Research Article

Bacterial Contamination of Clothes and Environmental Items in a Third-Level Hospital in Colombia

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Objective. This study evaluates the bacterial contamination rate of items in the hospital setting that are in frequent contact with patients and/or physicians. By determining the bacterial species and the associated antibiotic resistance that patients are exposed to. *Methods.* Hospital-based cross-sectional surveillance study of potential bacterial reservoirs. Cultures from 30 computer keyboards, 32 curtains, 40 cell phones, 35 white coats, and 22 ties were obtained. *Setting.* The study was conducted an urban academic 650-bed teaching hospital providing tertiary care to the city of Medellin, Colombia. *Results.* In total, 235 bacterial isolates were obtained from 159 surfaces sampled. 98.7% of the surfaces grew positive bacterial cultures with some interesting resistance profiles. *Conclusion.* There are significant opportunities to reduce patient exposure to frequently pathogenic bacteria in the hospital setting; patients are likely exposed to many bacteria through direct contact with white coats, curtains, and ties. They may be exposed to additional bacterial reservoirs indirectly through the hands of clinicians, using computer keyboards and cell phones.

1. Background

Antibiotic-resistant bacteria are implicated in an increasing amount of hospitalized patient infections worldwide. Among patients diagnosed with an infection, antibiotic resistance is associated with an increased length of hospital stay, health care costs, and patient morbidity, and mortality. Improved hand hygiene, environmental cleaning, and isolation of patients carrying pathogenic bacteria are the main methods for tackling the problem. Despite clear evidence that hygiene improves surgical outcomes, there remains considerable controversy over whether or not contaminated environmental surfaces contribute to transmission of healthcareassociated pathogens [1-8]. The risk of nosocomial infection depends on a number of factors. These include the ability of pathogens to remain viable on a surface, the rate at which contaminated surfaces are touched by patients and healthcare workers, the context in which the patient is exposed, and the levels of contamination that result in transmission to patients. Recent studies suggest that contaminated environmental surfaces may play an important role in transmission of healthcare-associated pathogens [9–23]. Clothing including white coats appears to be contaminated in the first several hours of use [24]. Other personnel effects with frequent hand contact such as pens, stethoscopes, and cell phones may have even higher levels of contamination [25].

This study demonstrates how cloth (white coats, curtains, and ties), computer keyboards, and cell phones may act as reservoirs for bacterial pathogens that may be associated with healthcare-associated infections.

2. Methods

2.1. Setting. The study was conducted in Hospital Universitario San Vicente Fundacion, an urban academic 650-bed teaching hospital providing tertiary care to the city of Medellin, Colombia. HCWs were randomly approached during routine daily patient care, and representative surfaces were randomly sampled during typical weekdays.

		Potentially clinically relevant microorganisms									Potentially clinically irrelevant microorganisms	
Type of surface	Number of samples per	Number of	Meticillin-resistant Staphylococcus sp.*		Meticillin-sensible Staphylococcus sp.+		Enterococcus sp.		Gram-negative rods [†]		Bacillus sp.	
	hospital area	Isolates	п	%	п	%	п	%	п	%	п	%
	Emergency room $(n = 7)$	8	0	0	3	37,5	0	0	2	25	3	37,5
	Adult surgical ICU $(n = 6)$	8	0	0	6	75	2	25	0	0	0	0
Keyboards	Adult medical ICU $(n = 6)$	9	0	0	3	33,3	1	11,1	1	11,1	4	44,4
(n = 30)	Adult special care unit (n = 6)	9	1	11,1	2	22,2	0	0	0	0	6	66,6
	Internal medicine ward $(n = 5)$	5	1	20	0	0	0	0	0	0	4	80
	Overall	39	2	5,1	14	35,9	3	7,7	3	7,7	17	43,5

TABLE 1: Distribution of bacterial isolates from keyboards.

* S. epidermidis. + S. epidermidis, S. aureus, S. warneri, S. haemolyticus. ^One was a E. faecium resistant to vancomycin. † Pantoea agglomerans, Escherichia hermannii, Leclercia adecarboxylata.

2.2. Sample Collection and Bacteriological Analysis. Samples were randomly collected from 30 keyboards, 32 curtains, 40 cell phones, 35 white coats, and 22 ties. At the time of the study, no active investigation was being performed for a nosocomial pathogen.

Curtains were sampled in a standardized aseptic fashion. The examiner first washed his hands and then put on a sterile surgical glove then swabbed the glove along a 25 cm² area on the lateral edge of the middle section of the curtain, because this is the area that HCWs most often contact with their hands when opening or closing the curtains. A hand imprint of the surgical glove was immediately printed onto a plate with blood agar for culture.

Ties and white coats were sampled in similar aseptic fashion by swabbing a sterile surgical glove along the whole cloth and then placing the glove onto a plate with blood agar for culture.

Keyboards and cell phones were sampled in a standardized aseptic fashion with sterile cotton-tipped applicators moisturized with Brain-Heart Infusion (BHI) liquid media. Then, the applicators were immediately used to inoculate BHI liquid transport media and sent directly to the laboratory for further procedures.

All liquid cultures were incubated for 24 hours at 35.5° C and then streaked on solid media culture plates, which were incubated for 48 hours at 35.5° C.

All isolates were Gram-stained, identification of the species and antibiotic resistance was performed by a Vitek Gram-positive and Gram-negative card (*bioMérieux SA*, *Marcy l'Etoile, France*) according to the manufacturer's recommendations.

3. Results

In June 2011, a total of 159 samples were collected from 30 keyboards, 32 curtains, 40 cell phones, 35 white coats, and

22 ties. From all surfaces, 98.7% had bacterial contamination, and a total of 235 unique colonies were obtained.

3.1. Keyboards. From 30 keyboards sampled, a total of 39 isolations were obtained, from those, 22 (56.4%) were considered potentially clinically relevant (Table 1), highlighting bacteria as *Escherichia hermannii*, Methicillin-resistant *S. epi-dermidis* (MRSE), *Enterococcus faecalis, Pantoea agglomerans,* and Vancomycin-resistant *Enterococcus faecium*.

3.2. Curtains. From 32 curtains sampled, a total of 59 isolations were obtained, from those, 47 (79.6%) were considered potentially clinically relevant (Table 2), highlighting bacteria as Methicillin-resistant *S. haemolyticus* (MRSH), Methicillin-resistant *S. cohnii* (MRSC), MRSE, Methicillin-resistant *S. saprophyticus* (MRSS), Moraxella sp., Acineto-bacter ursingii, AMP-C producer Pseudomonas oryzihabitans, Pantoea agglomerans, and Sphingomonas paucimobilis.

3.3. Cell Phones. From 40 cell phones sampled, a total of 58 isolations were obtained, from those, 51 (88%) were considered potentially clinically relevant (Table 3), highlighting bacteria as MRSH, MRSC, MRSE, Methicillin-resistant *S. hominis* (MRSh), *Pantoea agglomerans, Acinetobacter lwoffii* and *Sphingomonas paucimobilis.*

3.4. White Coats. From 35 with coats sampled, a total of 52 isolations were obtained, from those, 39 (75%) were considered potentially clinically relevant (Table 4), highlighting bacteria as *Pseudomonas oryzihabitans*, MRSE, MRSH, MRSh, and *Moraxella* sp.

3.5. Ties. From 22 ties sampled, a total of 27 isolations were obtained, from those, 18 (66.6%) were considered

Type of surface	Number of samples per Hospital area		Potentially clinically relevant microorganisms							ntially irrelevant rganisms
		Number of Isolates	Meticillin-resistant Staphylococcus sp.*		Meticillin-sensible <i>Staphylococcus</i> sp. ⁺		Gram-negative rods [†]		Bacillus sp.	
			п	%	n	%	п	%	n	%
	Emergency room $(n = 8)$	13	4	30,8	6	46,1	1	7,7	2	15,3
Curtains (<i>n</i> = 32)	Adult surgical ICU $(n = 5)$	8	4	50	3	37,5	0	0	1	12,5
	Adult medical ICU $(n = 2)$	2	1	50	1	50	0	0	0	0
	Adult special care unit $(n = 6)$	14	3	21,4	4	28,5	1	7,1	6	42,8
	Internal medicine ward $(n = 11)$	22	6	27,2	7	31,8	6	27,2	3	13,6
	Overall	59	18	30,5	21	35,5	8	13,5	12	20,3

TABLE 2: Distribution of bacterial isolates from curtains.

* S. epidermidis, S. haemolyticus, S. cohnii, S. saprophyticus, S. hominis. ⁺S. epidermidis, S. aureus, S. cohnii, S. hominis, S. haemolyticus, S. warneri, S. sciuri, S. saprophyticus. [†]Acinetobacter ursingii, Pantoea agglomerans, Moraxella sp., Pseudomonas oryzihabitans AMP-C producer, Sphingomonas paucimobilis, Pasteurella multocida.

TABLE 3: Distribution of bacterial isolates from cell phones.

				Potentially	rganisms		clinically	ntially virrelevant rganisms		
Type of surface	Number of samples per	Number of	Meticillin-resistant Staphylococcus sp.*		Meticillin-sensible <i>Staphylococcus</i> sp.+		Gram-negative rods [†]		Bacillus sp.	
	doctor specialty	Isolates	п	%	п	%	п	%	п	%
	General $(n = 7)$	11	2	18,1	8	72,7	1	9,1	0	0
	Internal medicine $(n = 8)$	9	2	22,2	5	55,5	1	11,1	1	11,1
	Clinical resident $(n = 6)$	9	3	33,3	4	44,4	1	11,1	1	11,1
Cell	Surgery $(n = 3)$	6	3	50	3	50	0	0	0	0
phones (<i>n</i> = 40)	Surgery resident $(n = 4)$	4	1	25	2	50	0	0	1	25
	Medical student $(n = 9)$	15	0	0	9	60	2	13,3	4	26,6
	Nurse $(n = 1)$	1	0	0	1	100	0	0	0	0
	Nutritionist $(n = 2)$	3	0	0	3	100	0	0	0	0
	Overall	58	11	19	35	60,3	5	8,6	7	12

* S. epidermidis, S. cohnii, S. hominis, S. haemolyticus. ⁺S. epidermidis, S. aureus, S. haemolyticus, S. hominis, S. warneri, S. chromogenes. [†]Acinetobacterlwoffii, Pantoea agglomerans, Aeromonas salmonicida, Sphingomonas paucimobilis.

potentially clinically relevant (Table 5), highlighting bacteria as Methicillin resistant *S. aureus* and MRSE.

4. Discussion

The prevalence of antibiotic resistant bacteria is a serious problem with important implications for hospital infection control. Some studies have found bacterial contamination in the community (of cell phones) to be nearly equivalent to hospital settings [15]. Yet antibiotic resistant bacteria remain more common in hospital settings. Although the geographic distribution of these bacteria is worldwide, the epidemiology and dissemination patterns appear to differ within and across regions [1–8]. In this study, we found an alarming number of potentially clinically relevant bacteria colonizing different surfaces, these bacterial reservoirs are a plausible source of

			Potentially clinically relevant microorganisms							ntially irrelevant rganisms
Type of surface	Number of samples per	Number of	Meticillin-resistant Staphylococcus sp.*		Meticillin-sensible Staphylococcus sp.+		Gram-negative rods [†]		Bacillus sp.	
	doctor specialty	Isolates	п	%	п	%	п	%	n	%
	General $(n = 4)$	9	0	0	5	55,5	1	11,1	3	33,3
	Internal medicine $(n = 12)$	19	1	5,2	11	57,9	1	5,2	6	31,5
	Clinical resident $(n = 7)$	7	2	28,5	4	57	0	0	1	14,2
White	Surgery $(n = 5)$	7	1	14,2	5	71	0	0	1	14,2
coats (<i>n</i> = 35)	Surgery resident $(n = 4)$	5	1	20	2	40	2	40	0	0
	Medical student $(n = 2)$	3	0	0	2	66,6	0	0	1	33,3
	Nutritionist $(n = 1)$	2	0	0	1	50	0	0	1	50
	Overall	52	5	9,6	30	57,7	4	7,7	13	25

TABLE 4: Distribution of bacterial isolates from white coats.

* S. epidermidis, S. haemolyticus. S. hominis. + S. capitis, S. aureus, S. warneri, S. epidermidis. + Pseudomonas oryzihabitans AMP-C producer, Moraxella sp.

			Pote	entially clinically	Potentially clinically irrelevant microorganisms				
Type of surface	Number of samples per	Number of Isolates	Meticillin- Staphyloco		Meticillin-sensible Staphylococcus sp. ⁺		Bacillus sp.		
	doctor specialty		п	%	п	%	п	%	
	General $(n = 4)$	6	0	0	4	66,6	2	33,3	
Ties (<i>n</i> = 22)	Internal medicine $(n = 10)$	13	2	15,3	6	46,1	5	38,4	
	Surgery $(n = 6)$	6	1	16,6	4	66,6	1	16,6	
	Medical student $(n = 2)$	2	0	0	1	50	1	50	
	Overall	27	3	11,1	15	55,5	9	33,3	

TABLE 5: Distribution of bacterial isolates from ties.

* S. aureus, S. epidermidis. + S. epidermidis, S. hominis, S. warneri, S. aureus.

infection for patients at this tertiary level hospital and likely any other hospital worldwide.

The most important implication of our study is to highlight the role of these items as bacterial reservoirs and how HCWs should perform hand hygiene after contact with any clothes or environmental item in agreement with the recommendation of the guideline on hand hygiene in healthcare settings [1, 3]. Some other strategies to reduce the potential for transmission of pathogens from this surfaces include improved or more frequent cleaning [4, 6–8].

In contrast to previous studies on the role of environmental colonization that were performed during nosocomial pathogen outbreaks [21, 24], our study was conducted when there was no outbreak and reflects the regular daily risk of colonization or infection from hospital fomites. Bacterial contamination of items in health care settings is likely ongoing as organisms such as *Staphylococci, E. coli*, and *P. aeruginosa* survive at least 3–6 months on dried blood or cotton and as long as four weeks on other surfaces [26, 27]. Unfortunately, we did not investigate other factors in the transmission route, such as HCWs' hand carriage and colonization of patients.

5. Conclusion

This hospital-based cross-sectional surveillance study demonstrates that a large proportion of health care workers' clothing and personal effects were contaminated with bacterial pathogens that can result in nosocomial infections. Further research is needed to evaluate strategies to minimize the risk of patient-to-patient transmission of pathogens from other contaminated items.

Conflict of Interests

All the authors report no conflict of interests relevant to this paper.

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