

Review Article

***Chlamydomphila pneumoniae* Infection and Its Role in Neurological Disorders**

Carlo Contini,¹ Silva Seraceni,¹ Rosario Cultrera,¹ Massimiliano Castellazzi,² Enrico Granieri,² and Enrico Fainardi³

¹ Section of Infectious Diseases, Department of Clinical and Experimental Medicine, National Institute of Neuroscience and Neuroscience Center, University of Ferrara, Via Fossato di Mortara, 23, 44100 Ferrara, Italy

² Section of Neurology, Department of Medical and Surgical Sciences of the Communication and Behaviour, Multiple Sclerosis Center, National Institute of Neuroscience and Neuroscience Center, University of Ferrara, Arcispedale S. Anna, Corso della Giovecca 203, 44100 Ferrara, Italy

³ Neuroradiology Unit, Department of Neurosciences and Rehabilitation, Azienda Ospedaliera-Universitaria, National Institute of Neuroscience and Neuroscience Center, University of Ferrara, Arcispedale S. Anna, Corso della Giovecca 203, 44100 Ferrara, Italy

Correspondence should be addressed to Carlo Contini, cnc@unife.it

Received 27 July 2009; Accepted 25 November 2009

Academic Editor: Guey Chuen Perng

Copyright © 2010 Carlo Contini et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Chlamydomphila pneumoniae is an intracellular pathogen responsible for a number of different acute and chronic infections. The recent deepening of knowledge on the biology and the use of increasingly more sensitive and specific molecular techniques has allowed demonstration of *C. pneumoniae* in a large number of persons suffering from different diseases including cardiovascular (atherosclerosis and stroke) and central nervous system (CNS) disorders. Despite this, many important issues remain unanswered with regard to the role that *C. pneumoniae* may play in initiating atheroma or in the progression of the disease. A growing body of evidence concerns the involvement of this pathogen in chronic neurological disorders and particularly in Alzheimer's disease (AD) and Multiple Sclerosis (MS). Monocytes may traffic *C. pneumoniae* across the blood-brain-barrier, shed the organism in the CNS and induce neuroinflammation. The demonstration of *C. pneumoniae* by histopathological, molecular and culture techniques in the late-onset AD dementia has suggested a relationship between CNS infection with *C. pneumoniae* and the AD neuropathogenesis. In particular subsets of MS patients, *C. pneumoniae* could induce a chronic persistent brain infection acting as a cofactor in the development of the disease. The role of Chlamydia in the pathogenesis of mental or neurobehavioral disorders including schizophrenia and autism is uncertain and fragmentary and will require further confirmation.

1. Introduction and Background

Chlamydiae were taxonomically categorised into their own order *Chlamydiales*, with one family, *Chlamydiaceae*, and a single genus, *Chlamydia* which included four species: *C. trachomatis*, *C. pneumoniae*, *C. psittaci*, and *C. pecorum*. Two of the species, *C. trachomatis* and *C. pneumoniae*, are common pathogens in humans, but the routes of transmission, susceptible populations, and clinical presentations differ markedly. The other species occur mainly in animals although *C. psittaci* may be also implicated in human respiratory diseases. In 1999, a new taxonomic classification was proposed, renaming *Chlamydia pneumoniae* as *Chlamydomphila pneumoniae* [1]. However, the proposal to change the

taxonomic nomenclature for the *Chlamydiae* family has not been universally accepted and both names are currently in use by different authors.

C. pneumoniae, a common cause of human respiratory disease, was first isolated from the conjunctiva of a child in Taiwan in 1965 but it was not until the early 1980s that it was scientifically identified as a distinct *Chlamydia* species and was established as a major respiratory pathogen in 1983 when it was isolated from the throat of a college student at the University of Washington. Most likely, *C. pneumoniae* is primarily transmitted from human to human by the respiratory tract without any animal reservoir [2, 3] and infection spreads slowly. The incubation period is several weeks, which is longer than that for many other

respiratory pathogens [4]. The time span of infection spread in families is shorter, however, ranging from 5 to 18 days [5]. As other *Chlamydia* species, *C. pneumoniae* has a unique biphasic life cycle with two forms that are functionally and morphologically distinct that undergo an orderly alternation: the elementary body (EB), infectious and metabolically inactive responsible for attaching to the target host cell and promoting its entry and the reticulate body (RB), an abortive non-infectious and metabolically active intracellular form which replicates by binary fission and reorganizes into EBs then released by cell lysis. In general, it is likely that this aberrant developmental step leads to the persistence of viable but non cultivable *Chlamydiae* within infected cells over long periods. In cell culture conditions, the duration of the developmental cycle is between 2 and 3 days, depending on the strain, when bacteria have differentiated back to EBs and are released in the extracellular medium. In natural infections, the situation is more complicated, and the normal development of *Chlamydia* is easily disturbed. Living separated from the host cell cytoplasm within its Chlamydial inclusion (a nonlysosomal vacuole), *C. pneumoniae* is able to create an intracellular niche from where it promotes host cell survival or death, modulates regulatory host cell signalling pathways, and bypasses the host cell's defence mechanisms. Thus, *C. pneumoniae* can induce a persistent infection due to the inability of the host to completely eliminate the pathogen [6–8]. The failure by the guest to eradicate the disease involves the establishment of a state of chronic infection in which *C. pneumoniae* after internalization into mononuclear cells, enters into a state of quiescence with intermittent periods of replication and characterized by antigenic variation, production of Heat Shock Proteins (HsPs) and proinflammatory cytokines capable of evading host defences and trigger tissue damage [8]. Chronic infection and clinical persistence are closely related. In chronic infections a different pathway is taken. Under pressure from host defences the metabolic processes of the organism are diminished. *C. pneumoniae* in this state is called the Cryptic Body (CB). This chronic unresolved infection, which can last for several decades, can also initiate the malign process of autoimmunity. To a large extent, the form of the disease may depend on the host's genetic inheritance. This is why many of the chronic disease forms caused by infections with *C. pneumoniae* tend to have inherited characteristics. It is thought that the host immunity together with individual traditional risk factors, serological markers of *C. pneumoniae* infection and genetic susceptibility, may play an important role in controlling Chlamydial infections. A chronic *C. pneumoniae* infection increases the expression of its own 60 kDa heat shock proteins (HsP60), especially when they are persistently elevated. The host immune response to microbial HsP60 may gradually lead or, contribute to autoimmunity to human HsP60 and, consequently, to the development of some chronic diseases such as asthma [9] atherosclerosis or clinical manifestations of Coronary Heart Disease (CHD) [4, 9–14]. Definitive pathogenesis and virulence investigations for the *Chlamydiae* have been hampered because methods for gene transfer have not yet been developed for these microorganisms. Despite this

critical experimental limitation, a great deal of information has been generated on intracellular Chlamydial growth and development, and the effects of chlamydial infection on host-cell physiology [15]. Like most intracellular pathogens, *C. pneumoniae* interferes with the normal apoptotic signalling pathways of these cells, perhaps contributing to long-term persistence and chronic inflammation in central nervous system (CNS) system. However, it is not clear how this happens: under some circumstances these microorganisms induce apoptosis and/or necrosis, but under other circumstances they inhibit apoptosis [15, 16]. The circumstances that dictate whether the *Chlamydiae* inhibit or activate host-cell death reflect several important pathogenic considerations, including whether an acute or chronic infection is in progress and whether intracellular chlamydial growth is programmed to go through a productive infectious cycle or is stalled under non-productive growth conditions [16]. The intracellular growth cycle of the *Chlamydiae* is complex and several growth options are possible, depending on the host-cell type, the particular environmental conditions in the host cell and the nature of the tissue that is being affected. It is possible that apoptotic activity is controlled to some extent by the intracellular growth status of the *Chlamydiae*, which can be influenced by any or all of these considerations [16].

Data on the distribution of seroprevalence reveal that the prevalence of *C. pneumoniae* infection increases with age. Antibodies against *C. pneumoniae* begin to appear at school age but are rare in children under the age of 5, except in developing and tropical countries. Antibody prevalence increases rapidly at ages 5 to 14, reaches 50% at the age of 20, and continues to increase slowly to 70% to 80% at ages 60 to 70 [4]. This seems to suggest that most people are infected and re-infected for life. *C. pneumoniae* accounts for 6%–20% of community acquired pneumoniae (CAP) in adults [3, 4], but participates in co-infection involving other bacterial agents in approximately 30% of adult cases of CAP [4]. Some studies have suggested a possible association of *C. pneumoniae* infection and acute exacerbations of asthma and chronic obstructive pulmonary diseases (COPD). In recent years, however, in addition to respiratory diseases, an increasing number of publications have been reported of detection of *C. pneumoniae* in chronic extra-respiratory diseases. In fact, the recent deepening of knowledge on the biology of *Chlamydia* and the use of increasingly more sensitive and specific molecular techniques, have allowed to demonstrate the presence of *C. pneumoniae* DNA in a large number of persons suffering from different diseases other than cardiovascular (atherosclerosis and stroke), such as osteoarthritis and CNS disorders. In this setting, the ability of *C. pneumoniae* to infect various human cells such as epithelial, endothelial, and smooth muscle cells as well as macrophages, monocytes, and lymphocytes, suggests a systemic dissemination following exposure to a respiratory infection. Certainly, the presence of *C. pneumoniae* DNA in peripheral blood mononuclear cells (PBMCs) strongly suggests that such dissemination can occur in a number of different tissues [17]. *C. pneumoniae* infection has been shown to promote the transmigration of monocytes through human brain endothelial cells, suggesting a mechanism by

which the organism may enter the CNS. This may account for the delivery of the organism to the CNS and result in chronic injury [18].

This review addresses the potential and the underlying mechanisms by which *C. pneumoniae* infections can play a role in different neurological diseases, examining the epidemiological and methodological findings among studies which will be highlighted. Suggestions for future studies and potential standardization of tools and protocols are proposed.

2. *C. pneumoniae* Infection, Atherosclerosis, and Cerebrovascular Diseases

2.1. *C. pneumoniae* and Atherosclerosis. If large is the amount of data that support the role of *C. pneumoniae* in the atherosclerosis, equally important is the weight that *C. pneumoniae* has recently taken in the cerebrovascular diseases. The mechanism of atherosclerotic disease and thrombosis is not completely known. The first report of an association between *C. pneumoniae* infection and atherosclerosis was by Saikku et al. [2]. Individuals with immunoglobulin G antibody specific for *C. pneumoniae* were shown to be at an increased risk for myocardial infarction and CHD, as there was a statistically significant difference in the frequency of antibody in patients versus controls. Subsequently, many other reports concerning the seroepidemiological association between specific antibodies to *C. pneumoniae* and atherosclerosis at several arterial sites using retrospective, cross-sectional and prospective studies, have confirmed a link between *C. pneumoniae* and atherosclerosis [19]. Overall, these seroepidemiological studies have shown some limitations because different laboratories have measured different antibody classes, used different criteria for determining seropositivity or measured antibodies that were cross-reactive with other chlamydial species. Although most of the studies have used the micro-immunofluorescence test (MIF) for measuring antibodies, inter-laboratory variation in interpreting the results remains a problem [20, 21]. However, a number of experimental data support the involvement of this pathogen in the pathogenesis of atheroma [11–13]. First, *C. pneumoniae* gains access the vasculature during local inflammation in lower respiratory tract infection. Second, the infected alveolar macrophages transmigrate through the mucosal barrier and give the pathogen access to the lymphatic system, systemic circulation, and atheromas. Third, *C. pneumoniae* can infect a variety of cells commonly found in atheromas, including coronary artery endothelial cells, macrophages, and aortic smooth muscle cells. Fourth, *C. pneumoniae* may influence atheroma biology by modulating macrophage-lipoprotein interactions. In this setting, chlamydial lipopolysaccharide (LPS), a potent endotoxin constituent of outer membrane of Gram negative bacteria induces the release of cytokines promoting leucocyte adherence, leucocyte migration, and intimal inflammation. LPS has also been demonstrated to have a crucial role in lipid metabolism [22] and it is involved in mediating the process of “catching” LDL cholesterol by

macrophages infected with *C. pneumoniae*, which is transformed into “foam cells,” the “key cells” of newly formed atherosclerotic lesions [23]. Moreover, in vitro studies have shown that upregulated molecules by *C. pneumoniae*, including α -2 and α -1-integrins, adhesion molecules (ICAM-1, VCAM-1), platelet derived growth factor (PDGF), tissue factor (TF), early growth response factor (EGR-1), appear to contribute to these events. Some of these, along with matrix metalloproteinases (MMPs) contribute to the destabilization of atheromas plaque, and the formation of thrombus [12] and consecutively may lead to the arterial thromboembolic complications. The link between *C. pneumoniae* and coronary atherosclerosis is also substantiated by studies in which antibodies against members of the HsPs family were detected in individuals with persistent *C. pneumoniae* infections [9–11, 24, 25]. Several findings indicate in fact that HsPs, which are evolutionary very conservative and are present in both microbial and host cells, seem to be important in the development of autoimmunity. *C. pneumoniae* HsPs are expressed in abundance in atherosclerotic lesions and a persistent *Chlamydia* infection is accompanied by an increased production of Hsp60. In particular, this protein, may represent a particular antigenic stimulus capable of eliciting strong humoral and cell-mediated immune responses with immunopathological sequelae of chronic chlamydial infections [9, 10, 12, 13, 26]. In this regard, *C. pneumoniae* Hsp60 has been demonstrated in patients with acute hemorrhagic evolution of the carotid plaque suggesting that *C. pneumoniae* might participate in the atherogenesis and to induce atherosclerosis complications by inflammatory pathways (activation of cytokines, endothelial factors and MMPs) [9]. At present, very little is known about genes predisposing the host to the persistent *C. pneumoniae* infection and its sequelae. HLA haplotype markers have been recently investigated in patients with coronary artery disease (CAD). In a recent study, a multiple logistic regression analysis showed HLA-B*35 allele as the strongest risk gene for the combined serological markers of *C. pneumoniae*. Male sex and smoking further strengthened the association between HLA-B35 and markers of *C. pneumoniae* infection. Interestingly, HLA-B*35 was also found to be associated with cardiovascular risk in Finnish patients [14].

2.2. *C. pneumoniae* and Stroke. The role of *C. pneumoniae* in the pathogenesis of ischemic stroke is still debated. Infection with *C. pneumoniae* may contribute to the risk of stroke by enhancing CAD, as addressed in several issues. Several published studies including the pilot case control “Northern Manhattan Stroke Study,” focused on the correlation between the *C. pneumoniae* infection, represented by the elevated serum levels of specific anti-*C. pneumoniae* (IgA, IgG, and IgM) antibodies at MIF test and stroke occurrence [26–31], in different race-ethnic groups and after adjusting for conventional risk factors. The MIF method was used in most of the previously published papers, as considered as a reference standard in the *C. pneumoniae* serology. The elevated titers of anti-*C. pneumoniae* IgA and IgG were more prevalent in subjects with acute ischemic stroke than in controls [26, 29–32]. With immunohistochemical

staining, other investigators demonstrated the presence of *C. pneumoniae* in more than 70% of the endarterectomy specimens from stroke patients with severe carotid stenosis [33]. Other authors assessed anti-*Chlamydia* IgA and IgG seropositivity using an ELISA test and found an increased risk of stroke in young patients seropositive to *C. pneumoniae* in the IgA antibody class rather than in IgG class [34]. This has suggested the possibility that IgA antibodies, which last only 3 to 5 days in the circulation, are a diagnostic marker of persistent, chronic infection (single IgA MIF titer $\geq 1:16$), whereas IgG antibodies, which are produced for 3 to 5 years, are a marker of older, inactive infection [14]. The association between IgA titers and risk of ischemic stroke was also stratified according to ischemic stroke subtype. In contrast with the less consistent evidence of an association of IgA titers and cardioembolic and cryptogenic stroke, there was a trend toward an association of IgA titers with large vessel atherosclerotic and lacunar stroke, strongly supporting evidence that *C. pneumoniae* contributes to atherosclerosis [30]. However, according to a previous consensus statement [35] and other authors [36, 37], there is not yet agreement that IgA titers are indicative of chronic, persistent infection. First, the half-life of specific *C. pneumoniae* is only a few days. Second, measurement of IgA antibodies may be complicated by cross-reacting *C. pneumoniae* IgG and rheumatoid factor or with antibodies to other chlamydial species and potentially other microorganisms. Third, MIF, the serologic “golden standard” which may have been overestimated in the past, is not standardized and there are interlaboratory variations in the performance of this test [38]. Finally, the hypothesis that *C. pneumoniae* infection, as indicated by elevated IgA and IgG antibody titers, may be not associated with an increased risk of ischemic stroke. This, however, may differ according to subtype of ischemic stroke, cut-off value of antibody titers, and gender. Anti-*C. pneumoniae* antibodies were also evaluated in HIV infected individuals with CAD. One study showed that both the IgA and IgG levels did not differ significantly and no subjects were positive for IgM, suggesting that the damage to the carotid wall in HIV-1 patients was not due to *C. pneumoniae* [39]. Other studies by contrast, found that *C. pneumoniae* represents a further risk factor for cardiovascular disease in HIV-positive patients with both low CD4 cell count and high HIV load [40] and that the coexistence of hypertriglyceridemia and *C. pneumoniae* infection significantly increases the risk of atherosclerosis up to three times [41].

A number of studies addressed the question of whether molecular tools and in particular PCR can be used to detect chlamydial DNA in cerebrovascular atherosclerotic lesions. The results provided from a consistent number of atherosclerotic lesion and blood specimens from more than 1500 patients analysed by different in-house PCRs, were found extremely variable. Being PCR not standardized, this technique has a tendency to produce false positive results. In this setting, there is the need for standardization of PCR methods and for assessing their sensitivity, specificity, and predictive values. In general, there is not consensus on how nested PCR (n-PCR) technology can be controlled [42]. PCRs have been shown to be predictive of CAD, when used

for detection of *C. pneumoniae* DNA in PBMC, suggesting that the detection of bacterial DNA in PBMC may be a valid surrogate marker to identify individual risk for endovascular chlamydial infection [43]. Recently, *C. pneumoniae* DNA and chlamydial LPS were measured during 12 months in 97 patients with acute CAD. *C. pneumoniae* DNA was detected in PBMC from 8 (8.2%) patients at the initial hospitalization during acute CAD and in 9 (10.6%) patients at 3 months. *C. pneumoniae* DNA declined in stable state and in the recovery period. These findings suggest a role of the bacterium in the acute phase of CAD [44]. Attempts to eradicate *C. pneumoniae* in patients with cardiovascular diseases including the “Azytromycin and Coronary Event Study” trial, have all failed [45–47]. As shown in previous studies the persistent state is completely refractory to antibiotic treatment [45] and first-choice antichlamydial drugs may even induce chlamydial persistence under certain conditions [46]. In the absence of a functional treatment strategy, the hypothesis of a chlamydial contribution to atherogenetic processes can thus neither be proved nor disproved by eradication studies, and a better understanding of chlamydial pathobiology in order to target specific chlamydial antigens is needed before implementing clinical studies with new effective antibiotic regimens.

In summary a causative role of *C. pneumoniae* infection in cardiovascular disease has not yet been firmly established. Despite the considerable laboratory and clinical research that has been done on the role of *C. pneumoniae* in the progression of atherosclerosis, several important questions remain unanswered. Most importantly, it is not known whether the *C. pneumoniae* bacterium is an innocent passenger aboard atheromas or whether it is actively involved in the initiation or progression of atherosclerotic disease. To answer this question, well-planned studies are needed to further characterize the molecular mechanisms that link *C. pneumoniae* to vascular disease. In particular, HsP60 needs to be explored further as a potential culprit and therapeutic target. The early results from antichlamydial intervention studies in humans are partially consistent with a causative role of *C. pneumoniae* in the disease process.

3. *C. pneumoniae*, Neurodegenerative, and Demyelinating Disorders

The ability of the *C. pneumoniae* to persist in monocytes and macrophages in tissues for long periods, to circumvent the mechanisms of bactericidal and oxidative stress, to activate the endothelial cells with production of adhesion molecules and cytokine overproduction, has suggested that it may participate in the development or progression of certain acute and chronic inflammatory diseases of the CNS. In recent years, in fact, a growing body of evidence concerning the involvement of *C. pneumoniae* in neurological diseases has been gradually increasing. This was supported in particular by the detection of genomic material of the microorganism into the cerebrospinal fluid (CSF) of patients with multiple sclerosis (MS), Alzheimer’s disease (AD), meningoencephalitis and neurobehavioral disorders [4, 6–8, 18].

3.1. Alzheimer's Disease. Alzheimer's disease (AD) is one of the most severe dementing illnesses that increases with the increasing age of the population. The disease is associated with atrophy/death of neurons in particular regions of the brain and occurs in two general forms: an early-onset form that is primarily genetically determined, and a far more common late-onset AD (LOAD), which is a non-familial, progressive neurodegenerative disease that is now the most common and severe form of dementia in the elderly. The defining neuropathology of both familial and sporadic AD includes the neuritic senile plaque (NSPs), consisting primarily of amyloid beta ($A\beta$) protein, and neurofibrillary tangles (NFTs), the major component being modified *tau* protein, that affect nerve synapses and nerve-nerve cell communication. Genetic, biochemical, and immunological analyses have in part provided a relatively detailed knowledge of these entities [48, 55]. The disease usually manifests initially as a gradual loss of short-term memory and later progresses to major cognitive dysfunction. The latter can take the form of various behavioural disorders, loss of orientation, difficulties with language, and impairment of daily living [56]. Estimates of crude incidence of AD range from 7.03 to 23.8 per 1000 person years [57, 58]. The range is likely attributable to different study populations and case investigating methods. The incidence of AD increases with age for both genders, but there is definite indication if there are gender differences. AD seems to be more common among women with approximately a third higher incidence and prevalence among females compared to males [59–61]. Using age-specific incidence rates one projection study in the U.S. predicted that incident cases would increase from the 360,000 cases in 1997 to 1.14 million in 2047 [61]. Despite the AD's discovery by Alois Alzheimer in 1907, the cause of this pathology and neurodegeneration is not unknown. Infections of the CNS have been shown to stimulate inflammatory responses that may result in neurodegeneration [62]. Several groups have investigated an association between various infectious agents and AD, but none of these has been accepted as either etiological for disease development, or worsening of neuropathology. Interesting perspectives came from one study which identified herpes simplex virus type 1 (HSV-1) infection as a risk factor for development of AD in subjects expressing APOE $\epsilon 4$ allele [63, 64]. Virus including measles virus, adenovirus, lentiviruses, and several other others were initially considered but discarded after [65, 66]. Bacterial pathogens including *C. trachomatis*, *Coxiella burnetii*, *Mycoplasma* spp and spirochetes [67, 68] have also been investigated and dismissed in relation to involvement in AD neuropathogenesis. Prions have been also taken into consideration but then excluded [69]. The first paper that reported an association between *C. pneumoniae* infection and LOAD was from Balin et al. [48]. He found that 90% of AD brains were positive for *C. pneumoniae* as assessed by highly sensitive and specific PCR and the organism was detected in various sections of brain (hippocampus, cerebellum, temporal cortex, and prefrontal cortex) that exhibited AD pathology of more or less variable intensity [55]. Electron microscopic (EM) results revealed *C. pneumoniae* like particles containing EBs and RBs in the brain

tissue and immunohistochemistry analysis indicated strong labelling in the sections of the brain most affected by AD among the cases, while no labelling in the controls. Moreover, a part the detection and visualization of *C. pneumoniae* within the cells of CNS that were associated with plaques and tangles, RNA transcripts of *C. pneumoniae* indicating metabolically active organisms were demonstrated by RT-PCR in frozen tissue samples and organisms were then isolated from the tissue and propagated in cell cultures. In that report, a strong association of APOE- $\epsilon 4$ genotype and *C. pneumoniae* infection was found in 58% (11/19) patients with AD suggesting, as shown in reactive arthritis, that the APOE- $\epsilon 4$ gene may promote some aspects of *C. pneumoniae* pathobiology in AD [48]. The Balin study report received a great deal of public and scientific attention and attempts to replicate the finding were conducted in mother reputable laboratories throughout the world. Two independent investigators (Ossewaarde et al., 2000 and Mahony et al., 2000, unpublished data) found *C. pneumoniae* in brains of AD patients with PCR and immunohistochemistry, validating Balin's results. However, conflicting results were found in subsequent studies by different authors using the same procedures but different protocols in paraffin-embedded brain tissues [49–53] attempting to revalidate the previous findings (Table 1). These studies have yielded contradictory results likely due to differences in diagnostic criteria and diagnostic tools. Demographic differences between the patient groups, such as geographic location, season of death and institutional history might also account for these opposing results. AD patients included in the Balin study, might have been recently exposed to *C. pneumoniae*, perhaps in an institutional setting, and therefore would have been at high risk of systemic spread from the respiratory tract to sites within the CNS, where advanced AD pathology already existed [70]. In an extension of these findings, Gerard, two years later, demonstrated *C. pneumoniae* in 80% of AD samples and 11.1% of the controls using primer targeting two *Chlamydia* (1046, 0695) genes. The AD cases (mean age 79.3 years) and controls (65.9 years) were age- and sex-matched as closely as possible, but the controls were younger and 22.7% were males [54]. The organism was again viable within the AD brain as assessed by culture of the organism from brain samples; moreover, RT-PCR analyses identified primary rRNA gene transcripts from *C. pneumoniae* indicating metabolic activity of the organism in those tissues. Interestingly, immunohistochemical analyses have also shown that astrocytes, microglia, and neurons all served as host cells for *C. pneumoniae* in the AD brain, and that infected cells were found in close proximity to both NSPs and NFTs in the AD brain (Figure 1). Recently, cultured astrocytes and microglia have also shown that *C. pneumoniae* displays an active, not a persistent, growth phenotype indicating probable concomitant destruction by lysis of some portion of host cells at the termination of that cycle [71]. In the years immediately after the study of Balin, some experimental discoveries have provided insights about the pathogenetic mechanisms of AD. First, a relationship exists between possession of the APOE- $\epsilon 4$ allele and the pathobiology of *C. pneumoniae* [72] and that

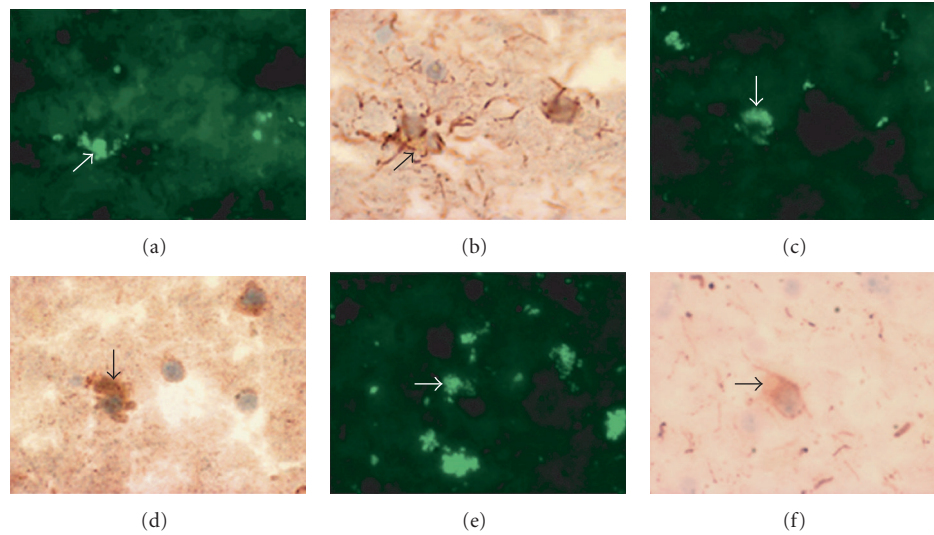


FIGURE 1: Representative images of double immunolabelling studies to demonstrate the infection of astrocytes, microglia, and neurons with *Chlamydia pneumoniae* in the AD brain. *Chlamydia pneumoniae*-infected cells were identified in all cases using the FITC-labelled monoclonal antibody targeting the *Chlamydia* LPS (Pathfinder TM; a, c, e). Astrocytes (b) and microglia (d) were identified by immunostaining using monoclonal antibodies targeting GFAP and iNOS, respectively. Neurons were identified by immunostaining with a monoclonal antibody targeting neuron-specific microtubule-associated protein (f). Images in all panels were obtained using a objective. In all panels, arrows indicate cells labelling with the Pathfinder TM and surface marker-specific monoclonal antibodies. (Reproduced from [54] with courtesy of Department of Immunology and Microbiology, Wayne State University School of Medicine, Detroit, MI, USA, and with permission of *FEMS Med Lett*).

TABLE 1: Studies demonstrating evidence or absence of *C. pneumoniae* in brain autoptic specimens from patients with Alzheimer disease.

| Author, year | No. of autoptic AD brain specimens examined | <i>C. pneumoniae</i> DNA detection and confirmation methods | Results of ^PCR in AD brain Positive % | Results of PCR in control brain Positive % | Study of APOEε4 gene expression |
|---------------------------|---|---|--|--|---------------------------------|
| Balin et al. 1998 [48] | 19 | §PCR, ¶RT-PCR, ^^IHM, *EM, Culture | 1790 | 15,2 | Yes |
| Nochlin et al. 1999 [49] | 13 | †PCR, IHM | 0 - | 0 - | Not done |
| Gieffers et al. 2000 [50] | 20 | °PCR, IHM | 0 - | 0 - | Not done |
| Ring and Lyons 2000 [51] | 15 | °°PCR, Culture | 0 - | 0 - | Not done |
| Taylor et al. 2002 [52] | 10 | *PCR, IHM | 0 - | 0 - | Not done |
| Wozniak et al. 2003 [53] | 4 | °°°PCR | 0 - | 0 - | Not done |
| Gérard et al. 2006 [54] | 27 | ‡PCR, RT-PCR, IHM, EM, Culture | 2580 | 311 | Yes |

^PCR, polymerase chain reaction; ¶RT-PCR, Reverse transcriptase PCR; ^^IHM, immunohistochemistry; *EM, electron microscopy; §primers targeting the 16S ribosomal RNA (rRNA) gene (Gaydos et al., *J Clin Microbiol* 1992; 30: 796–800) and the chlamydial major outer membrane protein (MOMP) gene (*ompA*) (Perez-Melgosa et al., *Infect Imm* 1991; 59: 2195–9); †PCR with seminested primer amplifying *C. pneumoniae*-specific DNA sequences of 437 bp (Kuo et al., *J Infect Dis* 1993; 167: 841–9); ° nested PCR (Maass et al., *Atherosclerosis* 1998; 140: S25–30); °° Primers targeting the MOMP gene (Perez-Melgosa et al., 1991; *Infect Imm* 1991; 59: 2195–9) *PCR using the 76 kDa protein gene [47], the *rpoB* gene encoding the RNA polymerase beta subunit (Ouchi et al., *J Med Microbiol* 1998; 47: 907–13), and the pan-*Chlamydia* primers targeting the 16S ribosomal RNA gene in *Chlamydia* and *Chlamydia*-like organisms (Ossewaarde et al., *Microbiology* 1999; 145: 411–17); °°°PCR with primer targeting the 16SrRNA gene (*J Clin Microbiol* 1992; 30: 796–800); ‡PCR-multiple assays targeting the Cpn1046 gene (aromatic amino acid hydroxylase) and Cpn0695 (MOMP)[54].

the *C. pneumoniae* load in the AD brain varies with APOE genotype [73]. Second, infection of human microvascular endothelial cells co-cultured with *C. pneumoniae* elicits increased expression of proteins relevant to access for the organism to CNS including N-cadherin and b-catenin [74].

Third, the expression of occludin, a protein associated with tight junctions, is attenuated in the *C. pneumoniae*-infected cells. Fourth, infection with *C. pneumoniae* through the olfactory pathways of nontransgenic young female BALB/c mice which usually do not develop AD, has shown to

promote the production of extracellular amyloidlike plaques ($A\beta$ 1-42) in the mouse's brain thus providing a unique model for the study of sporadic AD and suggesting that this model could be a primary trigger for AD pathology following a noninvasive route inoculation [75]. Since *C. pneumoniae* is harboured in the respiratory tract and has a predilection for infecting epithelial cells, the olfactory neuroepithelia in the nasal passages are a likely target for infection. Following entry into these epithelia, potential damage and/or cell death may occur in the main olfactory bulb and olfactory cortex, thereby setting the stage for further retrograde neuronal damage [55].

Because chlamydial chronic infections are characterized by the inaccessibility of the "chlamydial persistent state" to conventional antichlamydial agents, there are few clinical trials that have determined the effectiveness of antibiotic therapy against *C. pneumoniae* in AD. A first randomized, placebo controlled, multicentre clinical trial performed to determine whether a 3-month course of doxycycline and rifampin could reduce the decline of cognitive function in patients with AD showed significantly less cognitive decline at 6 months and less dysfunctional behaviour at 3 months, in the antibiotic group than in controls [76]. Although these observations do not demonstrate a causal relationship between CNS infection with *C. pneumoniae* in terms of eradication of chronic *C. pneumoniae* infection and the AD neuropathogenesis, they do open the way to further investigations. In this regard, animal modelling will be required to define in detail how chlamydial infection might result in AD-related pathological change in the CNS and to provide a better understanding of infection parameters. In vitro and mouse model studies have demonstrated that metal protein attenuating compounds (MPACs) promote the solubilisation and clearance of extracellular senile plaques comprised of beta-amyloid. The role of the antiprotozoal metal chelator clioquinol in AD, which has been reported to reduce beta-amyloid plaques, presumably by chelation associated with copper and zinc, is currently in clinical trials as potential for treatment of AD [77, 78]. The scientific knowledge surrounding Alzheimer's disease and infection by *C. pneumoniae* is still growing. Standardization of diagnostic techniques would certainly allow for better comparability of studies. However, other systemic infections as potential contributors to the pathogenesis of AD should be considered.

3.2. *C. pneumoniae* and AIDS Dementia. Many authors have explored the possibility that *C. pneumoniae* was involved in other neurodegenerative disorders other than Alzheimer's disease. The existing data are however few and not significant. Among those characterized by dementia, one study performed by our group explored the possible link between AIDS-dementia complex (ADC) and *C. pneumoniae* [79]. ADC is an HIV-derived neuropathological disorder characterised by infection of macrophages and microglia cells and release of proinflammatory cytokines into the parenchyma [80]. In this report, *C. pneumoniae* was identified in the CNS by PCR for *C. pneumoniae* MOMP and 16S rRNA gene in 4 (17.4%) out of 23 HIV-infected patients diagnosed as ADC

Stage 3 according to scheme for AIDS dementia complex and confirmed by autopsy. Sequence analysis revealed significant homologies with *C. pneumoniae* compared to *C. trachomatis* and *C. psittaci*. Moreover, high mean levels of CSF specific anti-*C. pneumoniae* antibodies and *C. pneumoniae* antibody specific index values significantly elevated were also found by ELISA in these patients. These findings suggest that although the low rate of isolation is not representative of the frequency with which *C. pneumoniae* is involved in the causation of CNS injury, in the late-stage HIV infection, an increase in "trafficking" of monocytes containing *C. pneumoniae* to the brain may carry this organism in the sites which are the major reservoirs of productive HIV replication and contribute to neuronal damage in HIV-infected patients [79]. Moreover, the possibility that may exist a patient's subgroup in whom this organism is not, as for atherosclerosis and other Chlamydial diseases, an "innocent bystander," but may survive and replicate in CNS macrophages cannot be excluded [8].

3.3. *C. pneumoniae* and Multiple Sclerosis. MS disease is a presumed autoimmune chronic inflammatory disease of the CNS of unknown aetiology triggered by an environmental factor in susceptible individuals. It generally affects 1 to 1.8 per 1,000 individuals and kills more than 3000 people each year, with a further estimated annual morbidity cost of over \$ 2.5 billion. In the United States, the prevalence of the diseases is 250,000 to 350,000 cases annually [97]. The pathological hallmark of multiple sclerosis (MS) is the demyelinating plaque that represents an area of demyelination and gliosis around blood vessels [98]. Acute lesions show perivascular lymphocytes and plasma cells along with the infiltration of macrophages and phagocytosis of myelin membranes. The continuous breakdown and regeneration of myelin has been demonstrated within the progressive MS plaque [99]. Toll-like receptors (TLR) are intimately involved in several neurodegenerative and demyelinating disorders including MS as demonstrated with the finding of a marked increase in TLR expression in MS lesions. PCR studies have shown that microglial cells from MS patients express TLRs 1-8 [100]. Moreover, while healthy white matter from MS patients does not contain TLRs, active lesions are associated with high expression of TLR3 and TLR4 on microglia and astrocytes. In contrast, late active lesions also contain astrocytes bearing surface TLR3 and TLR4 [100]. This suggests that early lesions are characterized by microglia infiltration, while astrocytes are also active in later MS lesions. However, the precise role of TLR3 and TLR4 activation in these lesions is yet unknown. TLRs have been shown to recognize highly conserved regions in various microorganisms (Pathogen-Associated Molecular Patterns) including *C. pneumoniae* and thus stimulate a potent inflammatory response contributing to the clearance of the pathogen [101]. Unpublished our findings have detected the major expression of mRNA TLR-2 and TLR-4 in peripheral blood but not in CSF from SM patients with RR forms, indicating that their combined activity might be crucial to modulate and activate the cellular-mediated immune response during chronic infections by *C. pneumoniae* [102]. Based on epidemiological observations, it

has been proposed that exposure to an environmental factor, such as an infectious agent, in combination with genetic predisposition could be implicated in MS pathogenesis [103]. The risk of MS is enhanced by the presence of specific genes on chromosome 6 in the area of MHC, Human Leukocyte Antigens (HLA) in humans. In particular, HLA-DR and HLA-DQ genes, which are involved in antigen presentation, are strongly associated to the development of the disease. However, although the risk of the disease is higher in monozygotic than in dizygotic twins (about 30% and 5%, resp.), the low concordance rate obtained in identical twins suggests that non-genetic factors can contribute to MS aetiology. In this setting, the aetiopathogenesis of MS disease is complex and still debated. So far, about 20 microorganisms including viruses have been associated with this disease [104]. The screening techniques in these studies varied from serology to PCR and quality and numbers of controls examined varied widely. The latest pathogen to be associated with MS is *C. pneumoniae* [105–110]. Sriram et al. reported the first evidence suggesting the potential role for *C. pneumoniae* as a candidate in MS pathogenesis [106]. One year later, a larger study from the same group strongly confirmed that CSF demonstration of *C. pneumoniae* was more frequent in MS patients than in control patients with other neurological disorders (OND) [107]. In particular, *C. pneumoniae* culture isolation was obtained in 24/37 (65%) MS and in 3/27 (11%) OND patients, CSF single polymerase chain reaction (PCR) for major outer-membrane protein (MOMP) was positive in 36/37 (97%) MS and in 5/27 (18%) OND patients, whereas CSF anti-*C. pneumoniae* IgG were detected by enzyme linked immunoadsorbent assay (ELISA) in 32/37 (86%) MS and in 0/27 (0%) OND patients. After this innovative publication, a number of studies have suggested that *C. pneumoniae* infection may be associated with MS, while other studies have found no association [108, 109]. During recent years, there have been many evidences of a possible role of *C. pneumoniae* involvement in MS disease supported in part by seroepidemiological, cultural, molecular, immunological and therapeutic studies. However, it is also true that there are not many studies that argue for a role of organism in MS. First, while some reports have documented that *C. pneumoniae* seropositivity was related to the risk of MS progressive forms (SP and PP), but only moderately linked to the risk of developing MS [110], others have not found association between serum titers of anti-*C. pneumoniae* antibodies and the risk for MS or, by contrast, a higher risk to develop MS in a subgroup of older patients after than before disease onset [111]. Second, the organism was found in course of MS relapses in the throat together with a rising serology [112]. Third, relapses of MS have long been noted to follow respiratory infections, including sore throat, or pneumoniae with a clinical pattern typical of respiratory infection caused by *C. pneumoniae*. The isolation of the pathogen, as assessed by culture assay in CSF and brain tissue failed repeatedly in MS patients [113–115] or was positive only in a small proportion of MS patients [81, 116]. Dong-Si et al. have noted gene transcription of messenger RNA by *C. pneumoniae* in CSF from MS patients suggesting active infection by this pathogen [91].

Recently, active transcription of DNA from the organism has been found in a persistent and metabolically active state in cultured CSF and PBMCs from MS patients, but not in controls [95]. Other investigators were able to culture and detect *C. pneumoniae* in buffy coat samples from a healthy blood donor population [117] demonstrating a *Chlamydia* carriage rate of 24.6%, within the WBC of the peripheral circulation. Because of the difficulties of isolating *C. pneumoniae* cultures, nucleic acid amplification methods such as PCR-based assays have become the method of choice for detection of this microorganism. However, PCR procedures often differ in several aspects which can affect sensitivity, reproducibility, and specificity when applied to direct testing of clinical specimens [86, 118, 119]. In this context, collaborative studies involving different laboratories in which the presence of *C. pneumoniae* was evaluated in blinded CSF samples, further underlined the lack of an accepted standardized PCR protocol [120, 121]. A number of PCR studies did not provide evidence of detection of *C. pneumoniae* DNA in CSF of MS patients. Most of these studies were performed using single or nested (n) PCR targeting either MOMP or 16S ribosomal (rRNA) chlamydial genes [11, 34, 114, 116, 122–124]. By contrast, a substantial body of work from around the world has provided clear evidence of the involvement of *C. pneumoniae* in MS. In this setting, a consistent number of studies did find PCR positive results with DNA or mRNA positive rates varying from 2.9% to 69% [81–91]; [94–96]; [117–126] as listed in Table 2. Some reports also demonstrated the more frequency of *C. pneumoniae* DNA in CSF of MS patients with Gd enhancing lesions on MRI scans [87, 89]. Moreover, CSF detection of heat-shock protein-60 messenger RNA (Hsp-60 mRNA) and 16S rRNA by Reverse-Transcriptase PCR (RT-PCR) was more frequent in MS patients than in controls signifying the presence of a high rate of gene transcription and, therefore, more active metabolism of *C. pneumoniae* in MS [91]. In 2004, our group developed a novel amplification program for MOMP gene by employing a “touchdown” technique and analyzing CSF samples from patients with MS, other inflammatory neurological disorders (OIND) and noninflammatory neurological disorders (NIND) and employed three gene targets (MOMP, 16S rRNA and HsP-70) in parallel to achieve a major sensitivity and specificity [92]. A PCR positivity for MOMP and 16S rRNA in CSF was present in a small proportion of MS (37%), OIND (28%) and NIND (37%) patients, without any differences between MS and controls. Furthermore, a PCR positivity for MOMP and 16S rRNA in CSF was more frequent in relapsing-remitting (RR) MS than in MS progressive forms (SP and in PP MS) as well as in clinically and magnetic resonance imaging (MRI) active than in clinically and MRI stable MS, whereas a CSF PCR positivity for HsP-70 was observed in only three active RR MS patients. Thus, it cannot be excluded that, in a particular subgroup of RR active MS patients, *C. pneumoniae* may enter into brain early in the course of the disease via transendothelial migration across the blood/brain barrier of activated infected blood-borne monocytes, resulting in ongoing inflammatory immune activation that takes place within the CNS. Alternatively, the presence of elevated rates

TABLE 2: Molecular protocols employed for detection of *C. pneumoniae* in clinical specimens from MS patients.

| MS patients (positive/total) | Controls (positive/total) | PCR assay | References |
|--------------------------------|-------------------------------|--|-----------------------------------|
| 7/30 (23%) | 0/56 (0%) | n-PCR [†] | Layh-Schmitt et al. 2000 [81] |
| 9/17 (52%) | 13/15 (86%) | PCR [†] | Li et al. 2000 [82] |
| 2/8 (25%) | | not reported [†] | Treib et al. 2000 [83] |
| 12/58 (21%) | 20/47 (43%) | n-PCR [†] | Gieffers et al. 2001 [84] |
| 3/32 (9%) | 0/30 (0%) | PCR [†] | Sotgiu et al. 2001 [85] |
| 11/16 (69%) | | PCR [†] | Ikejima et al. 2001 [86] |
| 9/28 (32%) | 2/15 (13%) | PCR [†] | Hao et al. 2002 [87] |
| 2/70 (2.9%) | 0/30 (0%) | n-PCR [†] | Chatzipanagiotou et al. 2003 [88] |
| 23/107 (21%) | 2/77 (3%) | Touchdown n-PCR [†] | Grimaldi et al. 2003 [89] |
| 2/25 (8%) | 3/28 (11%) | n-PCR and semi-n-PCR [†] | Rostasy et al. 2003 [90] |
| 42/84 (50%) | 25/89 (28%) | Touchdown n-PCR, RT-PCR [†] | Dong-Si et al. 2004 [91] |
| 26/71 (36.6%) | 24/72 (33.3%) | Touchdown n-PCR [†] | Contini et al. 2004 [92] |
| 12/20 (60%) | 1/12 (8%) | Touchdown n-PCR [†] | Sriram et al. 2005 [93] |
| 2/112 (2%) | 0/110 (0%) | Real-time PCR [‡] | Sessa et al. 2007 [94] |
| 7–9/14 (50–64.3%) [†] | 0–1/19 (0–5.2%) [†] | Touchdown n-PCR, | Contini et al. 2008 [95] |
| 5–7/14 (35.7–50%) [‡] | 0–2/19 (0–10.5%) [‡] | RT-PCR ^{†‡} , | |
| 64/80 (80%) | 5/57 (9%) | PCR-EIA, Touchdown n-PCR, Real Time-PCR [†] | Tang et al. 2009 [96] |

n-PCR: nested PCR; RT-PCR: reverse transcriptase PCR; PCR-EIA: PCR enzyme immunoassay; [†], CFS: cerebrospinal fluid; [‡], PBMC: peripheral blood mononuclear cells.

of *C. pneumoniae* DNA in CSF in this subset of MS patients could merely reflect the selective infiltration of monocytes which traffic into the brain after activation, thus suggesting a role for *C. pneumoniae* only as a silent passenger. In attempting to recover *C. pneumoniae* from cultured CSF and PBMC compartments with a PCR targeting multiple genes, a positivity for *C. pneumoniae* DNA and mRNA was recently detected in 64% of cocultured CSF and PBMCs of RR MS patients with evidence of disease activity, whereas only 3 controls were positive for Chlamydial DNA, suggesting that *C. pneumoniae* may occur in a persistent and metabolically active state at both peripheral and intrathecal levels in MS, but not in controls [95]. In this study the parallel molecular analysis of multiple Chlamydial target genes after co-culture of fresh CSF and PBMC specimens, has shown to enhance the sensitivity and specificity of molecular tools. Of note, as *C. pneumoniae* DNA was found in PBMCs which are able to cross the blood-brain barrier, these cells could be the source of intrathecally compartmentalized *C. pneumoniae* that, in turn, may induce a chronic persistent brain infection acting as a cofactor in the development of the disease. More recently, in a comparative study aimed to evaluate novel procedures for the detection of *C. pneumoniae* DNA in CSF, the qualitative colorimetric microtiter plate-based PCR-enzyme-immunoassay (PCR-EIA) has shown to be more sensitive

than a real-time quantitative PCR assay (TaqMan) and possessed a sensitivity that was equal to the nested-PCR [96]. In order to support the theory of an association between *C. pneumoniae* and MS, a number of studies did evaluate the presence of intrathecal IgG in the form of oligoclonal bands (OCB) in the CSF of MS patients. Their presence, as for other bacterial, viral, fungal, and parasitic diseases, would be of great evidence for an infectious cause of MS [127] and may reflect an antigen-driven immune response to infectious agents [128]. However, OCB are also detected other than in MS in 10% of patients with other inflammatory diseases of the CNS. In this regard, studies aimed to determine the CSF levels of anti-*C. pneumoniae* IgG in MS patients did result extremely variable (varying from 0% to 20%) producing any or scarce differences between MS and controls [81, 85, 87, 90, 122, 123, 126, 129, 130]. Recently, we found that an intrathecal synthesis of anti-*C. pneumoniae* IgG as evaluated by antibody specific index (ASI) was more frequent in MS (16.9%) and in OIND (21.6%) than in NIND (1.9%) patients and in patients with MS progressive forms (SP and PP MS) than in RR MS patients [131]. Moreover, among the patients with intrathecally produced anti-*C. pneumoniae* IgG, CSF *C. pneumoniae*-specific high-affinity antibodies, were demonstrated to be more frequent in a subset of patients with MS progressive forms (SP or

PP MS), than in OIND patients, and absent in RR MS and NIND patients. To further examine a possible relationship between *C. pneumoniae* infection and MS, Sriram published a study that examined autoptic samples of brain tissue and CSF using immunohistochemical staining with anti-*C. pneumoniae* monoclonal antibodies other than molecular and ultrastructural methods [93]. These techniques provided evidence of the presence of *C. pneumoniae* more commonly in MS patients (90%, 62%, and 55%, resp.) than in control patients. Using electron microscopy the authors first demonstrated the presence of immunogold-labeled objects of the morphology and size and of chlamydial EBs in the ependymal surfaces and periventricular regions in the CSF of four out of ten (40%) patients with MS but not in the CSF of control patients. Collectively taken, although MS patients were found to be more likely to have detectable levels of *C. pneumoniae* DNA in their CSF and intrathecally synthesized immunoglobulins, compared with patients with had neurological diseases, the overall findings examined in a review through 26 studies that considered 1332 MS patients and 1464 controls using random-effects methods and random-effects meta-regressions, adjusted for the confounding effect of gender differences, were insufficient to establish an etiologic relation between *C. pneumoniae* and MS [132].

The treatment directed against the inflammatory process is only partially efficacious on the MS disease course. In relapsing-remitting MS, such therapy slows the progression of disability, but in primary-progressive MS the same therapy has demonstrated to have little or no effect on the progression of disability. On the other hand, reports regarding the antimicrobial treatment of MS have provided conflicting results. In one trial the antibiotic minocycline resulted in a reduction in the number of gadolinium-enhancing MRI-detected lesions [133]. Another study showed that anti-chlamydial treatment reduced brain atrophy, but did not show any beneficial influence on the number of MRI Gd enhancing lesions [134]. The Vanderblit University group is currently treating patients with MS with a combination of bacterial protein synthesis inhibitors (doxycycline and azithromycin or roxithromycin or rifampicin) and then adding metronidazole to this. These studies will need to be validated by comprehensive multicenter trials of combined antibiotic treatment aimed at all phases of the organism's life-cycle [135].

From the data presented, there is strong evidence that *C. pneumoniae* has not a causal role in MS disease. Thus, the actual involvement of *C. pneumoniae* in MS still remains a matter of debate and requires further understanding through standardized cultural, molecular and ultrastructural protocols for *C. pneumoniae* in biological samples coming from MS patients and controls. While some studies suggest a role of *C. pneumoniae* only as a CNS innocent bystander epiphenomenon due to ongoing MS inflammation which favours a selective infiltration of infected-mononuclear cells within the CNS, others indicate a role of *C. pneumoniae* as a cofactor in development and progression of the disease by enhancing a pre-existing autoimmune response in a subset of MS patients, as supported by recent immunological and

molecular findings [95, 109, 136]. Recent our findings have demonstrated a possible association between *Parachlamydiae Like Organisms* and MS suggesting that these can act alone or together with *C. pneumoniae* as a cofactor in the development and progression of MS [137]. Although these data needs further assessment, their possible involvement in MS could be of great importance in public health.

Finally, we cannot exclude that other pathogens may be potentially involved in the development of MS disease. Virus have often been considered as potential candidates because they are known to cause demyelinating disease in experimental animals and man, and often cause disease with long periods of latency that presents clinically with relapsing, remitting symptoms [138]. To date, however, studies have failed to identify any single virus as playing a major role in MS. Among the virus suggested as MS cofactors, there are ubiquitous members of the family Herpesviridae, Human herpesvirus 6 (HHV-6), Epstein-Barr virus (EBV) [139–146]. As *Chlamydia*, these viruses can undergo an alternative infection cycle, entering a quiescent state (latency), with low grade viral infection that does not cause cell lysis, from which they subsequently can be reactivated. However, the cell type in which this occurs is usually not the same cell type in which the productive, cytotoxic infection occurs. The human MS-associated retrovirus (MSRV) belonging to endogenous retrovirus family, has been also described as potential pathogen in MS [147].

4. *C. pneumoniae* and Other Neurological Complications

A number of reports have focused on the involvement of *C. pneumoniae* in other CNS disorders and in particular in encephalitis or meningoencephalitis. We searched PubMed for articles on encephalitis and *Chlamydia pneumoniae* or *Chlamydophila pneumoniae* using keywords: encephalitis, meningoencephalitis, *Chlamydia pneumoniae*, *Chlamydophila pneumoniae*, and numerous additional keywords including “neurological complications” relevant to these topics. Due to the common usage of the previous genus name, “*Chlamydia*,” it was included in the search. The reported cases (Table 3) were not so frequent [148–159]. Most patients were young patients who presented with different neurological symptoms and/or neuro-radiological changes at CT or MRI scan and in most cases, there were also well defined accompanying respiratory symptoms, although these have in some cases preceded the onset of the neurological records. Three patients had cerebellar ataxia, acute demyelinating encephalitis (ADEM), and Guillain Barré syndrome. The detection of *Chlamydia* has been almost always done by serological methods based on detection of specific anti-*C. pneumoniae* antibodies of different classes by MIF (fourfold rise in the IgG titre) and ELISA techniques. One study detected the presence of IgA-type antibodies, suggesting a reinfection [158]. One note reported the use of PCR in a tracheal swab and increasing titres of *Chlamydia* IgM antibody [155]. These cases, and a review of the literature, suggest that *C. pneumoniae* infection in addition to

TABLE 3: Acute and chronic neurological complications preceding or following *C. pneumoniae* associated respiratory manifestations.

| Author, year | Patient n. | Sex/age range (yr) median | Clinical findings | <i>C. pneumoniae</i> detection | Neuroimaging | Treatment | Outcome |
|--------------------------------------|------------|---------------------------|--|--|---|--|----------------------------|
| Fryden et al. 1989 [148] | 1 | F/16 | Encephalitis, respiratory tract infection | Serum (4-fold rise) in IgG and IgM [§] CSF negative for IgG and IgM | Brain CT scan | Clorampheni-col, Steroids | Recovery 1 year later |
| Haidl et al. 1992 [149] | 1 | M/13 | Guillan-Barrè syndrome, parestesia, hyporeflexia, cough | Serum (4-fold rise) in IgG and IgM [§] | ND | Steroids | Recovery 5 weeks later |
| Michel et al. 1992 [150] | 1 | M/9 | Meningoradiculitis, cough, rhinitis, hyporeflexia, back stiffness | Serum 4-fold fall in IgM, CSF positive for total IgG | ND | ND | Recovery 6 months later |
| Sundelöf et al. 1993 [151] | 1 | M/18 | Meningoencephalitis, headache, diplopia paresthesia, fever, cough | Serum IgG (4-fold), IgM [§] CSF negative for IgG and IgM | Brain CT scan | Erythromycin, Cefotaxime, Acyclovir | Recovery <1 week later |
| Socan et al. 1994 [152] | 1 | M/18 | Diplopia, parestesia, meningeal syndrome, cough and pneumoniae | Serum IgM and IgG [§] CSF negative for IgG and IgM | Brain CT scan | Acyclovir, Cefotaxime, Erythromycin | Recovery <1 week later |
| Koskiniemi et al. 1997 [153] | 3 | F ² /5.6 | Pareses, sensory symptoms, convulsions, depression of consciousness | Serum IgM and IgG [§] | Brain CT, EEG ^{##} | Unspecified | ND |
| Korman et al. 1997 [154] | 1 | F/69 | Cerebellar ataxia, respiratory failure | Serum (4-fold rise) in IgG, elevated IgA [§] CSF negative for IgG and IgM | Brain CT | Erythromycin, Imipenem | Recovery <1 month later |
| Heick and Skriver 2000 [155] | 1 | F/18 | Coryzal illness, headache, dizziness, left-sided hemiparesis (†ADEM), ataxia, fever, dry cough | CKT [*] in serum serum IgM PCR in tracheal swab | Brain CT scan, cranial MR | Penicillin, Acyclovir, Gentamycin, Cefotaxime, Doxycycline, Methylprednisolone | Recovery 2 months |
| Guglielminotti et al. 2000 [156] | 1 | M/95 | Meningeal syndrome, sleepiness, dry cough | Serum (4-fold rise) in IgG and IgA [§] CSF IgG | Brain CT, EEG | Acyclovir, Amoxicillin, Erythromycin, Ofloxacin | died aspiration pneumoniae |
| Anton et al. 2000 [157] | 1 | F/16 | Hyporeflexia, bilateral nistagmus, sleepiness, motor weakness, rhinitis | Serum (4-fold rise) in IgG, IgM [§] CSF negative for IgG and IgM | Brain CT scan, Brain MR, Electrophysiological studies | Ceftriaxone, Methylprednisolone | Recovery 2 months later |
| Airas et al. 2001 [158] | 1 | F/33 | Sleepiness, memory loss, cough, pharyngitis | Serum IgA and IgG [†] | Brain MR | Acyclovir, Levofloxacin, Azithromycin | Partial improvement |
| Boschin-Crinquette et al. 2005 [159] | 1 | M/21 | Aphasia, meningeal syndrome, respiratory symptoms | Serum IgM [°] CSF negative for IgM, IgG and PCR | Brain CT scan, Brain MR | Acyclovir, Ofloxacin, Amoxicillin, Cefotaxime | Recovery <2 weeks later |

[§] MIF: microimmunofluorescence; [†]EIA: enzyme immunoassay; [°]ELISA: enzyme-linked immunoadsorbent assay; ^{*}CTK: *Chlamydia* Complement-binding Test; ^{##}EEG: electroencephalogram; [†]ADEM: acute disseminated encephalomyelitis.

other *Chlamydiae*, may present with significant neurological manifestations. Of interest, most of these patients did experience a favourable outcome after administration of antibiotic therapy with or without corticosteroid treatment, suggesting a strong etiologic link between the microorganism and encephalitis. Chlamydial infections along with *Mycoplasma* and legionella infections should be included in the differential diagnosis of respiratory infections with a neurologic presentation.

5. *C. pneumoniae* and Neurobehavioral Disorders

Although the limited data of literature, there is evidence that *Chlamydia* may be implicated in the pathogenesis of some mental or neurobehavioral disorders including autism and schizophrenia. Autism spectrum disorders (ASDs) are a group of neurobehavioral diseases of unknown aetiology, which include autism, attention deficit disorder, Asperger's syndrome, and so forth, which causes are unknown but appear to include genetic defects, heavy metal, and chemical and biological exposures [160]. Factors, such as geography, family socioeconomic status, vaccination records, and family educational levels may be also involved. They occur primarily in the young and are probably different in each patient. Such patients do not all share the same signs and symptoms but tend to share certain social, communication, motor, and sensory problems that affect their behaviour in predictable ways. In general, the criteria for diagnosis of ASD are the presence of a triad of impairments in social interaction, communication, and imagination [160]. These signs and symptoms are thought to be due to abnormalities in brain function or structure and are thought to have a genetic basis [161, 162]. There is growing awareness that ASD can have an infectious nature that may be a cofactor for the illness or can aggravate patient morbidity [163–165]. The appearance of infections and in particular *Mycoplasma* infections in children diagnosed with ASD has been also linked to the multiple vaccines received during childhood [136, 166]. In this setting, *C. pneumoniae* [167, 168] along with a number of systemic chronic infections, such as those by *Mycoplasma* species [169–172] and HHV-6 [171–173], have been identified in Gulf War veterans and in family members including their children, using highly sensitive PCR and confirming the results by Southern-blot and dot-blot hybridization. Interestingly, a number of these symptomatic children were diagnosed with autism or attention deficit disorder that fall under ASD [174]. Based on previous observations of persisting IgA titers in some patients with mental disorders, it has been hypothesized that Chlamydiaceae are main pathogenic factors in schizophrenia. Fellerhoff, using n-PCR, found a significant prevalence of *C. psittaci*, *C. pneumoniae*, and *C. trachomatis* (9/18, 50%), as compared to controls (8/115, 6.97%). Treatment with in vitro-activated immune cells together with antibiotic modalities showed sustained mental improvements in patients that did not depend on treatment with antipsychotic drugs [175].

6. Conclusions

C. pneumoniae is like a “New Bug that’s full of Surprises” [136]. This perfectly matches to the wide range of chronic diseases which can be sustained by this pathogen. Thanks to deep knowledge of the biology of *Chlamydia* and the use of increasingly sophisticated techniques than those traditionally used, the presence of *C. pneumoniae* genomic material was demonstrated in a large number of persons suffering from different acute and chronic diseases. Over the past 10 years, a growing number of reports have found a possible link between *C. pneumoniae* infection and atherosclerosis and CNS diseases including MS, AD other than a variety of neurobehavioral disorders. The main obstacles that have so far presented to support a definitive role of *C. pneumoniae* in chronic diseases are represented by the fact that no methods exist to safely and confidently diagnose chronic infection, and because chlamydial chronic infections are characterized by the inaccessibility of the “chlamydial persistent state” to conventional antichlamydial agents. A causative role of *C. pneumoniae* infection in cardiovascular disease has not yet been firmly established. Despite the molecular and genetic efforts that have been done on the role of *C. pneumoniae* in the progression of atherosclerosis, several important questions including whether the *C. pneumoniae* is an innocent passenger or whether it is actively involved in the initiation or progression of atherosclerotic disease urgently need an answer. In particular, *C. pneumoniae* HsP60 needs to be explored further as a potential culprit and therapeutic target [11–13]. Several drugs shown to be more or less effective in atherosclerotic disease are in the recent experiments, at the same time effective against *C. pneumoniae*. Statins, aspirin, and dietary polyphenolic compounds are among them. It is possible that the truly effective treatment targeting chronic *C. pneumoniae* infection will be found. At the same time, the development of efficacious vaccine should be continued [136]. The interpretation of the fact that astrocytes, microglia, and neurons are host cells for *C. pneumoniae* in the brain of AD patients, and that infected cells can be found in close proximity to both NSP and NFT, is hampered by the fact that most studies were done with different diagnostic methods, none of which still standardized. This has led a wide variation of interlaboratory test performance, even when the same test and the same criteria have been used. Thus, the actual involvement of *C. pneumoniae* in AD still remains a matter of debate and requires further understanding through standardized cultural, molecular protocols for *C. pneumoniae* in autoptic samples coming from AD patients and controls [51–53, 70]. The recent molecular, ultrastructural, and cultural advances that have provided evidence that *C. pneumoniae* is viable and metabolically active in different biological compartments such as CSF and PBMC from MS patients compared to controls, suggests an association between this pathogen and the disease, particularly in a subgroup of RR MS patients with clinical and MRI disease activity who experience the early inflammatory phase representing the development of the disease [89, 95, 108, 109]. However, the growing body of evidence suggests a role of *C. pneumoniae* only as a CNS innocent bystander

epiphenomenon due to ongoing MS inflammation or a cofactor in development and progression of the disease by enhancing a pre-existing autoimmune response in a subset of MS patients, as supported by the recent immunological and molecular findings [95, 96, 109]. Either for AD or MS there is urgency for further well-designed studies to determine both the importance of *C. pneumoniae* involvement in human diseases and the usefulness of antibiotic treatment. The role of *Chlamydia* in the pathogenesis of mental or neurobehavioral disorders including schizophrenia and autism is uncertain and fragmentary. However, the few existing reports suggest a potential involvement which will require further confirmation.

Acknowledgment

This work was in part supported by the Research Program Regione Emilia Romagna—University 2007–2009 (Innovative Research), entitled: “Regional Network for Implementing a Bank to Identify Biological Markers of Activity Disease to Clinical Variables in Multiple Sclerosis.”

References

- [1] K. D. E. Everett, R. M. Bush, and A. A. Andersen, “Emended description of the order *Chlamydiales*, proposal of *Parachlamydiaceae* fam. nov. and *Simkaniaceae* fam. nov., each containing one monotypic genus, revised taxonomy of the family *Chlamydiaceae*, including a new genus and five new species, and standards for the identification of organisms,” *International Journal of Systematic Bacteriology*, vol. 49, no. 2, pp. 415–440, 1999.
- [2] P. Saikku, M. Leinonen, K. Mattila, et al., “Serological evidence of an association of a novel *Chlamydia*, TWAR, with chronic coronary heart disease and acute myocardial infarction,” *The Lancet*, vol. 2, no. 8618, pp. 983–986, 1988.
- [3] M. Kleemola, P. Saikku, R. Visakorpi, S. P. Wang, and J. T. Grayston, “Epidemics of pneumoniae caused by TWAR, a new *Chlamydia* organism, in military trainees in Finland,” *Journal of Infectious Diseases*, vol. 157, no. 2, pp. 230–236, 1988.
- [4] C.-C. Kuo, L. A. Jackson, L. A. Campbell, and J. T. Grayston, “*Chlamydia pneumoniae* (TWAR),” *Clinical Microbiology Reviews*, vol. 8, no. 4, pp. 451–461, 1995.
- [5] C. H. Mordhorst, S. P. Wang, and J. T. Grayston, “Outbreak of *Chlamydia pneumoniae* infection in four farm families,” *European Journal of Clinical Microbiology and Infectious Diseases*, vol. 11, no. 7, pp. 617–620, 1992.
- [6] C. Yucesan and S. Sriram, “*Chlamydia pneumoniae* infection of the central nervous system,” *Current Opinion in Neurology*, vol. 14, no. 3, pp. 355–359, 2001.
- [7] C. W. Stratton and S. Sriram, “Association of *Chlamydia pneumoniae* with central nervous system disease,” *Microbes and Infection*, vol. 5, no. 13, pp. 1249–1253, 2003.
- [8] C. W. Stratton and W. M. Mitchell, “The immunopathology of chlamydial infections,” *Antimicrobics and Infectious Diseases Newsletter*, vol. 16, no. 12, pp. 89–94, 1997.
- [9] A. Ciervo, P. Visca, A. Petrucca, L. M. Biasucci, A. Maseri, and A. Cassone, “Antibodies to 60-kilodalton heat shock protein and outer membrane protein 2 of *Chlamydia pneumoniae* in patients with coronary heart disease,” *Clinical and Diagnostic Laboratory Immunology*, vol. 9, no. 1, pp. 66–74, 2002.
- [10] O. S. Mahdi, B. D. Horne, K. Mullen, J. B. Muhlestein, and G. I. Byrne, “Serum immunoglobulin G antibodies to chlamydial heat shock protein 60 but not to human and bacterial homologs are associated with coronary artery disease,” *Circulation*, vol. 106, no. 13, pp. 1659–1663, 2002.
- [11] M. Leinonen and P. Saikku, “Evidence for infectious agents in cardiovascular disease and atherosclerosis,” *Lancet Infectious Diseases*, vol. 2, no. 1, pp. 11–17, 2002.
- [12] L. A. Campbell and C.-C. Kuo, “*Chlamydia pneumoniae*: an infectious risk factor for atherosclerosis?” *Nature Reviews Microbiology*, vol. 2, no. 1, pp. 23–32, 2004.
- [13] F. F. Mussa, H. Chai, X. Wang, Q. Yao, A. B. Lumsden, and C. Chen, “*Chlamydia pneumoniae* and vascular disease: an update,” *Journal of Vascular Surgery*, vol. 43, no. 6, pp. 1301–1307, 2006.
- [14] A. Palikhe, M.-L. Lokki, P. Saikku, et al., “Association of *Chlamydia pneumoniae* infection with HLA-B*35 in patients with coronary artery disease,” *Clinical and Vaccine Immunology*, vol. 15, no. 1, pp. 55–59, 2008.
- [15] G. I. Byrne, “*Chlamydia* uncloaked,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 14, pp. 8040–8042, 2003.
- [16] G. I. Byrne and D. M. Ojcius, “*Chlamydia* and apoptosis: life and death decisions of an intracellular pathogen,” *Nature Reviews Microbiology*, vol. 2, no. 10, pp. 802–808, 2004.
- [17] J. Boman, S. Söderberg, J. Forsberg, et al., “High prevalence of *Chlamydia pneumoniae* DNA in peripheral blood mononuclear cells in patients with cardiovascular disease and in middle-aged blood donors,” *Journal of Infectious Diseases*, vol. 178, no. 1, pp. 274–277, 1998.
- [18] A. MacIntyre, R. Abramov, C. J. Hammond, et al., “*Chlamydia pneumoniae* infection promotes the transmigration of monocytes through human brain endothelial cells,” *Journal of Neuroscience Research*, vol. 71, no. 5, pp. 740–750, 2003.
- [19] C. C. Kuo and L. A. Campbell, “Chlamydial infections of the cardiovascular system,” *Frontiers in Bioscience*, vol. 8, pp. e36–e43, 2003.
- [20] R. W. Peeling, S.-P. Wang, J. T. Grayston, et al., “*Chlamydia pneumoniae* serology: interlaboratory variation in immunofluorescence assay results,” *Journal of Infectious Diseases*, vol. 181, no. 6, supplement 3, pp. S426–S429, 2000.
- [21] P. Apfalter, “*Chlamydia pneumoniae*, stroke, and serological associations: anything learned from the atherosclerosis-cardiovascular literature or do we have to start over again?” *Stroke*, vol. 37, no. 3, pp. 756–758, 2006.
- [22] N. Mackman, “Lipopolysaccharide induction of gene expression in human monocytic cells,” *Immunologic Research*, vol. 21, no. 2-3, pp. 247–251, 2000.
- [23] M. V. Kalayoglu, L. Indrawati, R. P. Morrison, S. G. Morrison, Y. Yuan, and G. I. Byrne, “Chlamydial virulence determinants in atherogenesis: the role of chlamydial lipopolysaccharide and heat shock protein 60 in macrophage-lipoprotein interactions,” *Journal of Infectious Diseases*, vol. 181, no. 6, supplement 3, pp. S483–S489, 2000.
- [24] I. W. Fong, B. Chiu, E. Viira, W. Tucker, H. Wood, and R. W. Peeling, “Chlamydial heat-shock protein-60 antibody and correlation with *Chlamydia pneumoniae* in atherosclerotic plaques,” *Journal of Infectious Diseases*, vol. 186, no. 10, pp. 1469–1473, 2002.
- [25] T. Huitinen, M. Leinonen, L. Tenkanen, et al., “Autoimmunity to human heat shock protein 60. *Chlamydia pneumoniae* infection, and inflammation in predicting coronary risk,” *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 22, no.

- 3, pp. 431–437, 2002.
- [26] M. L. J. Wimmer, R. Sandmann-Strupp, P. Saikku, and R. L. Haber, "Association of chlamydial infection with cerebrovascular disease," *Stroke*, vol. 27, no. 12, pp. 2207–2210, 1996.
- [27] M. S. V. Elkind, I.-F. Lin, J. T. Grayston, and R. L. Sacco, "Chlamydia pneumoniae and the risk of first ischemic stroke: the Northern Manhattan Stroke Study," *Stroke*, vol. 31, no. 7, pp. 1521–1525, 2000.
- [28] B. Piechowski-Jóźwiak, A. Mickielewicz, Z. Gaciong, H. Berent, and H. Kwieciński, "Elevated levels of anti-Chlamydia pneumoniae IgA and IgG antibodies in young adults with ischemic stroke," *Acta Neurologica Scandinavica*, vol. 116, no. 3, pp. 144–149, 2007.
- [29] J. G. Madre, J. L. R. Garcia, R. C. Gonzalez, et al., "Association between seropositivity to Chlamydia pneumoniae and acute ischaemic stroke," *European Journal of Neurology*, vol. 9, no. 3, pp. 303–306, 2002.
- [30] M. S. V. Elkind, M. L. C. Tondella, D. R. Feikin, B. S. Fields, S. Homma, and M. R. Di Tullio, "Seropositivity to Chlamydia pneumoniae is associated with risk of first ischemic stroke," *Stroke*, vol. 37, no. 3, pp. 790–795, 2006.
- [31] P. U. Heuschmann, D. Neureiter, M. Gesslein, et al., "Association between infection with Helicobacter pylori and Chlamydia pneumoniae and risk of ischemic stroke subtypes: results from a population-based case-control study," *Stroke*, vol. 32, no. 10, pp. 2253–2258, 2001.
- [32] A. Anzini, A. Cassone, M. Rasura, et al., "Chlamydia pneumoniae infection in young stroke patients: a case-control study," *European Journal of Neurology*, vol. 11, no. 5, pp. 321–327, 2004.
- [33] A. Vink, M. Poppen, A. H. Schoneveld, et al., "Distribution of Chlamydia pneumoniae in the human arterial system and its relation to the local amount of atherosclerosis within the individual," *Circulation*, vol. 103, no. 12, pp. 1613–1617, 2001.
- [34] K. Numazaki and S. Chibar, "Failure to detect Chlamydia pneumoniae in the central nervous system of patients with MS," *Neurology*, vol. 57, no. 4, p. 746, 2001.
- [35] S. F. Dowell, R. W. Peeling, J. Boman, et al., "Standardizing Chlamydia pneumoniae assays: recommendations from the Centers for Disease Control and Prevention (USA) and the Laboratory Centre for Disease Control (Canada)," *Clinical Infectious Diseases*, vol. 33, no. 4, pp. 492–502, 2001.
- [36] J. Boman and M. R. Hammerschlag, "Chlamydia pneumoniae and atherosclerosis: critical assessment of diagnostic methods and relevance to treatment studies," *Clinical Microbiology Reviews*, vol. 15, no. 1, pp. 1–20, 2002.
- [37] M. M. Ieven and V. Y. Hoymans, "Involvement of Chlamydia pneumoniae in atherosclerosis: more evidence for lack of evidence," *Journal of Clinical Microbiology*, vol. 43, no. 1, pp. 19–24, 2005.
- [38] A. J. Littman, L. A. Jackson, E. White, M. D. Thornquist, C. A. Gaydos, and T. L. Vaughan, "Interlaboratory reliability of microimmunofluorescence test for measurement of Chlamydia pneumoniae-specific immunoglobulin A and G antibody titers," *Clinical and Diagnostic Laboratory Immunology*, vol. 11, no. 3, pp. 615–617, 2004.
- [39] P. Maggi, R. Monno, A. Chirrianni, et al., "Role of Chlamydia infection in the pathogenesis of atherosclerotic plaques in HIV-1-positive patients," *In Vivo*, vol. 20, no. 3, pp. 409–414, 2006.
- [40] G. Tositti, M. Rassu, P. Fabris, et al., "Chlamydia pneumoniae infection in HIV-positive patients: prevalence and relationship with lipid profile," *HIV Medicine*, vol. 6, no. 1, pp. 27–32, 2005.
- [41] V. Gaona-Flores, G. García-Elorriaga, M. Valerio-Minero, et al., "Anti-Chlamydia pneumoniae antibodies as associated factor for carotid atherosclerosis in patients with AIDS," *Current HIV Research*, vol. 6, no. 3, pp. 267–271, 2008.
- [42] P. Apfalter, O. Assadian, F. Blasi, et al., "Reliability of nested PCR for detection of Chlamydia pneumoniae DNA in atheromas: results from a multicenter study applying standardized protocols," *Journal of Clinical Microbiology*, vol. 40, no. 12, pp. 4428–4434, 2002.
- [43] R. Sessa, M. Di Pietro, G. Schiavoni, et al., "Chlamydia pneumoniae DNA in patients with symptomatic carotid atherosclerotic disease," *Journal of Vascular Surgery*, vol. 37, no. 5, pp. 1027–1031, 2003.
- [44] A. Palikhe, T. Tirola, M. Puolakkainen, et al., "Chlamydia pneumoniae DNA is present in peripheral blood mononuclear cells during acute coronary syndrome and correlates with chlamydial lipopolysaccharide levels in serum," *Scandinavian Journal of Infectious Diseases*, vol. 41, no. 3, pp. 201–205, 2009.
- [45] J. Gieffers, H. Füllgraf, J. Jahn, et al., "Chlamydia pneumoniae infection in circulating human monocytes is refractory to antibiotic treatment," *Circulation*, vol. 103, no. 3, pp. 351–356, 2001.
- [46] J. Gieffers, J. Rupp, A. Gebert, W. Solbach, and M. Klinger, "First-choice antibiotics at subinhibitory concentrations induce persistence of Chlamydia pneumoniae," *Antimicrobial Agents and Chemotherapy*, vol. 48, no. 4, pp. 1402–1405, 2004.
- [47] J. T. Grayston, R. A. Kronmal, L. A. Jackson, et al., "Azithromycin for the secondary prevention of coronary events," *The New England Journal of Medicine*, vol. 352, no. 16, pp. 1637–1645, 2005.
- [48] B. J. Balin, H. C. Gérard, E. J. Arking, et al., "Identification and localization of Chlamydia pneumoniae in the Alzheimer's brain," *Medical Microbiology and Immunology*, vol. 187, no. 1, pp. 23–42, 1998.
- [49] D. Nochlin, C. M. Shaw, L. A. Campbell, and C.-C. Kuo, "Failure to detect Chlamydia pneumoniae in brain tissues of Alzheimer's disease," *Neurology*, vol. 53, no. 8, p. 1888, 1999.
- [50] J. Gieffers, E. Reusche, W. Solbach, and M. Maass, "Failure to detect Chlamydia pneumoniae in brain sections of Alzheimer's disease patients," *Journal of Clinical Microbiology*, vol. 38, no. 2, pp. 881–882, 2000.
- [51] R. H. Ring and J. M. Lyons, "Failure to detect Chlamydia pneumoniae in the late-onset Alzheimer's brain," *Journal of Clinical Microbiology*, vol. 38, no. 7, pp. 2591–2594, 2000.
- [52] G. S. Taylor, I. B. Vipond, I. D. Paul, S. Matthews, G. K. Wilcock, and E. O. Caul, "Failure to correlate C. pneumoniae with late onset Alzheimer's disease," *Neurology*, vol. 59, no. 1, pp. 142–143, 2002.
- [53] M. A. Wozniak, A. Cookson, G. K. Wilcock, and R. F. Itzhaki, "Absence of Chlamydia pneumoniae in brain of vascular dementia patients," *Neurobiology of Aging*, vol. 24, no. 6, pp. 761–765, 2003.
- [54] H. C. Gérard, U. Dreses-Werringloer, K. S. Wildt, et al., "Chlamydia (Chlamydia) pneumoniae in the Alzheimer's brain," *FEMS Immunology and Medical Microbiology*, vol. 48, no. 3, pp. 355–366, 2006.
- [55] B. J. Balin and D. M. Appelt, "Role of infection in Alzheimer's disease," *Journal of the American Osteopathic Association*, vol. 101, no. 12, supplement 1, pp. S1–S6, 2001.
- [56] R. W. Keefover, "The clinical epidemiology of Alzheimer's

- disease," *Neurologic Clinics*, vol. 14, no. 2, pp. 337–351, 1996.
- [57] C. Kawas, S. Gray, R. Brookmeyer, J. Fozard, and A. Zonderman, "Age-specific incidence rates of Alzheimer's disease: the Baltimore Longitudinal Study of Aging," *Neurology*, vol. 54, no. 11, pp. 2072–2077, 2000.
- [58] A. Di Carlo, M. Baldereschi, L. Amaducci, et al., "Incidence of dementia, Alzheimer's disease, and vascular dementia in Italy. The ILSA study," *Journal of the American Geriatrics Society*, vol. 50, no. 1, pp. 41–48, 2002.
- [59] G. Ravaglia, P. Forti, F. Maioli, et al., "Incidence and etiology of dementia in a large elderly Italian population," *Neurology*, vol. 64, no. 9, pp. 1525–1530, 2005.
- [60] D. A. Evans, D. A. Bennett, R. S. Wilson, et al., "Incidence of Alzheimer disease in a biracial urban community: relation to apolipoprotein E allele status," *Archives of Neurology*, vol. 60, no. 2, pp. 185–189, 2003.
- [61] R. Brookmeyer, S. Gray, and C. Kawas, "Projections of Alzheimer's disease in the United States and the public health impact of delaying disease onset," *American Journal of Public Health*, vol. 88, no. 9, pp. 1337–1342, 1998.
- [62] L.-F. Lue, L. Brachova, W. H. Civin, and J. Rogers, "Inflammation, A β deposition, and neurofibrillary tangle formation as correlates of Alzheimer's disease neurodegeneration," *Journal of Neuropathology and Experimental Neurology*, vol. 55, no. 10, pp. 1083–1088, 1996.
- [63] R. F. Itzhaki, W.-R. Lin, D. Shang, G. K. Wilcock, B. Faragher, and G. A. Jamieson, "Herpes simplex virus type 1 in brain and risk of Alzheimer's disease," *The Lancet*, vol. 349, no. 9047, pp. 241–244, 1997.
- [64] R. F. Itzhaki, C. B. Dobson, W.-R. Lin, and M. A. Wozniak, "Association of HSV1 and apolipoprotein E- ϵ 4 in Alzheimer's disease," *Journal of Neurovirology*, vol. 7, no. 6, pp. 570–571, 2001.
- [65] B. G. T. Pogo, J. Casals, and T. S. Elizan, "A study of viral genomes and antigens in brains of patients with Alzheimer's disease," *Brain*, vol. 110, no. 4, pp. 907–915, 1987.
- [66] R. P. Friedland, C. May, and J. Dahlberg, "The viral hypothesis of Alzheimer's disease: absence of antibodies to lentiviruses," *Archives of Neurology*, vol. 47, no. 2, pp. 177–178, 1990.
- [67] E. B. Renvoize, I. O. Awad, and M. H. Hambling, "A sero-epidemiological study of conventional infectious agents in Alzheimer's disease," *Age and Ageing*, vol. 16, no. 5, pp. 311–314, 1987.
- [68] J. Miklossy, "Alzheimer's disease—a spirochetosis?" *Neuro-Report*, vol. 4, no. 7, pp. 841–848, 1993.
- [69] W. B. Matthews, "Unconventional virus infection and neurological disease," *Neuropathology and Applied Neurobiology*, vol. 12, no. 2, pp. 111–116, 1986.
- [70] S. R. Robinson, C. Dobson, and J. Lyons, "Challenges and directions for the pathogen hypothesis of Alzheimer's disease," *Neurobiology of Aging*, vol. 25, no. 5, pp. 629–637, 2004.
- [71] U. Dreses-Werringloer, M. Bhuiyan, Y. Zhao, H. C. Gérard, J. A. Whittum-Hudson, and A. P. Hudson, "Initial characterization of *Chlamydia* (*Chlamydia*) *pneumoniae* cultured from the late-onset Alzheimer brain," *International Journal of Medical Microbiology*, vol. 299, no. 3, pp. 187–201, 2009.
- [72] H. C. Gérard, G. F. Wang, B. J. Balin, H. R. Schumacher, and A. P. Hudson, "Frequency of apolipoprotein E (APOE) allele types in patients with *Chlamydia*-associated arthritis and other arthritides," *Microbial Pathogenesis*, vol. 26, no. 1, pp. 35–43, 1999.
- [73] H. C. Gérard, K. L. Wildt, J. A. Whittum-Hudson, Z. Lai, J. Ager, and A. P. Hudson, "The load of *Chlamydia pneumoniae* in the Alzheimer's brain varies with APOE genotype," *Microbial Pathogenesis*, vol. 39, no. 1-2, pp. 19–26, 2005.
- [74] A. MacIntyre, C. J. Hammond, C. S. Little, D. M. Appelt, and B. J. Balin, "*Chlamydia pneumoniae* infection alters the junctional complex proteins of human brain microvascular endothelial cells," *FEMS Microbiology Letters*, vol. 217, no. 2, pp. 167–172, 2002.
- [75] C. S. Little, C. J. Hammond, A. MacIntyre, B. J. Balin, and D. M. Appelt, "*Chlamydia pneumoniae* induces Alzheimer-like amyloid plaques in brains of BALB/c mice," *Neurobiology of Aging*, vol. 25, no. 4, pp. 419–429, 2004.
- [76] M. B. Loeb, D. W. Molloy, M. Smieja, et al., "A randomized, controlled trial of doxycycline and rifampin for patients with Alzheimer's disease," *Journal of the American Geriatrics Society*, vol. 52, no. 3, pp. 381–387, 2004.
- [77] A. E. Finefrock, A. I. Bush, and P. M. Doraiswamy, "Current status of metals as therapeutic targets in Alzheimer's disease," *Journal of the American Geriatrics Society*, vol. 51, no. 8, pp. 1143–1148, 2003.
- [78] L. Cahoon, "The curious case of cloquinol," *Nature Medicine*, vol. 15, no. 4, pp. 356–359, 2009.
- [79] C. Contini, E. Fainardi, S. Seraceni, E. Granieri, M. Castellazzi, and R. Cultrera, "Molecular identification and antibody testing of *Chlamydia pneumoniae* in a subgroup of patients with HIV-associated dementia complex. Preliminary results," *Journal of Neuroimmunology*, vol. 136, no. 1-2, pp. 172–177, 2003.
- [80] S. Gartner, "HIV infection and dementia," *Science*, vol. 287, no. 5453, pp. 602–604, 2000.
- [81] G. Layh-Schmitt, C. Bendl, U. Hildt, et al., "Evidence for infection with *Chlamydia pneumoniae* in a subgroup of patients with multiple sclerosis," *Annals of Neurology*, vol. 47, no. 5, pp. 652–655, 2000.
- [82] W. P. Li, X. Ming, S. Cook, B. Blumberg, and P. Dowling, "*Chlamydia pneumoniae* sequence frequently present in both MS and control spinal fluid," *Neurology*, vol. 54, supplement 3, p. A165, 2000.
- [83] J. Treib, A. Haaß, W. Stille, et al., "Multiple sclerosis and *Chlamydia pneumoniae*," *Annals of Neurology*, vol. 47, no. 3, p. 408, 2000.
- [84] J. Gieffers, D. Pohl, J. Treib, et al., "Presence of *Chlamydia pneumoniae* DNA in the cerebral spinal fluid is a common phenomenon in a variety of neurological diseases and not restricted to multiple sclerosis," *Annals of Neurology*, vol. 49, no. 5, pp. 585–589, 2001.
- [85] S. Sotgiu, A. Piana, M. Pugliatti, et al., "*Chlamydia pneumoniae* in the cerebrospinal fluid of patients with multiple sclerosis and neurological controls," *Multiple Sclerosis*, vol. 7, no. 6, pp. 371–374, 2001.
- [86] H. Ikejima, S. Haranaga, H. Takemura, et al., "PCR-based method for isolation and detection of *Chlamydia pneumoniae* DNA in cerebrospinal fluids," *Clinical and Diagnostic Laboratory Immunology*, vol. 8, no. 3, pp. 499–502, 2001.
- [87] Q. Hao, N. Miyashita, M. Matsui, H.-Y. Wang, T. Matsushima, and T. Saida, "*Chlamydia pneumoniae* infection associated with enhanced MRI spinal lesions in multiple sclerosis," *Multiple Sclerosis*, vol. 8, no. 5, pp. 436–440, 2002.

- [88] S. Chatzipanagiotou, C. Tsakanikas, M. Anagnostouli, M. Rentzos, A. Ioannidis, and C. Nicolaou, "Detection of *Chlamydia pneumoniae* in the cerebrospinal fluid of patients with multiple sclerosis by combination of cell culture and PCR. No evidence for possible association," *Molecular Diagnosis*, vol. 7, no. 1, pp. 41–43, 2003.
- [89] L. M. E. Grimaldi, A. Pincherle, F. Martinelli-Boneschi, et al., "An MRI study of *Chlamydia pneumoniae* infection in Italian multiple sclerosis patients," *Multiple Sclerosis*, vol. 9, no. 5, pp. 467–471, 2003.
- [90] K. Rostasy, H. Reiber, D. Pohl, et al., "*Chlamydia pneumoniae* in children with MS: frequency and quantity of intrathecal antibodies," *Neurology*, vol. 61, no. 1, pp. 125–128, 2003.
- [91] T. Dong-Si, J. Weber, Y. B. Liu, et al., "Increased prevalence of and gene transcription by *Chlamydia pneumoniae* in cerebrospinal fluid of patients with relapsing-remitting multiple sclerosis," *Journal of Neurology*, vol. 251, no. 5, pp. 542–547, 2004.
- [92] C. Contini, R. Cultrera, S. Seraceni, M. Castellazzi, E. Granieri, and E. Fainardi, "Cerebrospinal fluid molecular demonstration of *Chlamydia pneumoniae* DNA is associated to clinical and brain magnetic resonance imaging activity in a subset of patients with relapsing-remitting multiple sclerosis," *Multiple Sclerosis*, vol. 10, no. 4, pp. 360–369, 2004.
- [93] S. Sriram, A. Ljunggren-Rose, S.-Y. Yao, and W. O. Whetsell Jr., "Detection of chlamydial bodies and antigens in the central nervous system of patients with multiple sclerosis," *Journal of Infectious Diseases*, vol. 192, no. 7, pp. 1219–1228, 2005.
- [94] R. Sessa, G. Schiavoni, G. Borriello, et al., "Real time PCR for detection of *Chlamydia pneumoniae* in peripheral blood mononuclear cells of patients with multiple sclerosis," *Journal of Neurology*, vol. 254, no. 9, pp. 1293–1295, 2007.
- [95] C. Contini, S. Seraceni, M. Castellazzi, E. Granieri, and E. Fainardi, "*Chlamydia pneumoniae* DNA and mRNA transcript levels in peripheral blood mononuclear cells and cerebrospinal fluid of patients with multiple sclerosis," *Neuroscience Research*, vol. 62, no. 1, pp. 58–61, 2008.
- [96] Y.-W. Tang, S. Sriram, H. Li, et al., "Qualitative and quantitative detection of *Chlamydia pneumoniae* DNA in cerebrospinal fluid from multiple sclerosis patients and controls," *PLoS ONE*, vol. 4, no. 4, article e5200, 2009.
- [97] D. W. Anderson, J. H. Ellenberg, C. M. Leventhal, S. C. Reingold, M. Rodriguez, and D. H. Silberberg, "Revised estimate of the prevalence of multiple sclerosis in the United States," *Annals of Neurology*, vol. 31, no. 3, pp. 333–336, 1992.
- [98] M. H. Barnett and I. Sutton, "The pathology of multiple sclerosis: a paradigm shift," *Current Opinion in Neurology*, vol. 19, no. 3, pp. 242–247, 2006.
- [99] J. W. Princeas, E. E. Kwon, E.-S. Cho, and L. R. Sharer, "Continual breakdown and regeneration of myelin in progressive multiple sclerosis plaques," *Annals of the New York Academy of Sciences*, vol. 436, pp. 11–32, 1984.
- [100] M. Bsibsi, R. Ravid, D. Gveric, and J. M. van Noort, "Broad expression of Toll-like receptors in the human central nervous system," *Journal of Neuropathology and Experimental Neurology*, vol. 61, no. 11, pp. 1013–1021, 2002.
- [101] A. G. Joyee and X. Yang, "Role of toll-like receptors in immune responses to chlamydial infections," *Current Pharmaceutical Design*, vol. 14, no. 6, pp. 593–600, 2008.
- [102] S. Seraceni, L. Badia, A. Grilli, E. Fainardi, and C. Contini, "Role of toll like receptors in patients with multiple sclerosis and *Chlamydia pneumoniae* infection," in *Proceedings of the European Congress of Clinical Microbiology and Infectious Diseases*, Helsinki, Finland, May 2009, abstract no. R2174.
- [103] D. H. Gilden, "Infectious causes of multiple sclerosis," *Lancet Neurology*, vol. 4, no. 3, pp. 195–202, 2005.
- [104] I. Steiner, P. Nisipianu, and I. Wirguin, "Infection and the etiology and pathogenesis of multiple sclerosis," *Current Neurology and Neuroscience Reports*, vol. 1, no. 3, pp. 271–276, 2001.
- [105] D. H. Gilden, "*Chlamydia*: a role for multiple sclerosis or more confusion," *Annals of Neurology*, vol. 46, no. 1, pp. 4–5, 1999.
- [106] S. Sriram, W. Mitchell, and C. Stratton, "Multiple sclerosis associated with *Chlamydia pneumoniae* infection of the CNS," *Neurology*, vol. 50, no. 2, pp. 571–572, 1998.
- [107] S. Sriram, C. W. Stratton, S.-Y. Yao, et al., "*Chlamydia pneumoniae* infection of the central nervous system in multiple sclerosis," *Annals of Neurology*, vol. 46, no. 1, pp. 6–14, 1999.
- [108] C. W. Stratton and D. B. Wheldon, "Multiple sclerosis: an infectious syndrome involving *Chlamydia pneumoniae*," *Trends in Microbiology*, vol. 14, no. 11, pp. 474–479, 2006.
- [109] E. Fainardi, M. Castellazzi, S. Seraceni, E. Granieri, and C. Contini, "Under the microscope: focus on *Chlamydia pneumoniae* infection and multiple sclerosis," *Current Neurovascular Research*, vol. 5, no. 1, pp. 60–70, 2008.
- [110] K. L. Munger, R. W. Peeling, M. A. Hernán, et al., "Infection with *Chlamydia pneumoniae* and risk of multiple sclerosis," *Epidemiology*, vol. 14, no. 2, pp. 141–147, 2003.
- [111] K. L. Munger, G. N. DeLorenze, L. I. Levin, et al., "A prospective study of *Chlamydia pneumoniae* infection and risk of MS in two US cohorts," *Neurology*, vol. 62, no. 10, pp. 1799–1803, 2004.
- [112] D. Buljevac, R. P. Verkooyen, B. C. Jacobs, et al., "*Chlamydia pneumoniae* and the risk for exacerbation in multiple sclerosis patients," *Annals of Neurology*, vol. 54, no. 6, pp. 828–831, 2003.
- [113] M. R. Hammerschlag, Z. Ke, F. Lu, P. Roblin, J. Boman, and B. Kalman, "Is *Chlamydia pneumoniae* present in brain lesions of patients with multiple sclerosis?" *Journal of Clinical Microbiology*, vol. 38, no. 11, pp. 4274–4276, 2000.
- [114] J. Boman, P. M. Roblin, P. Sundström, M. Sandström, and M. R. Hammerschlag, "Failure to detect *Chlamydia pneumoniae* in the central nervous system of patients with MS," *Neurology*, vol. 54, no. 1, p. 265, 2000.
- [115] S. D. Poland and G. P. A. Rice, "*Chlamydia pneumoniae* and multiple sclerosis," *Neurology*, vol. 54, supplement 3, p. A165, 2000.
- [116] S. J. Furrows, J. C. Hartley, J. Bell, et al., "*Chlamydia pneumoniae* infection of the central nervous system in patients with multiple sclerosis," *Journal of Neurology, Neurosurgery and Psychiatry*, vol. 75, no. 1, pp. 152–154, 2004.
- [117] F. Cirino, W. C. Webley, C. West, N. L. Croteau, C. Andrezejewski Jr., and E. S. Stuart, "Detection of *Chlamydia* in the peripheral blood cells of normal donors using in vitro culture, immunofluorescence microscopy and flow cytometry techniques," *BMC Infectious Diseases*, vol. 6, article 23, 2006.
- [118] Y. Yamamoto, "PCR in diagnosis of infection: detection of bacteria in cerebrospinal fluids," *Clinical and Diagnostic Laboratory Immunology*, vol. 9, no. 3, pp. 508–514, 2002.
- [119] J. B. Mahony, S. Chong, B. K. Coombes, M. Smieja, and A. Petrich, "Analytical sensitivity, reproducibility of results, and clinical performance of five PCR assays for detecting *Chlamydia pneumoniae* DNA in peripheral blood

- mononuclear cells," *Journal of Clinical Microbiology*, vol. 38, no. 7, pp. 2622–2627, 2000.
- [120] M. Kaufman, C. A. Gaydos, S. Sriram, J. Boman, M. L. Tondella, and H. J. Norton, "Is *Chlamydia pneumoniae* found in spinal fluid samples from multiple sclerosis patients? Conflicting results," *Multiple Sclerosis*, vol. 8, no. 4, pp. 289–294, 2002.
- [121] S. Sriram, S.-Y. Yao, C. Stratton, et al., "Comparative study of the presence of *Chlamydia pneumoniae* in cerebrospinal fluid of patients with clinically definite and monosymptomatic multiple sclerosis," *Clinical and Diagnostic Laboratory Immunology*, vol. 9, no. 6, pp. 1332–1337, 2002.
- [122] A. Saiz, M. A. Marcos, F. Graus, J. Vidal, and M. T. Jimenez de Anta, "No evidence of CNS infection with *Chlamydia pneumoniae* in patients with multiple sclerosis," *Journal of Neurology*, vol. 248, no. 7, pp. 617–618, 2001.
- [123] T. Derfuss, R. Gürkov, F. T. Bergh, et al., "Intrathecal antibody production against *Chlamydia pneumoniae* in multiple sclerosis is part of a polyspecific immune response," *Brain*, vol. 124, no. 7, pp. 1325–1335, 2001.
- [124] F. Budak, S. Keçeli, H. Efendi, F. Budak, and H. Vahaboğlu, "The investigation of *Chlamydia pneumoniae* in patients with multiple sclerosis," *International Journal of Neuroscience*, vol. 117, no. 3, pp. 409–415, 2007.
- [125] S. A. Morré, C. J. A. de Groot, J. Killestein, et al., "Is *Chlamydia pneumoniae* present in the central nervous system of multiple sclerosis patients?" *Annals of Neurology*, vol. 48, no. 3, p. 399, 2000.
- [126] E. Pucci, C. Taus, E. Cartechini, et al., "Lack of *Chlamydia* infection of the central nervous system in multiple sclerosis," *Annals of Neurology*, vol. 48, no. 3, pp. 399–400, 2000.
- [127] E. J. Thompson and M. S. Freedman, "Cerebrospinal fluid analysis in the diagnosis of multiple sclerosis," *Advances in Neurology*, vol. 98, pp. 147–160, 2006.
- [128] T. Smith-Jensen, M. P. Burgoon, J. Anthony, H. Kraus, D. H. Gilden, and G. P. Owens, "Comparison of immunoglobulin G heavy-chain sequences in MS and SSPE brains reveals an antigen-driven response," *Neurology*, vol. 54, no. 6, pp. 1227–1232, 2000.
- [129] D. Krametter, G. Niederwieser, A. Berghold, et al., "*Chlamydia pneumoniae* in multiple sclerosis: humoral immune responses in serum and cerebrospinal fluid and correlation with disease activity marker," *Multiple Sclerosis*, vol. 7, no. 1, pp. 13–18, 2001.
- [130] D. Franciotta, E. Zardini, R. Bergamaschi, L. M. Grimaldi, L. Andreoni, and V. Cosi, "Analysis of *Chlamydia pneumoniae*-specific oligoclonal bands in multiple sclerosis and other neurologic diseases," *Acta Neurologica Scandinavica*, vol. 112, no. 4, pp. 238–241, 2005.
- [131] E. Fainardi, M. Castellazzi, I. Casetta, et al., "Intrathecal production of *Chlamydia pneumoniae*-specific high-affinity antibodies is significantly associated to a subset of multiple sclerosis patients with progressive forms," *Journal of the Neurological Sciences*, vol. 217, no. 2, pp. 181–188, 2004.
- [132] P. G. Bagos, G. Nikolopoulos, and A. Ioannidis, "*Chlamydia pneumoniae* infection and the risk of multiple sclerosis: a meta-analysis," *Multiple Sclerosis*, vol. 12, no. 4, pp. 397–411, 2006.
- [133] L. M. Metz, Y. Zhang, M. Yeung, et al., "Minocycline reduces gadolinium-enhancing magnetic resonance imaging lesions in multiple sclerosis," *Annals of Neurology*, vol. 55, no. 5, p. 756, 2004.
- [134] S. Sriram, S. Y. Yao, C. Stratton, H. Moses, P. A. Narayana, and J. S. Wolinsky, "Pilot study to examine the effect of antibiotic therapy on MRI outcomes in RRMS," *Journal of the Neurological Sciences*, vol. 234, no. 1–2, pp. 87–91, 2005.
- [135] C. W. Stratton and D. B. Wheldon, "Antimicrobial treatment of multiple sclerosis," *Infection*, vol. 35, no. 5, pp. 383–385, 2007.
- [136] C. Contini, "International conference on chlamydial and *Mycoplasma* human infections," *Future Microbiology*, vol. 2, no. 4, pp. 373–376, 2007.
- [137] C. Contini, S. Seraceni, R. Cultrera, M. Castellazzi, E. Granieri, and E. Fainardi, "Molecular detection of *Parachlamydia*-like organisms in cerebrospinal fluid of patients with multiple sclerosis," *Multiple Sclerosis*, vol. 14, no. 4, pp. 564–566, 2008.
- [138] D. H. Gilden, M. E. Devlin, M. P. Burgoon, and G. P. Owens, "The search for virus in multiple sclerosis brain," *Multiple Sclerosis*, vol. 2, no. 4, pp. 179–183, 1996.
- [139] R. Álvarez-Lafuente, V. De las Heras, M. Bartolomé, J. J. Picazo, and R. Arroyo, "Relapsing-remitting multiple sclerosis and human herpesvirus 6 active infection," *Archives of Neurology*, vol. 61, no. 10, pp. 1523–1527, 2004.
- [140] F. G. A. Moore and C. Wolfson, "Human herpes virus 6 and multiple sclerosis," *Acta Neurologica Scandinavica*, vol. 106, no. 2, pp. 63–83, 2002.
- [141] T. Christensen, "The role of EBV in MS pathogenesis," *International MS Journal*, vol. 13, no. 2, pp. 52–57, 2006.
- [142] D. Clark, "Human herpesvirus type 6 and multiple sclerosis," *Herpes*, vol. 11, supplement 2, pp. 112A–119A, 2004.
- [143] G. N. DeLorenze, K. L. Munger, E. T. Lennette, N. Orentreich, J. H. Vogelstein, and A. Ascherio, "Epstein-Barr virus and multiple sclerosis: evidence of association from a prospective study with long-term follow-up," *Archives of Neurology*, vol. 63, no. 6, pp. 839–844, 2006.
- [144] P. Höllsberg, M. Kusk, E. Bech, H. J. Hansen, J. Jakobsen, and S. Haahr, "Presence of Epstein-Barr virus and human herpesvirus 6B DNA in multiple sclerosis patients: associations with disease activity," *Acta Neurologica Scandinavica*, vol. 112, no. 6, pp. 395–402, 2005.
- [145] J. M. van Noort, J. J. Bajramovic, A. C. Plomp, and M. J. B. van Stipdonk, "Mistaken self, a novel model that links microbial infections with myelin-directed autoimmunity in multiple sclerosis," *Journal of Neuroimmunology*, vol. 105, no. 1, pp. 46–57, 2000.
- [146] S. Cepok, D. Zhou, R. Srivastava, et al., "Identification of Epstein-Barr virus proteins as putative targets of the immune response in multiple sclerosis," *Journal of Clinical Investigation*, vol. 115, no. 5, pp. 1352–1360, 2005.
- [147] A. Dolei, "MSRV/HERV-W/syncytin and its linkage to multiple sclerosis: the usability and the hazard of a human endogenous retrovirus," *Journal of NeuroVirology*, vol. 11, no. 2, pp. 232–235, 2005.
- [148] A. Fryden, E. Kihlstrom, R. Maller, K. Persson, V. Romanus, and S. Ansehn, "A clinical and epidemiological study of "ornithosis" caused by *Chlamydia psittaci* and *Chlamydia pneumoniae* (strain TWAR)," *Scandinavian Journal of Infectious Diseases*, vol. 21, no. 6, pp. 681–691, 1989.
- [149] S. Haidl, S. Ivarsson, I. Bjerre, and K. Persson, "Guillain-Barré syndrome after *Chlamydia pneumoniae* infection," *The New England Journal of Medicine*, vol. 326, no. 8, pp. 576–577, 1992.

- [150] D. Michel, J. C. Antoine, B. Pozzetto, O. G. Gaudin, and F. Lucht, "Lumbosacral meningoradiculitis associated with *Chlamydia pneumoniae* infection," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 55, no. 6, p. 511, 1992.
- [151] B. Sundelöf, H. Gnärpe, and J. Gnärpe, "An unusual manifestation of *Chlamydia pneumoniae* infection: meningitis, hepatitis, iritis and atypical erythema nodosum," *Scandinavian Journal of Infectious Diseases*, vol. 25, no. 2, pp. 259–261, 1993.
- [152] M. Socan, B. Beovic, and D. Kese, "*Chlamydia pneumoniae* and meningoencephalitis," *The New England Journal of Medicine*, vol. 331, no. 6, p. 406, 1994.
- [153] M. Koskiniemi, M. Korppi, K. Mustonen, et al., "Epidemiology of encephalitis in children. A prospective multicentre study," *European Journal of Pediatrics*, vol. 156, no. 7, pp. 541–545, 1997.
- [154] T. M. Korman, J. D. Turnidge, and M. L. Grayson, "Neurological complications of chlamydial infections: case report and review," *Clinical Infectious Diseases*, vol. 25, no. 4, pp. 847–851, 1997.
- [155] A. Heck and E. Skriver, "*Chlamydia pneumoniae*-associated ADEM," *European Journal of Neurology*, vol. 7, no. 4, pp. 435–438, 2000.
- [156] J. Guglielminotti, N. Lellouche, E. Maury, M. Alzieu, B. Guidet, and G. Offenstadt, "Severe meningoencephalitis: an unusual manifestation of *Chlamydia pneumoniae* infection," *Clinical Infectious Diseases*, vol. 30, no. 1, pp. 209–210, 2000.
- [157] E. Anton, A. Otegui, and A. Alonso, "Meningoencephalitis and *Chlamydia pneumoniae* infection," *European Journal of Neurology*, vol. 7, no. 5, p. 586, 2000.
- [158] L. Airas, P. Kotilainen, R. Vainionpää, and R. J. Marttila, "Encephalitis associated with *Chlamydia pneumoniae*," *Neurology*, vol. 56, no. 12, pp. 1778–1779, 2001.
- [159] C. Boschín-Crinquette, A. Kreisler, L. Legout, P. Charpentier, A. Destée, and L. Defebvre, "Can meningoencephalitis be caused by *Chlamydia pneumoniae*?" *Revue Neurologique*, vol. 161, no. 10, pp. 979–983, 2005.
- [160] L. Wing, S. R. Leekam, S. J. Libby, J. Gould, and M. Larcombe, "The diagnostic interview for social and communication disorders: background, inter-rater reliability and clinical use," *Journal of Child Psychology and Psychiatry and Allied Disciplines*, vol. 43, no. 3, pp. 307–325, 2002.
- [161] S. E. Folstein and B. Rosen-Sheidley, "Genetics of autism: complex aetiology for a heterogeneous disorder," *Nature Reviews Genetics*, vol. 2, no. 12, pp. 943–955, 2001.
- [162] J. Veenstra-vanderweele, E. H. Cook Jr., and P. J. Lombroso, "Genetics of childhood disorders: XLVI. Autism, part 5: genetics of autism," *Journal of the American Academy of Child and Adolescent Psychiatry*, vol. 42, no. 1, pp. 116–118, 2003.
- [163] H. Takahashi, S. Arai, K. Tanaka-Taya, and N. Okabe, "Autism and infection/immunization episodes in Japan," *Japanese Journal of Infectious Diseases*, vol. 54, no. 2, pp. 78–79, 2001.
- [164] Y. Yamashita, C. Fujimoto, E. Nakajima, T. Isagai, and T. Matsuishi, "Possible association between congenital cytomegalovirus infection and autistic disorder," *Journal of Autism and Developmental Disorders*, vol. 33, no. 4, pp. 455–459, 2003.
- [165] J. E. Libbey, T. L. Sweeten, W. M. McMahon, and R. S. Fujinami, "Autistic disorder and viral infections," *Journal of NeuroVirology*, vol. 11, no. 1, pp. 1–10, 2005.
- [166] D. H. Thornton, "A survey of *mycoplasma* detection in veterinary vaccines," *Vaccine*, vol. 4, no. 4, pp. 237–240, 1986.
- [167] J. K. S. Chia and L. Y. Chia, "Chronic *Chlamydia pneumoniae* infection: a treatable cause of chronic fatigue syndrome," *Clinical Infectious Diseases*, vol. 29, no. 2, pp. 452–453, 1999.
- [168] R. Nicolson and P. Szatmari, "Genetic and neurodevelopmental influences in autistic disorder," *Canadian Journal of Psychiatry*, vol. 48, no. 8, pp. 526–537, 2003.
- [169] W. Huang, D. See, and J. Tiles, "The prevalence of *Mycoplasma incognitus* in the peripheral blood mononuclear cells of normal controls or patients with AIDS or chronic fatigue syndrome," *Journal of Clinical Microbiology*, vol. 231, pp. 457–467, 1998.
- [170] J. Nijs, G. L. Nicolson, P. De Becker, D. Coomans, and K. De Meirleir, "High prevalence of *Mycoplasma* infections among European chronic fatigue syndrome patients. Examination of four *Mycoplasma* species in blood of chronic fatigue syndrome patients," *FEMS Immunology and Medical Microbiology*, vol. 34, no. 3, pp. 209–214, 2002.
- [171] G. L. Nicolson, R. Gan, and J. Haier, "Multiple co-infections (*Mycoplasma*, *Chlamydia*, human herpes virus-6) in blood of chronic fatigue syndrome patients: association with signs and symptoms," *APMIS: Acta Pathologica, Microbiologica et Immunologica Scandinavica*, vol. 111, no. 5, pp. 557–566, 2003.
- [172] G. L. Nicolson, "Chronic bacterial and viral infections in neurodegenerative and neurobehavioral diseases," *Laboratory Medicine*, vol. 39, no. 5, pp. 291–299, 2008.
- [173] G. Campadelli-Fiume, P. Mirandola, and L. Menotti, "Human herpesvirus 6: an emerging pathogen," *Emerging Infectious Diseases*, vol. 5, no. 3, pp. 353–366, 1999.
- [174] D. Keen and S. Ward, "Autistic spectrum disorder: a child population profile," *Autism*, vol. 8, no. 1, pp. 39–48, 2004.
- [175] B. Fellerhoff, B. Laumbacher, and R. Wank, "High risk of schizophrenia and other mental disorders associated with chlamydial infections: hypothesis to combine drug treatment and adoptive immunotherapy," *Medical Hypotheses*, vol. 65, no. 2, pp. 243–252, 2005.