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Original article

4-[1-(Substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzenesulfonic acids: Synthesis, antimicrobial activity, QSAR studies, and antiviral evaluation

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ARTICLE INFO

Article history:

Received 13 August 2010

Received in revised form

24 September 2010

Accepted 28 September 2010

Available online 7 October 2010

Keywords:

Benzimidazoles

QSAR

Antimicrobial

Antiviral

ABSTRACT

A series of 4-[1-(substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzene sulphonic acids (**1-20**) was synthesized and evaluated, *in vitro*, for their antimicrobial activity and the results indicated that compounds 4-[1-(4-Nitrobenzoyl)-1H-benzimidazol-2-yl]-benzenesulfonic acid (**9**) and 4-(1-octadec-9-enoyl-1H-benzimidazol-2-yl)-benzenesulfonic acid (**18**) were found to be the most active ones. QSAR investigations indicated that the multi-target QSAR model was effective in describing the antimicrobial activity over the one-target QSAR models. Further the mt-QSAR model indicated the importance of the topological parameter, Balaban index (J) followed by the electronic parameter, LUMO and topological parameter, valence second order molecular connectivity index ($^2\chi^v$) in describing the antimicrobial activity of synthesized compounds (**1-20**).

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1. Introduction

The increasing incidence of infection caused by the rapid development of microbial resistance to most of the known antibiotics is a serious health problem. There are a number of factors responsible for mutations in the microbial genomes. As multidrug-resistant microbial strains proliferate, the necessity for effective therapy has stimulated research on the design and synthesis of novel antimicrobial molecules [1].

In spite of nearly three decades of intensive research toward the treatment and eradication of HIV/AIDS, the disease remains a global health problem. The advent of highly active antiretroviral therapy (HAART) in the mid 1990s, which employed various combinations of reverse transcriptase and protease inhibitors, led to marked improvements of clinical outcomes. While the diversity of drugs now available to the clinician has substantially improved HIV treatment, still significant challenges remain. Of particular concern are long-term treatment side effects and the emergence of drug-resistant viral strains [2].

The heterocyclic molecule, benzimidazole, is isosteric with indole and purine nuclei, which are present in a number of fundamental

cellular components and bioactive compounds which makes the benzimidazole molecule endowed with a variety of biological properties [3]. The study of recent literature reveals that benzimidazole is reported to have number of biological activities *viz.* antiviral [4], antifungal [5], anthelmintic [6], antiprotozoal [7], psychotropic [8], anti-HIV [9], antitumor [10], antioxidant [11] and anticoagulant [12] activities.

Indeed, a number of important drugs used in different therapeutic areas contain the benzimidazole ring, as proton pump inhibitors (omeprazole), antihypertensives (candesartan, telmisartan), antihistaminics (astemizole), anthelmintics (albendazole, mebendazole), as well as several other kinds of still investigational therapeutic agents, including antitumorals and antivirals [1].

QSAR refers to a discipline in computational chemistry that addresses the modeling of biological activities or chemical reactivity based on the quantitative description for the chemical structure of molecules. QSAR relies on the basic assumption that molecules with similar physicochemical properties or structures will have similar activities [13]. Quantitative structure–activity relationship (QSAR) is one of the most important areas in chemometrics, and is a valuable tool that is used extensively in drug design and medicinal chemistry. Once a reliable QSAR model is established, we can predict the activities of molecules, and know which structural features play an important role in biological processes [14].

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Bearing these results in mind and in continuation of our research efforts in searching novel antiviral agents [15,16], we hereby present the synthesis, antimicrobial and antiviral activity and QSAR modeling of 4-[1-(substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzenesulfonic acids.

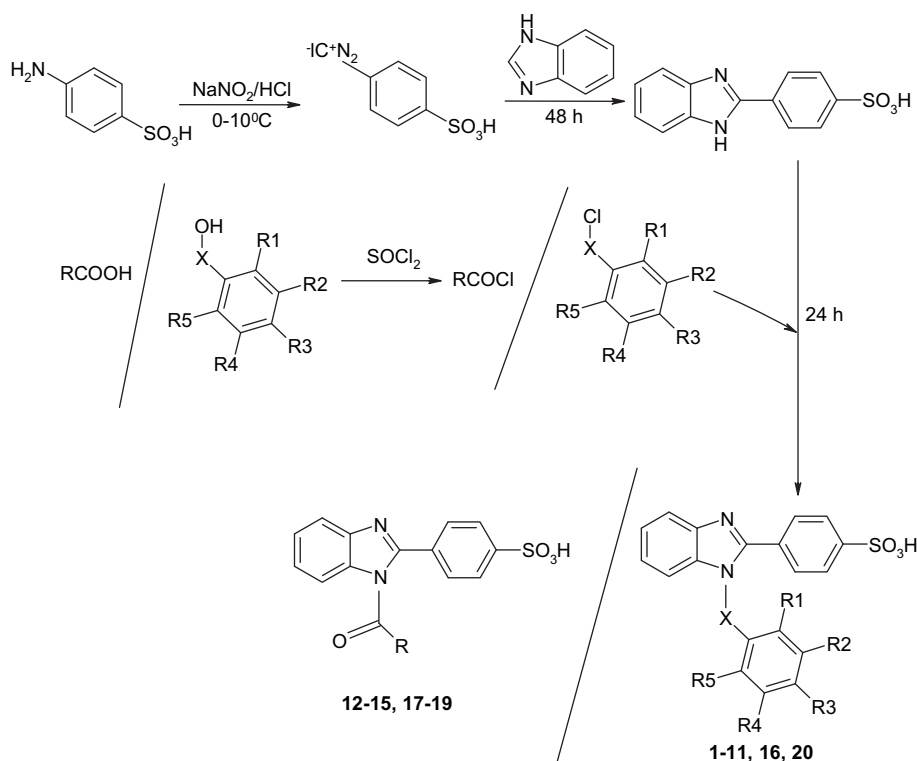
2. Chemistry

The synthesis of compounds **1-20** followed the general pathway elicited in Scheme 1. The key intermediate, 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid was prepared by the condensation of benzimidazole with aryldiazonium chloride of 4-amino-benzenesulfonic acid, which in turn prepared by the diazotization of 4-amino-benzenesulfonic acid [17]. However, based on our experience, the application of the cupric chloride for the condensation of aryldiazonium chloride with benzimidazole as suggested by Dahiya and Pathak [18] resulted in resinous (sticky) products. Therefore, the coupling was carried out by using sodium acetate along with stirring at cold conditions for the initial 3 h followed by 48 h stirring at room temperature which resulted in a solid product. For the synthesis of 4-[1-(substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzenesulfonic acids, the key intermediates 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid has been reacted with corresponding aryl/alkyl acid chlorides which were formed by the reaction of corresponding aryl/alkyl acids with thionyl chloride. It is important to note here that the yield of most of the synthesized benzimidazoles were below 50%. The low yield of synthetic compounds may be attributed to any one or more of the following reasons [17]: a). The reaction may be reversible and position of equilibrium is unfavorable to the product; b). The incursion of side reactions leading to the formation of by-products; c). The premature work-up of the reaction before its completion; d). The volatilization of products during reaction or work-up; e). The loss of product due to incomplete extraction, inefficient crystallization or other work-up procedures; f). The presence of contaminants in the

reactants or reagents leading to a less efficient reaction. The physicochemical characteristics of the synthesized compounds are presented in Table 1.

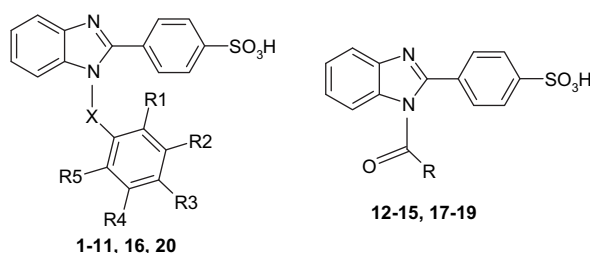
The structures of compounds **1-20** were assigned by IR and ^1H NMR spectroscopic data, which are consistent with the proposed molecular structures. The primary amino group in compound **1** is depicted by the presence of NH asymmetric stretch at 3481.81 cm^{-1} . The IR bending vibration corresponding to OCH_3 of compound **8** appeared at 1425.61 cm^{-1} . The presence of heterocyclic pyridine moiety in compound **15** is demonstrated by the presence of CH out of plane bending at 829.04 cm^{-1} . The presence of an additional S–N stretch at 808.61 cm^{-1} apart from the SO_3 asymmetric stretch in the region of $1260\text{--}1150\text{ cm}^{-1}$ indicated that compound **20** contains a link between SO_2 and N_1 of benzimidazole. The appearance of C=O stretch in the range of $1630\text{--}1600\text{ cm}^{-1}$ indicated the formation of tertiary amides (**1-20**) by the reaction of acid chlorides with the 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid. Formation of tertiary amide is further evidenced by the absence of IR bands at $1725\text{--}1700\text{ cm}^{-1}$ and $1700\text{--}1680\text{ cm}^{-1}$ corresponding to the alkyl and aryl acids, respectively. The appearance of SO_3 asymmetric stretch in the region of $1260\text{--}1150\text{ cm}^{-1}$ demonstrated the presence of SO_3H group of benzenesulfonic acid attached to the 2nd position of benzimidazole.


The appearance of singlet around $2.5\text{ }\delta$ ppm corresponds to the proton of SO_3H in the NMR of all the compounds indicated the presence of para sulfonyl phenyl nucleus to the 2nd position of synthesized benzimidazoles (**11-20**). Further the multiplet corresponds to $6.9\text{--}8.6\text{ }\delta$ ppm confirmed the presence of protons of benzimidazole and aryl nucleus. The appearance of δ at $8.46\text{--}9.66$ confirmed the presence of the pyridine ring system in compound **15**. The appearance doublet at $3.66\text{--}3.72\text{ }\delta$ ppm (CH_2 of $\text{CH}=\text{CH}_2$) and $3.79\text{--}3.85\text{ }\delta$ ppm (CH of $\text{CH}=\text{CH}_2$) indicated the formation of compound **14** by the reaction of 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid and acid



Scheme 1. Scheme for the synthesis of 4-[1-(substituted aryl/alkyl)-benzimidazol-2-yl]-benzenesulfonic acids.

Table 1
Physicochemical characteristics of synthesized benzimidazole derivatives.



Comp.	R ₁	R ₂	R ₃	R ₄	R ₅	X	Mol. Formula	Mol. Wt.	m.p.	Rf value (CHCl ₃)	Yield %	
1	NH ₂	H	H	H	H	C=O	C ₂₀ H ₁₅ N ₃ O ₄ S	393.42	116--118	0.33	70.9	
2	H	H	H	H	H	C=O	C ₂₀ H ₁₄ N ₂ O ₄ S	378.40	93--95	0.49	43.9	
3	H	H	NH ₂	H	H	C=O	C ₂₀ H ₁₅ N ₃ O ₄ S	393.42	212--214	0.43	99.0	
4	Cl	H	H	H	H	C=O	C ₂₀ H ₁₃ ClN ₂ O ₄ S	412.85	128--130	0.48	37.5	
5	H	H	Cl	H	H	C=O	C ₂₀ H ₁₃ ClN ₂ O ₄ S	412.85	65--67	0.44	63.5	
6	H	OH	H	H	H	C=O	C ₂₀ H ₁₄ N ₂ O ₅ S	394.40	98--100	0.41	101.6	
7	OH	H	OH	H	H	C=O	C ₂₀ H ₁₄ N ₂ O ₆ S	410.40	163--165	0.38	70.6	
8	H	H	OCH ₃	H	H	C=O	C ₂₁ H ₁₆ N ₂ O ₅ S	408.43	90--92	0.36	60.9	
9	H	H	NO ₂	H	H	C=O	C ₂₀ H ₁₃ N ₃ O ₆ S	423.40	92--94	0.34	46.3	
10	CH ₃	H	H	H	H	C=O	C ₂₁ H ₁₆ N ₂ O ₄ S	392.43	70--72	0.35	37.0	
11	H	H	CH ₃	H	H	C=O	C ₂₁ H ₁₆ N ₂ O ₄ S	392.43	95--97	0.45	28.1	
16	OH	H	H	H	H	C=O	C ₂₀ H ₁₄ N ₂ O ₅ S	394.40	78--80	0.77	60.0	
20	H	H	CH ₃	H	H	SO ₂	C ₂₀ H ₁₆ N ₂ O ₅ S ₂	428.48	156--158	0.40	35.8	
12	R											
12	-CH=CH-CH ₃						C ₁₇ H ₁₄ N ₂ O ₄ S	342.37	112--114	0.82	30.2	
13	-CH=CH ₂						C ₁₆ H ₁₂ N ₂ O ₄ S	328.34	100--102	0.56	32.8	
14	-C(CH ₃)=CH ₂						C ₁₇ H ₁₄ N ₂ O ₄ S	342.37	102--104	0.43	33.5	
15								C ₁₉ H ₁₃ N ₃ O ₄ S	379.39	110--112	0.96	44.0
17	-CH ₂ -(CH ₂) ₂ -CH ₃						C ₁₈ H ₁₈ N ₂ O ₄ S	358.41	107--109	0.42*	33.8	
18	-CH ₂ -(CH ₂) ₆ -CH=CH-(CH ₂) ₇ -CH ₃						C ₃₁ H ₄₂ N ₂ O ₄ S	538.74	109--111	0.75	31.7	
19	-CH ₂ -(CH ₂) ₁₃ -CH ₃						C ₂₉ H ₄₀ N ₂ O ₄ S	512.70	115--117	0.83*	61.3	

*TLC mobile phase – Benzene.

chloride of methacrylic acid. Further the coupling constant value of 18 Hz in compound **14** indicated that the nature of the double bond in the synthesized compound **14** is of trans nature. Compound **8** showed a singlet at δ 3.78 ppm corresponding to a proton of the OCH₃. This confirmed the attachment of the para methoxy benzoyl group to the N₁ position of benzimidazole. The absence of a singlet at δ 11 ppm in the NMR spectra of compounds **1-20** indicated the absence of the free COOH group. This confirms that the compounds **1-20** are tertiary amides and not the physical mixture of aryl/alkyl acids and 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid. Therefore, this assures the reaction of aryl/alkyl acid chlorides with the secondary nitrogen of the benzimidazole nucleus.

3. Results and discussion

3.1. Antimicrobial activity

The synthesized compounds were evaluated for their *in vitro* antimicrobial activities against two Gram-positive bacteria – *Staphylococcus aureus*, *Bacillus subtilis*; the Gram-negative bacterium – *Escherichia coli* and the fungal strains – *Aspergillus niger* and *Candida albicans* by tube dilution method [19] using ciprofloxacin and fluconazole as control drugs for antibacterial and antifungal activity, respectively. The results of the antimicrobial studies are presented in Table 2. In general the compounds showed an improved antibacterial activity when compared to their antifungal activity. The deduced

patterns of antimicrobial activity of the synthesized benzimidazoles are in the following order: Antibacterial activity > antifungal activity.

The compound **9** is the most effective compound against *S. aureus* with pMIC₅₀ value of 2.43 (Table 2), respectively. For activity against *B. subtilis* the compounds **7** and **9** yielded better activity (Table 2) in comparison to other compounds synthesized. The antimicrobial spectrum of synthesized benzimidazoles against *E. coli* demonstrated that compounds **4**, **5**, **7** and **9** were the most active ones with pMIC₅₀ values of 2.42, 2.42, 2.42 and 2.43 (Table 2) respectively. From the above discussion it is evident that compound **9** emerged as the most active antibacterial benzimidazole. Compounds **9** and **18** emerged as the most effective antifungal agents against *C. albicans* whereas compound **18** was most active against *A. niger* (Table 2). From the above discussion it is evident that compound **18** emerged as the most effective antifungal benzimidazole.

The minimum bactericidal concentration/minimum fungicidal concentration (MBC/MFC) (Table 3) determination results revealed that the synthesized compounds were fungistatic against both fungi and bacteriostatic against *S. aureus*, *B. subtilis* but bacteriocidal against *E. coli*. In general, the MBC and MFC of synthesized benzimidazole derivatives were 3-fold higher than their MIC values, which indicated that they were bacteriostatic and fungistatic in action except for *E. coli* (a drug is considered to be bacteriostatic/fungistatic when its MBC and MFC values are 3-fold higher than its MIC values) [20]. In the case of *S. aureus* compounds

Table 2
Antimicrobial activity of synthesized benzimidazole derivatives.

Comp.	For ot-QSAR				For mt-QSAR			
	pMICsa	pMICbs	pMICec	pMICca	pMICan	pMICab	pMICaf	pMICam
1	1.80	2.03	2.40	1.70	1.50	2.08	1.60	1.89
2	2.08	2.12	2.12	1.78	1.48	2.11	1.63	1.92
3	2.10	1.80	2.10	1.20	1.50	2.00	1.35	1.74
4	2.09	1.82	2.42	1.52	1.22	2.11	1.37	1.81
5	2.12	2.13	2.42	1.52	1.22	2.22	1.37	1.88
6	2.10	2.15	2.40	1.50	1.20	2.22	1.35	1.87
7	2.12	2.42	2.42	1.52	1.22	2.32	1.37	1.94
8	2.12	2.12	2.12	1.51	1.21	2.12	1.36	1.82
9	2.43	2.43	2.43	2.13	1.53	2.43	1.83	2.19
10	2.10	2.10	2.10	1.50	1.20	2.10	1.35	1.80
11	2.10	1.95	2.10	1.50	0.89	2.05	1.20	1.71
12	1.85	1.85	1.85	1.44	0.84	1.85	1.14	1.57
13	1.72	2.02	2.32	1.42	1.12	2.02	1.27	1.72
14	1.74	1.74	2.04	1.44	1.14	1.84	1.29	1.62
15	2.13	2.13	2.39	1.75	1.48	2.22	1.62	1.98
16	2.10	2.10	2.26	1.50	1.50	2.15	1.50	1.89
17	1.76	1.76	1.76	1.46	1.16	1.76	1.31	1.58
18	2.24	2.24	1.85	1.94	1.63	2.11	1.79	1.98
19	2.23	2.23	1.61	1.61	1.61	2.02	1.61	1.86
20	1.84	1.84	1.54	1.54	1.23	1.74	1.39	1.60
SD ^a	0.19	0.20	0.28	0.20	0.22	0.18	0.19	0.16
Std.	2.61*	2.61*	2.61*	2.64**	2.64**	2.61	2.64	2.62

*Ciprofloxacin, **Fluconazole.

^a Standard deviation.

10 and **17** and for *Bacillus subtilis* compounds **11** and **17** were found to be active bacteriocidal agents.

3.2. QSAR studies

3.2.1. Development of one-target QSAR model

In order to understand the experimental antimicrobial data on a theoretical basis, we established a quantitative structure activity relationship (QSAR) between the *in vitro* antimicrobial activity of synthesized benzimidazole derivatives and descriptors coding for lipophilic, electronic, steric and topological properties of the molecules under consideration using the linear free energy relationship model (LFER) described by Hansch and Fujita [21]. Biological activity data determined as MIC values were first transformed into pMIC

values and used as dependent variables in a QSAR study. The different molecular descriptors (independent variables) like log of octanol-water partition coefficient (log P), molar refractivity (MR), Kier's molecular connectivity ($^0\chi$, $^0\chi^v$, $^1\chi$, $^1\chi^v$, $^2\chi$, $^2\chi^v$) and shape (κ_1 , κ_1) topological indices, Randic topological index (R), Balaban topological index (J), Wiener topological index (W), Total energy (Te), energies of highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO), dipole moment (μ), electronic energy (Ele.E), nuclear energy (Nu.E) and molecular surface area (SA) calculated for of 4-[1-(substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzenesulfonic acids are used as independent variables and are presented in Table 4 [22–27].

In the present study, a data set of 20 benzimidazole derivatives was subjected to linear free energy regression analysis for model generation. The reference drugs were not included in model development as they belong to different structural series. Preliminary

Table 3
MBC/MFC of synthesized benzimidazole derivatives.

Comp.	MBC ($\mu\text{g/ml}$)			MFC ($\mu\text{g/ml}$)	
	<i>S. aureus</i>	<i>B. subtilis</i>	<i>E. coli</i>	<i>C. albicans</i>	<i>A. niger</i>
1	50	>50	1.56	50	>50
2	12.5	>50	1.56	50	>50
3	12.5	>50	25	>50	>50
4	25	>50	1.56	>50	>50
5	25	>50	1.56	>50	>50
6	12.5	>50	3.12	50	>50
7	12.5	25	1.56	12.5	>50
8	>50	>50	3.12	>50	>50
9	50	50	1.56	25	>50
10	6.25	>50	3.12	50	>50
11	12.5	6.25	25	50	>50
12	12.5	50	12.5	>50	>50
13	6.25	12.5	1.56	>50	>50
14	6.25	12.5	12.5	>50	>50
15	12.5	>50	1.56	>50	>50
16	25	25	12.5	50	>50
17	6.25	6.25	6.25	12.5	>50
18	12.5	50	3.12	50	>50
19	6.25	12.5	12.5	>50	>50
20	6.25	12.5	12.5	50	>50
Std.	0.019	0.019	0.019	0.040	0.040

Table 4
Values of selected parameters used in regression analysis.

Comp.	log P	MR	$^2\chi^v$	R	J	Te	NE	LUMO
1	3.07	104.11	7.69	13.36	1.36	-4861.42	32108.70	-1.28
2	3.87	99.29	7.47	12.95	1.34	-4640.27	29585.80	-1.37
3	3.07	104.11	7.72	13.34	1.34	-4861.39	30708.30	-1.13
4	4.43	103.90	8.02	13.36	1.36	-5000.25	32054.10	-1.34
5	4.43	103.90	8.08	13.34	1.34	-5000.34	31558.70	-1.43
6	3.48	101.10	7.65	13.34	1.34	-4955.73	31701.40	-1.43
7	3.09	102.92	7.80	13.75	1.37	-5281.53	34388.40	-1.41
8	3.74	106.54	7.83	13.88	1.33	-5116.18	32616.70	-1.18
9	3.16	106.32	7.90	14.25	1.33	-5470.94	36375.50	-1.85
10	4.36	105.19	7.92	13.36	1.36	-4796.08	32177.00	-1.34
11	4.36	105.19	7.97	13.34	1.34	-4796.16	30732.20	-1.18
12	3.02	90.36	6.63	11.41	1.54	-4256.52	24803.10	-1.40
13	2.65	84.44	6.34	10.91	1.53	-4100.52	22838.50	-1.45
14	3.00	88.48	6.88	11.29	1.56	-4256.30	25159.20	-1.41
15	2.53	97.76	7.34	12.95	1.34	-4705.22	29639.90	-1.43
16	3.48	101.10	7.62	13.36	1.36	-4960.77	32297.30	-1.32
17	3.46	93.10	7.29	11.91	1.54	-4440.90	27410.30	-1.40
18	8.56	154.26	11.52	18.41	1.24	-6438.19	48954.00	-1.39
19	8.05	143.70	11.18	17.41	1.29	-6155.08	46304.10	-1.39
20	3.96	108.96	9.97	13.68	1.39	-5182.61	34218.00	-1.41

Table 6
Correlation of antimicrobial activity of benzimidazole derivatives with studied molecular descriptors.

	pMICsa	pMICbs	pMICec	pMICca	pMICan	pMICab	pMICaf	pMICam
log P	0.420	0.261	-0.484	0.306	0.389	-0.014	0.402	0.182
MR	0.562	0.416	-0.391	0.429	0.574	0.148	0.580	0.376
$^0\chi^v$	0.530	0.387	-0.438	0.414	0.546	0.099	0.555	0.332
$^1\chi^v$	0.474	0.346	-0.499	0.398	0.544	0.031	0.545	0.281
$^2\chi^v$	0.438	0.294	-0.529	0.363	0.504	-0.018	0.503	0.227
κ_1	0.576	0.472	-0.375	0.467	0.591	0.182	0.610	0.414
κ_2	0.497	0.413	-0.414	0.435	0.563	0.110	0.576	0.350
$\kappa\alpha_1$	0.527	0.427	-0.418	0.438	0.568	0.124	0.581	0.361
$\kappa\alpha_2$	0.458	0.377	-0.443	0.410	0.542	0.067	0.550	0.308
R	0.669	0.531	-0.277	0.493	0.634	0.291	0.650	0.506
J	-0.824	-0.610	-0.144	-0.457	-0.632	-0.606	-0.630	-0.707
W	0.525	0.439	-0.385	0.458	0.577	0.146	0.597	0.383
Te	-0.699	-0.585	0.221	-0.517	-0.621	-0.352	-0.655	-0.549
ELE	-0.661	-0.551	0.279	-0.517	-0.636	-0.295	-0.664	-0.515
Nu. E	0.657	0.547	-0.284	0.516	0.637	0.289	0.664	0.511
LUMO	-0.242	-0.458	-0.172	-0.656	-0.165	-0.354	-0.454	-0.453
HOMO	-0.234	-0.179	0.172	-0.379	-0.127	-0.054	-0.284	-0.173
μ	0.118	-0.150	-0.137	-0.122	-0.135	-0.088	-0.143	-0.127

predict accurately the biological activities of compounds. As the observed and predicted values are close to each other (Table 7), the QSAR model for *A. niger* [Eq. (7)] is a valid one.

It is important to note that all the equations were derived using the entire data set as there were no outliers in the data set. Generally for QSAR studies, the biological activities of compounds should span 2–3 orders of magnitude. But in the present study the range of antimicrobial activities of the synthesized compounds is within one order of magnitude. But it is important to note that the predictability of the QSAR models developed in the present study is highly evidenced by the low residual values. This is in accordance with results suggested by Bajaj et al. [30], who stated that the reliability of the QSAR model lies in its predictive ability even though the activity data are in the narrow range. Further, recent literature reveals that the QSAR have been applied to describe the relationship between narrow range of biological activity and physicochemical properties of the molecules [31–33]. When biological activity data lies in the narrow range, the presence of minimum standard deviation of the biological activity justifies its use in QSAR studies [34,35]. The minimum standard deviation

(Table 2) observed in the antimicrobial activity data justifies its use in QSAR studies.

3.2.2. Development of multi-target QSAR model

According to the above ot-QSAR models one should use five different equations to predict the activity of a new compound against the five microbial species. The ot-QSAR models, which are almost in all the literature, become unpractical or complicated to use when we have to predict each compound results for more than one-target. In these cases we have to develop one ot-QSAR for each target. However, very recently the interest has been increased in development of multi-target QSAR (mt-QSAR) models. In opposition to ot-QSAR, the mt-QSAR model is a single equation that considers the nature of molecular descriptors which are common and essential for describing the antimicrobial activity [36–40].

In the present study we have attempted to develop three different types of mt-QSAR models viz. mt-QSAR model for describing antibacterial activity of synthesized compounds against *S. aureus*, *B. subtilis* and *E. coli*, mt-QSAR model for describing antifungal activity of synthesized compounds against *C. albicans*

Table 7
Comparison of observed and predicted antibacterial and antifungal activity obtained by ot-QSAR model.

Comp.	pMICsa (Eq. (3))			pMICbs (Eq. (4))			pMICec (Eq. (5))			pMICca (Eq. (6))			pMICan (Eq. (7))		
	Obs.	Pre.	Res.	Obs.	Pre.	Res.	Obs.	Pre.	Res.	Obs.	Pre.	Res.	Obs.	Pre.	Res.
1	1.80	2.04	-0.24	2.03	2.01	0.02	2.40	2.22	0.18	1.70	1.50	0.20	1.50	1.37	0.13
2	2.08	2.14	-0.06	2.12	2.11	0.01	2.12	2.38	-0.26	1.78	1.62	0.16	1.48	1.19	0.29
3	2.10	2.03	0.07	1.80	1.93	-0.13	2.10	2.20	-0.10	1.20	1.38	-0.18	1.50	1.31	0.19
4	2.09	2.06	0.03	1.82	2.05	-0.23	2.42	2.16	0.26	1.52	1.55	-0.03	1.22	1.25	-0.03
5	2.12	2.15	-0.03	2.13	2.15	-0.02	2.42	2.27	0.15	1.52	1.67	-0.15	1.22	1.23	-0.01
6	2.10	2.15	-0.05	2.15	2.14	0.01	2.40	2.35	0.05	1.50	1.67	-0.17	1.20	1.31	-0.11
7	2.12	2.09	0.03	2.42	2.10	0.32	2.42	2.24	0.18	1.52	1.62	-0.10	1.22	1.46	-0.24
8	2.12	2.06	0.06	2.12	1.98	0.14	2.12	2.22	-0.10	1.51	1.44	0.07	1.21	1.33	-0.12
9	2.43	2.35	0.08	2.43	2.47	-0.04	2.43	2.54	-0.11	2.13	2.10	0.03	1.53	1.54	-0.01
10	2.10	2.06	0.04	2.10	2.05	0.05	2.10	2.19	-0.09	1.50	1.56	-0.06	1.20	1.26	-0.06
11	2.10	2.03	0.07	1.95	1.96	-0.01	2.10	2.15	-0.05	1.50	1.43	0.07	0.89	1.20	-0.31
12	1.85	1.76	0.09	1.85	1.83	0.02	1.85	2.03	-0.18	1.44	1.41	0.03	0.84	1.06	-0.22
13	1.72	1.81	-0.09	2.02	1.88	0.14	2.32	2.15	0.17	1.42	1.47	-0.05	1.12	1.01	0.11
14	1.74	1.71	0.03	1.74	1.80	-0.06	2.04	1.92	0.12	1.44	1.39	0.05	1.14	1.08	0.06
15	2.13	2.17	-0.04	2.13	2.15	-0.02	2.39	2.44	-0.05	1.75	1.68	0.07	1.48	1.31	0.17
16	2.10	2.06	0.04	2.10	2.03	0.07	2.26	2.25	0.01	1.50	1.54	-0.04	1.50	1.34	0.16
17	1.76	1.74	0.02	1.76	1.83	-0.07	1.76	1.88	-0.12	1.46	1.41	0.05	1.16	1.14	0.02
18	2.24	2.24	0.00	2.24	2.27	-0.03	1.85	1.72	0.13	1.94	1.76	0.18	1.63	1.60	0.03
19	2.23	2.16	0.07	2.23	2.20	0.03	1.61	1.68	-0.07	1.61	1.70	-0.09	1.61	1.53	0.08
20	1.84	1.98	-0.14	1.84	2.06	-0.22	1.54	1.68	-0.14	1.54	1.59	-0.05	1.23	1.38	-0.15

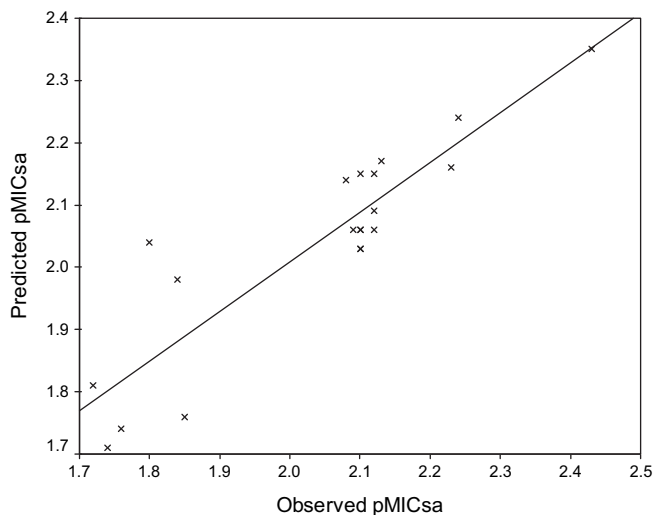


Fig. 1. Plot of predicted $pMIC_{sa}$ against the experimental $pMIC_{sa}$ for the MLR model developed by Eq. (3).

and *A. niger* as well a common mt-QSAR model for describing the antimicrobial activity of synthesized benzimidazole derivatives against all the above mentioned microorganisms.

In order to develop mt-QSAR models, we have calculated the average antibacterial activity [$pMIC_b = (pMIC_{sa} + pMIC_{bs} + pMIC_{ec})/3$], antifungal activity [$pMIC_f = (pMIC_{ca} + pMIC_{an})/2$] and antimicrobial activity values [$pMIC_{am} = (pMIC_{sa} + pMIC_{bs} + pMIC_{ec} + pMIC_{ca} + pMIC_{an})/5$] of benzimidazole derivatives which are presented in Table 2. These average activity values were also correlated with the molecular descriptors of synthesized compounds (Table 6). The LR model derived for the antibacterial activity of benzimidazole derivatives indicated the importance of topological parameter, Balaban index (J) [Eq. (8)].

mt-QSAR model for antibacterial activity

$$pMIC_{ab} = -1.197 J + 3.726 \quad (8)$$

$$n = 20 \quad r = 0.605 \quad q^2 = 0.245 \quad s = 0.144 \quad F = 10.44$$

The addition of electronic parameter, LUMO and topological parameter, valence second order molecular connectivity index (${}^2\chi^v$)

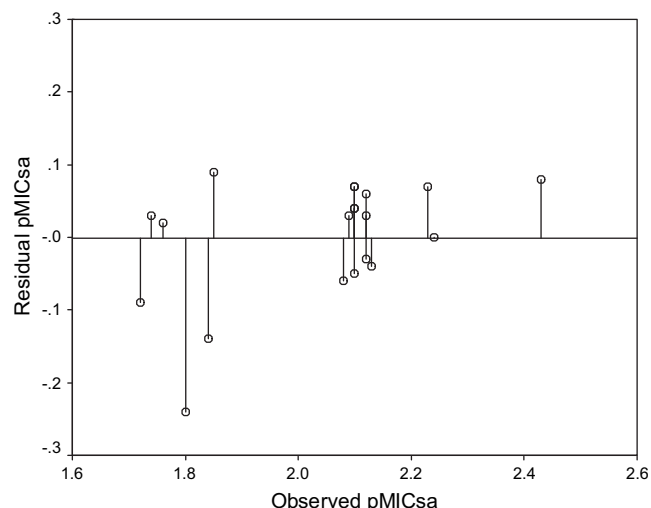


Fig. 2. Plot of residual $pMIC_{sa}$ against the experimental $pMIC_{sa}$ by Eq. (3).

improved the correlation to 0.732 [Eq. (9)] and 0.805 [Eq. (10)] respectively.

mt-QSAR model for antibacterial activity

$$pMIC_{ab} = -1.271 J - 0.505 \text{ LUMO} + 3.134 \quad (9)$$

$$n = 20 \quad r = 0.732 \quad q^2 = 0.280 \quad s = 0.127 \quad F = 9.82$$

mt-QSAR model for antibacterial activity

$$pMIC_{ab} = -0.091 {}^2\chi^v - 2.077 J + 5.677 \quad (10)$$

$$n = 20 \quad r = 0.805 \quad q^2 = 0.541 \quad s = 0.110 \quad F = 15.74$$

Based on the above-improved correlations, we decided to study the addition of electronic parameter, LUMO and topological parameter, valence second order molecular connectivity index (${}^2\chi^v$) to the topological parameter, Balaban index (J). This addition resulted in appreciable improvement in the statistical parameters as depicted in [Eq. (11)].

mt-QSAR model for antibacterial activity

$$pMIC_{ab} = -0.097 {}^2\chi^v - 2.219 J - 0.559 \text{ LUMO} + 5.151 \quad (11)$$

$$n = 20 \quad r = 0.925 \quad q^2 = 0.768 \quad s = 0.073 \quad F = 31.61$$

The mt-QSAR MLR model for antifungal activity revealed the importance of electronic parameter, LUMO and the topological parameter, Balaban index (J) in describing antifungal activity [Eq. (12)].

mt-QSAR model for antifungal activity

$$pMIC_{af} = -1.417 J - 0.669 \text{ LUMO} + 2.472 \quad (12)$$

$$n = 20 \quad r = 0.812 \quad q^2 = 0.567 \quad s = 0.115 \quad F = 16.47$$

The mt-QSAR MLR model for antimicrobial activity revealed the importance of the topological parameter, Balaban index (J) followed by the electronic parameter, LUMO and topological parameter, valence second order molecular connectivity index (${}^2\chi^v$) [Eq. (13)].

mt-QSAR model for antimicrobial activity

$$pMIC_{am} = -0.052 {}^2\chi^v - 1.831 J - 0.597 \text{ LUMO} + 3.942 \quad (13)$$

$$n = 20 \quad r = 0.940 \quad q^2 = 0.809 \quad s = 0.057 \quad F = 41.10$$

The predictability of Eqs. (11)–(13) was verified by calculating the antibacterial, antifungal and antimicrobial activities for substituted benzimidazoles respectively and the low residual activity values indicated the high predictability of above mentioned equations (Table 8). Further the plot of observed $pMIC_{am}$ and predicted $pMIC_{am}$ (Fig. 3) also in favor of the mt-QSAR target model for antimicrobial activity. The propagation of activity values on both sides of zero, in the plot of observed $pMIC_{am}$ against residual $pMIC_{am}$, indicated that there was no systemic error in the development of mt-QSAR model (Fig. 4). The other statistically significant ot-QSAR and mt-QSAR models derived are presented in Table 9.

The Balaban index J is a variant of connectivity index and represents extended connectivity. It is a good descriptor for the shape of molecules and is one of the widely used topological indices for QSAR and QSPR studies. The Balaban index of a connected (molecular) graph G is defined as [41]:

$$J = (m/\mu + 1) \sum_{uv \in E(G)} (D_u D_v)^{-1/2}$$

where, m is the number of edges, μ is the cyclomatic number, D_u is the sum of distances between vertex u and all other vertices of G , and the summation goes over all edges from the edge set $E(G)$. The

Table 8
Comparison of observed and predicted antibacterial, antifungal and antimicrobial activity obtained by mt-QSAR model.

Comp.	pMIC _{ab} (Eq. (11))			pMIC _{af} (Eq. (12))			pMIC _{am} (Eq. (13))		
	Obs.	Pre.	Res.	Obs.	Pre.	Res.	Obs.	Pre.	Res.
1	2.08	2.09	-0.01	1.60	1.40	0.20	1.89	1.81	0.08
2	2.11	2.22	-0.11	1.63	1.49	0.14	1.92	1.92	0.00
3	2.00	2.06	-0.06	1.35	1.33	0.02	1.74	1.76	-0.02
4	2.11	2.09	0.02	1.37	1.43	-0.06	1.81	1.83	-0.02
5	2.22	2.19	0.03	1.37	1.53	-0.16	1.88	1.92	-0.04
6	2.22	2.22	0.00	1.35	1.52	-0.17	1.87	1.94	-0.07
7	2.32	2.15	0.17	1.37	1.48	-0.11	1.94	1.88	0.06
8	2.12	2.09	0.03	1.36	1.37	-0.01	1.82	1.80	0.02
9	2.43	2.46	-0.03	1.83	1.82	0.01	2.19	2.20	-0.01
10	2.10	2.10	0.00	1.35	1.43	-0.08	1.80	1.83	-0.03
11	2.05	2.05	0.00	1.20	1.36	-0.16	1.71	1.77	-0.06
12	1.85	1.87	-0.02	1.14	1.23	-0.09	1.57	1.61	-0.04
13	2.02	1.95	0.07	1.27	1.27	0.00	1.72	1.68	0.04
14	1.84	1.80	0.04	1.29	1.20	0.09	1.62	1.56	0.06
15	2.22	2.27	-0.05	1.62	1.53	0.09	1.98	1.97	0.01
16	2.15	2.12	0.03	1.50	1.42	0.08	1.89	1.84	0.05
17	1.76	1.81	-0.05	1.31	1.22	0.09	1.58	1.58	0.00
18	2.11	2.05	0.06	1.79	1.64	0.15	1.98	1.90	0.08
19	2.02	1.98	0.04	1.61	1.58	0.03	1.86	1.84	0.02
20	1.74	1.88	-0.14	1.39	1.44	-0.05	1.60	1.72	-0.12

electronic parameter LUMO, which denotes the energy of lowest unoccupied molecular orbital directly relates to the electron affinity and characterizes the sensibility of the molecule towards an attack by nucleophile. The contribution of LUMO in describing antimicrobial activity may be attributed to the interaction of benzimidazole derivatives with nucleophilic amino acid residue like cysteine of microorganisms [42]. The topological descriptor, i.e. the valence second order molecular connectivity index encodes information related to the degree of stargraph likeness and takes a large value for more linear molecule [43]. The trend in Eq. (13) indicated that higher the values of J , LUMO and ${}^2\chi^v$ lower will be the antimicrobial activity of benzimidazole derivatives.

The residual activity values in the case of mt-QSAR model for antimicrobial activity are less when compared to the residuals of one-target models (Table 7) as well the multi-target models for antibacterial and antifungal activities (Table 8). This mt-QSAR equation (Eq. (13)) can be used to predict the activity of substituted benzimidazoles against different microbial species.

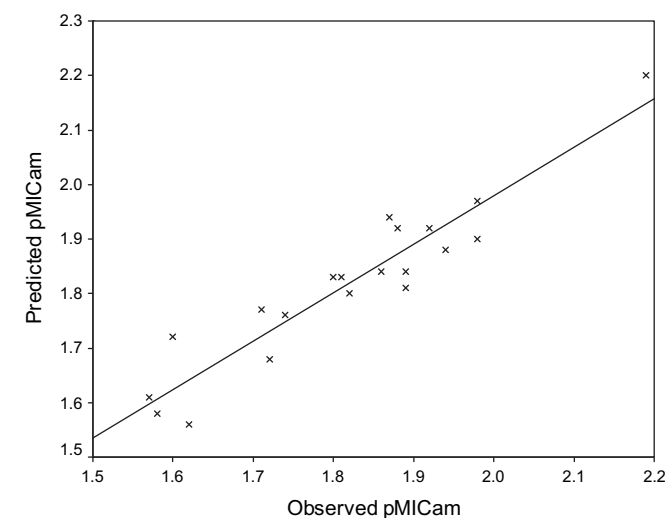


Fig. 3. Plot of predicted pMIC_{am} against the experimental pMIC_{am} values for the MLR model developed by Eq. (13).

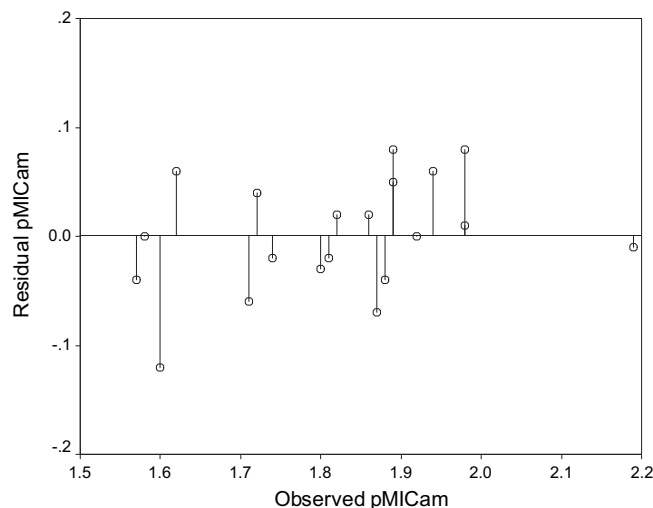


Fig. 4. Plot of predicted pMIC_{am} values against the experimental pMIC_{am} values for the MLR model developed by Eq. (13).

Furthermore, the topological parameter, Balaban index (J) followed by the electronic parameter, LUMO and topological parameter, valence second order molecular connectivity index (${}^2\chi^v$) is describing the antimicrobial activity of 4-[1-(substituted aryl/alkyl carbonyl)-benzoimidazol-2-yl]-benzenesulfonic acids (1-20).

It is important to mention here that the development of multi-QSAR target models exhibited the following advantages over one-target QSAR models: (a). There is an improvement in statistical parameters when compared to one-target models (improvement in r , q^2 , etc. cf. Eq. (13) with Eqs. (1)–(7)). (b). No change in the trend of molecular descriptors has been observed, i.e. all the three parameters, i.e. J , LUMO and ${}^2\chi^v$ showed the same negative correlation with antimicrobial activity as observed in the case of one-target models. (c). There was a significant decrease in residual values when compared to the residual values observed in one-target QSAR models, i.e. mt-QSAR model has better predictability than the ot-QSAR model.

Table 9

Regression analysis and quality of correlation for modeling antibacterial and antifungal activity of synthesized Benzimidazole derivatives.

S. No.	QSAR model	n	r	q ²	s	F
<i>B. subtilis</i>						
1	pMIC _{bs} = -1.383J + 3.959	20	0.610	0.269	0.164	10.66
<i>E. coli</i>						
2	pMIC _{ec} = -0.112 ${}^2\chi^v$ + 3.034	20	0.528	0.108	0.247	6.97
3	pMIC _{ec} = -0.224 ${}^2\chi^v$ + 2.622J + 7.559	20	0.823	0.558	0.170	17.90
<i>C. albicans</i>						
4	pMIC _{ca} = -0.923 LUMO + 0.302	20	0.656	0.237	0.157	13.60
<i>A. niger</i>						
5	pMIC _{an} = 0.00002 NE + 0.556	20	0.636	0.314	0.177	12.28
Antifungal activity						
6	pMIC _{af} = -1.319J + 3.257	20	0.629	0.278	0.149	11.81
Antimicrobial activity						
7	pMIC _{am} = -1.244J + 3.535	20	0.707	0.419	0.113	18.03
8	pMIC _{am} = -1.370J - 0.568 LUMO + 2.869	20	0.877	0.705	0.079	28.60
9	pMIC _{am} = -0.045 ${}^2\chi^v$ - 1.680 LUMO + 4.503	20	0.767	0.456	0.106	12.15

Table 10

Anti-Feline Corona Virus (FIPV) and anti-Feline Herpes Virus activity and cytotoxicity of synthesized benzimidazole derivatives in CRFK cell cultures.

Comp.	CC ₅₀ ^a (μg/mL)	EC ₅₀ ^b (μg/mL)	
		Feline Herpes Virus (FIPV)	Feline Corona Virus
1	11	>4	>4
2	>100	>100	>100
3	>100	>100	>100
4	>100	>100	>100
5	>100	>100	>100
6	>100	>100	>100
7	>100	>100	>100
8	>100	>100	>100
9	>100	>100	>100
10	>100	>100	>100
11	>100	>100	>100
12	>100	>100	>100
13	82	>20	>20
14	>100	>100	>100
15	>100	>100	>100
16	>100	>100	>100
17	>100	>100	>100
18	>100	>100	>100
19	>100	>100	>100
20	>100	>100	>100
HHA	>100	32.6	2.0
UDA	14	1.5	0.4
Ganciclovir (μM)	>100	>100	1.7

CRFK cells: Crandell–Rees Feline Kidney cells.

^a 50% Cytotoxic concentration, as determined by measuring the cell viability with the colorimetric formazan-based MTS assay.^b 50% Effective concentration, or compound concentration resulting in 50% inhibition of the virus-induced cytopathic effect, as determined by measuring the cell viability with the colorimetric formazan-based MTS assay.

3.3. Antiviral activity

A broad antiviral screening of the 4-[1-(substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzenesulfonic acids (**1–20**) was performed against a variety of DNA and RNA viruses (Tables 10–14). None of the compounds were inhibitory to the virus-induced cytopathicity at subtoxic concentrations.

3.4. Structure activity relationship

From the results of antimicrobial and antiviral activities, the following structure activity relationship can be drawn:

1. The results of antibacterial activity indicated that compound **9**, 4-[1-(4-Nitrobenzoyl)-1H-benzimidazol-2-yl]-benzenesulfonic acid was effective against all the three bacterial species under test. It may be due to the presence of the electron withdrawing nitro group. The role of electron withdrawing group in increasing the antimicrobial activity is similar to the results of Sharma et al. [44].
2. The results of antifungal activity indicated that compound **18**, 4-(1-octadec-9-enoyl-1H-benzimidazol-2-yl)-benzenesulfonic acid was effective against both tested fungal strains.
3. The comparison of most effective antibacterial and antifungal compound gave information that the aromatic acid substitution at N₁ of benzimidazole is required for antibacterial activity and aliphatic acid substitution at N₁ of benzimidazole is required for antifungal activity. This indicated that different structural requirements are essential for binding of drug to bacterial or fungal targets respectively [45].
4. None of the compounds were endowed with antiviral activity at subtoxic concentrations.

Table 11

Cytotoxicity and antiviral activity of synthesized benzimidazole derivatives in HEL cell cultures.

Comp.	Minimum cytotoxic concentration ^a (μg/mL)	EC ₅₀ ^b (μg/mL)				
		Herpes simplex virus-1 (KOS)	Herpes simplex virus-2 (G)	Vaccinia virus	Vesicular stomatitis virus	Herpes simplex virus-1 TK ⁻ KOS ACV ^r
1	>100	>100	>100	>100	>100	>100
2	>100	>100	>100	>100	>100	>100
3	>100	>100	>100	>100	>100	>100
4	>100	>100	>100	>100	>100	>100
5	>100	>100	>100	>100	>100	>100
6	>100	>100	>100	>100	>100	>100
7	100	>20	>20	>20	>20	>20
8	>100	>100	>100	>100	>100	>100
9	>100	>100	>100	>100	>100	>100
10	>100	>100	>100	>100	>100	>100
11	>100	>100	>100	>100	>100	>100
12	>100	>100	>100	>100	>100	>100
13	100	>20	>20	>20	>20	>20
14	>100	>100	>100	>100	>100	>100
15	>100	>100	>100	>100	>100	>100
16	>100	>100	>100	>100	>100	>100
17	>100	>100	>100	>100	>100	>100
18	>100	>100	>100	>100	>100	>100
19	>100	>100	>100	>100	>100	>100
20	>100	>100	>100	>100	>100	>100
Brivudin (μM)	>250	0.08	250	112	>250	>250
Ribavirin (μM)	>250	1	2	10	>250	2
Cidofovir (μM)	>250	0.4	0.2	>250	>250	>250
Ganciclovir (μM)	>100	0.03	0.03	>100	>100	58

^a Required to cause a microscopically detectable alteration of normal cell morphology.^b Required to reduce virus-induced cytopathogenicity by 50%.

Table 12
Cytotoxicity and antiviral activity of synthesized benzimidazole derivatives in HeLa cell cultures.

Comp.	Cytotoxicity ($\mu\text{g/ml}$)		EC_{50}^c ($\mu\text{g/ml}$)					
	CC_{50}^a	Minimum cytotoxic concentration ^b	Vesicular stomatitis virus		Coxsackie virus B4		Respiratory syncytial virus	
			visual CPE score	MTS	visual CPE score	MTS	visual CPE score	MTS
1	>100	≥ 20	>20	>20	>20	>20	>20	>20
2	>100	>100	>100	>100	>100	>100	>100	>100
3	>100	>100	>100	>100	>100	>100	>100	>100
4	>100	>100	>100	>100	>100	>100	>100	>100
5	>100	>100	>100	>100	>100	>100	>100	>100
6	>100	>100	>100	>100	>100	>100	>100	>100
7	>100	>100	>100	>100	>100	>100	>100	>100
8	>100	>100	>100	>100	>100	>100	>100	>100
9	>100	>100	>100	>100	>100	>100	>100	>100
10	>100	>100	>100	>100	>100	>100	>100	>100
11	>100	>100	>100	>100	>100	>100	>100	>100
12	>100	>100	>100	>100	>100	>100	>100	>100
13	>100	>100	>100	>100	>100	>100	>100	>100
14	>100	>100	>100	>100	>100	>100	>100	>100
15	>100	>100	>100	>100	>100	>100	>100	>100
16	>100	>100	>100	>100	>100	>100	>100	>100
17	>100	>100	>100	>100	>100	>100	>100	>100
18	>100	>100	>100	>100	>100	>100	>100	>100
19	>100	>100	>100	>100	>100	>100	>100	>100
20	>100	>100	>100	>100	>100	>100	>100	>100
DS-5000	>100	>100	>100	>100	20	80.5	4	3.6
(S)-DHPA (μM)	>250	>250	>250	>250	>250	>250	>250	>250
Ribavirin (μM)	>250	>250	10	3.6	50	14	6	2.8

^a 50% Cytotoxic concentration, as determined by measuring the cell viability with the colorimetric formazan-based MTS assay.

^b Minimum compound concentration that causes a microscopically detectable alteration of normal cell morphology.

^c 50% Effective concentration, or concentration resulting in 50% inhibition of virus-induced cytopathic effect, as determined by visual scoring of the CPE, or by measuring the cell viability with the colorimetric formazan-based MTS assay.

Table 13
Cytotoxicity and antiinfluenza activity of synthesized benzimidazole derivatives in MDCK cell cultures.

Comp.	Cytotoxicity ($\mu\text{g/ml}$)		EC_{50}^c ($\mu\text{g/ml}$)					
	CC_{50}^a	MCC ^b	Influenza A H1N1 subtype		Influenza A H3N2 subtype		Influenza B	
			visual CPE score	MTS	visual CPE score	MTS	visual CPE score	MTS
1	4.1	≥ 4	>4	>4	>4	>4	>4	>4
2	>100	>100	>100	>100	>100	>100	>100	>100
3	46.8	20	>4	>4	>4	>4	>4	>4
4	>100	>100	>100	>100	>100	>100	>100	>100
5	>100	>100	>100	>100	>100	>100	>100	>100
6	>100	100	>20	>20	>20	>20	>20	>20
7	54.0	20	>4	>4	>4	>4	>4	>4
8	>100	>100	>100	>100	>100	>100	>100	>100
9	>100	>100	>100	>100	>100	>100	>100	>100
10	>100	>100	>100	>100	>100	>100	>100	>100
11	>100	>100	>100	>100	>100	>100	>100	>100
12	>100	>100	>100	>100	>100	>100	>100	>100
13	14.1	≥ 0.8	>0.8	>0.8	>0.8	>0.8	>0.8	>0.8
14	>100	>100	>100	>100	>100	>100	>100	>100
15	>100	>100	>100	>100	>100	>100	>100	>100
16	>100	100	>20	>20	>20	>20	>20	>20
17	>100	>100	>100	>100	>100	>100	>100	>100
18	>100	>100	>100	>100	>100	>100	>100	>100
19	>100	>100	>100	>100	>100	>100	>100	>100
20	>100	>100	>100	>100	>100	>100	>100	>100
Oseltamivir carboxylate (μM)	>100	>100	12	28	9	6.9	>100	>100
Ribavirin (μM)	>100	>100	9	12	9	8.4	7	4.3
Amantadine (μM)	>200	>200	18	44	0.3	0.2	>200	>200
Rimantadine (μM)	>200	>200	4	12	0.03	0.02	>200	>200

^a 50% Cytotoxic concentration, as determined by measuring the cell viability with the colorimetric formazan-based MTS assay.

^b Minimum cytotoxic concentration, i.e. Minimum compound concentration that causes a microscopically detectable alteration of normal cell morphology.

^c 50% Effective concentration, or compound concentration resulting in 50% inhibition of the virus-induced cytopathic effect, as determined by visual scoring of the CPE, or by measuring the cell viability with the colorimetric formazan-based MTS assay.

Table 14
Cytotoxicity and antiviral activity of synthesized benzimidazole derivatives in in Vero cell cultures.

Comp.	Minimum cytotoxic concentration ^a (μg/ml)	EC ₅₀ ^b (μg/ml)				
		Para-influenza-3 virus	Reovirus-1	Sindbis virus	Coxsackie virus B4	Punta Toro virus
1	20	>4	>4	>4	>4	>4
2	>100	>100	>100	>100	>100	>100
3	>100	>100	>100	>100	>100	>100
4	>100	>100	>100	>100	>100	>100
5	>100	>100	>100	>100	>100	>100
6	>100	>100	>100	>100	>100	>100
7	>100	>100	>100	>100	>100	>100
8	>100	>100	>100	>100	>100	>100
9	>100	>100	>100	>100	>100	>100
10	>100	>100	>100	>100	>100	>100
11	>100	>100	>100	>100	>100	>100
12	>100	>100	>100	>100	>100	>100
13	>100	>100	>100	>100	>100	>100
14	>100	>100	>100	>100	>100	>100
15	>100	>100	>100	>100	>100	>100
16	>100	>100	>100	>100	>100	>100
17	>100	>100	>100	>100	>100	>100
18	>100	>100	>100	>100	>100	>100
19	>100	>100	>100	>100	>100	>100
20	>100	>100	>100	>100	>100	>100
DS-5000	>100	>100	>100	100	58	100
(S)-DHPA (μM)	>250	>250	>250	>250	>250	>250
Ribavirin (μM)	>250	95	112	>250	250	112

^a Required to cause a microscopically detectable alteration of normal cell morphology.

^b Required to reduce virus-induced cytopathogenicity by 50%.

4. Conclusion

A series of 4-[1-(substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzenesulfonic acids (**1**–**20**), was synthesized by the reaction of 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid with corresponding aryl/alkyl acid chlorides. The synthesized compounds were characterized by physicochemical and spectral means and the IR and NMR spectral data are found in agreement with the assigned molecular structures. Further, the synthesized compounds were evaluated for their *in vitro* antimicrobial activities against the bacterial strains *S. aureus*, *B. subtilis*; *E. coli* and fungal strains – *A. niger* and *C. albicans* by tube dilution method. The antimicrobial screening results indicated that compounds 4-[1-(4-Nitrobenzoyl)-1H-benzimidazol-2-yl]-benzenesulfonic acid (**9**) and 4-(1-octadec-9-enoyl-1H-benzimidazol-2-yl)-benzenesulfonic acid (**18**) were the most active ones. The antiviral evaluations revealed no specific antiviral activity for the test compounds. The QSAR study carried out to find the relationship between physicochemical parameters and antimicrobial activity of benzimidazole derivatives indicated the importance of the topological parameter, Balaban index (J) followed by the electronic parameter, LUMO and the topological parameter, valence second order molecular connectivity index ($^2\chi^v$) in describing the antimicrobial activity of 4-[1-(substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzenesulfonic acids (**1**–**20**).

4.1. Synthesis of 4-[1-(2-amino-benzoyl)-1H-benzimidazol-2-yl]-benzenesulfonic acid (**1**)

4-Amino-benzenesulfonic acid (0.13 mol) in hydrochloric acid/water mixture (1:1) was diazotized using solution of sodium nitrite at 0–10 °C. To the diazotized mixture, benzimidazole (0.004 mol) was added with vigorous shaking. A solution of sodium acetate (40 g in 100 ml) was added drop wise to the above mixture by

maintaining temperature at 5–10 °C. The above solution was stirred initially for 3 h at cold condition followed by continuation of stirring at room temperature for 48 h. The product, 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid obtained was filtered, dried and recrystallized using alcohol.

A solution of 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid (0.002 mol) in diethyl ether (50 ml) was added with a solution of acid chloride of anthranilic acid (0.002 mol) in diethyl ether (50 ml). The above mixture was stirred for 24 h at room temperature. The resultant product, 4-[1-(2-amino-benzoyl)-1H-benzimidazol-2-yl]-benzenesulfonic acid was isolated by evaporation of ether and purified by recrystallisation with methanol.

The compounds **2**–**20** are synthesized by the similar procedure followed for compound **1** using corresponding aryl/alkyl acid chlorides with 4-(1H-benzimidazol-2-yl)-benzenesulfonic acid.

Compound 1: Mp (°C) 116–118; Yield-70.9%; ¹H NMR (DMSO) δ ppm: 6.54–8.56 (m, 12H, Ar–H of ArNH₂, ArSO₃H and benzimidazole), 2.50 (s, 1H, OH of SO₃H); IR (KBr pellets, cm⁻¹): 1618.29 (C=O str., tertiary amide), 755.51 (OCN bending), 1442.0 (C=C str., skeletal vibration of phenyl nucleus), 1233.60 (SO₃ asymmetric str.), 3481.81 (NH asymmetric str., primary amine).

Compound 6: Mp (°C) 98–100; Yield-101.6%; ¹H NMR (DMSO) δ ppm: 7.0–8.11 (m, 12H, Ar–H of ArOH ArSO₃H and benzimidazole), 2.50 (s, 1H, OH of SO₃H); IR (KBr pellets, cm⁻¹): 1610.90 (C=O str., tertiary amide), 750.67 (OCN bending), 1452.01 (C=C str., skeletal vibration of phenyl nucleus), 1106.82 (SO₃ asymmetric str.), 1715.01 (CH str., aromatic), 991.75 (C–C out of plane bending; 1,3-disubstituted benzene).

Compound 8: Mp (°C) 90–92; Yield-60.9%; ¹H NMR (DMSO) δ ppm: 6.95–8.10 (m, 12H, Ar–H of ArOCH₃, ArSO₃H and benzimidazole), 3.78 (s, 3H, OH, OCH₃); IR (KBr pellets, cm⁻¹): 1601.31 (C=O str., tertiary amide), 751.4 (OCN bending), 1454.21 (C=C str., skeletal vibration of phenyl nucleus), 1258.08 (SO₃ asymmetric str.), 1687.12 (CH str., aromatic), 1425.61 (CH₃ bending of OCH₃).

Compound 14: Mp (°C) 102–104; Yield-33.5%; ¹H NMR (DMSO) δ ppm: 6.90–7.84 (m, 8H, Ar–H of ArSO₃H and benzimidazole), 3.66–3.72 (d, 2H, CH₂ of C=CH₂; *J*_{trans} = 18 Hz); IR (KBr pellets, cm⁻¹): 1613.57 (C=O str., tertiary amide), 755.49 (OCN bending), 1450.12 (C=C str., skeletal vibration of phenyl nucleus), 1233.56 (SO₃ asymmetric stretch), 1004.74 (C–H out of plane bending of C(CH₃) = CH₂), 1384.75 (CH in plane bending of C(CH₃) = CH₂), 1531.84 (C=C str.), 1262.16 (CH in plane bending of phenyl ring).

Compound 15: Mp (°C) 110–112; Yield-44.0%; ¹H NMR (DMSO) δ ppm: 7.27–8.12 (m, 8H, Ar–H of ArSO₃H and benzimidazole), 2.50 (s, 1H, OH of SO₃H), 8.46–9.66 (m, 4H, H of pyridine ring); IR (KBr pellets, cm⁻¹): 1617.65 (C=O str., tertiary amide), 751.40 (OCN bending), 1450.12 (C=C str., skeletal vibration of phenyl nucleus), 1233.56 (SO₃ asymmetric stretch), 1531.84 (C=C str.), 1266.25 (CH in plane bending of phenyl ring), 1392.92 (C–N str. of tertiary amide), 829.04 (C–H out of plane bending of 3-substituted pyridine).

Compound 20: Mp (°C) 156–158; Yield-35.8%; ¹H NMR (DMSO) δ ppm: 7.12–7.88 (m, 12H, Ar–H of ArCH₃, ArSO₃H and benzimidazole), 2.50 (s, 3H, CH₃ of ArCH₃); IR (KBr pellets, cm⁻¹): 1621.74 (C=O str., tertiary amide), 1450.12 (C=C str., skeletal vibration of phenyl nucleus), 1192.7 (SO₃ asymmetric stretch), 808.61 (S–N str.), 1033.34 (SO₂ symmetric str.), 747.32 (OCN bending).

4.2. Evaluation of antimicrobial activity

4.2.1. Determination of minimum inhibitory concentration

The antimicrobial activity was performed against Gram-positive bacteria: *S. aureus*, *Bacillus subtilis*, Gram-negative bacterium: *E. coli* and fungal strains: *C. albicans* and *A. niger* by tube dilution method. Dilutions of test and standard compounds [ciprofloxacin (antibacterial) and fluconazole (antifungal)] were prepared in double strength nutrient broth – I.P. (bacteria) and Sabouraud dextrose broth I.P. (fungi) [46]. The samples were incubated at 37 °C for 24 h (bacteria), at 25 °C for 7 d (*A. niger*) and at 37 °C for 48 h (*C. albicans*), respectively, and the results were recorded in terms of MIC (the lowest concentration of test substance which inhibited the growth of microorganisms).

4.2.2. Determination of minimum bactericidal/fungicidal concentration (MBC/MFC)

The minimum bactericidal concentration (MBC) and fungicidal concentration (MFC) were determined by subculturing 100 μL of culture from each tube that remained clear in the MIC determination into fresh medium. MBC and MFC values represent the lowest concentration of compound that produces a 99.9% end point reduction [47].

4.3. QSAR studies

The structures of substituted benzimidazole derivatives are first pre-optimized with the Molecular Mechanics Force Field (MM+) procedure included in Hyperchem 6.03 [48] and the resulting geometries are further refined by means of the semiempirical method PM3 (parametric Method-3). We chose a gradient norm limit of 0.01 kcal/Å° for the geometry optimization. The lowest energy structure was used for each molecule to calculate physico-chemical properties using TSAR 3.3 software for Windows [49]. Further, the regression analysis was performed using the SPSS software package [50].

The predictive powers of the equations were validated by leave one out (LOO) cross validation method, where a model is built with N-1 compounds and Nth compound is predicted. Each compound is left out of the model derivation and predicted in turn. An indication

of the performance is obtained from cross validated q^2 method which is defined as

$$q^2 = 1 - \frac{\sum Y_{\text{predicted}} - Y_{\text{actual}}^2}{\sum Y_{\text{actual}} - Y_{\text{mean}}^2}$$

where, $Y_{\text{predicted}}$, Y_{actual} and Y_{mean} are predicted, actual and mean values of target property (pMIC) respectively. $\sum (Y_{\text{predicted}} - Y_{\text{actual}})^2$ is predictive residual error sum of squares [51].

4.4. Evaluation of antiviral activity

4.4.1. Antiviral assays

The antiviral screening of the 4-[1-(substituted aryl/alkyl carbonyl)-benzimidazol-2-yl]-benzenesulfonic acids (**1-20**) was performed against Feline Corona virus (FIPV), Feline Herpes virus in CRFK cell cultures; Herpes simplex virus-1 (KOS) [HSV-1 KOS], Herpes simplex virus-2 (G) [HSV-2G], Vaccinia virus [VV], Vesicular stomatitis virus [VSV], Herpes simplex virus-1 TK⁻ KOS ACV^r [HSV-1 TK⁻ KOS ACV^r] in HEL cell cultures; Vesicular stomatitis virus (VSV), Coxsackie virus B4, Respiratory syncytial virus in HeLa cell cultures; Para-influenza-3 virus, Reovirus-1, Sindbis virus, Coxsackie virus B4, Punta Toro virus in Vero cell cultures; Influenza A virus H1N1 subtype, Influenza A virus H3N2 subtype and influenza B virus in MDCK cell cultures, human immunodeficiency virus type 1 (HIV-1)(III_B) and HIV-2(ROD) in MT-4 cell cultures and the results were expressed as the 50% effective concentration (EC₅₀) or drug concentration required to inhibit virus-induced cytopathicity by 50%. Read-out was through microscopical inspection or the MTS viability staining method. Cells, grown in 96-well plates, were infected with 100 CCID₅₀ of virus, one CCID₅₀ being the 50% cell culture infective dose in the presence of serial dilutions of the compounds. The cultures were further incubated at 37 °C for several (2–4) days, until complete cytopathicity was observed in the infected and untreated virus control.

4.4.2. Cytotoxic assays

The cytotoxicity of the compounds was evaluated in parallel with their antiviral activity in uninfected cell cultures, and is expressed as the minimum cytotoxic concentration (MCC) that causes a microscopically detectable alteration of normal cell morphology (HEL, HeLa, CRFK, MDCK and Vero cells).

Acknowledgements

We would like to thank Mrs. Leentje Persoons, Frieda De Meyer, Kristien Erven and Kris Uyttersprot, Rega Institute for Medical Research, Belgium for their excellent technical assistance in the evaluation of antiviral activity. The antiviral evaluations were supported by the K.U. Leuven (GOA no. 10/014).

References

- [1] M. Saleh, S. Abbott, V.C. Lauzon, C. Penney, B. Zacharie, *Bioorg. Med. Chem. Lett.* 20 (2010) 945–949.
- [2] J.F. Miller, E.M. Turner, K.S. Gudmundsson, S. Jenkinson, A. Spaltenstein, M. Thomson, P. Wheelan, *Bioorg. Med. Chem. Lett.* 20 (2010) 2125–2128.
- [3] M. Tonelli, M. Simone, B. Tasso, F. Novelli, V. Boido, F. Sparatore, G. Paglietti, S. Pricl, G. Giliberti, S. Blois, C. Ibba, G. Sanna, R. Loddo, P.L. Colla, *Bioorg. Med. Chem.* 18 (2010) 2937–2953.
- [4] X. Luo, Z. Zhang, Y. Yang, F. Xue, N. Xiu, Y. She, *Front. Chem. Eng. China* 3 (3) (2009) 305–309.
- [5] V.S. Pilyugin, S.L. Kuznetsova, Y.E. Sapozhnikov, G.E. Chikisheva, G.V. Kiseleva, T.P. Vorob'eva, E.V. Klimakova, N.A. Sapozhnikova, R.D. Davletov, Z.B. Galeeva, *Russ. J. Gen. Chem.* 78 (3) (2008) 446–450.
- [6] V.S. Pilyugin, A.N. Mikhailyuk, V.M. Kosareva, G.E. Chikisheva, G.V. Kiseleva, S.L. Kuznetsova, T.P. Vorobyeva, E.V. Klimakova, *Russ. J. Org. Chem.* 39 (8) (2003) 1154–1160.
- [7] L. Chomicz, M. Padzik, A.E. Laudy, M. Kozłowska, A. Pietruczuk, J. Piekarczyk, N. Godineau, G. Oledzka, Z. Kazimierczuk, *Acta Parasitol.* 54 (2) (2009) 165–171.

- [8] A.I. Bokanov, M.I. Evstratova, K.F. Turchin, V.G. Granik, N.I. Andreeva, V.V. Asnina, S.M. Golovina, M.D. Mashkovskii, *Pharm. Chem. J.* 31 (10) (1997) 27–29.
- [9] S.M. Rida, S.A.M. EL-Hawash, H.T.Y. Fahmy, A.A. Hazzaa, M.M.M. El-Meligy, *Arch. Pharm. Res.* 29 (10) (2006) 826–833.
- [10] S.R. Ranganatha, C.V. Kavitha, K. Vinaya, D.S. Prasanna, S. Chandrappa, S.C. Raghavan, K.S. Rangappa, *Arch. Pharm. Res.* 32 (10) (2009) 1335–1343.
- [11] V.M. Dianov, *Pharm. Chem. J.* 41 (6) (2007) 20–21.
- [12] H.L. Kou, J.C. Lien, C.H. Chung, C.H. Chang, S.C. Lo, I.C. Tsai, H.C. Peng, S.C. Kuo, T.F. Huang, *Naunyn-Schmied Arch. Pharmacol* 381 (6) (2010) 495–505.
- [13] C. Huang, M.J. Embrechts, N. Sukumar, C.M. Breneman, *Artif. Neural Networks. ICANN 4668* (2007) 628–637.
- [14] J. Mungalpara, A. Pandey, V. Jain, C.G. Mohan, *J. Mol. Model.* 16 (2010) 629–644.
- [15] D. Sharma, B. Narasimhan, P. Kumar, V. Judge, R. Narang, E.D. Clercq, J. Balzarini, *Eur. J. Med. Chem.* 44 (2009) 2347–2353.
- [16] D. Kumar, V. Judge, R. Narang, S. Sangwan, E.D. Clercq, J. Balzarini, B. Narasimhan, *Eur. J. Med. Chem.* 45 (2010) 2806–2816.
- [17] B.S. Furniss, A.J. Hannaford, P.W.G. Smith, A.R. Tatchell, *Vogel's Textbook of Practical Organic Chemistry*. Addison Wesley Longman Inc., California, 1998, pp. 34, 951.
- [18] R. Dahiya, D. Pathak, *Eur. J. Med. Chem.* 42 (2007) 772–798.
- [19] J.G. Cappucino, N. Sherman, *Microbiology – A Laboratory Manual*. Addison Wesley Longman Inc, California, 1999, pp. 263.
- [20] S. Emami, M. Falhati, A. Banifafemi, A. Shafiee, *Bioorg. Med. Chem.* 12 (2004) 5881–5889.
- [21] C. Hansch, T. Fujita, *J. Am. Chem. Soc.* 86 (1964) 1616–1626.
- [22] C. Hansch, A. Leo, S.H. Unger, K.H. Kim, D. Nikaitani, E.J. Lien, *J. Med. Chem.* 16 (11) (1973) 1207–1216.
- [23] L.B. Kier, L.H. Hall, *Molecular Connectivity in Chemistry and Drug Research*. Academic Press, New York, 1976.
- [24] M. Randic, *J. Am. Chem. Soc.* 97 (1975) 6609–6615.
- [25] A.T. Balaban, *Chem. Phys. Lett.* 89 (1982) 399–404.
- [26] H. Wiener, *J. Am. Chem. Soc.* 69 (1947) 17–20.
- [27] M. Randic, *Croat. Chem. Acta* 66 (1993) 289–312.
- [28] J.M. Heravi, A. Kyani, *J. Chem. Inf. Comput. Sci.* 44 (2005) 1328–1335.
- [29] A. Golbraikh, A. Tropsha, *J. Mol. Graph. Model.* 20 (4) (2002) 269–276.
- [30] S. Bajaj, S.S. Sambhi, A.K. Madan, *Croat. Chem. Acta* 78 (2) (2005) 165–174.
- [31] A. Kumar, P. Sharma, V.K. Gurram, N. Rane, *Biorg. Med. Chem. Lett.* 16 (2006) 2484–2491.
- [32] P. Sharma, A. Kumar, M. Sharma, *Eur. J. Med. Chem.* 41 (2006) 833–840.
- [33] S.A. Hatya, E. Aki-sener, B. Tekiner-Gulbas, I. Yildiz, O. Temiz-Arpaci, I. Yalcin, N. Altanlar, *Eur. J. Med. Chem.* 41 (2006) 1398–1404.
- [34] A. Kumar, B. Narasimhan, D. Kumar, *Bioorg. Med. Chem.* 15 (2007) 4113–4124.
- [35] B. Narasimhan, V. Judge, R. Narang, S. Ohlan, R. Ohlan, *Bioorg. Med. Chem. Lett.* 17 (2007) 5836–5845.
- [36] F.J. Prado-Prado, H. Gonzalez-Diaz, O.M. de la Vega, F.M. Ubeira, K.C. Chou, *Bioorg. Med. Chem.* 16 (11) (2008) 5871–5880.
- [37] H. Gonzalez-Diaz, F.J. Prado-Prado, *J. Comput. Chem* 29 (4) (2008) 656–667.
- [38] M. Cruz-Montegudo, H. Gonzalez-Diaz, G. Agüero-Chapin, L. Santana, F. Borges, E.R. Dominguez, G. Podda, E. Uriarte, *J. Comput. Chem.* 28 (11) (2007) 1909–1923.
- [39] H. Gonzalez-Diaz, S. Vilar, L. Santana, E. Uriarte, *Curr. Top. Med. Chem.* 7 (10) (2007) 1015–1029.
- [40] H. Gonzalez-Diaz, Y. Gonzalez-Diaz, L. Santana, F.M. Ubeira, E. Uriarte, *Proteomics* 8 (4) (2008) 750–778.
- [41] B. Zhou, N. Trinajstić, *Croat. Chem. Acta* 81 (2) (2008) 319–323.
- [42] P. Kumar, B. Narasimhan, D. Sharma, *ARKIVOC* xiii (2008) 159–178.
- [43] D. Sharma, B. Narasimhan, P. Kumar, A. Jalbout, *Eur. J. Med. Chem.* 44 (3) (2009) 1119–1127.
- [44] P. Sharma, N. Rane, V.K. Gurram, *Bioorg. Med. Chem. Lett.* 14 (2004) 4185–4190.
- [45] M. Sortino, P. Delgado, S. Jaurez, J. Quiroga, R. Abonia, B. Insuasey, M. Noguera, L. Rodero, F.M. Garibotto, R.D. Enriz, S.A. Zacchino, *Bioorg. Med. Chem. Lett.* 15 (2007) 484–494.
- [46] *Pharmacopoeia of India, vol. II*, Ministry of Health Department: Govt. of India, New Delhi, 1996, pp. A-88.
- [47] M.C. Rodriguez-Argüelles, E.C. Lopez- Silva, J. Sanmartin, P. Pelagatti, F. Zani, *J. Inorg. Biochem.* 99 (2005) 2231–2239.
- [48] *Hyperchem 6.0*, Hypercube, Inc., Florida, 1993.
- [49] *TSAR 3D Version 3.3*, Oxford Molecular Limited, 2000.
- [50] *SPSS for Windows Version 10.05*, SPSS Inc., Bangalore, India, 1999.
- [51] I.V. Tetko, V.Y. Tanchuk, A.E. Villa, *J. Chem. Inf. Comput. Sci.* 41 (2001) 1407–1421.