



Evaluation of Serum Concentration of the Myokine Irisin (FNDC5) in Patients with Age-Related **Macular Degeneration**

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Abstract

Objectives: This study evaluated the serum irisin level of patients with age-related macular degeneration (ARMD) and compared it with that of healthy individuals.

Methods: The serum irisin level of 15 healthy controls (Group 1) and 15 dry ARMD patients (Group 2) and 15 wet ARMD patients (Group 3) were measured using the enzyme-linked immunosorbent assay (ELISA) method and compared. **Results:** There was no statistically significant difference between the groups in terms of age or gender (p>0.05). The mean serum irisin levels of Group 1, Group 2, and Group 3 were 25.81±24.04 ng/mL, 22.93±19.05 ng/mL, and 12.38±8.16 ng/mL, respectively. Although the mean irisin level in the wet ARMD patients was lower than that of the control and dry ARMD groups, there was no statistically significant difference between the groups (p>0.05).

Conclusion: The results suggest that the serum irisin level in ARMD patients is not different from that of healthy individuals. Studies of larger groups that examine the irisin level in the vitreous and neovascular membranes will further elucidate any role in the pathogenesis of ARMD.

Keywords: Age-related macular degeneration, dry, FNDC5, irisin, serum level, wet

Introduction

Age-related macular degeneration (ARMD) is a chronic and progressive disease of the macula that causes vision loss in people over 50 years old in developed countries. In ARMD, neurodegeneration which affects photoreceptor cells, retina pigment epithelium (RPE), Bruch membrane, and choriocapillaris in the macular region develops (1, 2). ARMD is

divided into two clinical types as dry (non-neovascular and non-exudative) and wet (neovascular and exudative) ARMD. Dry ARMD (d-ARMD) includes macular RPE changes and drusen in early stages and macular atrophy in late stages. In wet ARMD (w-ARMD), the choroidal neovascular membrane (CNVM) and RPE detachment are developed. The w-ARMD, which is seen in 10-15% of patients with ARMD,

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is responsible for about 90% of visual loss due to the complications associated with neovascularization (NV) (1, 2). Vascular endothelial growth factor-A (VEGF-A) is the main factor responsible for NV and increased vascular permeability in w-ARMD (3-6). At present, intravitreal administration of various anti-VEGF agents is the most important treatment in ARMD treatment, and it has been widely demonstrated that vision loss due to w-ARMD could be markedly prevented by this intervention (7).

Irisin is an exercise-induced, glycosylated protein that is defined as myokine and adipokine. It is formed by the proteolytic cleavage of fibronectin type III domain-containing protein 5 (FNDC5) in muscle tissue. Irisin turns white adipose tissue cells into brown adipose tissue cells through exercise. Elevated irisin level causes an increase in energy metabolism, weight loss, and heat in the body and improves glucose tolerance (8-10). Irisin was first discovered from mouse skeletal muscle, and it was shown that FNDC5 mRNA, the precursor of the irisin, was existed in various tissues or organs. In addition, irisin is released highly in the optic nerve and lower concentration in the retina (11). It has been reported that irisin immunoreactivity is present in the porcupine neuroretina (12). In another study, it has been demonstrated that irisin immunoreactivity was found in all layers of the retina excluding the outer nuclear layer in hamsters (13).

Recent reports demonstrated that serum or plasma levels of irisin are decreased in many patients with metabolic syndromes and related diseases (14). High plasma irisin levels are associated with the stage of diabetic retinopathy (DR) (15). It has been also suggested that irisin might protect against DR with potential anti-IL-17A effects (16). In addition, it has been reported that the patients with proliferative DR had decreased serum and vitreous irisin levels compared with the control group and type 2 diabetic patients without DR and that irisin levels were associated with the presence of diabetic nephropathy and DR (17).

To PubMed search, we could not detect a report on the serum irisin levels in any ophthalmic disease in the literature. In light of the recent data, we hypothesized that irisin might play a role in ARMD pathogenesis, and our study, we aimed to evaluate the levels of irisin in the serum in patients with ARMD.

Methods

This pilot study was conducted in full accordance with the Declaration of Helsinki and approved by the ethics committee of the university (Date: 30.03.2017, Number: 16). All the study subjects were provided informed consent.

A full ocular examination containing best-corrected visual acuity, slit-lamp biomicroscopic examination, ocular tonometry, indirect ophthalmoscopy, fundus fluorescein angiography (FFA, Zeiss FF 450 IR plus), and optic coherence tomography (OCT, Zeiss Cirrus HD-5000) were performed for all study subjects. ARMD classification as d-ARMD or w-ARMD was performed by a single retina specialist.

The study included three groups: Group I included I5 sex- and age-matched healthy control subjects without drusen, RPE changes, or CNVM. Group 2 included I5 dry ARMD patients having macular RPE changes and/or drusen or geographic atrophy without CNVM detected by FFA and OCT. Group 3 included I5 wet ARMD patients with CNVM or disciform scar detected by FFA and OCT.

The patients with morbid obesity, and/or chronic alcohol abuse, and any hematological, psychiatric, immune, malignant, obvious inflammatory, systemic infectious, connective tissue, cardiac, renal, hepatic, cerebrovascular disease, cardiovascular disease (such as diabetes mellitus, systemic hypertension, etc.), and the patients with ocular infection and inflammation, retinal vaso-occlusive disease, and patients having retinal laser photocoagulation or intravitreal injection were not admitted to the study.

The Assay of Serum Irisin Concentrations (18)

Blood samples were taken from patients and healthy controls to measure irisin levels at 09.00 h after overnight fasting in all subjects. All subjects were rested for 15 min before the blood collection process. Samples were delivered to the laboratory within 20 min. They were centrifuged in 2000 turns for 10 min at 4°C. The sera are stored at -80°C until biochemical assay. For serum irisin measurements, a commercial kit (Sunred Bio, Baoshan, Shanghai) was used. Serum irisin levels were assayed by enzyme-linked immunosorbent test (ELISA) following the manufacturer's instructions. The minimum detectable level (sensitivity) was less than 0.157 ng/mL and the assay range for irisin was 0.2-60 ng/mL. Intra- and interassay CVs were less than 10% and 12%, respectively. All samples were measured spectrophotometrically using ELx-800TM Absorbance Microplate Reader (BioTek Instruments, Inc., Winooski, VT, USA) at 450 nm. The biochemist blindly assayed samples. The results are presented as ng/mL.

Statistical Analysis

Results are given as means \pm SD. The data provided from the subjects were entered into the Statistical Package for the Social Sciences, version 11.0 (SPSS Inc., Chicago, IL) for statistical analysis. Individual group parameters were assessed with the one-sample Kolmogorov–Smirnov Z test and were found to be abnormally distributed (p<0.05). Hence, statistical comparisons between groups were performed by the non-parametric Kruskal–Wallis and the Mann–Whitney U-test. For all comparisons, statistical significance was defined by p<0.05.

Results

There was no statistically significant difference between the groups in terms of age and gender (p>0.05). The mean serum irisin levels of Group I, Group 2, and Group 3 were found 25.81±24.04 ng/mL, 22.93±19.05 ng/mL, and 12.38±8.16 ng/mL, respectively. Although the mean irisin levels in w-ARMD patients seem to be numerically lower than those in the control and d-ARMD groups, there was no statistically significant difference between the groups (p>0.05) (Table I).

Discussion

Although there is some evidence that irisin promotes angiogenesis through stimulation endothelium cells migration and tubule formation (19) and these limited data are controversial, it has been speculated that possible protective mechanisms of irisin on the retina are local delivery of FNDC5, then irisin production, and transport into the central nervous system and brain-derived neurotrophic factor (BDN-F)-producing retina cells of irisin, subsequently, the induction of BDNF expression through a Pgc1alpha pathway and last TrkB activation (20-22).

Recent studies have reported that irisin secreted the following exercise might be a beneficial role in brain function in neurodegenerative diseases such as Alzheimer's disease (23-27).

Irisin is a peptide secreted by muscle and regulates energy metabolism. It has been well-known that BDNF plays an important role in synaptic functions and neuronal survival. It has been reported that exercise-induced irisin increases the efficacy of BDNF in the hippocampus. On the other hand, irisin prevents the ischemia-induced neuronal injury due to oxidative stress through a reduction in the secretion of a pro-inflammatory cytokine such as tumor necrosis factor- α . In addition, a recent study suggested that irisin has a neuroprotective effect through the suppression of ROS-NLRP3 inflammatory signaling, expression of interleukin-1 β , and the activation of caspase I signaling in ischemic conditions (28-39).

Recently, Guler et al. (40) reported that irisin levels increased in the aqueous humor of patients with pseudoexfoliative glaucoma. In addition, we recently reported that serum irisin levels are not different in various open-angle glaucoma types (17). As it has been considered that low levels of chronic inflammation in choroid and complement system pathologies were played role in ARMD pathogenesis, irisin may be a potential candidate for ARMD treatment through the suppression of inflammatory processing (41-46).

In the setting of our study, we hypothesized that the levels of irisin in the serum in ARMD patients may be lower than those of healthy controls because of the possible tissue-protective effects of irisin on the retina and choroid. We found that there was no statistically significant difference between the groups concerning the mean serum irisin levels. Although its levels in w-ARMD patients seem to be numerically lower than those in the control and d-ARMD groups.

To the best of our knowledge, it seems like the first study investigating serum irisin level in ARMD patients. Although the main limitation of our study is the lower cohort number, our results suggest that serum irisin levels do not change in patients with ARMD. The insignificant results concerning the levels of serum irisin in our study may be due to a local neurodegeneration nature of ARMD. However, yet, in the light of literature, we speculate that irisin may be a protective myokine against ARMD by possible effects including suppression of the inflammation or neuroprotection on neural and vascular tissues. Thus, as a next step, the measurement of both vitreous and aqueous humor levels of irisin in ARMD patients with and without treatment may obtain detailed and realistic comments about the effects of irisin in ARMD patients. Further researches are needed to have more information on the effects of irisin and investigate the levels of free and bound irisin and determine the exact role of irisin in the pathogenesis of ARMD. Studies in which large groups are formed and iris levels in vitreous and neovascular membranes are measured are necessary to reveal whether iris plays a role in ARMD pathogenesis.

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Table 1. Comparison of serum irisin levels between the groups					
Study groups	Subject Number	Serum Irisin (ng/mL)	Standard deviation	p-value	Significance (Bonferroni corrected Mann–Whitney U test)
Control	15	25.81	24.04	>0.05	d-ARMD versus w-ARMD
d-ARMD	15	22.93	19.05		Control versus d-ARMD
w-ARMD	15	12.38	8.16		Control versus w-ARMD

d-ARMD: Dry age-related macular degeneration; w-ARMD: Wet age-related macular degeneration.

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Disclosures

Ethics Committee Approval: Firat University Faculty of Medicine Ethics Committee, protocol number: 16, Date: 30/03/2017.

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Conflict of Interest: None declared.

Authorship Contributions: Involved in design and conduct of the study (BT, KM, NI, SSA); preparation and review of the study (BT, KM, SSA); data collection (BT, KM, SSA); and statistical analysis (BT, SSA).

References

- Kniggendorf V, Dreyfuss JL, Regatieri CV. Age-related macular degeneration: a review of current therapies and new treatments. Arq Bras Oftalmol 2020;83:552–61.
- 2. Ambati J, Fowler BJ. Mechanisms of age-related macular degeneration. Neuron 2012;75:26–39.
- Wu J, Sun X. Complement system and age-related macular degeneration: drugs and challenges. Drug Des Devel Ther 2019;13:2413–25.
- Tsai DC, Charng MJ, Lee FL, Hsu WM, Chen SJ. Different plasma levels of vascular endothelial growth factor and nitric oxide between patients with choroidal and retinal neovascularization. Ophthalmologica 2006;220:246–51.
- Penn JS, Madan A, Caldwell RB, Bartoli M, Caldwell RW, Hartnett ME. Vascular endothelial growth factor in eye disease. Prog Retin Eye Res 2008;27:331–71.
- Wallace GR, John Curnow S, Wloka K, Salmon M, Murray PI. The role of chemokines and their receptors in ocular disease. Prog Retin Eye Res 2004;23:435–48.
- Kniggendorf V, Dreyfuss JL, Regatieri CV. Age-related macular degeneration: a review of current therapies and new treatments. Arq Bras Oftalmol 2020;83:552-61.
- Teufel A, Malik N, Mukhopadhyay M, Westphal H. Frcp1 and Frcp2, two novel fibronectin type III repeat containing genes. Gene 2002;297:79–83.
- Boström P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, et al. A PGC1-α-dependent myokine that drives brown-fat-like development of white fat and thermogenesis. Nature 2012;481:463– 8.
- Schumacher MA, Chinnam N, Ohashi T, Shah RS, Erickson HP. The structure of irisin reveals a novel intersubunit β-sheet fibronectin type III (FNIII) dimer: implications for receptor activation. J Biol Chem 2013;288:33738–44.
- 11. Huh JY, Panagiotou G, Mougios V, Brinkoetter M, Vamvini MT,

Schneider BE et al. FNDC5 and irisin in humans: I. Predictors of circulating concentrations in serum and plasma and II. mRNA expression and circulating concentrations in response to weight loss and exercise. Metabolism 2012;61:1725–38.

- Gençer Tarakçı B, Girgin A, Timurkaan S, Yalçın MH, Gür FM, Karan M. Immunohistochemical localization of irisin in skin, eye, and thyroid and pineal glands of the crested porcupine (Hystrix cristata). Biotech Histochem 2016;91:423–7.
- Gür FM, Timurkaan S, Gençer Tarakçi B, Yalçin MH, Özkan ZE, Baygeldi SB, et al. Identification of immunohistochemical localization of irisin in the dwarf hamster (Phodopus roborovskii) tissues. Anat Histol Embryol 2018;47:174–9.
- Gouveia MC, Vella JP, Cafeo FR, Affonso Fonseca FL, Bacci MR. Association between irisin and major chronic diseases: a review. Eur Rev Med Pharmacol Sci 2016;20:4072–7.
- Tarboush NA, Abu-Yaghi NE, Al Ejeilat LH, Wahed RKA, Jeris IN. Association of irisin circulating level with diabetic retinopathy: a case-control study. Exp Clin Endocrinol Diabetes 2021;129:36–42.
- 16. Wang C, Wang L, Liu J, Song J, Sun Y, Lin P, et al. Irisin modulates the association of interleukin-17A with the presence of non-proliferative diabetic retinopathy in patients with type 2 diabetes. Endocrine 2016;53:459–64.
- Hu W, Wang R, Li J, Zhang J, Wang W. Association of irisin concentrations with the presence of diabetic nephropathy and retinopathy. Ann Clin Biochem 2016;53:67–74.
- Turgut B, Mercan K, Ilhan N, Çatak O. Serum irisin levels in chronic open-angle glaucoma. Adv Ophthalmol Vis Syst 2019;9:159–63.
- Yosten GLC, Stein LM, Samson WK. Chapter 5 Novel small peptide hormones. In: Schisler JC, Lang CH, Willis MS, editors. Endocrinology of the Heart in Health and Disease. Cambridge, Massachusetts: Academic Press; 2017. p. 115–35.
- 20. Xu B. BDNF (I)rising from exercise. Cell Metab 2013;18:612-4.
- Albrecht E, Norheim F, Thiede B, Holen T, Ohashi T, Schering L, et al. Irisin – a myth rather than an exercise-inducible myokine. Sci Rep 2015;5:8889.
- Pardue MT, Chrenek MA, Schmidt RH, Nickerson JM, Boatright JH. Potential role of exercise in retinal health. Prog Mol Biol Transl Sci 2015;134:491–502.
- Erickson HP. Irisin and FNDC5 in retrospect: An exercise hormone or a transmembrane receptor? Adipocyte 2013;2:289–93.
- Okonkwo OC, Schultz SA, Oh JM, Larson J, Edwards D, Cook D, et al. Physical activity attenuates age-related biomarker alterations in preclinical AD. Neurology 2014;83:1753–60.
- 25. Walker JM, Klakotskaia D, Ajit D, Weisman GA, Wood WG, Sun GY, et al. Beneficial effects of dietary EGCG and voluntary exercise on behavior in an Alzheimer's disease mouse model. J Alzheimers Dis 2015;44:561–72.
- 26. Kim OY, Song J. The role of irisin in alzheimer's disease. J Clin

Med 2018;7:407.

- Jin Y, Sumsuzzman DM, Choi J, Kang H, Lee SR, Hong Y. Molecular and functional interaction of the myokine irisin with physical exercise and Alzheimer's disease. Molecules 2018;23:3229.
- Grygiel-Gorniak B., Puszczewicz M. A review on irisin, a new protagonist that mediates muscle-adipose-bone-neuron connectivity. Eur Rev Med Pharmacol Sci 2017;21:4687–93.
- 29. Diniz BS, Teixeira AL. Brain-derived neurotrophic factor and Alzheimer's disease: Physiopathology and beyond. Neuromol Med 2011;13:217–22.
- Wrann CD, White JP, Salogiannnis J, Laznik-Bogoslavski D, Wu J, Ma D, et al. Exercise induces hippocampal BDNF through a PGC-1 /FNDC5 pathway. Cell Metab 2013;18:649–59.
- 31. Islam MR, Young MF, Wrann CD. The Role of FNDC5/Irisin in the nervous system and as a mediator for beneficial effects of exercise on the brain. In: Spiegelman B, editor. Hormones, metabolism and the benefits of exercise research and perspectives in endocrine interactions. Cham: Springer; 2017. p. 93–102.
- 32. Li DJ, Li YH, Yuan HB, Qu LF, Wang P. The novel exercise-induced hormone irisin protects against neuronal injury via activation of the Akt and ERK1/2 signaling pathways and contributes to the neuroprotection of physical exercise in cerebral ischemia. Metab Clin Exp 2017;68:31–42.
- Peng J, Deng X, Huang W, Yu JH, Wang JX, Wang JP, et al. Irisin protects against neuronal injury induced by oxygen-glucose deprivation in part depends on the inhibition of ROS-NLRP3 inflammatory signaling pathway. Mol Immunol 2017;91:185–94.
- Ohtaki H. Irisin. In: Takei Y, Ando H, Tsutsui K, editors. Handbook of hormones. Cambridge, Massachusetts: Academic Press; 2016. p. 329–30.
- 35. Panati K, Suneetha Y, Narala VR. Irisin/FNDC5--An updated review. Eur Rev Med Pharmacol Sci 2016;20:689–97.
- 36. Quinn LS, Anderson BG, Conner JD, Wolden-Hanson T. Cir-

culating irisin levels and muscle FNDC5 mRNA expression are independent of IL-15 levels in mice. Endocrine 2015;50:368–77.

- 37. Albayrak S, Atci IB, Kalayci M, Yilmaz M, Kuloglu T, Aydin S, et al. Effect of carnosine, methylprednisolone and their combined application on irisin levels in the plasma and brain of rats with acute spinal cord injury. Neuropeptides 2015;52:47–54.
- Dun SL, Lyu RM, Chen YH, Chang JK, Luo JJ, Dun NJ. Irisin-immunoreactivity in neural and non-neural cells of the rodent. Neuroscience 2013;240:155–62.
- Wang K, Li H, Wang H, Wang JH, Song F, Sun Y. Irisin exerts neuroprotective effects on cultured neurons by regulating astrocytes. Mediators Inflamm 2018;2018:9070341.
- Güler M, Aydın S, Urfalıoğlu S, Yardım M. Aqueous humor heatshock protein 70, periostin, and irisin levels in patients with pseudoexfoliation syndrome. Arq Bras Oftalmol 2020;83:378– 82.
- Moschos MM, Nitoda E, Chatziralli IP, Demopoulos CA. Age-related macular degeneration: pathogenesis, genetic background, and the role of nutritional supplements. J Chem 2014;2014:317536.
- 42. Patel M, Chan CC. Immunopathological aspects of age-related macular degeneration. Semin Immunopathol 2008;30:97–110.
- 43. Ambati J, Atkinson JP, Gelfand BD. Immunology of age-related macular degeneration. Nat Rev Immunol 2013;13:438–51.
- 44. Copland DA, Theodoropoulou S, Liu J, Dick AD. A perspective of AMD through the eyes of immunology. Invest Ophthalmol Vis Sci 2018;59:AMD83–AMD92.
- 45. Kauppinen A, Paterno JJ, Blasiak J, Salminen A, Kaarniranta K. Inflammation and its role in age-related macular degeneration. Cell Mol Life Sci 2016;73:1765–86.
- Knickelbein JE, Chan CC, Sen HN, Ferris FL, Nussenblatt RB. Inflammatory mechanisms of age-related macular degeneration. Int Ophthalmol Clin 2015;55:63–78.