



Non-*fumigatus* *Aspergillus* Infection Associated with a Negative *Aspergillus* Precipitin Test in Patients with Chronic Pulmonary Aspergillosis

Keita Takeda,^a Junko Suzuki,^a Akira Watanabe,^b Osamu Narumoto,^a Masahiro Kawashima,^a Yuka Sasaki,^a Hideaki Nagai,^a Katsuhiko Kamei,^b Hirotohi Matsui^a

^aCenter for Pulmonary Diseases, National Hospital Organization Tokyo National Hospital, Tokyo, Japan

^bDivision of Clinical Research, Medical Mycology Research Center, Chiba University, Chiba, Japan

ABSTRACT *Aspergillus* antibody testing is key for the clinical diagnosis of chronic pulmonary aspergillosis (CPA) with high sensitivity. However, false-negative results in patients with CPA might be obtained, depending on the *Aspergillus* species. The aim of this study was to investigate which factors are associated with false-negative results in *Aspergillus* precipitin tests and whether the sensitivity of precipitin tests in CPA is influenced by *Aspergillus fumigatus* and non-*fumigatus* *Aspergillus* species. Between February 2012 and December 2020, 116 consecutive antifungal treatment-naïve patients with CPA were identified and included in this retrospective chart review. *Aspergillus* species isolated from the respiratory tract of patients were identified by DNA sequencing. Characteristics of patients with positive and negative results for *Aspergillus* precipitin tests were compared. The sensitivity of the *Aspergillus* precipitin tests was compared between patients with *A. fumigatus*-associated CPA and non-*fumigatus* *Aspergillus*-associated CPA. A non-*fumigatus* *Aspergillus* species was the only factor significantly associated with negative *Aspergillus* precipitin test results in patients with CPA in the multivariate analysis (hazard ratio, 8.3; 95% confidence interval, 3.2 to 22.1; $P < 0.0001$). The positivity of the *Aspergillus* precipitin test for patients with non-*fumigatus* *Aspergillus*-associated CPA was lower than that for patients with *A. fumigatus*-associated CPA (84.8% versus 37.9%; $P < 0.0001$). These results revealed that the presence of non-*fumigatus* *Aspergillus*-associated CPA should be considered with a negative *Aspergillus* precipitin test; this finding may prevent diagnostic delay or misdiagnosis for CPA.

KEYWORDS *Aspergillus fumigatus*, *Aspergillus* antibody test, sensitivity

Chronic pulmonary aspergillosis (CPA) is a refractory fungal disease occurring as a complication of various pulmonary diseases (1–4); it has an estimated global prevalence of approximately 3 million (5). CPA is diagnosed based on clinical, microbiological, radiological, serological, and histopathological findings (1–3). The microbiological tests are important in the definite diagnosis of CPA (1–3). However, the culture-positive rate in CPA remains relatively low (6). When microbiological tests are negative, serological assays, particularly *Aspergillus* antibody testing, are key diagnostic tests with high sensitivity for CPA.

The *Aspergillus* precipitin test and *Aspergillus*-specific IgG assay are two *Aspergillus* antibody testing methods utilized commonly for diagnosing CPA. Among precipitin tests, the Ouchterlony double immunodiffusion technique has been performed to diagnose aspergillosis since the 1960s (7); this precipitin test is still used worldwide, although the efficacy of *Aspergillus*-specific IgG assays has been reported to be higher than that of precipitin tests (8, 9).

In previous studies, the sensitivity of the precipitin test has ranged from 59% to 88% in patients with CPA (8, 10). It was suggested that the false-negative results in

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Address correspondence to Keita Takeda, takeda.keita.ax@mail.hosp.go.jp.

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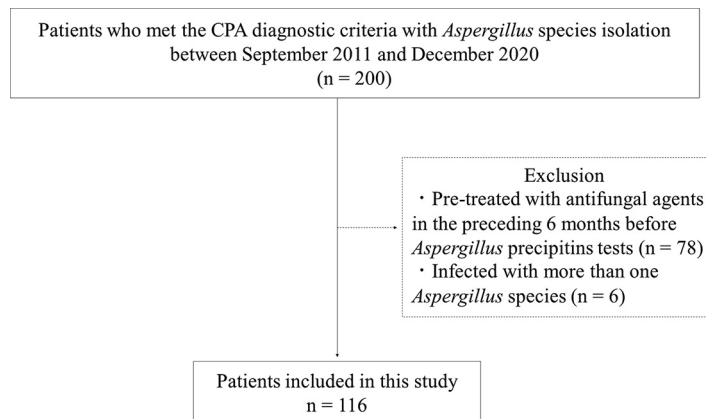


FIG 1 Patient selection flowchart. A total of 116 patients were included in this study after the exclusion of cases that did not fulfil the selection criteria.

Aspergillus antibody tests were caused by the host immune deficiency (11) and *Aspergillus* species (12). The causative organism associated most commonly with CPA is *Aspergillus fumigatus*, and the efficacy of *Aspergillus* antibody testing has been proven in most cases of *A. fumigatus*-associated CPA (8, 13). Conversely, the disease may be caused by other members of the genus *Aspergillus*, i.e., non-*fumigatus* *Aspergillus*, such as *Aspergillus niger*, *Aspergillus tubingensis*, *Aspergillus welwitschiae*, *Aspergillus flavus*, and *Aspergillus terreus* (14). The sensitivity of *Aspergillus* antibody testing for patients with non-*fumigatus* *Aspergillus*-associated CPA remains unknown.

The false-negative results in *Aspergillus* antibody tests might lead to a misdiagnosis or diagnostic delay in patients with CPA, which is a potential risk for poor prognosis due to treatment delay. Therefore, in this study, we aimed to investigate which factors are associated with false-negative results in *Aspergillus* precipitin tests and whether the sensitivity of precipitin tests in CPA is influenced by the species of *Aspergillus*, comparing between *A. fumigatus* and non-*fumigatus* *Aspergillus* species.

MATERIALS AND METHODS

Diagnostic criteria of CPA. CPA was diagnosed based on chronic pulmonary symptoms lasting for a few months and thoracic imaging showing cavitation, pleural thickening, pericavitary infiltrates, a fungal ball, or parenchymal destruction or fibrosis, as specified in practice guidelines by the Infectious Diseases Society of America (2) and the joint guidelines of the European Society for Clinical Microbiology and Infectious Diseases and the European Respiratory Society (3).

Study participants. This study was a retrospective chart review. The criteria for patient selection are shown in Fig. 1. Between February 2012 and December 2020, we identified 200 consecutive patients who met the CPA criteria described above, with *Aspergillus* species isolated from the patient respiratory tract at the NHO Tokyo National Hospital, Tokyo, Japan. A total of 116 patients with CPA were included in the study following the exclusion of (i) 78 patients who had undergone antifungal treatment in the preceding 6 months, as this treatment could possibly influence the values of *Aspergillus* antibodies in the precipitin test, and (ii) 6 patients who were suspected of being coinfecting with more than 1 *Aspergillus* species within 6 months before and after CPA diagnosis to avoid confounding effects of coinfection on the antibody test.

Patient characteristics, including age; sex; underlying lung diseases; comorbidity due to diabetes mellitus, human immunodeficiency virus seropositive (HIV), malignant diseases, and atopic diseases; use of corticosteroids and/or immunosuppressants; type of CPA; overlap allergic bronchopulmonary aspergillosis; and laboratory findings were noted.

The Institutional Review Board of NHO Tokyo National Hospital (approval number 200048) approved this study and waived the requirement for written informed consent.

***Aspergillus* species identification.** Lower respiratory tract samples, i.e., sputum, bronchoalveolar lavage fluid, endotracheal aspirate, and surgical samples, were cultured on Sabouraud dextrose agar (Kanto Kagaku, Tokyo, Japan) or potato dextrose agar (Kanto Kagaku), as described previously (15, 16). *Aspergillus* species were identified genetically using DNA sequencing of the domain 1/domain 2, internally transcribed spacer regions 1 and 2, and β -tubulin and calmodulin genes, based on previously described methods (17–19).

***Aspergillus* precipitin test.** The *Aspergillus* precipitin test was performed within 1 month of CPA diagnosis using the FSK1 *Aspergillus* immunodiffusion system (Microgen, Surrey, UK), according to the

manufacturer's instructions (20). The results were recorded as "1+" to "3+" based on the precipitation arc. The results of 1+ or 2+ were recorded according to the number of precipitation arcs. The result of 3+ was recorded when the number of precipitation arcs was >2.

Statistical analysis. The chi-square test for frequency measurements and the *t* test for continuous variables were used to verify statistical differences in characteristics between patients with positive and negative *Aspergillus* precipitin test results and in the sensitivity of the *Aspergillus* precipitin test between patients with *A. fumigatus*-associated CPA and those with non-*fumigatus Aspergillus*-associated CPA. For the multivariate analysis, logistic regression analysis followed by stepwise selection (inclusion criteria, $P < 0.25$; exclusion criteria, $P < 0.10$) was used to detect factors associated with a negative result on the *Aspergillus* precipitin tests. A *P* value of <0.05 was considered statistically significant. Statistical analyses were performed using JMP 13.00 (SAS Institute Inc., Cary, NC, USA).

RESULTS

Patient characteristics. Table 1 indicates all patient characteristics, including the comparison of patients with positive and negative *Aspergillus* precipitin tests. The proportion of cases caused by non-*fumigatus Aspergillus* species was significant for patients with negative *Aspergillus* precipitin test results ($P < 0.0001$). There were no significant differences between the two groups in terms of the other characteristics.

The presence of non-*fumigatus Aspergillus* was the only significant factor associated with negative results of the *Aspergillus* precipitin test in the multivariate analysis ($P < 0.0001$; hazard ratio, 8.3; 95% confidence interval, 3.2 to 22.1) (Table 1).

Species identification. Results of *Aspergillus* species identification among the patients are shown in Fig. 2. *Aspergillus fumigatus sensu stricto* was the most frequently isolated species ($n = 87$, 75.0%) followed by *A. tubingensis* ($n = 13$, 11.2%), *A. welwitschiae* ($n = 7$, 6.0%), *A. flavus* ($n = 4$, 3.4%), *A. terreus* ($n = 3$, 2.6%), *A. niger* ($n = 1$, 0.9%), and *Aspergillus luchuensis* ($n = 1$, 0.9%).

Results of *Aspergillus* precipitin tests. Table 2 shows the results of the *Aspergillus* serological tests for patients with *A. fumigatus*-associated CPA and those with non-*fumigatus Aspergillus*-associated CPA. The positivity of the *Aspergillus* precipitin test for patients with non-*fumigatus Aspergillus*-associated CPA (37.9%) was lower than that for patients with *A. fumigatus*-associated CPA (81.6%) ($P < 0.0001$). The values of the semiquantitative analysis for the *Aspergillus* precipitin tests were also lower for patients with non-*fumigatus Aspergillus*-associated CPA than for patients with *A. fumigatus*-associated CPA ($P < 0.0001$).

Precipitin test results based on *Aspergillus* species. Table 3 shows the results of the *Aspergillus* precipitin test for patients according to each *Aspergillus* species. The positivity rates of the precipitin test for patients infected with the *Aspergillus* species varied from 0% to 100%.

DISCUSSION

In this study, non-*fumigatus Aspergillus* infection was observed to be the cause of negative *Aspergillus* precipitin test results for patients with CPA. The sensitivity and semiquantitative analysis values of the *Aspergillus* precipitin test were significantly lower for patients infected with non-*fumigatus Aspergillus* than those for patients infected with *A. fumigatus*.

The *Aspergillus* precipitin test was positive for approximately 40% of patients with non-*fumigatus Aspergillus*-associated CPA with a low reaction level in the semiquantitative analysis. The low sensitivity of the *Aspergillus* precipitin test for patients with non-*fumigatus Aspergillus*-associated CPA might have occurred because the antigens detected using the test kit had been obtained from the culture of *A. fumigatus* (20). Nevertheless, the reactions might have occurred owing to (i) cross-reactions between *A. fumigatus* and other *Aspergillus* species (21, 22) or (ii) mixed infections with *A. fumigatus* (23). Although patients infected with more than one *Aspergillus* species were excluded from the culture study, those infected with *A. fumigatus* might have been included in the non-*fumigatus Aspergillus* group because of the low sensitivity of the culture tests. *Aspergillus* precipitin tests using antigens obtained from each *Aspergillus* species were not performed in this study.

In the *Aspergillus* antibody testing, the strength of cross-reactions between *A. fumigatus* and other *Aspergillus* species might vary among different *Aspergillus* species. It

TABLE 1 Characteristics of patients with CPA and factors associated with negative *Aspergillus* precipitin tests^a

Characteristic	Data ^b for:			Univariate analysis ^c P value	Multivariate analysis	
	All patients (n = 116)	CPA patients with positive <i>Aspergillus</i> precipitin tests (n = 82)	CPA patients with negative <i>Aspergillus</i> precipitin tests (n = 34)		Adjusted OR (95% CI)	P value
Age (yrs)	69.0 ± 12.0	68.7 ± 12.3	69.9 ± 11.2	0.62		
Male/female	78 (67.2)/38 (32.8)	57 (69.5)/25 (30.5)	21 (61.8)/13 (38.2)	0.52		
<i>Aspergillus</i> species				<0.0001		
<i>Aspergillus fumigatus</i>	87 (75.0)	71 (87.8)	16 (47.1)			
non- <i>fumigatus Aspergillus</i>	29 (25.0)	11 (14.1)	18 (52.9)		8.3 (3.2–22.1)	<0.0001
Underlying pulmonary diseases ^d				0.81		
Prior pulmonary tuberculosis	38 (32.8)	30 (36.6)	8 (23.5)			
Nontuberculous pulmonary infection	29 (25.0)	20 (24.4)	9 (26.5)			
Chronic obstructive pulmonary disease	24 (20.7)	15 (18.3)	9 (26.5)			
Interstitial lung disease	11 (9.5)	8 (9.8)	3 (8.8)			
Bronchiectasis	10 (8.6)	6 (7.3)	4 (11.8)			
History of thoracic surgery	9 (7.8)	7 (8.5)	2 (5.9)			
Lung cancer	6 (5.2)	5 (6.1)	1 (2.9)			
Others ^e	10 (8.6)	8 (9.8)	2 (5.9)			
Comorbidities						
Diabetes mellitus	11 (9.5)	8 (9.8)	3 (8.8)	>0.99		
HIV	0	0	0	>0.99		
Malignant diseases ^f	8 (6.9)	6 (7.3)	2 (5.9)	>0.99		
Atopic diseases ^g	11 (9.5)	9 (11.0)	2 (2.9)	0.50		
Use of corticosteroids and/or immunosuppressants	10 (8.6)	7 (8.5)	3 (8.8)	>0.99		
Type of CPA				0.19		
Simple aspergilloma	4 (3.4)	1 (1.2)	3 (8.8)			
Chronic cavitary pulmonary aspergillosis	83 (71.6)	59 (72.0)	24 (70.6)			
Chronic fibrosing pulmonary aspergillosis	18 (15.5)	13 (15.8)	5 (14.7)			
Subacute invasive aspergillosis	11 (9.5)	9 (11.0)	2 (5.9)			
<i>Aspergillus</i> nodule	0	0	0			
ABPA overlap	1 (0.9)	1 (1.2)	0	>0.99		
Laboratory findings at diagnosis						
White blood cell count (cells/ μ L)	8,400 ± 3,968	8,439 ± 3,856	8,150 ± 4,211	0.72		
Lymphocyte count (cells/ μ L)	1,281 ± 617	1,227 ± 547	1,399 ± 745	0.17		
Eosinophil count (cells/ μ L)	283 ± 479	329 ± 554	210 ± 213	0.23		
CRP (mg/dL)	6.3 ± 7.4	6.7 ± 7.1	4.7 ± 7.8	0.18		

^aCPA, chronic pulmonary aspergillosis; HIV, human immunodeficiency virus; ABPA, allergic bronchopulmonary aspergillosis; CRP, C-reactive protein; OR, odds ratio; CI, confidence interval.

^bData are presented as mean ± SD or n (%).

^cComparison between CPA patients with positive and negative precipitin tests.

^dIncluding duplicate cases.

^eIncluding asbestosis, pneumoconiosis, bronchial asthma, and alveolar proteinosis.

^fIncluding lung cancer and gastric cancer.

^gIncluding bronchial asthma, atopic dermatitis, allergic rhinitis, and atopic conjunctivitis.

has been reported that cross-reactivity is higher in patients with *A. flavus*-associated pulmonary aspergillosis than that in patients with *A. niger*- and *A. terreus*-associated pulmonary aspergillosis (21, 22). Correspondingly, the positivity in our study was 100% in four patients with *A. flavus*-associated CPA. The positivity was 0 to 42.9% for *Aspergillus* section *Nigri*, namely, *A. tubingensis*, *A. welwitschia*, *A. niger*, and *A. luchuensis*, and 66.7% for *A. terreus*.

The sensitivity in patients with *A. fumigatus*-associated CPA was approximately 80%. In other words, approximately 20% of the tests revealed false-negative results. The reasons for the false-negative results were presumed to be as follows. (i) One reason is the

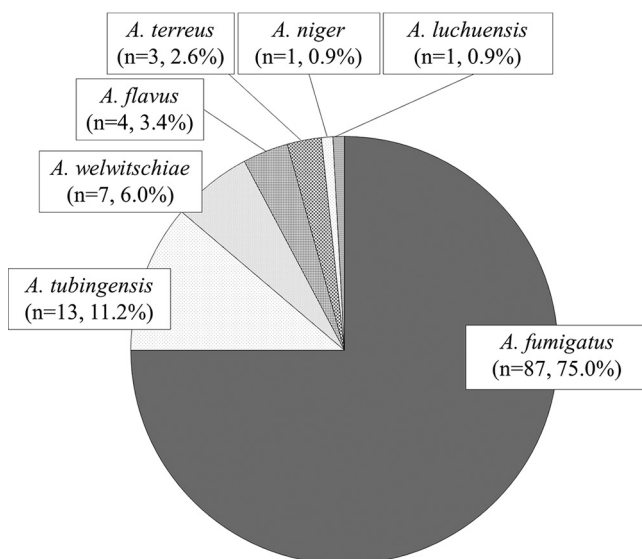


FIG 2 *Aspergillus* species isolated from the patients with chronic pulmonary aspergillosis (CPA). *Aspergillus fumigatus sensu stricto* was the most frequently isolated species, followed by *A. tubingenis*, *A. welwitschiae*, *A. flavus*, and *A. terreus*.

sensitivity of the precipitin test, as Page et al. suggested that this sensitivity was lower than that of *Aspergillus*-specific IgG tests (8). The precipitin test is a subjective test dependent partly on the testers; therefore, very low reaction levels may not be detected. (ii) Another reason is early-stage infection (20), as antibodies against *Aspergillus* infection might develop during disease progression. Therefore, antibody tests during the early stage of CPA might give false-negative results.

Aspergillus-specific IgG assays have recently become one of the best practices for diagnosing CPA. However, there has been no study regarding the efficacy of the test in patients with non-*fumigatus* *Aspergillus*-associated CPA. Guo et al. suggested that *Aspergillus*-specific IgG might be low for patients with non-*fumigatus* *Aspergillus*-associated CPA (12). This low sensitivity might be because most *Aspergillus*-specific IgG tests use recombinant proteins produced from *A. fumigatus* as antigens (9). Consequently, it is necessary to verify the sensitivity of *Aspergillus*-specific IgG assays for patients with non-*fumigatus* *Aspergillus*-associated CPA by comparing them with the *Aspergillus*-specific IgG assays for each *Aspergillus* species. In addition, new diagnostic tests will be required to increase the diagnostic sensitivity for CPA patients with non-*fumigatus* *Aspergillus* by using a common antigen of *Aspergillus* species.

TABLE 2 *Aspergillus* serological tests

Test	Data ^a for:			P value ^b
	All patients (n = 116)	Patients with <i>Aspergillus</i> <i>fumigatus</i> -associated CPA ^c (n = 87)	Patients with non- <i>fumigatus</i> <i>Aspergillus</i> -associated CPA (n = 29)	
Positive <i>Aspergillus</i> precipitin test	82 (70.7)	71 (81.6)	11 (37.9)	<0.0001
Semiquantitative results of <i>Aspergillus</i> precipitin test				<0.0001
Negative	34 (29.3)	16 (18.4)	18 (62.1)	
1+	20 (17.2)	13 (14.9)	7 (24.1)	
2+	12 (10.3)	10 (11.5)	2 (6.9)	
3+	50 (43.1)	48 (55.2)	2 (6.9)	

^aData are presented as n (%).

^bComparison between patients with CPA by *A. fumigatus* and non-*fumigatus* *Aspergillus*.

^cCPA, chronic pulmonary aspergillosis.

TABLE 3 *Aspergillus* precipitin test in patients with CPA^a according to each non-*fumigatus Aspergillus* species

Test	n (%) by species					
	<i>A. tubingensis</i> (n = 13)	<i>A. welwitschiae</i> (n = 7)	<i>A. flavus</i> (n = 4)	<i>A. terreus</i> (n = 3)	<i>A. niger</i> (n = 1)	<i>A. luchuensis</i> (n = 1)
Positive <i>Aspergillus</i> precipitin test	2 (15.4)	3 (42.9)	4 (100)	2 (66.7)	0	0
Semiquantitative results						
Negative	11 (84.6)	4 (57.1)	0	0	1 (100)	1 (100)
1+	1 (7.7)	3 (42.9)	2 (50.0)	1 (33.3)	0	0
2+	0	0	1 (25.0)	1 (33.3)	0	0
3+	1 (7.7)	0	1 (25.0)	0	0	0

^aCPA, chronic pulmonary aspergillosis.

Disregarding *Aspergillus* species, it was suggested that host immunodeficiency may influence the positivity of *Aspergillus* antibody tests (11). In this study, to the best of our knowledge, there were no immunodeficiency factors associated with false-negative results in the *Aspergillus* precipitin tests. The patients were evaluated regarding diabetes mellitus, malignant diseases, and use of corticosteroids and/or immunosuppressants; in addition, white blood cell and lymphocyte counts were determined. Other immune factors, such as immunoglobulin levels, could not be evaluated in this study. Further study will be needed to assess the influence of host immunodeficiency on *Aspergillus* antibody tests.

Our study has the following limitations. First, the number of *Aspergillus* species in patients with non-*fumigatus Aspergillus*-associated CPA varied and was small. This result warrants an investigation of the sensitivity of patients with CPA to each *Aspergillus* species. Second, this study was a single-center analysis; a multicenter study is required to validate our findings. Third, the *Aspergillus* precipitin test shows subjective semiquantitative values. A quantitative analysis is required to verify the interpretation of our study.

In conclusion, the sensitivity of the *Aspergillus* precipitin test for patients with CPA depends on the causative *Aspergillus* species. To prevent diagnostic delay, the presence of non-*fumigatus Aspergillus*-associated CPA should be considered in cases of suspected CPA with a negative *Aspergillus* precipitin test.

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REFERENCES

- Kohno S, Tamura K, Niki Y, Izumikawa K, Oka S, Ogawa K, Kadota J, Kamei K, Kanda Y, Kiuchi T, Shibuya K, Takakura S, Takata T, Takesue Y, Teruya K, Tokimatsu I, Fukuda T, Maesaki S, Makimura K, Mikamo H, Mitsutake K, Miyazaki Y, Mori M, Yasuoka A, Yano K, Yamanaka N, Yoshida M. 2016. Executive summary of Japanese Domestic Guidelines for Management of Deep-Seated Mycosis 2014. *Med Mycol J* 57:E117–E163. <https://doi.org/10.3314/mmj.16-00010>.
- Patterson TF, Thompson GR, III, Denning DW, Fishman JA, Hadley S, Herbrecht R, Kontoyiannis DP, Marr KA, Morrison VA, Nguyen MH, Segal BH, Steinbach WJ, Stevens DA, Walsh TJ, Wingard JR, Young JA, Bennett JE. 2016. Practice guidelines for the diagnosis and management of aspergillosis: 2016 update by the Infectious Diseases Society of America. *Clin Infect Dis* 63:e1–e60. <https://doi.org/10.1093/cid/ciw326>.
- Denning DW, Cadranet J, Beigelman-Aubry C, Ader F, Chakrabarti A, Blot S, Ullmann AJ, Dimopoulos G, Lange C, European Society for Clinical Microbiology and Infectious Diseases and European Respiratory Society. 2016. Chronic pulmonary aspergillosis: rationale and clinical guidelines for diagnosis and management. *Eur Respir J* 47:45–68. <https://doi.org/10.1183/13993003.00583-2015>.
- Maitre T, Cottenet J, Godet C, Roussot A, Abdoul Carime N, Ok V, Parrot A, Bonniaud P, Quantin C, Cadranet J. 2021. Chronic pulmonary aspergillosis: prevalence, favouring pulmonary diseases and prognosis. *Eur Respir J* 58:2003345. <https://doi.org/10.1183/13993003.03345-2020>.
- Barac A, Kosmidis C, Alastruey-Izquierdo A, Salzer HJF, CPAnet. 2019. Chronic pulmonary aspergillosis update: a year in review. *Med Mycol* 57: S104–S109. <https://doi.org/10.1093/mmy/myy070>.
- Vergidis P, Moore CB, Novak-Frazer L, Rautemaa-Richardson R, Walker A, Denning DW, Richardson MD. 2020. High-volume culture and quantitative real-time PCR for the detection of *Aspergillus* in sputum. *Clin Microbiol Infect* 26:935–940. <https://doi.org/10.1016/j.cmi.2019.11.019>.
- Longbottom JL, Pepys J, Clive FT. 1964. Diagnostic precipitin test in *Aspergillus* pulmonary mycetoma. *Lancet* 283:588–589. [https://doi.org/10.1016/S0140-6736\(64\)91335-2](https://doi.org/10.1016/S0140-6736(64)91335-2).

8. Page ID, Richardson MD, Denning DW. 2016. Comparison of six *Aspergillus*-specific IgG assays for the diagnosis of chronic pulmonary aspergillosis (CPA). *J Infect* 72:240–249. <https://doi.org/10.1016/j.jinf.2015.11.003>.
9. Richardson MD, Page ID. 2017. *Aspergillus* serology: have we arrived yet? *Med Mycol* 55:48–55. <https://doi.org/10.1093/mmy/myw116>.
10. Kohno S, Izumikawa K, Ogawa K, Kurashima A, Okimoto N, Amitani R, Takeya H, Niki Y, Miyazaki Y, Japan Chronic Pulmonary Aspergillosis Study Group (JCPASG). 2010. Intravenous micafungin versus voriconazole for chronic pulmonary aspergillosis: a multicenter trial in Japan. *J Infect* 61:410–418. <https://doi.org/10.1016/j.jinf.2010.08.005>.
11. Hunter ES, Wilopo B, Richardson MD, Kosmidis C, Denning DW. 2021. Effect of patient immunodeficiencies on the diagnostic performance of serological assays to detect *Aspergillus*-specific antibodies in chronic pulmonary aspergillosis. *Respir Med* 178:106290. <https://doi.org/10.1016/j.rmed.2020.106290>.
12. Guo Y, Bai Y, Yang C, Gu L. 2019. Evaluation of *Aspergillus* IgG, IgM antibody for diagnosing in chronic pulmonary aspergillosis: a prospective study from a single center in China. *Medicine (Baltimore, MD)* 98:e15021. <https://doi.org/10.1097/MD.00000000000015021>.
13. Sehgal IS, Choudhary H, Dhooria S, Aggarwal AN, Garg M, Chakrabarti A, Agarwal R. 2018. Diagnostic cut-off of *Aspergillus fumigatus*-specific IgG in the diagnosis of chronic pulmonary aspergillosis. *Mycoses* 61:770–776. <https://doi.org/10.1111/myc.12815>.
14. Godet C, Alastruey-Izquierdo A, Flick H, Hennequin C, Mikilps-Mikgelbs R, Munteanu O, Page I, Seidel D, Salzer HJF, CPAnet. 2018. A CPAnet consensus statement on research priorities for chronic pulmonary aspergillosis: a neglected fungal infection that requires attention. *J Antimicrob Chemother* 73:280–286. <https://doi.org/10.1093/jac/dkx390>.
15. Takeda K, Suzuki J, Watanabe A, Matsuki M, Higa K, Inoue E, Akashi S, Shimada M, Kawashima M, Ohshima N, Fukami T, Masuda K, Yamane A, Tamura A, Nagai H, Matsui H, Tohma S, Kamei K. 2020. Species identification, antifungal susceptibility, and clinical feature association of *Aspergillus* section Nigri isolates from the lower respiratory tract. *Med Mycol* 58:310–314. <https://doi.org/10.1093/mmy/myz072>.
16. Takeda K, Suzuki J, Watanabe A, Arai T, Koiwa T, Shinfuku K, Narumoto O, Kawashima M, Fukami T, Tamura A, Nagai H, Matsui H, Kamei K. 2021. High detection rate of azole-resistant *Aspergillus fumigatus* after treatment with azole antifungal drugs among patients with chronic pulmonary aspergillosis in a single hospital setting with low azole resistance. *Med Mycol* 59:327–334. <https://doi.org/10.1093/mmy/myaa052>.
17. Yaguchi T, Horie Y, Tanaka R, Matsuzawa T, Ito J, Nishimura K. 2007. Molecular phylogenetics of multiple genes on *Aspergillus* section Fumigati isolated from clinical specimens in Japan. *Nihon Ishinkin Gakkai Zasshi* 48:37–46. <https://doi.org/10.3314/jjmm.48.37>.
18. O'Donnell K. 1993. *Fusarium* and its near relatives, p 225–236. In Reynolds DR, Taylor JW (ed), *The fungal holomorph: mitotic, meiotic, and pleomorphic speciation in fungal systematics*. CAB International, Wallingford, UK.
19. White TJ, Bruns T, Lee S, Taylor J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics, p 315–322. In Innis MA, Gelfand DH, Sninsky JJ, White TJ (ed), *PCR protocols: a guide to methods and applications*. Academic Press, London, UK.
20. Key Diagnostics. FSK1 *Aspergillus* immunodiffusion system package insert. Key Diagnostics, Gympsea, Australia. [http://www.keydiagnostics.com.au/images/PDF/new/Aspergillus%20Immunodiffusion%20System/CEFSK1%20Aspergillus%20\(Eng,Ital,Span,Ger,Fre\)%20KD%2012-13.pdf](http://www.keydiagnostics.com.au/images/PDF/new/Aspergillus%20Immunodiffusion%20System/CEFSK1%20Aspergillus%20(Eng,Ital,Span,Ger,Fre)%20KD%2012-13.pdf).
21. Harada K, Oguma T, Saito A, Fukutomi Y, Tanaka J, Tomomatsu K, Taniguchi M, Asano K. 2018. Concordance between *Aspergillus*-specific precipitating antibody and IgG in allergic bronchopulmonary aspergillosis. *Allergol Int* 67:S12–S17. <https://doi.org/10.1016/j.alit.2018.04.009>.
22. Sehgal IS, Choudhary H, Dhooria S, Aggarwal AN, Bansal S, Garg M, Behera D, Chakrabarti A, Agarwal R. 2019. Prevalence of sensitization to *Aspergillus flavus* in patients with allergic bronchopulmonary aspergillosis. *Med Mycol* 57:270–276. <https://doi.org/10.1093/mmy/myy012>.
23. Orzechowski XM, Pasqualotto AC, Uchoa SMP, Bittencourt SC, Peixoto CJJ, Severo LC. 2008. Invasive pulmonary aspergillosis due to a mixed infection caused by *Aspergillus flavus* and *Aspergillus fumigatus*. *Rev Iberoaam Micol* 25:176–178. [https://doi.org/10.1016/S1130-1406\(08\)70041-X](https://doi.org/10.1016/S1130-1406(08)70041-X).