} Over and over again, a positive feedback circulatory mechanism is formed between ENaC and arteriosclerosis, which finally leads to the gradual increase of blood pressure. However, the specific mechanism of up-regulation of ENaC expression in specific sclerotic vessels has not been clarified, which needs further study. The study of ENaC is mainly in cerebral vessels and renal vessels, but virtually not in pulmonary vessels. ENaC is not excluded in the formation of PH, so it can be utilized as a research direction for the pathogenesis of PH.

NCX. NCX is encoded by the SLC8 gene family and has three subtypes: NCX1, NCX2 and NCX3. NCX2 and NCX3 were noted in the brain and skeletal muscles. NCX1 is widespread in mammalian cells.¹⁴⁹ NCX consists of 938 amino acid residues. In NCX, three Na⁺ and one Ca²⁺ bind to the centre of the protein. NCX is symmetrical and can produce two-way ion exchange reaction.¹⁵⁰ NCX is divided into forward mode and reverse mode. In the forward mode, three Na⁺ are passed into the cell and one Ca²⁺ is transferred out of the cell at the same time. In cardiomyocytes, this function is very important for the timely excretion of diastolic calcium ions. Under some conditions (injury, ischemia, hypoxia, etc.), NCX can be reversed, namely reverse mode. In the reverse mode, three Na⁺ were transferred out of the cell, while one Ca²⁺ was transferred into the cell, which increased ([Ca²⁺]_i), mediated vascular contraction and remodelling, and finally led to the increase of vascular resistance.^{151,152} At present, NCX1 is widely studied in blood vessels. In order to study the role of the reverse mode of NCX in the proliferation of VSMC, Liu et al. found that the expression of NCX1 was up-regulated and neointima was formed in the model of vascular injury with balloon. Inhibition or knockout of NCX1 could inhibit neointimal formation, VSMC proliferation, SOCE, proliferating cell nuclear antigen (PCNA) and Ki-67 expression (Fig. 3).¹⁵³ Reverse mode of NCX participates in the regulation of calcium concentration in PASMCM of patients with IPAH. Zhang et al found that the expression of NCX1 in PASMCM of IPAH patients was up-regulated, the reverse pattern of NCX was enhanced, and [Ca²⁺]_i was increased, thus promoting the contraction and proliferation of PASMCM.¹⁵² NCX can mediate inflammation related to

macrophages, dendritic cells and B cells. NCX is also engaged in insulin secretion, calcium reabsorption in the kidney, and learning and memory related to the hippocampus.^{151,154} We will not elaborate on it here.

NHE. The NHE family is composed of at least 10 known isomers, and the expression is tissue-specific. NHE-1 phenotype is mainly expressed in vascular tissues, while other subtypes can be expressed in gastrointestinal tract, kidney, brain tissue, germ cells, etc.^{155–158} Na⁺-H⁺ exchange channels play a major role in vasoconstriction and remodelling. It was noted that the activity of NHEs in aortic smooth muscle cells of spontaneously hypertensive rats was significantly higher than that in the control group.¹⁵⁷ In the hypoxia-induced model, it was found that the right ventricular systolic pressure, right ventricular weight and the inner wall thickness of pulmonary arterioles in NHE1 knockout mice were significantly higher than those in the control group.¹⁵⁹ In the pathogenesis of PH, it is considered that NHE-1 affects intracellular pH (pHi). Some previous studies have shown that NHE-1 activity increases and pHi shows alkaline shift in PASMCM in chronic hypoxia-induced PH model.¹⁶⁰ When the cells are alkalinized, IP3R, LTCC and SOCE can be activated, thus increasing [Ca²⁺]_i and inhibiting potassium influx, which leads to vasoconstriction and remodelling (Fig. 3).^{161–163} It was found that after inhibiting the effect of buffer pair (CO₂/HCO₃⁻) on pH, NHE-1 gene knockout led to cell acidification, and then decreased arterial tension, arterial media remodelling, blood pressure and Rho-mediated VSMCM sensitivity to calcium ions.¹⁶⁴ In PASMCM, HIF-1 could up-regulate NHE-1, and thus up-regulate pHi, cause alkaline drift of cells, promote cell proliferation and mediate vascular remodelling.¹⁶⁵ In view of the fact that ET-1 can up-regulate the expression of HIF-1, it is speculated that ET-1 can further enhance the activity of NHE-1.¹⁶⁶ NHE1 (-/-) mice down-regulate Rho kinases (ROCK1 and ROCK2) and cyclinD1 and up-regulate p27 expression (Fig. 3).¹⁵⁹ Interestingly, Huetsch et al. recently used the SuHx model (the model constructed by vascular endothelial growth factor receptor inhibitor SU5416 combined with chronic hypoxia exposure, which is more suitable for the pathological development of PH). The results showed that the activity of NHE in PASMCM increased in SuHx model, and PASMCM still proliferated and migrated obviously when pHi was unchanged. The degree of proliferation and migration reduced after the pharmacological inhibition of NHE, which is contradictory to the fact that NHE regulates cell contraction and proliferation by changing pHi.¹⁶⁷ Considering that NHE is not the only substance that regulates intracellular acid and base, there are other buffer pairs. Therefore, the result of constant pHi in the laboratory does not rule out the interference of other regulatory substances, which can be further verified after the strict control of other ion buffer substances. In short, the regulatory mechanism of NHE in the formation

of PH cannot be ignored and can be used as a new target for the treatment of PH in the future.

Cl⁻

Cl⁻ is the most abundant anion in vivo, and its extracellular concentration is higher than that of intracellular fluid. Cl⁻ accounts for about 70% and 65% of the total anions in tissue fluid and plasma, respectively.¹⁶⁸ Cl⁻ is involved in a variety of physiological functions, including cell volume regulation, proliferation, membrane potential changes, signal transduction, acidolysis balance, blood pressure regulation, etc. Cl⁻ channels are widely distributed, which are classified as calcium-activated chloride channels (CaCCs), volume-or swelling- sensitive Cl⁻ (Clswell), cysticfibrosis transmembrane conductance regulator, P64-related chloride channels and ligand-gated chloride channels. The chloride channels distributed in smooth muscle cells are mainly CaCCs and Clswell channels.¹⁶⁹

CaCC. CaCC includes at least two protein families, TMEM16 and bestrophins (corresponding to BEST1-4 in humans). TMEM16 is divided into TMEM16A and TMEM16B (corresponding to ANO1 and ANO2 in humans, respectively). Later studies have found that TMEM16A is a potential CaCC.^{170,171} CaCC is not only permeable to chloride ions, but also permeable to univalent anions such as I⁻, Br⁻, F⁻, SCN⁻, etc.¹⁷² CaCC is a group of heterochannel proteins that are sensitive to anions and dependent on Ca²⁺.¹⁷³ The increase of [Ca²⁺]_i can trigger CaCC activity, cause chloride ion outflow, form inward current and cause cell depolarization (Fig. 4). The factors regulating CaCC gating are [Ca²⁺]_i, membrane depolarization, extracellular anions and intracellular protons. Among them, Ca²⁺ plays a key role in the activation of CaCC. TMEM16 is the pore-forming subunit of CaCC. There are conservative regulatory calcium binding sites in the pore region of the TMEM16A structure, in which the sixth transmembrane segment is the main gated part of the channel activated by Ca²⁺, and the sixth transmembrane segment has pore lining residues and Ca²⁺ binding sites. After binding with calcium ions, the transmembrane segment was activated after conformational change and maintained the stability of the current activated state.¹⁷⁴ It was found that the activation of TMEM16A was regulated by binding to at least two Ca²⁺ in a certain order and coupled with cell membrane voltage.¹⁷⁵ The coupling relationship between Ca²⁺ and membrane voltage changes at different concentrations, and the coupling intensity is higher at a low concentration, but when it is increased to a certain concentration, the coupling disappears.¹⁷⁶ With the increase of [Ca²⁺]_i, the first Ca²⁺ binds to the corresponding binding site, and then the anion is allowed to enter the channel, and the channel structure rapidly undergoes a voltage-dependent conformational change, making the channel open, thus mediating the anion current, and then the

second Ca²⁺ binds to the calcium binding site. The secondary binding of Ca²⁺ is mainly to completely activate the channel and maintain the stability of the current conformation.¹⁷⁴ The source of calcium ion can be released through VDCC, IP3R, Na⁺-Ca²⁺ reverse exchange, SOCE, etc.¹⁷⁷⁻¹⁸¹ CaCC is widely expressed in different tissues, causing different physiological functions, such as cell contraction/proliferation, regulation of blood pressure, heart/neuron excitation, glandular secretion, olfactory signal transduction, etc.¹⁸²⁻¹⁸⁶ In pulmonary vessels, CaCC can cause SMC proliferation, contraction and increase of PAP. Through the study of 40 healthy donor lungs and 38 IPA donor lungs, Papp et al found that the expression of TMEM16A gene in PASM of IPA patients was significantly higher than that of the control group, and Cl⁻ efflux increased, resulting in cell depolarization. Knockout or inhibition of TMEM16A can repolarize cells, inhibit cell proliferation and alleviate vascular remodeling.¹⁸⁷ TMEM16A could be used as a novel target for reversing PH. In PH, TMEM16A can up-regulate the expression of cyclin. Shang et al studied the mechanism of TMEM16A promoting cell proliferation in PH rat model, and found that the expression of cyclin E, D1 and TMEM16A were up-regulated in PH group. Knockout of TMEM16A gene could reverse the expression of cyclin.¹⁸⁶ However, there were some contradictions in the results of TMEM16A. Zeng et al. found that the overexpression of TMEM16A in basilar artery SMC could down-regulate the activity of TGF-β1/Smad3, TGF-β1/ERK and JNK1 pathway in Ang II-induced hypertension model. Tissue inhibitors of metalloproteinases (TIMP-1 and TIMP-2) were down-regulated when TMEM16A was overexpressed, and matrix metalloproteinases (MMP-2, MMP-9 and MMP-14) were down-regulated by inhibiting the activity of WNK1 (reduced Cl⁻-sensitive serine/threonine kinase).¹⁸⁸ It is speculated that the overexpression of TMEM16A can reduce the deposition of arterial matrix and play a protective role in vascular remodelling (Fig. 4). It is thought that there are differences in the role of TMEM16A in different types of arteries. It is worth noting that after specific knockout of TMEM16A in uterine SMC of mice, Qu et al. found that the calcium signal pattern, size and overall uterine contraction induced by agonists (potassium chloride, oxytocin and prostaglandin) in the gene knockout group had no significant change compared with the control group. It is speculated that TMEM16A may have no obvious effect on uterine SMC contraction, but it does not rule out the difference of experimental consequences caused by the difference of experimental detection methods.¹⁸⁹ Some studies have found that specific knockout of TMEM16A in VSMC does not cause complete ablation of TMEM16A protein, and TMEM16A protein still exists in VSMC in the form of truncation after gene knockout. This situation does not rule out the incomplete knockout of genes, but we still need to pay attention to it when doing experiments.¹⁹⁰ TMEM16A also regulates with some inflammatory factors

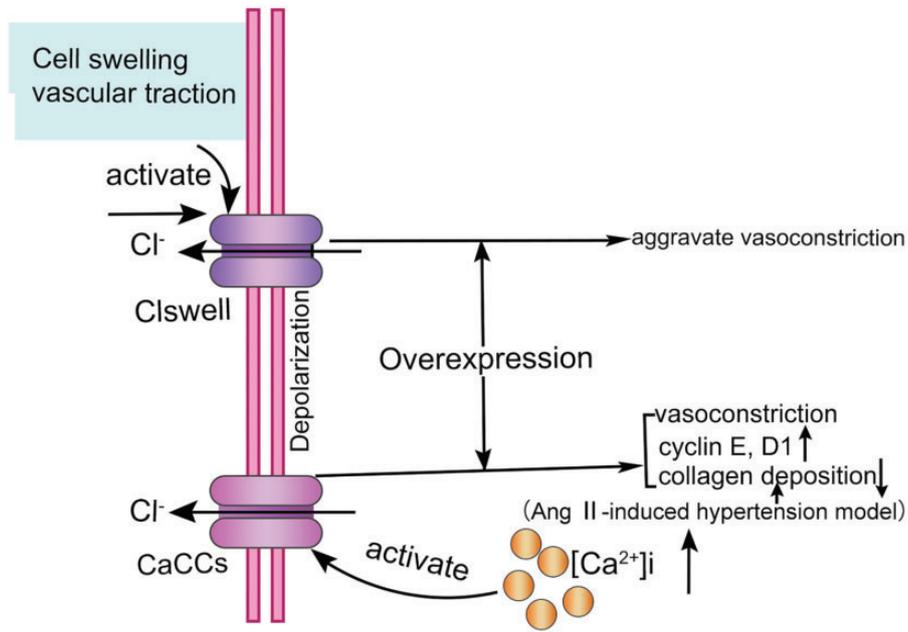


Fig. 4. Regulation of chloride channel in vasoconstriction and remodeling. The increase of $[Ca^{2+}]_i$ can activate calcium-activated chloride channels (CaCC), to cause Cl^- outflow, form inward current, cause cell depolarization, up-regulate cyclin E and D1, thus promote vasoconstriction and remodeling. However, the overexpression of CaCC can reduce the deposition of vascular matrix in Ang II-induced hypertension model. Volume- or swelling-sensitive Cl^- (Clswell) is activated when the cell volume increases and the blood vessel is pulled, which further aggravates the vasoconstriction.

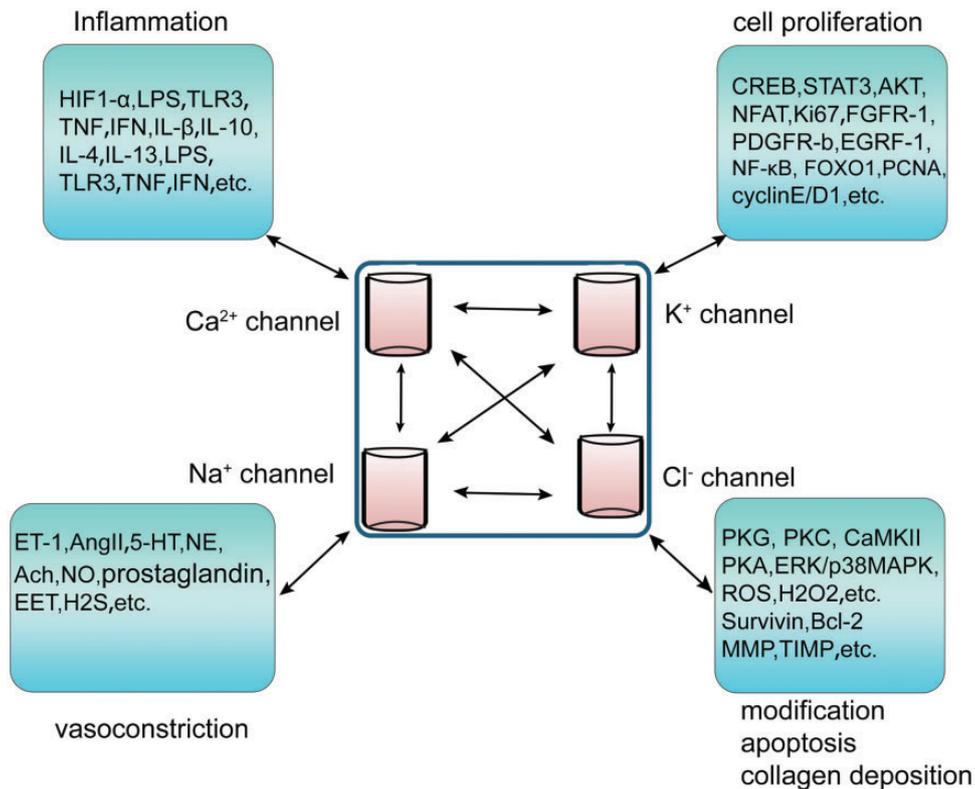


Fig. 5. The interaction between ion channels and different substances is shown. Ions play a major role in the pathogenesis of pulmonary arterial hypertension (PH). Many different chemicals (vasoactive substances, inflammatory mediators, transcription inducing factors, apoptosis factors, translation modifiers, etc.) and mechanical stimulation mediate vascular tension and vascular remodeling through the activity of various ion channels. On the contrary, ions can also regulate the above diverse chemical substances and physical stimuli. The ions and their channels are not independent of each other, but influence each other.

(IL-4, IL-13, IL, etc.) and signal molecules (RhoA/ROCK, AKT, ERK1/2, SRC, CaMKII, EGFR, etc.), thus participating in inflammation, cell proliferation and signal transduction.¹⁹¹ All in all, TMEM16A protein plays an important role in the pathogenesis of PH. The differences of experimental results suggest that TMEM16A may have distinct functions in different tissues, so its mechanism needs to be further clarified. At the same time, it should be noted that the expression of the protein should be detected after knocking out TMEM16A.

Clswell. As the name implies, Clswell channels are regulated by cell volume and are activated when the cell expands and the volume increases. It can participate in cell proliferation/apoptosis, angiogenesis, pH regulation, LTCC/SOCC activation, etc.¹⁹² In SMC, Clswell is activated when sensing changes in cell volume caused by mechanical stretching and osmotic pressure.¹⁹³ By stimulating α 1-adrenergic receptor to regulate the change of arteriole pressure, it was found that the chloride current induced by cell swelling (IClswell) was sensitive to the change of transmural pressure and mechanical traction. The greater the pressure, the higher the traction force and the gradual enhancement of IClswell, which depolarizes the cells and further contracts the blood vessels (Fig. 2). The use of Clswell blockers can inhibit the spontaneous myogenic tension of blood vessels. However, Ca^{2+} -activated chloride current (IClCa²⁺) mainly maintained the continuous contraction of arteries, and its ability to maintain vasoconstriction decreased with the increase of transmural pressure.¹⁹⁴ It is assumed that Clswell further increases blood pressure in the process of rising blood pressure. Angiotensin and epinephrine can contract blood vessels by acting on Clswell. Clswell can be used as a research direction to control the further rise of PAP, but at present, there are few studies on its role in pulmonary vessels, which need to be further explored.

Conclusions

To sum up, we know that various ions play an important role in the pathogenesis of PH, such as pulmonary vasoconstriction or relaxation, cell proliferation and apoptosis, myogenic reaction, inflammation, maintenance of cell volume, collagen deposition, fibrosis and so on. The ions do not function independently, but interact with each other, mediating the opening or closing of the ion channels, thus regulating the ion concentration on both sides of the membrane. In addition to the interaction between ions, ion channels can be regulated with different chemicals, such as vasoactive substances, inflammatory mediators, transcription-inducing factors, apoptosis factors, translation modification factors, etc. (Fig. 5). Vascular mechanical traction stimulation can also regulate the activity of ion channels. Ions participate in vascular tension, vascular remodelling and systemic blood pressure regulation through the above interactions. In addition to the vascular system,

ions can also regulate the nervous system, endocrine system, metabolism, acid–base balance and so on. In short, ion channels are widely distributed, participate in different functions of the human body, and play an important role in human physiology and pathology. After years of research, it has been found that many substances use ions as targets to achieve related functions, which show that ions are in the transportation hub of various signal pathways. At present, various laboratories have studied the pathogenesis of PH from diverse aspects, and developed related drugs. However, the efficacy of drugs is poor, considering that a sole drug only acts on one aspect, while PH is formed under the interaction of different stimulating factors, not a single factor. Therefore, the combination of drugs is imperative, or finding the common target of various stimulating factors, such as ions, will achieve twice the result with half the effort. Recently, it has been proposed that energy metabolism may be a universal feature in the pathogenesis of PH, and almost all functions of the human body cannot be achieved without energy supply, which should also be paid attention to. It is worth noting that there are different subtypes of ion channels, and there are differences in channel subtypes and their expression in different tissues. Distinct subtypes or the same subtypes may have different functions in different tissues and conditions. Therefore, in drug research and development, we should pay attention to the differences of ion channel subtypes, distribution tissue and function, so as to achieve accurate targeted therapy and avoid some side effects. At present, many drugs that can be effective in animal models are not applicable in human experiments, so we pay attention to the differences of target genes, ion channel subtypes and drug doses among different species in drug research and development. In a word, as the target of various substances involved in the pathogenesis of PH, ions can be used as an important direction in the treatment of PH.

At present, the treatment of PH is mainly focused on ET-1, NO, PDE5, guanylate cyclase, prostacyclin and so on. It is pointed out that the combination of drugs can improve the symptoms and delay the progress of PH, but the effect is still not as expected or cured.^{195–197} We need to find some new therapeutic targets in different directions. In our description of ions and their channels, the above-mentioned ions and their channels can be used as research targets for the treatment of PH. As we have previously described, in hereditary or non-hereditary PH patients, the gene KCNK3 encoding K_{2P} channel is mutated or down-regulated, suggesting that gene targeting therapy can be undertaken. In addition to KCNK3 gene, there are also some studies on ion channel gene targeting therapy in Ca_v1.2, Kv1.5 and BK channels.^{92,198–200} However, at present, there are still some problems in the therapeutic drugs related to ion channels. Some ion channel inhibitors have different selectivity at different concentrations. However, there are still some problems in the therapeutic drugs related

to ion channels. Some ion channel inhibitors have different selectivity at different concentrations; some are proved to be effective in vitro, but they cannot function effectively because of some reactions and interference in vivo; some ion channel blockers take longer time to spread, so the time to take effect is slower, increasing the concentration of the inhibitor can accelerate the time to take effect, but the drug appears non-targeted effect, which affects the curative effect or produces some side effects. Some showed inhibitory or excitatory effects at different concentrations, while some inhibitors had poor specificity and could inhibit different channels at the same time.²⁰¹ Therefore, at present, there are still some problems in the drug treatment of ion channels, and we are still not fully clear that under some conditions, the change in channel expression is the result, or the inducement, or just a compensatory change in the progression of the disease. The phenotypes of ion channels are diverse, and many different subtypes may be distributed in the same vessel, and there may be significant differences in the function of different subtypes. At the same time, in the whole between different ion channels, different cells can influence each other, which can significantly reduce our drug selectivity, accompanied by certain side effects, so it is difficult to achieve accurate and effective treatment. Therefore, it will be an interesting topic to take gene targeting therapy as the key research direction of PH therapy in the future.

Authors' contributions

GL drafted the manuscript; DF and HT revised the manuscript; AD approved the final version of the manuscript.

Conflict of interest

The author(s) declare that there is no conflict of interest.

Funding

This article is supported by the National Natural Research Foundation of China (81570052).

Guarantor

AD.

ORCID iD

Heshen Tian  <https://orcid.org/0000-0002-3037-7293>

References

1. Simonneau G, Montani D, Celermajer DS, et al. Haemodynamic definitions and updated clinical classification of pulmonary hypertension. *Eur Respir J* 2019; 53: 1801913.
2. Sommer N, Ghofrani HA, Pak O, et al. Current and future treatments of pulmonary arterial hypertension. *Br J Pharmacol* 2020; 178: 6–30.
3. Galiè N, Humbert M, Vachiery JL, et al. 2015 ESC/ERS Guidelines for the diagnosis and treatment of pulmonary hypertension: The Joint Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS): Endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC), International Society for Heart and Lung Transplantation (ISHLT). *Eur Heart J* 2016; 37: 67–119.
4. Berridge MJ, Bootman MD and Roderick HL. Calcium signalling: Dynamics, homeostasis and remodelling. *Nat Rev Mol Cell Biol* 2003; 4: 517–529.
5. Lewis RS and Cahalan MD. Mitogen-induced oscillations of cytosolic Ca^{2+} and transmembrane Ca^{2+} current in human leukemic T cells. *Cell Regul* 1989; 1: 99–112.
6. Hoth M and Penner R. Depletion of intracellular calcium stores activates a calcium current in mast cells. *Nature* 1992; 355: 353–356.
7. Lewis RS. Store-operated calcium channels: From function to structure and back again. *Cold Spring Harb Perspect Biol* 2020; 12: a035055.
8. Prakriya M and Lewis RS. Store-operated calcium channels. *Physiol Rev* 2015; 95: 1383–1436.
9. Feske S, Gwack Y, Prakriya M, et al. A mutation in Orai1 causes immune deficiency by abrogating CRAC channel function. *Nature* 2006; 441: 179–185.
10. Vig M, Peinelt C, Beck A, et al. CRACM1 is a plasma membrane protein essential for store-operated Ca^{2+} entry. *Science* 2006; 312: 1220–1223.
11. Parekh AB. Slow feedback inhibition of calcium release-activated calcium current by calcium entry. *J Biol Chem* 1998; 273: 14925–14932.
12. Lacruz RS and Feske S. Diseases caused by mutations in ORAI1 and STIM1. *Ann N Y Acad Sci* 2015; 1356: 45–79.
13. Xu YJ, Elimban V and Dhalla NS. Reduction of blood pressure by store-operated calcium channel blockers. *J Cell Mol Med* 2015; 19: 2763–2770.
14. Calderón-Sánchez EM, Ávila-Medina J, Callejo-García P, et al. Role of Orai1 and L-type $Ca_v1.2$ channels in endothelin-1 mediated coronary contraction under ischemia and reperfusion. *Cell Calcium* 2020; 86: 102157.
15. Song S, Carr SG, McDermott KM, et al. STIM2 (stromal interaction molecule 2)-mediated increase in resting cytosolic free Ca^{2+} concentration stimulates PSMC proliferation in pulmonary arterial hypertension. *Hypertension* 2018; 71: 518–529.
16. Fan G, Baker ML, Wang Z, et al. Gating machinery of InsP3R channels revealed by electron cryomicroscopy. *Nature* 2015; 527: 336–341.
17. Marks AR. Calcium channels expressed in vascular smooth muscle. *Circulation* 1992; 86: Iii61–Iii67.
18. Ottolini M, Hong K and Sonkusare SK. Calcium signals that determine vascular resistance. *Wiley Interdiscip Rev Syst Biol Med* 2019; 11: e1448.
19. Ng LC, Wilson SM, McAllister CE, et al. Role of InsP3 and ryanodine receptors in the activation of capacitative Ca^{2+} entry by store depletion or hypoxia in canine pulmonary arterial smooth muscle cells. *Br J Pharmacol* 2007; 152: 101–111.
20. Yadav VR, Song T, Mei L, et al. PLC γ 1-PKC ϵ -IP(3)R1 signaling plays an important role in hypoxia-induced calcium response in pulmonary artery smooth muscle cells. *Am J Physiol Lung Cell Mol Physiol* 2018; 314: L724–L735.

21. Patel S, Joseph SK and Thomas AP. Molecular properties of inositol 1,4,5-trisphosphate receptors. *Cell Calcium* 1999; 25: 247–264.
22. Vermassen E, Fissore RA, Nadif Kasri N, et al. Regulation of the phosphorylation of the inositol 1,4,5-trisphosphate receptor by protein kinase C. *Biochem Biophys Res Commun* 2004; 319: 888–893.
23. Mikoshiba K. Role of IP₃ receptor signaling in cell functions and diseases. *Adv Biol Regul* 2015; 57: 217–227.
24. Hedgepeth SC, Garcia MI, Wagner LE 2nd, et al. The BRCA1 tumor suppressor binds to inositol 1,4,5-trisphosphate receptors to stimulate apoptotic calcium release. *J Biol Chem* 2015; 290: 7304–7313.
25. Ivanova H, Vervliet T, Missiaen L, et al. Inositol 1,4,5-trisphosphate receptor-isoform diversity in cell death and survival. *Biochim Biophys Acta* 2014; 1843: 2164–2183.
26. Vanderheyden V, Devogelaere B, Missiaen L, et al. Regulation of inositol 1,4,5-trisphosphate-induced Ca²⁺ release by reversible phosphorylation and dephosphorylation. *Biochim Biophys Acta* 2009; 1793: 959–970.
27. Drawnel FM, Wachten D, Molkentin JD, et al. Mutual antagonism between IP₃RII and miRNA-133a regulates calcium signals and cardiac hypertrophy. *J Cell Biol* 2012; 199: 783–798.
28. Bánsághi S, Golenár T, Madesh M, et al. Isoform- and species-specific control of inositol 1,4,5-trisphosphate (IP₃) receptors by reactive oxygen species. *J Biol Chem* 2014; 289: 8170–8181.
29. Berridge MJ. Smooth muscle cell calcium activation mechanisms. *J Physiol* 2008; 586: 5047–5061.
30. Bernier S and Guillemette G. Increased inositol 1,4,5-trisphosphate binding capacity in vascular smooth muscle of spontaneously hypertensive rats. *Am J Hypertens* 1993; 6: 217–225.
31. Foskett JK, White C, Cheung KH, et al. Inositol trisphosphate receptor Ca²⁺ release channels. *Physiol Rev* 2007; 87: 593–658.
32. Bezprozvanny I, Watras J and Ehrlich BE. Bell-shaped calcium-response curves of Ins(1,4,5)P₃- and calcium-gated channels from endoplasmic reticulum of cerebellum. *Nature* 1991; 351: 751–754.
33. Lock JT, Smith IF and Parker I. Spatial-temporal patterning of Ca²⁺ signals by the subcellular distribution of IP₃ and IP₃ receptors. *Semin Cell Dev Biol* 2019; 94: 3–10.
34. Wilson C, Saunter CD, Girkin JM, et al. Pressure-dependent regulation of Ca²⁺ signalling in the vascular endothelium. *J Physiol* 2015; 593: 5231–5253.
35. Mountian II, Baba-Aïssa F, Jonas JC, et al. Expression of Ca²⁺ transport genes in platelets and endothelial cells in hypertension. *Hypertension* 2001; 37: 135–141.
36. Yuan Q, Yang J, Santulli G, et al. Maintenance of normal blood pressure is dependent on IP₃R1-mediated regulation of eNOS. *Proc Natl Acad Sci U S A* 2016; 113: 8532–8537.
37. Shibata A, Uchida K, Kodo K, et al. Type 2 inositol 1,4,5-trisphosphate receptor inhibits the progression of pulmonary arterial hypertension via calcium signaling and apoptosis. *Heart Vessels* 2019; 34: 724–734.
38. Sampieri A, Santoyo K, Asanov A, et al. Association of the IP₃R to STIM1 provides a reduced intraluminal calcium microenvironment, resulting in enhanced store-operated calcium entry. *Sci Rep* 2018; 8: 13252.
39. Ji HL, Zhao RZ, Chen ZX, et al. δ ENaC: A novel divergent amiloride-inhibitable sodium channel. *Am J Physiol Lung Cell Mol Physiol* 2012; 303: L1013–L1026.
40. Hofmann T, Schaefer M, Schultz G, et al. Transient receptor potential channels as molecular substrates of receptor-mediated cation entry. *J Mol Med (Berl)* 2000; 78: 14–25.
41. Earley S and Brayden JE. Transient receptor potential channels in the vasculature. *Physiol Rev* 2015; 95: 645–690.
42. Jiang Y, Zhou Y, Peng G, et al. Topotecan prevents hypoxia-induced pulmonary arterial hypertension and inhibits hypoxia-inducible factor-1 α and TRPC channels. *Int J Biochem Cell Biol* 2018; 104: 161–170.
43. Hofmann T, Schaefer M, Schultz G, et al. Subunit composition of mammalian transient receptor potential channels in living cells. *Proc Natl Acad Sci U S A* 2002; 99: 7461–7466.
44. Zhu X, Jiang M, Peyton M, et al. trp, a novel mammalian gene family essential for agonist-activated capacitative Ca²⁺ entry. *Cell* 1996; 85: 661–671.
45. Bavencoffe A, Zhu MX and Tian JB. New aspects of the contribution of ER to SOCE regulation: TRPC proteins as a link between plasma membrane ion transport and intracellular Ca²⁺ stores. *Adv Exp Med Biol* 2017; 993: 239–255.
46. Lin MJ, Leung GP, Zhang WM, et al. Chronic hypoxia-induced upregulation of store-operated and receptor-operated Ca²⁺ channels in pulmonary arterial smooth muscle cells: A novel mechanism of hypoxic pulmonary hypertension. *Circ Res* 2004; 95: 496–505.
47. Rohacs T and Nilius B. Regulation of transient receptor potential (TRP) channels by phosphoinositides. *Pflugers Arch* 2007; 455: 157–168.
48. Rohacs T. Phosphoinositide regulation of TRP channels. *Handb Exp Pharmacol* 2014; 223: 1143–1176.
49. Trebak M, Vazquez G, Bird GS, et al. The TRPC3/6/7 subfamily of cation channels. *Cell Calcium* 2003; 33: 451–461.
50. Shi J, Miralles F, Birnbaumer L, et al. Store-operated interactions between plasmalemmal STIM1 and TRPC1 proteins stimulate PLC β 1 to induce TRPC1 channel activation in vascular smooth muscle cells. *J Physiol* 2017; 595: 1039–1058.
51. Ambudkar IS, de Souza LB and Ong HL. TRPC1, Orai1, and STIM1 in SOCE: Friends in tight spaces. *Cell Calcium* 2017; 63: 33–39.
52. DeHaven WI, Jones BF, Petranka JG, et al. TRPC channels function independently of STIM1 and Orai1. *J Physiol* 2009; 587: 2275–2298.
53. Earley S, Heppner TJ, Nelson MT, et al. TRPV4 forms a novel Ca²⁺ signaling complex with ryanodine receptors and BKCa channels. *Circ Res* 2005; 97: 1270–1279.
54. Cussac LA, Cardouat G, Tiruchellvam Pillai N, et al. TRPV4 channel mediates adventitial fibroblast activation and adventitial remodeling in pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol* 2020; 318: L135–L146.
55. Zamponi GW, Striessnig J, Koschak A, et al. The physiology, pathology, and pharmacology of voltage-gated calcium channels and their future therapeutic potential. *Pharmacol Rev* 2015; 67: 821–870.
56. Catterall WA, Perez-Reyes E, Snutch TP, et al. International Union of Pharmacology. XLVIII. Nomenclature and

- structure-function relationships of voltage-gated calcium channels. *Pharmacol Rev* 2005; 57: 411–425.
57. Ávila-Medina J, Calderón-Sánchez E, González-Rodríguez P, et al. Orail and TRPC1 proteins co-localize with $\text{Ca}_v1.2$ channels to form a signal complex in vascular smooth muscle cells. *J Biol Chem* 2016; 291: 21148–21159.
 58. Fan G, Kaßmann M, Hashad AM, et al. Differential targeting and signalling of voltage-gated T-type $\text{Ca}_v3.2$ and L-type $\text{Ca}_v1.2$ channels to ryanodine receptors in mesenteric arteries. *J Physiol* 2018; 596: 4863–4877.
 59. Fan G, Cui Y, Gollasch M, et al. Elementary calcium signalling in arterial smooth muscle. *Channels (Austin)* 2019; 13: 505–519.
 60. Navedo MF, Amberg GC, Votaw VS, et al. Constitutively active L-type Ca^{2+} channels. *Proc Natl Acad Sci U S A* 2005; 102: 11112–11117.
 61. Santana LF, Navedo MF, Amberg GC, et al. Calcium sparklets in arterial smooth muscle. *Clin Exp Pharmacol Physiol* 2008; 35: 1121–1126.
 62. Ghosh D, Nieves-Cintrón M, Tajada S, et al. Dynamic L-type $\text{Ca}_v1.2$ channel trafficking facilitates $\text{Ca}_v1.2$ clustering and cooperative gating. *Biochim Biophys Acta Mol Cell Res* 2018; 1865: 1341–1355.
 63. Ghosh D, Syed AU, Prada MP, et al. Calcium channels in vascular smooth muscle. *Adv Pharmacol* 2017; 78: 49–87.
 64. Wan J, Yamamura A, Zimnicka AM, et al. Chronic hypoxia selectively enhances L- and T-type voltage-dependent Ca^{2+} channel activity in pulmonary artery by upregulating $\text{Ca}_v1.2$ and $\text{Ca}_v3.2$. *Am J Physiol Lung Cell Mol Physiol* 2013; 305: L154–L164.
 65. Yang H, Kuang SJ, Rao F, et al. Species-specific differences in the role of L-type Ca^{2+} channels in the regulation of coronary arterial smooth muscle contraction. *Naunyn Schmiedeberg Arch Pharmacol* 2016; 389: 151–157.
 66. Zhu Y, Ye P, Chen SL, et al. Functional regulation of large conductance Ca^{2+} -activated K^+ channels in vascular diseases. *Metabolism* 2018; 83: 75–80.
 67. Weigand L, Foxson J, Wang J, et al. Inhibition of hypoxic pulmonary vasoconstriction by antagonists of store-operated Ca^{2+} and nonselective cation channels. *Am J Physiol Lung Cell Mol Physiol* 2005; 289: L5–L13.
 68. Wang Y, Deng X, Mancarella S, et al. The calcium store sensor, STIM1, reciprocally controls Orail and $\text{Ca}_v1.2$ channels. *Science* 2010; 330: 105–109.
 69. Park CY, Shcheglovitov A and Dolmetsch R. The CRAC channel activator STIM1 binds and inhibits L-type voltage-gated calcium channels. *Science* 2010; 330: 101–105.
 70. Lambert M, Capuano V, Olschewski A, et al. Ion channels in pulmonary hypertension: A therapeutic interest? *Int J Mol Sci* 2018; 19: 3162.
 71. López-López JR, Ciudad P and Pérez-García MT. K_v channels and vascular smooth muscle cell proliferation. *Microcirculation* 2018; 25: e12427.
 72. Platoshyn O, Yu Y, Golovina VA, et al. Chronic hypoxia decreases K_v channel expression and function in pulmonary artery myocytes. *Am J Physiol Lung Cell Mol Physiol* 2001; 280: L801–L812.
 73. Yuan JX, Aldinger AM, Juhaszova M, et al. Dysfunctional voltage-gated K^+ channels in pulmonary artery smooth muscle cells of patients with primary pulmonary hypertension. *Circulation* 1998; 98: 1400–1406.
 74. McCormack T, McCormack K, Nadal MS, et al. The effects of Shaker beta-subunits on the human lymphocyte K^+ channel $\text{K}_v1.3$. *J Biol Chem* 1999; 274: 20123–20126.
 75. Beeton C, Wulff H, Standifer NE, et al. $\text{K}_v1.3$ channels are a therapeutic target for T cell-mediated autoimmune diseases. *Proc Natl Acad Sci U S A* 2006; 103: 17414–17419.
 76. Ciudad P, Moreno-Domínguez A, Novensá L, et al. Characterization of ion channels involved in the proliferative response of femoral artery smooth muscle cells. *Arterioscler Thromb Vasc Biol* 2010; 30: 1203–1211.
 77. Ciudad P, Miguel-Velado E, Ruiz-McDavitt C, et al. $\text{K}_v1.3$ channels modulate human vascular smooth muscle cells proliferation independently of mTOR signaling pathway. *Pflugers Arch* 2015; 467: 1711–1722.
 78. Neylon CB. Potassium channels and vascular proliferation. *Vascul Pharmacol* 2002; 38: 35–41.
 79. Lv Y, Fu L, Zhang Z, et al. Increased expression of microRNA-206 inhibits potassium voltage-gated channel subfamily A member 5 in pulmonary arterial smooth muscle cells and is related to exaggerated pulmonary artery hypertension following intrauterine growth retardation in rats. *J Am Heart Assoc* 2019; 8: e010456.
 80. Pardo LA. Voltage-gated potassium channels in cell proliferation. *Physiology (Bethesda)* 2004; 19: 285–292.
 81. Zhang S, Liu B, Fan Z, et al. Targeted inhibition of survivin with YM155 promotes apoptosis of hypoxic human pulmonary arterial smooth muscle cells via the upregulation of voltage-dependent K^+ channels. *Mol Med Rep* 2016; 13: 3415–3422.
 82. McMurtry MS, Archer SL, Altieri DC, et al. Gene therapy targeting survivin selectively induces pulmonary vascular apoptosis and reverses pulmonary arterial hypertension. *J Clin Invest* 2005; 115: 1479–1491.
 83. Lasch M, Caballero Martinez A, Kumaraswami K, et al. Contribution of the potassium channels $\text{K}_v1.3$ and $\text{K}_{Ca}3.1$ to smooth muscle cell proliferation in growing collateral arteries. *Cells* 2020; 9: 913.
 84. Bobi J, Garabito M, Solanes N, et al. $\text{K}_v1.3$ blockade inhibits proliferation of vascular smooth muscle cells in vitro and intimal hyperplasia in vivo. *Transl Res* 2020; 224: 40–54.
 85. Gardos G. The function of calcium in the potassium permeability of human erythrocytes. *Biochim Biophys Acta* 1958; 30: 653–654.
 86. Adelman JP, Maylie J and Sah P. Small-conductance Ca^{2+} -activated K^+ channels: Form and function. *Annu Rev Physiol* 2012; 74: 245–269.
 87. Mandegar M, Remillard CV and Yuan JX. Ion channels in pulmonary arterial hypertension. *Prog Cardiovasc Dis* 2002; 45: 81–114.
 88. Clements RT, Terentyev D and Sellke FW. Ca^{2+} -activated K^+ channels as therapeutic targets for myocardial and vascular protection. *Circ J* 2015; 79: 455–462.
 89. Latorre R, Castillo K, Carrasquel-Ursulaez W, et al. Molecular determinants of BK channel functional diversity and functioning. *Physiol Rev* 2017; 97: 39–87.
 90. Xiong Y, Aroor AR, Ramirez-Perez FI, et al. Western diet induces renal artery endothelial stiffening that is dependent

- on the epithelial Na⁺ channel. *Am J Physiol Renal Physiol* 2020; 318: F1220–F1228.
91. Chen L, Bi D, Lu ZH, et al. Distinct domains of the β 1-subunit cytosolic N terminus control surface expression and functional properties of large-conductance calcium-activated potassium (BK) channels. *J Biol Chem* 2017; 292: 8694–8704.
 92. Joseph BK, Thakali KM, Moore CL, et al. Ion channel remodeling in vascular smooth muscle during hypertension: Implications for novel therapeutic approaches. *Pharmacol Res* 2013; 70: 126–138.
 93. Ledoux J, Werner ME, Brayden JE, et al. Calcium-activated potassium channels and the regulation of vascular tone. *Physiology (Bethesda)* 2006; 21: 69–78.
 94. Hu XQ and Zhang L. Function and regulation of large conductance Ca²⁺-activated K⁺ channel in vascular smooth muscle cells. *Drug Discov Today* 2012; 17: 974–987.
 95. Singh H, Stefani E and Toro L. Intracellular BK(Ca) (iBK (Ca)) channels. *J Physiol* 2012; 590: 5937–5947.
 96. Hou S, Heinemann SH and Hoshi T. Modulation of BKCa channel gating by endogenous signaling molecules. *Physiology (Bethesda)* 2009; 24: 26–35.
 97. Yan J, Chen R, Liu P, et al. Docosahexaenoic acid attenuates hypoxic pulmonary vasoconstriction by activating the large conductance Ca²⁺-activated K⁺ currents in pulmonary artery smooth muscle cells. *Pulm Pharmacol Ther* 2014; 28: 9–16.
 98. Gessner G, Heller R, Hoshi T, et al. The amiodarone derivative 2-methyl-3-(3,5-diiodo-4-carboxymethoxybenzyl)benzofuran (KB130015) opens large-conductance Ca²⁺-activated K⁺ channels and relaxes vascular smooth muscle. *Eur J Pharmacol* 2007; 555: 185–193.
 99. Plüger S, Faulhaber J, Fürstenau M, et al. Mice with disrupted BK channel beta1 subunit gene feature abnormal Ca²⁺ spark/STOC coupling and elevated blood pressure. *Circ Res* 2000; 87: E53–E60.
 100. Si H, Heyken WT, Wölfle SE, et al. Impaired endothelium-derived hyperpolarizing factor-mediated dilations and increased blood pressure in mice deficient of the intermediate-conductance Ca²⁺-activated K⁺ channel. *Circ Res* 2006; 99: 537–544.
 101. Minami K, Hirata Y, Tokumura A, et al. Protein kinase C-independent inhibition of the Ca²⁺-activated K⁺ channel by angiotensin II and endothelin-1. *Biochem Pharmacol* 1995; 49: 1051–1056.
 102. Peng W, Michael JR, Hoidal JR, et al. ET-1 modulates KCa-channel activity and arterial tension in normoxic and hypoxic human pulmonary vasculature. *Am J Physiol* 1998; 275: L729–L739.
 103. Guo S, Shen Y, He G, et al. Involvement of Ca²⁺-activated K⁺ channel 3.1 in hypoxia-induced pulmonary arterial hypertension and therapeutic effects of TRAM-34 in rats. *Biosci Rep* 2017; 37: BSR20170763.
 104. Ketchum KA, Joiner WJ, Sellers AJ, et al. A new family of outwardly rectifying potassium channel proteins with two pore domains in tandem. *Nature* 1995; 376: 690–695.
 105. Sepúlveda FV, Pablo Cid L, Teulon J, et al. Molecular aspects of structure, gating, and physiology of pH-sensitive background K_{2P} and Kir K⁺-transport channels. *Physiol Rev* 2015; 95: 179–217.
 106. Feliciangeli S, Chatelain FC, Bichet D, et al. The family of K_{2P} channels: Salient structural and functional properties. *J Physiol* 2015; 593: 2587–2603.
 107. Olschewski A, Li Y, Tang B, et al. Impact of TASK-1 in human pulmonary artery smooth muscle cells. *Circ Res* 2006; 98: 1072–1080.
 108. Fink M, Duprat F, Lesage F, et al. Cloning, functional expression and brain localization of a novel unconventional outward rectifier K⁺ channel. *EMBO J* 1996; 15: 6854–6862.
 109. Lesage F, Guillemare E, Fink M, et al. TWIK-1, a ubiquitous human weakly inward rectifying K⁺ channel with a novel structure. *EMBO J* 1996; 15: 1004–1011.
 110. Nielsen G, Wandall-Frostholm C, Sadda V, et al. Alterations of N-3 polyunsaturated fatty acid-activated K_{2P} channels in hypoxia-induced pulmonary hypertension. *Basic Clin Pharmacol Toxicol* 2013; 113: 250–258.
 111. Antigny F, Hautefort A, Meloche J, et al. Potassium channel subfamily K member 3 (KCNK3) contributes to the development of pulmonary arterial hypertension. *Circulation* 2016; 133: 1371–1385.
 112. Lambert M, Boet A, Rucker-Martin C, et al. Loss of KCNK3 is a hallmark of RV hypertrophy/dysfunction associated with pulmonary hypertension. *Cardiovasc Res* 2018; 114: 880–893.
 113. Lambert M, Capuano V, Boet A, et al. Characterization of Kenk3-mutated rat, a novel model of pulmonary hypertension. *Circ Res* 2019; 125: 678–695.
 114. Han L, Song N, Hu X, et al. Inhibition of RELM- β prevents hypoxia-induced overproliferation of human pulmonary artery smooth muscle cells by reversing PLC-mediated KCNK3 decline. *Life Sci* 2020; 246: 117419.
 115. Aziz Q, Thomas AM, Gomes J, et al. The ATP-sensitive potassium channel subunit, Kir6.1, in vascular smooth muscle plays a major role in blood pressure control. *Hypertension* 2014; 64: 523–529.
 116. Nichols CG. K_{ATP} channels as molecular sensors of cellular metabolism. *Nature* 2006; 440: 470–476.
 117. Bohnen MS, Ma L, Zhu N, et al. Loss-of-function ABCC8 mutations in pulmonary arterial hypertension. *Circ Genom Precis Med* 2018; 11: e002087.
 118. Tinker A, Aziz Q, Li Y, et al. ATP-sensitive potassium channels and their physiological and pathophysiological roles. *Compr Physiol* 2018; 8: 1463–1511.
 119. Aziz Q, Li Y and Tinker A. Endothelial biology and ATP-sensitive potassium channels. *Channels (Austin)* 2018; 12: 45–46.
 120. Quayle JM, Nelson MT and Standen NB. ATP-sensitive and inwardly rectifying potassium channels in smooth muscle. *Physiol Rev* 1997; 77: 1165–1232.
 121. Shi WW, Yang Y, Shi Y, et al. K_{ATP} channel action in vascular tone regulation: From genetics to diseases. *Sheng Li Xue Bao* 2012; 64: 1–13.
 122. McClenaghan C, Woo KV and Nichols CG. Pulmonary hypertension and ATP-sensitive potassium channels. *Hypertension* 2019; 74: 14–22.
 123. Liu X, Duan P, Hu X, et al. Altered K_{ATP} channel subunits expression and vascular reactivity in spontaneously hypertensive rats with age. *J Cardiovasc Pharmacol* 2016; 68: 143–149.
 124. Hayabuchi Y, Davies NW and Standen NB. Angiotensin II inhibits rat arterial K_{ATP} channels by inhibiting steady-state

- protein kinase A activity and activating protein kinase C. *J Physiol* 2001; 530: 193–205.
125. Park WS, Ko EA, Han J, et al. Endothelin-1 acts via protein kinase C to block K_{ATP} channels in rabbit coronary and pulmonary arterial smooth muscle cells. *J Cardiovasc Pharmacol* 2005; 45: 99–108.
 126. Zhu R, Bi LQ, Wu SL, et al. Iptakalim attenuates hypoxia-induced pulmonary arterial hypertension in rats by endothelial function protection. *Mol Med Rep* 2015; 12: 2945–2952.
 127. Xie W, Wang H, Wang H, et al. Effects of iptakalim hydrochloride, a novel K_{ATP} channel opener, on pulmonary vascular remodeling in hypoxic rats. *Life Sci* 2004; 75: 2065–2076.
 128. Friedel HA and Brogden RN. Pinacidil. A review of its pharmacodynamic and pharmacokinetic properties, and therapeutic potential in the treatment of hypertension. *Drugs* 1990; 39: 929–967.
 129. Jahangir A and Terzic A. K_{ATP} channel therapeutics at the bedside. *J Mol Cell Cardiol* 2005; 39: 99–112.
 130. Taggart M. The knack of resisting increases in intravascular pressure: The potential role for epithelial Na^+ channel protein subunits in arterial myogenic reactivity. *Exp Physiol* 2012; 97: 474–475.
 131. Canessa CM, Schild L, Buell G, et al. Amiloride-sensitive epithelial Na^+ channel is made of three homologous subunits. *Nature* 1994; 367: 463–467.
 132. Li Q and Fung E. Multifaceted functions of epithelial Na^+ channel in modulating blood pressure. *Hypertension* 2019; 73: 273–281.
 133. Drummond HA, Price MP, Welsh MJ, et al. A molecular component of the arterial baroreceptor mechanotransducer. *Neuron* 1998; 21: 1435–1441.
 134. Drummond HA, Welsh MJ and Abboud FM. ENaC subunits are molecular components of the arterial baroreceptor complex. *Ann N Y Acad Sci* 2001; 940: 42–47.
 135. Amin MS, Wang HW, Reza E, et al. Distribution of epithelial sodium channels and mineralocorticoid receptors in cardiovascular regulatory centers in rat brain. *Am J Physiol Regul Integr Comp Physiol* 2005; 289: R1787–R1797.
 136. Drummond HA, Gebremedhin D and Harder DR. Degenerin/epithelial Na^+ channel proteins: Components of a vascular mechanosensor. *Hypertension* 2004; 44: 643–648.
 137. Lin W, Finger TE, Rossier BC, et al. Epithelial Na^+ channel subunits in rat taste cells: Localization and regulation by aldosterone. *J Comp Neurol* 1999; 405: 406–420.
 138. Korte S, Sträter AS, Drüppel V, et al. Feedforward activation of endothelial ENaC by high sodium. *FASEB J* 2014; 28: 4015–4025.
 139. Oberleithner H, Riethmüller C, Schillers H, et al. Plasma sodium stiffens vascular endothelium and reduces nitric oxide release. *Proc Natl Acad Sci U S A* 2007; 104: 16281–16286.
 140. Oberleithner H, Riethmüller C, Ludwig T, et al. Aldosterone remodels human endothelium. *Acta Physiol (Oxf)* 2006; 187: 305–312.
 141. Pérez FR, Venegas F, González M, et al. Endothelial epithelial sodium channel inhibition activates endothelial nitric oxide synthase via phosphoinositide 3-kinase/Akt in small-diameter mesenteric arteries. *Hypertension* 2009; 53: 1000–1007.
 142. Jernigan NL and Drummond HA. Vascular ENaC proteins are required for renal myogenic constriction. *Am J Physiol Renal Physiol* 2005; 289: F891–F901.
 143. Kim EC, Choi SK, Lim M, et al. Role of endogenous ENaC and TRP channels in the myogenic response of rat posterior cerebral arteries. *PLoS One* 2013; 8: e84194.
 144. Kim EC, Ahn DS, Yeon SI, et al. Epithelial Na^+ channel proteins are mechanotransducers of myogenic constriction in rat posterior cerebral arteries. *Exp Physiol* 2012; 97: 544–555.
 145. Choi SK, Yeon SI, Kwon Y, et al. Involvement of epithelial Na^+ channel in the elevated myogenic response in posterior cerebral arteries from spontaneously hypertensive rats. *Sci Rep* 2017; 7: 45996.
 146. Li YL, Zhang D, Tu H, et al. Altered ENaC is associated with aortic baroreceptor dysfunction in chronic heart failure. *Am J Hypertens* 2016; 29: 582–589.
 147. Baldin JP, Barth D and Fronius M. Epithelial Na^+ channel (ENaC) formed by one or two subunits forms functional channels that respond to shear force. *Front Physiol* 2020; 11: 141.
 148. Jia G, Habibi J, Aroor AR, et al. Epithelial sodium channel in aldosterone-induced endothelium stiffness and aortic dysfunction. *Hypertension* 2018; 72: 731–738.
 149. Philipson KD and Nicoll DA. Sodium-calcium exchange: A molecular perspective. *Annu Rev Physiol* 2000; 62: 111–133.
 150. Liao J, Li H, Zeng W, et al. Structural insight into the ion-exchange mechanism of the sodium/calcium exchanger. *Science* 2012; 335: 686–690.
 151. Khananshvili D. The SLC8 gene family of sodium-calcium exchangers (NCX) - Structure, function, and regulation in health and disease. *Mol Aspects Med* 2013; 34: 220–235.
 152. Zhang S, Dong H, Rubin LJ, et al. Upregulation of Na^+/Ca^{2+} exchanger contributes to the enhanced Ca^{2+} entry in pulmonary artery smooth muscle cells from patients with idiopathic pulmonary arterial hypertension. *Am J Physiol Cell Physiol* 2007; 292: C2297–C2305.
 153. Liu B, Zhang B, Huang S, et al. Ca^{2+} entry through reverse mode Na^+/Ca^{2+} exchanger contributes to store operated channel-mediated neointima formation after arterial injury. *Can J Cardiol* 2018; 34: 791–799.
 154. Neubert P, Homann A, Wendelborn D, et al. NCX1 represents an ionic Na^+ sensing mechanism in macrophages. *PLoS Biol* 2020; 18: e3000722.
 155. Orłowski J, Kandasamy RA and Shull GE. Molecular cloning of putative members of the Na/H exchanger gene family. cDNA cloning, deduced amino acid sequence, and mRNA tissue expression of the rat Na/H exchanger NHE-1 and two structurally related proteins. *J Biol Chem* 1992; 267: 9331–9339.
 156. Wang Z, Orłowski J and Shull GE. Primary structure and functional expression of a novel gastrointestinal isoform of the rat Na/H exchanger. *J Biol Chem* 1993; 268: 11925–11928.
 157. Lucchesi PA, DeRoux N and Berk BC. Na^+-H^+ exchanger expression in vascular smooth muscle of spontaneously hypertensive and Wistar-Kyoto rats. *Hypertension* 1994; 24: 734–738.

158. Huetsch J and Shimoda LA. Na^+/H^+ exchange and hypoxic pulmonary hypertension. *Pulm Circ* 2015; 5: 228–243.
159. Yu L, Quinn DA, Garg HG, et al. Deficiency of the NHE1 gene prevents hypoxia-induced pulmonary hypertension and vascular remodeling. *Am J Respir Crit Care Med* 2008; 177: 1276–1284.
160. Rios EJ, Fallon M, Wang J, et al. Chronic hypoxia elevates intracellular pH and activates Na^+/H^+ exchange in pulmonary arterial smooth muscle cells. *Am J Physiol Lung Cell Mol Physiol* 2005; 289: L867–L874.
161. Wakabayashi I, Marumo M and Sotoda Y. Intracellular alkalinization augments capacitative Ca^{2+} entry in vascular smooth muscle cells. *J Cardiovasc Pharmacol* 2003; 41: 903–907.
162. Tsukioka M, Iino M and Endo M. pH dependence of inositol 1,4,5-trisphosphate-induced Ca^{2+} release in permeabilized smooth muscle cells of the guinea-pig. *J Physiol* 1994; 475: 369–375.
163. Wakabayashi I, Sakamoto K, Hatake K, et al. Potentiating effect of NH_4Cl on vasoconstriction in rat aorta. *Biochem Biophys Res Commun* 1991; 178: 808–814.
164. Boedtker E, Damkier HH and Aalkjaer C. NHE1 knockout reduces blood pressure and arterial media/lumen ratio with no effect on resting pH_i in the vascular wall. *J Physiol* 2012; 590: 1895–1906.
165. Shimoda LA, Fallon M, Pisarcik S, et al. HIF-1 regulates hypoxic induction of NHE1 expression and alkalinization of intracellular pH in pulmonary arterial myocytes. *Am J Physiol Lung Cell Mol Physiol* 2006; 291: L941–L949.
166. Pisarcik S, Maylor J, Lu W, et al. Activation of hypoxia-inducible factor-1 in pulmonary arterial smooth muscle cells by endothelin-1. *Am J Physiol Lung Cell Mol Physiol* 2013; 304: L549–L561.
167. Huetsch JC, Jiang H, Larrain C, et al. The Na^+/H^+ exchanger contributes to increased smooth muscle proliferation and migration in a rat model of pulmonary arterial hypertension. *Physiol Rep* 2016; 4: e12729.
168. Okada Y, Okada T, Sato-Numata K, et al. Cell volume-activated and volume-correlated anion channels in mammalian cells: Their biophysical, molecular, and pharmacological properties. *Pharmacol Rev* 2019; 71: 49–88.
169. Jentsch TJ, Stein V, Weinreich F, et al. Molecular structure and physiological function of chloride channels. *Physiol Rev* 2002; 82: 503–568.
170. Kunzelmann K, Kongsuphol P, Chootip K, et al. Role of the Ca^{2+} -activated Cl^- channels bestrophin and anoctamin in epithelial cells. *Biol Chem* 2011; 392: 125–134.
171. Kamaledin MA. Molecular, biophysical, and pharmacological properties of calcium-activated chloride channels. *J Cell Physiol* 2018; 233: 787–798.
172. Qu Z and Hartzell HC. Anion permeation in Ca^{2+} -activated Cl^- channels. *J Gen Physiol* 2000; 116: 825–844.
173. Pusch M. Ca^{2+} -activated chloride channels go molecular. *J Gen Physiol* 2004; 123: 323–325.
174. Peters CJ, Gilchrist JM, Tien J, et al. The sixth transmembrane segment is a major gating component of the TMEM16A calcium-activated chloride channel. *Neuron* 2018; 97: 1063–1077.e4.
175. Contreras-Vite JA, Cruz-Rangel S, De Jesús-Pérez JJ, et al. Revealing the activation pathway for TMEM16A chloride channels from macroscopic currents and kinetic models. *Pflugers Arch* 2016; 468: 1241–1257.
176. Cruz-Rangel S, De Jesús-Pérez JJ, Aréchiga-Figueroa IA, et al. Extracellular protons enable activation of the calcium-dependent chloride channel TMEM16A. *J Physiol* 2017; 595: 1515–1531.
177. Large WA and Wang Q. Characteristics and physiological role of the Ca^{2+} -activated Cl^- conductance in smooth muscle. *Am J Physiol* 1996; 271: C435–C454.
178. Yuan XJ. Role of calcium-activated chloride current in regulating pulmonary vasomotor tone. *Am J Physiol* 1997; 272: L959–L968.
179. Hogg RC, Wang Q and Large WA. Action of niflumic acid on evoked and spontaneous calcium-activated chloride and potassium currents in smooth muscle cells from rabbit portal vein. *Br J Pharmacol* 1994; 112: 977–984.
180. Leblanc N and Leung PM. Indirect stimulation of Ca^{2+} -activated Cl^- current by $\text{Na}^+/\text{Ca}^{2+}$ exchange in rabbit portal vein smooth muscle. *Am J Physiol* 1995; 268: H1906–H1917.
181. Forrest AS, Angermann JE, Raghunathan R, et al. Intricate interaction between store-operated calcium entry and calcium-activated chloride channels in pulmonary artery smooth muscle cells. *Adv Exp Med Biol* 2010; 661: 31–55.
182. Matchkov VV, Boedtker DM and Aalkjaer C. The role of Ca^{2+} activated Cl^- channels in blood pressure control. *Curr Opin Pharmacol* 2015; 21: 127–137.
183. Duan D. Phenomics of cardiac chloride channels: The systematic study of chloride channel function in the heart. *J Physiol* 2009; 587: 2163–2177.
184. Sun Y, Birnbaumer L and Singh BB. TRPC1 regulates calcium-activated chloride channels in salivary gland cells. *J Cell Physiol* 2015; 230: 2848–2856.
185. Hengl T, Kaneko H, Dauner K, et al. Molecular components of signal amplification in olfactory sensory cilia. *Proc Natl Acad Sci U S A* 2010; 107: 6052–6057.
186. Shang L, Wang K, Liu D, et al. TMEM16A regulates the cell cycle of pulmonary artery smooth muscle cells in high-flow-induced pulmonary arterial hypertension rat model. *Exp Ther Med* 2020; 19: 3275–3281.
187. Papp R, Nagaraj C, Zabini D, et al. Targeting TMEM16A to reverse vasoconstriction and remodeling in idiopathic pulmonary arterial hypertension. *Eur Respir J* 2019; 53: 1800965.
188. Zeng XL, Sun L, Zheng HQ, et al. Smooth muscle-specific TMEM16A expression protects against angiotensin II-induced cerebrovascular remodeling via suppressing extracellular matrix deposition. *J Mol Cell Cardiol* 2019; 134: 131–143.
189. Qu M, Lu P, Bellve K, et al. Smooth muscle cell-specific TMEM16A deletion does not alter Ca^{2+} signaling, uterine contraction, gestation length, or litter size in mice. *Biol Reprod* 2019; 101: 318–327.
190. Heinze C, Seniuk A, Sokolov MV, et al. Disruption of vascular Ca^{2+} -activated chloride currents lowers blood pressure. *J Clin Invest* 2014; 124: 675–686.
191. Ji Q, Guo S, Wang X, et al. Recent advances in TMEM16A: Structure, function, and disease. *J Cell Physiol* 2019; 234: 7856–7873.

192. Nilius B, Eggermont J, Voets T, et al. Properties of volume-regulated anion channels in mammalian cells. *Prog Biophys Mol Biol* 1997; 68: 69–119.
193. Greenwood IA and Large WA. Properties of a Cl⁻ current activated by cell swelling in rabbit portal vein vascular smooth muscle cells. *Am J Physiol* 1998; 275: H1524–H1532.
194. Remillard CV, Lupien MA, Crépeau V, et al. Role of Ca²⁺- and swelling-activated Cl⁻ channels in alpha1-adrenoceptor-mediated tone in pressurized rabbit mesenteric arterioles. *Cardiovasc Res* 2000; 46: 557–568.
195. Bhogal S, Khraisha O, Al Madani M, et al. Sildenafil for pulmonary arterial hypertension. *Am J Ther* 2019; 26: e520–e526.
196. Lau EMT, Giannoulatou E, Celermajer DS, et al. Epidemiology and treatment of pulmonary arterial hypertension. *Nat Rev Cardiol* 2017; 14: 603–614.
197. Dodson MW, Brown LM and Elliott CG. Pulmonary arterial hypertension. *Heart Fail Clin* 2018; 14: 255–269.
198. Rhee SW, Stimers JR, Wang W, et al. Vascular smooth muscle-specific knockdown of the noncardiac form of the L-type calcium channel by microRNA-based short hairpin RNA as a potential antihypertensive therapy. *J Pharmacol Exp Ther* 2009; 329: 775–782.
199. Hu Z, Li G, Wang JW, et al. Regulation of blood pressure by targeting Ca_v1.2-galectin-1 protein interaction. *Circulation* 2018; 138: 1431–1445.
200. Zhancheng W, Wenhui J, Yun J, et al. The dominant models of KCNJ11 E23K and KCNMB1 E65K are associated with essential hypertension (EH) in Asian: Evidence from a meta-analysis. *Medicine (Baltimore)* 2019; 98: e15828.
201. Tykocki NR, Boerman EM and Jackson WF. Smooth muscle ion channels and regulation of vascular tone in resistance arteries and arterioles. *Compr Physiol* 2017; 7: 485–581.