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Data Article

# Synthesis of practical red fluorescent probe for cytoplasmic calcium ions with greatly improved cell-membrane permeability



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## ABSTRACT

In this data article, we described the detailed synthetic procedure and the experimental data for the synthesis of a red-fluorescent probe for calcium ions ( $Ca^{2+}$ ) with improved water solubility. This  $Ca^{2+}$  red-fluorescent probe CaTM-3 AM could be applied to fluorescence imaging of physiological  $Ca^{2+}$  concentration changes in not only live cells, but also brain slices, with high cell-membrane permeability leading to bright fluorescence in biosamples. The data provided herein are in association with the research article "The Development of Practical Red Fluorescent Probe for Cytoplasmic Calcium Ions with Greatly Improved Cell-membrane Permeability" in Cell Calcium (Hirabayashi et al., 2016) [1].

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Subject area	Chemistry
More specific sub- ject area	Synthesis of fluorescent probes
Type of data	Synthetic schemes, experimental synthesis protocols, NMR and MS spectra, HPLC chromatogram
How data was acquired	NMR: JNM-LA300 (JEOL) or JNM-LA400 (JEOL), mass spectroscopy: JMS-T100LC AccuTOF (JEOL), HPLC analyses: Inertsil ODS-3 (4.6 × 250 mm) column (GL Sciences Inc.) using a HPLC system composed of a pump (PU-2080, JASCO) and a detector (MD-2018 or FP-2025, JASCO)
Data format	Analyzed
Experimental factors	Starting compounds were either purchased or synthesized using already pub- lished synthetic protocols
Experimental features	Compounds were synthesized and their structures were identified by NMR and mass spectrometry
Data source location	Tokyo, Japan
Data accessibility	Data are provided with this article

## Specifications table

## Value of the data

- The data allows to reproduce the experiments described in the research article Ref. [1].
- The synthesized compound, CaTM-3 AM, could be used to fluorescence imaging of the physiological Ca<sup>2+</sup> concentration change.
- The data provides an opportunity to synthesize red fluorescent probes based on TokyoMagenta (TM) derivatives.

## 1. Data

NMR spectra were recorded on a JEOL JNM-LA300 instrument at 300 MHz for <sup>1</sup>H NMR and at 75 MHz for <sup>13</sup>C NMR or JEOL JNM-LA400 instrument at 100 MHz for <sup>13</sup>C NMR. Mass spectra (ESI<sup>+</sup>) were measured with a JEOL JMS-T100LC AccuTOF for ESI. HPLC analyses were performed on an Inertsil ODS-3 ( $4.6 \times 250$  mm) column (GL Sciences Inc.) using a HPLC system composed of a pump (PU-2080, JASCO) and a detector (MD-2018 or FP-2025, JASCO).

#### 2. Experimental design, materials and methods

#### 2.1. Synthetic materials and instrumentation

General chemicals were of the best grade available, supplied by Tokyo Chemical Industries, Wako Pure Chemical, Aldrich Chemical Co., Kanto Chemical Co., Inc., Toronto Research Chemicals Inc., Watanabe Chemical Industries, Applied Biosystems, Dojindo and Invitrogen Corp., and were used without further purification. All solvents were used after appropriate distillation or purification. Preparative HPLC were performed on an Inertsil ODS-3 ( $10 \times 250$  mm) column (GL Sciences Inc.) using a HPLC system composed of a pump (PU-2080, JASCO) and a detector (MD-2015 or FP-2025, JASCO) or a HPLC system composed of a pump (PU-2086, JASCO) and a detector (MD-2018, JASCO).

#### 2.2. Synthesis and characterization of compounds

#### 2.2.1. Synthesis of compound 1



This compound was synthesized according to Ref. [2].

## 2.2.2. Synthesis of di-tert-butyl 4-bromoisophthalate (2)



To a mixture of 4-bromoisophthalic acid (4.90 g, 20.0 mmol), *N*,*N*-dicyclohexylcarbodiimide (10.0 g, 46.5 mmol) and *N*,*N*-dimethylaminopyridine (500 mg, 4.10 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (100 mL), *tert*-butyl alcohol (50 mL) was added. The mixture was stirred at room temperature overnight. The precipitate was filtered off, and the filtrate was evaporated to dryness. The residue was purified by column chromatography (silica gel, 1/1 CH<sub>2</sub>Cl<sub>2</sub>/hexane) to give di-*tert*-butyl 4-bromoisophthalate (**2**) (4.32 g, 61% yield). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  1.60 (s, 9H), 1.62 (s, 9H), 7.67 (d, 1H, *J* = 8.1 Hz), 7.86 (dd, 1H, *J* = 8.1, 2.2 Hz), 8.25 (d, 1H, *J* = 2.2 Hz); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  28.1, 81.9, 83.0, 125.7, 131.1, 131.6, 132.2, 134.1, 134.4, 164.3, 165.1; HRMS (ESI<sup>+</sup>): Calcd for [M+Na]<sup>+</sup>, 379.0521; Found, 379.0552 (+3.1 mmu).

2.2.3. Synthesis of 2,4-diCOOH DCTM



To a flame-dried flask flushed with argon, di-*tert*-butyl 4-bromoisophthalate (**2**) (357 mg, 1.00 mmol) and anhydrous THF (5 mL) were added. The solution was cooled to -78 °C, and 1 M sec-BuLi (0.70 mL, 0.70 mmol) was slowly added to it. The mixture was stirred at -78 °C for 10 seconds, then compound **1** (50.0 mg, 0.0705 mmol) dissolved in anhydrous THF (5 mL) was slowly added. The resulting mixture was warmed to room temperature, then stirred for 1 h, and 2 N HCl aq. (2.0 mL) was added to it. Stirring was continued for 20 min, then the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated to dryness. The residue was dissolved in TFA (5.0 mL) and the solution was stirred for 1 h, then evaporated to dryness, and the resulting residue was purified by HPLC to give 2,4-diCOOH DCTM (18.1 mg, 53% yield). <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD):  $\delta$  0.84 (s, 3H), 0.99 (s, 3H), 6.87 (d, 2H, *J*=8.8 Hz), 6.91 (d, 2H, *J* = 8.8 Hz), 7.05 (d, 1H, *J* = 8.1 Hz), 8.22 (dd, 1H, *J* = 8.1, 1.5 Hz), 8.49 (d, 1H, *J* = 1.5 Hz); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD):  $\delta$  -0.1, 0.5, 91.4, 119.7, 124.0, 124.9, 127.8, 128.0, 128.3, 133.5, 135.6, 135.9, 137.5, 154.4, 162.3, 167.7, 171.9; HRMS (ESI<sup>+</sup>): Calcd for [M+H]<sup>+</sup>, 487.0171, Found, 487.0186 (+1.5 mmu).

2.2.4. Synthesis of 5-amino-BAPTA-tetramethylester (3)



5-Amino-BAPTA-tetramethylester (3) was synthesized according to reference [3,4].

2.2.5. Synthesis of CaTM-3



To a solution of 5-amino-BAPTA-tetramethylester (**3**) (27.4 mg, 0.0500 mmol) in DMF (3.0 mL), HATU (37.0 mg, 0.100 mmol), HOBt (15.3 mg, 0.100 mmol) and 2,4-diCOOH DCTM (9.8 mg, 0.020 mmol) were added. The mixture was stirred at room temperature overnight, then 2 N HCl aq. (2 mL) was added to it. The whole was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated to dryness. Then, 2 N NaOH aq. (3.0 mL) and MeOH (3.0 mL) were added to the residue. The mixture was stirred at room temperature for 4 h, then neutralized with 2 N HCl aq., and purified by HPLC to give CaTM-3 (4.6 mg, 24% yield). <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD):  $\delta$  0.84 (s, 3H), 1.00 (s, 3H), 3.74 (s, 4H), 3.77 (s, 4H), 4.38 (s, 4H), 6.84–6.94 (m, 9H), 7.09 (d, 1H, *J* = 8.1 Hz), 7.33 (d, 1H, *J* = 7.3 Hz), 7.39 (s, 1H), 8.14 (d, 1H, *J* = 8.1 Hz), 8.46 (s, 1H); HRMS (ESI<sup>+</sup>): Calcd for [M + H]<sup>+</sup>, 960.1606, Found, 960.1616 (+1.0 mmu). HPLC analysis: retention time 13.7 min (purity, 99.7% integrated intensity); eluent: A (H<sub>2</sub>O, 0.1 M TEAA (triethylammonium acetate)), B (80% acetonitrile/H<sub>2</sub>O, 0.1 M TEAA); gradient: A : B = 80 : 20 to 0 : 100 (15 min); flow rate: 1.0 mL/min; detection wavelength, 280 nm.

2.2.6. Synthesis of 5-amino-BAPTA-tetraacetoxymethylester (4)



AM = acetoxymethyl ester

5-Amino-BAPTA-tetraacetoxymethylester (4) was synthesized according to Ref. [2].







Scheme 2. Synthetic scheme for CaTM-3.



Scheme 3. Synthetic Scheme for CaTM-3 AM (AM = acetoxymethyl ester).

#### 2.2.7. Synthesis of CaTM-3 AM



AM = acetoxymethyl ester

To a solution of 5-amino-BAPTA-tetraacetoxymethylester (**4**) (31.2 mg, 0.0400 mmol) in DMF (2.0 mL), HATU (18.5 mg, 0.0500 mmol), HOBt (7.7 mg, 0.050 mmol) and 2,4-diCOOH DCTM (9.8 mg, 0.020 mmol) were added. The mixture was stirred at room temperature overnight, then neutralized with AcOH aq., and purified by HPLC to give CaTM-3 AM (8.6 mg, 34% yield). <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD):  $\delta$  0.84 (s, 3H), 1.00 (s, 3H), 2.00 (s, 6H), 2.02 (s, 6H), 4.17 (s, 8H), 4.30 (s, 4H), 5.58 (s, 4H), 5.59 (s, 4H), 6.83–7.09 (m, 9H), 7.08 (d, 1H, *J* = 8.1 Hz), 7.17 (dd, 1H, *J* = 8.8, 2.2 Hz), 7.46 (d, 1H, *J* = 2.2 Hz), 8.11 (dd, 1 H, *J* = 8.1, 1.5 Hz), 8.44 (s, 1 H); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD):  $\delta$  -0.1, 0.5, 20.5, 54.6, 54.7, 68.6, 68.7, 80.6, 80.6, 91.4, 108.5, 114.9, 115.0, 119.7, 120.7, 120.8, 122.5, 123.8, 124.1, 124.9, 126.1, 127.9, 127.9, 134.8, 135.8, 135.9, 137.1, 137.8, 140.21, 152.1, 154.4, 161.2, 166.5, 171.1, 171.7, 177.8, 172.1; HRMS (ESI<sup>+</sup>): Calcd for [M + H]<sup>+</sup>, 1248.2451, Found, 1248.2420 (-3.1 mmu). (Schemes 1–3).

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#### Transparency document. Supporting information

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