1 2 3 4 5	Repeated presentation of visual threats drives innate fear habituation and is modulated by environmental and physiological factors
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# 44 Abstract:

45 To survive predation, animals must be able to detect and appropriately respond to predator 46 threats in their environment. Such defensive behaviors are thought to utilize hard-wired neural 47 circuits for threat detection, sensorimotor integration, and execution of ethologically relevant 48 behaviors. Despite being hard-wired, defensive behaviors (i.e. fear responses) are not fixed, but 49 rather show remarkable flexibility, suggesting that extrinsic factors such as threat history, 50 environmental contexts, and physiological state may alter innate defensive behavioral 51 responses. The goal of the present study was to examine how extrinsic and intrinsic factors 52 influence innate defensive behaviors in response to visual threats. In the absence of a 53 protective shelter, our results indicate that mice showed robust freezing behavior following both 54 looming (proximal) and sweeping (distal) threats, with increased behavioral vigor in response to 55 looming stimuli, which represent a higher threat imminence. Repeated presentation of looming 56 or sweeping stimuli at short inter-trial intervals resulted in robust habituation of freezing, which 57 was accelerated at longer inter-trial intervals, regardless of contextual cues. Finally, 58 physiological factors such as acute stress further disrupted innate freezing habituation, resulting 59 in a delayed habituation phenotype, consistent with a heightened fear state. Together, our 60 results indicate that extrinsic factors such as threat history, environmental familiarity, and 61 physiological stressors have robust and diverse effects on defensive behaviors, highlighting the

62 behavioral flexibility in how mice respond to predator threats.

63

### 64 Introduction:

65 The ability of animals to accurately and appropriately respond to predator threats in the 66 environment is critical for survival. As such, antipredator defensive behaviors are observed 67 across evolutionary history (D. C. Blanchard & Blanchard, 2008; Kavaliers & Choleris, 2001; 68 Kikuchi et al., 2023; LeDoux, 2012). Such antipredator defensive behaviors form the basis of 69 unconditioned or innate fear responses, which are distinct from conditioned fear, in that they do 70 not require previous associative learning; suggesting the presence of hard-wired, dedicated 71 neural circuitry for the detection, integration, and execution of appropriate behavioral responses 72 (Carrive, 1993; De Franceschi et al., 2016; Evans et al., 2018; Keay & Bandler, 2001; LeDoux, 73 2012; B. A. Silva et al., 2016; Yilmaz & Meister, 2013; Zhang et al., 1990). For example, mice 74 show a dynamic repertoire of defensive behaviors which are differentially engaged depending 75 on the nature of the threat (De Franceschi et al., 2016; Fanselow, 1991, 1994; Tafreshiha et al., 76 2021). These observations have informed the development of the threat imminence model, in 77 which specific sensory stimuli are ethologically matched to appropriate defensive behaviors 78 (Bolles, 1970; Fanselow, 2018; Fanselow & Lester, 1988; Perusini & Fanselow, 2015). For 79 instance, sweeping visual stimuli that mimic a distal aerial predator engage freezing behaviors 80 to avoid detection. Conversely, looming visual stimuli, which mimic a proximal aerial threat, 81 engage more active defensive strategies such as flight to a shelter (De Franceschi et al., 2016; 82 Liu et al., 2022; Solomon et al., 2023; Yilmaz & Meister, 2013)

83 Consistent with the threat imminence theory, freezing and flight behaviors are thought to be 84 differentially engaged by distinct rostro-caudal columns in the midbrain periaqueductal gray 85 (Bandler et al., 1985, 2000; Bandler & Shipley, 1994; Carrive, 1993; Keay & Bandler, 2001; 86 Toyote et al., 2016; Zhang et al., 1990). More specifically, activation of the ventrolateral column 87 of the periaqueductal gray (vIPAG) results in robust freezing behaviors, whereas activation of 88 the dorsolateral periaqueductal gray (dIPAG) results in active avoidance strategies, such as 89 flight (Bandler & Shipley, 1994; Carrive, 1993; La-Vu et al., 2022; Tovote et al., 2016; Vaaga et 90 al., 2020; Zhang et al., 1990). Despite this theoretical and neural framework, innate fear 91 behaviors are not fixed responses, and therefore may be modulated by environmental and 92 physiological variables. For example, looming threats can elicit *freezing* in experimental 93 conditions without a protective shelter (De Franceschi et al., 2016; Yilmaz & Meister, 2013). 94 This observation raises the possibility that other environmental factors, such as threat history, 95 environmental familiarity, or physiological factors, such as exposure to acute stress, may 96 similarly alter innate fear responses (Hassien et al., 2020; Lenzi et al., 2022; Perusini &

97 Fanselow, 2015; Rau et al., 2005; Tafreshiha et al., 2021). However, one limitation of the threat 98 imminence model is that behavioral variables such as response vigor are often inferred by the 99 defensive strategy employed, limiting direct comparisons. As such, understanding how environmental and physiological variables impact innate fear behavior has been difficult to 101 assess. Of particular interest is how such variables contribute to behavioral flexibility, as 102 inflexible fear responses are observed in disorders such as post-traumatic stress disorder 103 (PTSD; Friedman et al., 2011; Iqbal et al., 2023; Koenen et al., 2017)

104 To begin to understand how intrinsic and extrinsic factors influence innate fear and behavioral 105 flexibility, we exposed mice to looming and sweeping threats in an arena without a shelter, in an 106 attempt to limit the available defensive behavioral repertoire. We demonstrate that under such 107 conditions, both sweeping and looming threats engage immobility behavior, although threat 108 imminence is still encoded by response vigor (i.e. freezing duration). Repeated threat 109 presentation resulted in a progressive reduction in immobility (see also (Lenzi et al., 2022), 110 independent of the nature of the visual stimulus. Furthermore, the rate, but not degree, of 111 habituation significantly varied with changes in environmental condition and/or physiological 112 stressors, suggesting that threat habituation is a key variable in the innate fear response.

# 113 Methods:

114 Ethical Note: All experimental procedures were conducted in accordance with institutional 115 guidelines regarding the ethical use of animals. All experimental methods were approved by 116 Northwestern University (protocol IS00014844, IMR) and Colorado State University Institutional 117 Animal Care and Use Committees (protocol 3836, CEV). The study utilized (6-12 week) adult 118 wild-type mice, purchased from a commercial supplier (Jackson Laboratories). Experiments 119 involved non-invasive behavioral observations of animals exposed to visual stimuli mimicking 120 predators. All reasonable efforts were made to increase scientific transparency and openness. 121 All original data, python code, and digital research materials are available upon reasonable 122 request. The study design and analysis were not pre-registered.

123 *Animals*. Adult male and female C57BI6/J wild type mice were used for the study, in sex

balanced cohorts. Cohorts of 10 wildtype mice (5 male and 5 female) were purchased from

125 Jackson Laboratories (Strain: 000664) at 4-6 weeks of age and allowed to recover from

126 transport stress in the animal facility for at least 2 weeks prior to behavioral testing. Mice were

socially housed (2-5 mice per cage) on a 12:12 hour light:dark cycle with *ad libitum* access tofood and water.

- 129 At least 1 week prior to behavioral testing, mice were transported to a satellite housing facility 130 located in the same building as the behavioral testing suite to reduce daily transport stress. 131 Much of the data was collected during the animal light cycle, although in a subset of cohorts. 132 testing was performed during the animal dark cycle. No differences in behavior were observed 133 across the light cycle, so data were pooled. To reduce potential circadian effects on arousal and 134 behavioral responses, animals tested during the light cycle were allowed to acclimate to the 135 dark behavioral testing suite for at least 30 minutes prior to testing. Unless otherwise noted, two 136 days prior to behavioral testing, all animals underwent at least 2 days of handling and 137 behavioral familiarization in the experimental chamber for at least 10 minutes each day. 138 Following familiarization trials, mice were placed in a temporary holding cage before all mice 139 were returned to their home cage.
- 140 Innate Fear Paradigm: The experimental
- 141 setup consisted of a 25 x 25 x 25 cm
- 142 acrylic behavioral chamber with 3 light grey
- 143 walls and 1 transparent wall to facilitate
- 144 video recordings of animal behavior. Visual
- 145 stimuli were presented using an LCD
- 146 monitor placed 40 cm above the arena
- 147 floor. The sweeping visual stimulus (Figure
- 148 1B, left) consisted of a high contrast disk
- 149 (5° visual angle) which traversed the
- 150 screen and returned to its original position
- 151 in a total of 8 seconds (De Franceschi et
- al., 2016). The looming visual stimulus
- 153 (Figure 1B, right) consisted of a high
- 154 contrast rapidly expanding disk which
- 155 expanded to 20° visual angle in 333 ms
- and repeated 5 times in a total of 6
- 157 seconds (Yilmaz & Meister, 2013).



**Figure 1: Experimental Methodology.** (A) Timeline of typical experiment. Mice were familiarized with the behavioral arena for at least two days prior to behavioral testing. (B) Schematics illustrating the sweeping (left) and looming (right) visual stimuli used to elicit innate fear responses. (C) Diagram of the behavioral chamber, indicating relative position of the stimulus display and video camera. (D) Exemplar output from DeepLabCut illustrating average animal position within the behavioral chamber during the baseline period and the 10 seconds immediately after presentation of a looming stimulus. The data point from each frame is pseudo-colored by velocity.

158 Both the stimulus presentation and video acquisition were controlled using custom-written 159 python modules and a raspberry pi system. Briefly, videos were recorded using an infrared 160 raspberry pi camera module at 15 or 30 fps. Because the recordings were done in the dark, the 161 only ambient light was from the overhead monitor. Additional infrared lights were used to evenly 162 illuminate the behavioral arena. Visual stimuli were manually triggered using a Master-8 163 programmable pulse generator. Stimuli were triggered after 2-3 minutes of baseline activity to 164 allow the mice to refamiliarize themselves with the behavioral chamber. Furthermore, attempts 165 were made to trigger the stimuli during periods of movement, to adequately capture freezing 166 behaviors. For most experiments, mice were exposed to three identical stimuli in a single 167 behavioral session, separated by an inter-trial interval of ~5 minutes. In some experiments, the 168 inter-trial interval was increased to 24 hours, to test longer term behavioral habituation.

169 Acute Stress Paradigm: To test for effects of acute stress on innate fear, mice were exposed to 170 a modified stress-enhanced fear learning paradigm (Hassien et al., 2020; Perusini et al., 2016; 171 Rau et al., 2005; Rau & Fanselow, 2009). After 2 days of familiarization in the open field arena, 172 mice were placed in a novel context fear conditioning chamber to undergo an acute stress 173 paradigm. After approximately 1 minute, mice were given 4 unconditioned, unpredictable foot 174 shocks (2 sec duration, 1 mA) at an interval of 60-80 seconds (Hassien et al., 2020). Mice were 175 then allowed to recover for either 1 hour or 24 hours before being placed in the open field arena 176 for innate fear testing.

177 Data Analysis: Videos were initially analyzed using DeepLabCut marker-less pose estimation to 178 track animal position. Subsequent data analysis was performed using custom python code. 179 First, for each frame, the x- and y- position of the mouse's center of mass was identified (Figure 180 1D). Animal speed was calculated frame-by-frame by dividing the change in animal position by 181 the interval frame rate. Velocity data was smoothed using a rolling average across 10 frames, 182 and an immobility (freezing) epoch was defined as any 500 ms period in which the animal 183 velocity was less than 2 cm/sec. To facilitate data presentation, velocity traces were then filtered 184 to only show periods of immobility. Percent immobility was calculated within the 20 second 185 period after the onset of the visual stimulus.

Statistical Testing: Data are reported as mean±S.E.M unless otherwise noted. Data analysis
 and statistical testing was performed in GraphPad Prism software. Statistical comparisons
 between two groups were calculated using either a two-sample paired or unpaired t-test, as
 indicated in the text. For experiments in which we compared immobility across trials, data was

analyzed using a one-way repeated measures ANOVAs with a Tukey post-hoc comparison.
 Comparisons of normalized innate fear habituation were compared using a ordinary one-way
 repeated measures ANOVA. Datasets were compared using a two-way repeated measure
 ANOVA.

194 As stated above, all experiments were performed with sex-balanced cohorts, which, unless 195 otherwise indicated, were pooled together for analysis. To further reduce bias, animals were 196 randomly assigned to experimental cohorts. Data was analyzed using a pipeline to reduce 197 experimenter bias. The n values reported reflect the number of animals in each experiment. 198 Sample sizes were determined using a power analysis with preliminary data, which indicated 199 that a sample size of 10 mice per group was sufficient to detect a biologically relevant effect size 200 of ~20% with a statistical power ( $\beta$ ) of 0.8 and at a type I error rate ( $\alpha$ ) of 0.05. Exclusion criteria 201 included low baseline movement, which would occlude the ability to detect freezing behavior; 202 however, no animals were excluded from the dataset using this criterion. One animal was 203 excluded, as described in the text, from further analysis because its response was greater than

three times the standard deviation of the population response.

### 205 **Results:**

# 206 Freezing responses to sweeping and looming visual stimuli

207 Under experimental conditions where animals have access to a shelter, looming (proximal) and 208 sweeping (distal) threats engage distinct active and passive coping behavioral strategies, 209 respectively (De Franceschi et al., 2016; Evans et al., 2018; Tafreshiha et al., 2021; Yilmaz & 210 Meister, 2013). Such distinct behavioral responses to proximal vs. distal threats have precluded 211 testing whether and how mice differentially encode threat imminence via changes in response 212 vigor. We therefore sought to directly compare behavioral responses to looming and sweeping 213 visual threats under conditions in which the available defensive strategies are limited due to the 214 absence of a protective shelter.

To examine the behavioral response to distal threats, mice were exposed to a sweeping visual stimulus (De Franceschi et al., 2016). Consistent with previous results, mice exposed to sweeping stimuli engaged in passive avoidance strategies, namely immobility to avoid detection (Figure 2A). At the population level, mice showed a robust increase in immobility in the first 20 seconds after the onset of the sweeping stimulus (Figure 2B; baseline:  $12.7 \pm 3.4\%$  immobility; response:  $34.2 \pm 3.3\%$  immobility; paired t-test: p < 0.0003, t = 4.448, df = 19, n = 20 mice). We

- 221 observed no differences in stimulus-evoked immobility across sex (Figure 2C; males: 35.5 ±
- 222 6.0% immobility; females:  $33.0 \pm 2.9\%$  immobility; unpaired t-test: p = 0.71, t = 0.376, df = 18).
- 223 Despite the lack of sex differences in overall freezing responses, males showed a significantly
- more variable response to innate threat, contrary to behavioral observations in response to
- conditioned threat (Gruene et al., 2015; F test: p = 0.04, F = 4.33, df = 9).
- 226 In the absence of a 227 shelter, we reasoned that 228 looming threats may result 229 in one of two behavioral 230 responses. One possibility 231 is that mice will engage in 232 an un-directed darting 233 strategy, as has been 234 observed, preferentially in 235 females, in response to 236 conditioned footshocks 237 (Gruene et al., 2015). 238 Alternatively, mice may 239 respond to looming 240 threats with increased 241 immobility (De Franceschi 242 et al., 2016), in an attempt 243 to avoid detection. In a 244 separate cohort of mice, 245 animals responded to 246 looming threats with a 247 robust increase in 248 immobility (Figure 2D, E; 249 baseline: 16.8 ± 3.4% 250 immobility; response: 87.7 251 ± 2.3% immobility; paired 252 t-test: p < 0.0001, t = 253 15.75, df = 38), which was



**Figure 2: Freezing responses are elicited by looming and sweeping visual stimuli.** (A) Stimulus evoked immobility responses triggered by sweeping visual stimuli in male and female mice. Periods of immobility (red, velocity < 2 cm/sec) are indicated for each animal. For clarity, the analysis window (grey box) and stimulus duration (black bar) are also indicated. (B) Sweeping visual stimuli elicit a significant increase in immobility in the 20 seconds after stimulus presentation. (C) There was no difference in the immobility responses across sex. (D) Immobility responses to looming visual stimuli. (E,F) Looming stimuli significantly increased the percent time immobile (E) with no difference observed across sex (F). (G, H) Looming stimuli elicited more robust freezing behaviors, as measured by the percent time freezing (G) and the total time immobile (H). (I) Average velocity as a function of time for all animals for sweeping (left) and looming (right) stimuli. Looming stimulus: n=19 mice, 9 males, 10 females; Sweeping stimulus: n=20 mice, 10 males, 10 females.

254not significantly different between males and females (Figure 2F; males:  $85.5 \pm 3.8\%$  immobility;255females:  $88.8 \pm 2.6\%$  immobility; unpaired t-test: p = 0.48, t = 0.712, df = 37). These data256suggest that under behavioral conditions in which flight to a shelter is not possible, male and

- 257 female mice similarly engage in defensive *immobility* to both proximal and distal threats.
- 258 Interestingly, when directly comparing behavior across stimuli, we observed significantly more 259 immobility to the looming as compared to the sweeping stimulus (Figure 2G; sweeping: 34.2 ± 260 3.3% immobility, n = 20 mice; looming:  $87.2 \pm 2.3\%$  immobility; unpaired t-test: p < 0.0001, t = 261 13.48, df = 57). Considering that the % immobility measurement only considers the 20 second 262 window after stimulus presentation, we additionally calculated the total time each animal 263 engaged in immobility. This measurement accounts for immobility across the entire behavioral 264 trial (i.e. not limited to the first 20 seconds). This measurement similarly showed comparatively 265 more immobility in response to the looming stimulus, suggesting that the differences in 266 immobility were not restricted to the 20 seconds immediately following the stimulus (Figure 2H; 267 sweeping:  $9.9 \pm 0.9$  seconds, n = 20 mice; looming:  $35.0 \pm 2.7$  seconds, n = 19 mice, unpaired 268 t-test: p < 0.0001, t = 6.46, df = 57).

269 To more fully capture the dynamic responses of animals to threatening stimuli, we averaged the 270 velocity across animals thereby avoiding the categorical classification of animal behavior. In 271 agreement with the above data, looming stimuli resulted in a comparatively more robust and 272 prolonged decrease in velocity (Figure 2I). Interestingly, however, in response to both looming 273 and sweeping stimuli, mice showed a transient elevation in velocity prior to freezing (looming 274 stimulus:  $31.3 \pm 4.6$  cm/s; sweeping stimulus:  $20.1 \pm 2.8$  cm/s, n = 20 mice) which was not 275 significantly different across stimulus type (unpaired t-test: p = 0.10, t = 1.657, df = 57). This 276 transient increase in velocity may represent an initial orienting behavior to assess if an active 277 defensive strategy, such as flight, is a viable response (Evans et al., 2018). These data indicate 278 that in response to both proximal and distal innate threats, mice engage in the optimal 279 behavioral strategy available (in this case immobility), and that they encode threat imminence 280 (i.e. threat proximity) with a significant increase in behavioral vigor.





**Figure 3: Repeated threat presentation results in robust habituation of freezing behavior.** (A, B) Repeated presentation of sweeping (A) or looming (B) visual stimuli at inter-trial intervals as short as 5 minutes results in a gradual reduction in immobility across trials. (C, D) Quantification of immobility across trials for sweeping (C) and looming (D) stimuli. (E, F) Repeated presentation of sweeping and looming stimuli results in equivalent levels of total habituation (E) as well as similar rates of habituation across trials. (F) Normalized immobility responses across trials reveals linear change in immobility. Looming stimulus: n=19 mice, 9 males, 10 females; Sweeping stimulus: n=20 mice, 10 males.

282

# 283 Responses to repeated visual stimuli:

- Fear responses, including innate defensive behaviors, may be modulated by a variety of
- 285 intrinsic and extrinsic factors such as environmental context, threat history, and internal state
- 286 (De Franceschi et al., 2016; Hassien et al., 2020; Tafreshiha et al., 2021; Yilmaz & Meister,
- 287 2013). We therefore sought to test whether repeated presentation of threatening stimuli (i.e.

differential threat history) may alter fear responses at short and long-time scales. We
 considered two possibilities: either mice will show stable immobility across trials or will show a
 progressive habituation (Rankin et al., 2009), which may be differentially engaged depending on
 the proximity (i.e. imminence) of the threat.

292 To begin to test this, in a single behavioral session, we exposed mice to a series of three 293 repeated sweeping or looming stimuli, each separated by ~5 minutes. Contrary to our 294 hypothesis, repeated presentation of both stimuli resulted in a gradual decrease in immobility 295 across trials (Figure 3 A, B). In mice exposed to sweeping stimuli, a repeated measures one-296 way ANOVA revealed a significant decrease in freezing responses across trials (Figure 3C; RM 297 one-way ANOVA: p = 0.0007, F(1.765, 33.54) = 9.869, n = 20 mice), indicative of fear 298 suppression or habituation of immobility. Similarly, looming stimuli elicited robust habituation 299 across trials (Figure 3D; RM one-way ANOVA: p = < 0.0001, F(1.634, 29.40) = 15.79). To 300 measure the total degree of habituation, we used a post-hoc multiple comparison test to 301 compare the freezing response on trial 1 and trial 3 in mice exposed to either looming or 302 sweeping threats. Both stimuli resulted in significant habituation on trial 3 (sweeping: trial 1 vs 303 trial 3: Tukey multiple comparison t-test: p = 0.0015; looming: trial 1 vs trial 3: Tukey multiple 304 comparison t-test: p < 0.0001). To facilitate more direct comparisons across both datasets, we 305 calculated the habituation index as a normalized metric of total habituation in each animal, 306 thereby correcting for differences in overall freezing observed across stimuli. These data 307 indicate that the degree of habituation did not differ as a function of stimulus type (Figure 3E; 308 sweeping: habituation index: 0.57±0.08; looming: habituation index: 0.59±0.08; unpaired t-test: 309 p = 0.83 t = 0.214, df = 37). Additionally, we compared the rate of habituation by normalizing 310 immobility within each animal to their response on stimulus 1. Our results indicate that in naïve 311 mice, the rate of habituation across trials was well-fit by a linear regression. Furthermore, the 312 overall rate of habituation did not significantly differ between looming (Figure 3F; slope =  $-0.20 \pm$ 313 0.05) and sweeping (slope =  $-0.22 \pm 0.03$ ; p = 0.91, F(1,113) = 0.012). Taken together, these 314 results indicate that freezing behaviors habituate across repeated trials, regardless of the 315 stimulus. Furthermore, the observed linear decrease in immobility occurs on a relatively rapid 316 timescale, suggestive of rapid circuit-level changes in sensorimotor processing.

We next wondered whether the observed habituation was dependent on the relatively short time between threatening stimuli. To test this, mice were exposed to an identical stimulus paradigm (i.e. three presentations of a looming visual stimulus) but each trial was separated by 24 hours rather than 5 minutes. If habituation resulted from a reduction in threat salience at relatively 321 short time scales, then we would predict more stable immobility across repeated trials at longer 322 time scales (i.e. 24 hours). Contrary to this prediction, looming stimuli presented at intervals of 323 24 hours, resulted in an enhanced habituation across trials (Figure 4A; RM one-way ANOVA: 324 F(1.631, 30.99) = 38.06, p < 0.0001, n = 20 mice). Interestingly, increasing the interval between 325 stimuli appeared to alter the overall pattern of habituation across trials. To quantify these 326 changes, we first used a two-way repeated measures ANOVA to compare the overall pattern of 327 habituation in mice exposed to looming stimuli separated by 5 minutes and 24 hours. As 328 expected, there was a significant main effect of stimulus number (p < 0.0001, F(1.913, 70.76) = 329 4.05). Additionally, there was a significant interaction between stimulus number and inter-trial 330 interval (p = 0.02, F(2,74) = 4.05), suggesting that the overall pattern of habituation differed as a 331 function of inter-trial interval. The observed change in habituation pattern resulted from a non-332 linear change in habituation across trials. More specifically, at 24-hour inter-trial intervals, we 333 observed a significant decrease in freezing between trials 1 and 2 (Tukey's multiple 334 comparisons test: adjusted p: <0.0001) but no difference in immobility between trials 2 and 3 335 (Tukey's multiple comparisons test: adjusted p: 0.841). This pattern suggests that habituation is 336 accelerated at longer inter-trial intervals. To quantify the accelerated habituation, we compared



**Figure 4:** Accelerated pattern of habituation when the looming visual stimuli was separated by 24 hour intertrial intervals. (A) Immobility response to repeated looming visual stimuli presented at 24 hour inter-trial intervals in a behavioral chamber with identical contextual cues across trials. (B) Comparing the change in immobility from the first and second stimulus presentation (Trial 2-Trial 1) between animals that received the repeated looming visual stimuli with ~5 minute inter-trial intervals (Blue) or ~24 hour inter-trial intervals (Green) revealed an accelerated rate of habituation in 3x24 animals. (C) Normalized rate of habituation in 3x5 and 3x24 animals showed similar rates of overall innate fear behavioral habituation. (D) Immobility responses at inter-trial intervals of 24 hours in distinct behavioral contexts was similar to those observed in the same context. 3x5 min: n=19 mice, 9 males, 10 females; 3x24 SC: n=20 mice, 10 males, 10 females; 3x24 DC: n=10 mice, 5 males, 5 females.

the change in freezing between trials 1 and 2 at both inter-trial intervals. Consistent with the

- 338 observation of accelerated habituation, the change in the freezing duration between trials 1 and
- 339 2 was significantly larger at 24 hours (-22.24  $\pm$  2.5%, n = 20 mice) than at 5 minutes (12.25  $\pm$
- 2.9%, n = 19 mice; unpaired t-test: p = 0.012, t=2.617, df = 37). However, despite the
- 341 accelerated habituation, the overall degree of habituation measured on trial 3 was similar across
- inter-trial intervals (Figure 4C; 5 minute interval: habituation index:  $0.60 \pm 0.08$ ; 24 hour interval:
- habituation index:  $0.47 \pm 0.05$ ; unpaired t-test: p = 0.19, t = 1.336, df = 37). Together, these data
- indicate that the overall rate of habituation depends on the interval between trials, however, the
- total degree of habituation is consistent regardless of inter-trial interval.

346 The observed accelerated habituation at longer inter-trial intervals could be explained, in part,

- 347 by contextual fear learning extinction (Maren et al., 2013), as stimuli were presented in identical
- 348 contexts across all three days. To explicitly test whether the accelerated habituation was
- 349 mediated by contextual cues, we repeated the experiment in a new cohort of mice, in which the
- 350 context was varied between trials, with an inter-trial interval of 24 hours. Under these
- 351 experimental conditions, the overall pattern of habituation was similar to that observed at 24
- hour inter-trial intervals in a single context. More specifically, we observed non-linear,
- accelerated habituation, similar to that observed in a single context (Figure 4D; RM one-way
- ANOVA: p = 0.0007, F(1.912, 17.21) = 11.7, which was not significantly different than that
- 355 observed in a single context (two-way ANOVA interaction: p = 0.61 F(2,56) = 0.505). Together,
- 356 these results indicate that contextual cues were insufficient to explain the accelerated
- 357 habituation at extended inter-trial intervals. Furthermore, our results suggest that the neural
- 358 mechanisms underlying behavioral habituation may be distinct at short and long-time scales.
- 359 Freezing responses are diminished in novel contexts.

360 We next sought to determine whether innate freezing differs in mice that have been exposed to 361 the behavioral arena through familiarization versus mice in a completely new environment. We 362 reasoned that in novel environments, mice may show increased behavioral vigilance resulting in 363 increased fear responses, as they explore the novel environment. Alternatively, mice may 364 disregard potentially threatening stimuli as they familiarize themselves with their surroundings, 365 which would result in a decreased freezing behavior. To test this, mice were not familiarized 366 with the open field chamber prior to behavioral testing. To test whether environmental novelty 367 impacted behavioral habituation, we presented mice with 3 looming stimuli separated by 5 368 minutes, as in previous datasets. In response to the initial stimulus presentation, mice in novel



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*Figure 5: Freezing response in a Familiar (blue) and novel (light grey) environment* (A) Immobility response to a looming visual stimulus in mice that were not familiarized with the testing environment before behavioral testing. (B) Comparison of percent time immobile between familiarized (Blue) and unfamiliarized (Grey) animals in response to a looming visual threat. (C) Total time immobile in seconds during the duration of video recording after looming stimulus onset, exceeding the 20 second analysis window used to calculate percent time immobile. (D) Repeated presentations of the looming visual stimulus at inter-trial intervals of 5 minutes results in a reduced freezing response in mice within a novel environment. (E) Comparison of normalized habituation in mice exposed to looming stimuli under novel and familiar environmental conditions. Familiarized: n=20 mice, 10 males, 10 females; Novel: n=19 mice, 9 males, 10 females.

- 370 environment showed a small yet statistically significant immobility response (Figure 5A;
- baseline: 15.5 ± 3.6% time immobile; response: 28.2 ± 4% time immobile, paired Student's t-
- test: p = 0.01, t = 3.1, df = 19). However, when compared to a separate cohort of mice that
- 373 underwent chamber familiarization (as in previous cohorts), mice exposed to looming stimuli in
- novel environments showed a significantly attenuated fear response (Figure 5B; novel: 28.2 ± 4
- 375 % time immobile; familiar: 72.2 ± 5.2% time immobile; unpaired Student's t-test: p < 0.0001, t =
- 6.8, df = 38). Similar results were obtained when comparing the total time immobile in the
- minute following stimulus presentation (Figure 5C; novel: 9.6 ± 1.6 seconds; familiar: 23.7 ± 2.2
- 378 seconds; unpaired Student's t-test: p < 0.0001, t = 5.13, df = 38).

379 Despite the lower overall freezing behavior in response to a single looming stimulus, mice in
 380 novel environments still showed significant habituation across repeated trials, as there was a

- 381 significant main effect of trial number on freezing responses in a repeated measure ANOVA
- 382 (Figure 5D; RM one-way ANOVA: F(1.6, 31.05) = 3.7, p = 0.07; trial 1 vs. trial 3: Tukey post-hoc
- 383 comparison: p = 0.015). Compared to mice in familiar environments, the rate of habituation was
- 384 delayed in the novel environment, as there was no significant difference in freezing across trials
- 1 and 2 (Figure 5C; Tukey's post-hoc comparison: p = 0.47). However, the total degree of
- 386 habituation was not significantly different in mice exposed to looming stimuli novel versus
- 387 familiar environments (Figure 5E; novel: 0.66 ± 0.57; familiar: 0.53 ± 0.36; unpaired Student's t-
- test: p = 0.38, t = 0.9, df = 37, n = 19 mice (novel), 20 mice (familiar), one mouse was removed

389 from analysis in the novel condition as an outlier exceeding >3 standard deviations from the

390 population mean). These results indicate that mice in novel environments demonstrate both a

391 reduced overall fear response and a delayed habituation profile across repeated trials.

392 Effects of acute stress on innate freezing:

393 In addition to extrinsic factors, such as the nature of the visual stimulus, environmental context, 394 or context familiarity, fear responses can also be modulated by physiological factors such as 395 internal state. For example, in conditioned fear paradigms, exposure to intensely threatening 396 acute stress has been shown to sensitize both associative and non-associative fear learning 397 (Hassien et al., 2020; Perusini et al., 2016; Rau et al., 2005; Rau & Fanselow, 2009). We 398 therefore sought to test whether similar acute stress paradigms resulted in changes in innate 399 fear responses. To do this, mice were exposed to a set of four unconditioned footshocks 400 (amplitude: 1 mA, duration: 2 seconds, inter-trial interval: randomly applied between 60-90 401 seconds, as in (Hassien et al., 2020)) in a distinct behavioral context either 1 hour (Figure 6A) or 402 24 hours (Figure 6B) prior to exposure to looming threats. We first compared the immobility 403 response on the first of three visual stimuli and found no significant effect of acute stress on

initial innate fear responses (Figure 6C; Ordinary one-way ANOVA: F(2,54) = 2.06, p = 0.14).

405 We next examined the pattern of habituation across trials, which together demonstrated that the 406 rate of innate fear habituation was significantly delayed at 1 hour and 24 hours after acute 407 stress. At both time points, a repeated measure one-way ANOVA revealed a significant main 408 effect of trial number on freezing (Figure 6D-E 1 hour post-stress: RM one-way ANOVA: 409 F(1.505, 27.09) = 10.62, p = 0.001; 24 hours post-stress: F(1.575, 28.36) = 10.46, p = 0.0009). 410 Furthermore, in both datasets mice showed a significant decrease in freezing when comparing 411 trials 1 and 3 (Figure 6D-E; 1 hour post-stress: Tukey post-hoc comparison: adjusted p = 412 0.0062: Figure 6E: 24 hours post-stress: Tukey post-hoc comparison: adjusted p = 0.0005). 413 indicating intact habituation across trials. However, there was no statistically significant 414 difference in freezing between trials 1 and 2 (1 hour post-stress: Tukey post-hoc comparison: 415 adjusted p = 0.051; 24 hours post-stress: Tukey post-hoc comparison: adjusted p = 0.26), 416 suggesting that the habituation was significantly delayed. Finally, we compared the overall 417 degree of habituation across naïve (unstressed) animals and animals exposed to stress and 418 found no significant difference across all three datasets (Figure 6F; ordinary one-way ANOVA: 419 F(2.53) = 1.514, p = 0.23) indicating that stress does not significantly impact the overall degree 420 of innate fear habituation. Together, these data suggest that changes in internal state, such as



**Figure 6:** Acute stress effects on innate fear behaviors. (A,B) Immobility responses (red, velocity <2 cm/sec) triggered by Looming visual stimuli in male and female mice. Mice were exposed to acute footshock stress either 1 hour (A) or 24 hours (B) before exposure to innate fear paradigm. (C) Comparison of the freezing response on trial 1 across naïve and stressed animals. (D, E) Quantification of freezing behavior across trials in mice exposed to footshock stress 1 hour (D) or 24 hours (E) before the innate fear paradigm. (F) Comparison of the overall level of habituation across all three cohorts (naïve and stressed). Naïve: n=19 mice, 9 males, 10 females; Acute stress 1 hour: n=19 mice, 9 males, 10 females; Acute stress 24 hours: n=19 mice, 10 males, 9 females (one mouse was removed from analysis in the novel condition as an outlier exceeding >2.5 standard deviations from the population mean).

- 421 those following an unpredicted stressor, can significantly delay the rate of innate fear
- 422 habituation, but have little effect on overall freezing levels or the total degree of habituation
- 423 across trials.

### 424 **Discussion**:

- 425 Here, we set out to determine whether and how threat imminence is encoded in behavioral
- 426 action following innate predator threats, specifically under conditions in which the available
- 427 defensive reactions are limited. Our results indicate that under experimental conditions in which
- 428 escape behaviors are disfavored, namely in the absence of a protective shelter, mice responded
- 429 to both looming and sweeping stimuli with robust immobility. Both male and female mice

430 showed comparatively increased immobility to the looming stimulus, which, ethologically, 431 represents a more proximal threat. These results suggest that mice encode threat imminence 432 not only in behavioral action selection, but also in response vigor. Furthermore, our results 433 indicate that in response to repeated presentation of looming and sweeping threats, mice 434 consistently reduce immobility across trials, reflecting habituation. However, the overall pattern 435 of fear suppression differed across experimental manipulations, suggesting that innate fear 436 suppression is subject to modulation by environmental (extrinsic) and physiological (intrinsic) 437 factors.

# 438 Threat imminence theory and species-specific defense reactions

439 One predominant and unifying theory of innate (and conditioned) fear, is that fear serves to 440 restrict the behavioral repertoire of prey animals to a circumscribed, species-specific set of 441 defensive reactions designed to promote survival (Bolles, 1970; Crawford & Masterson, 1982; 442 Fanselow, 2018; Fanselow & Lester, 1988). This theory has been further expanded upon with 443 the development of the threat imminence theory (R. J. Blanchard & Blanchard, 1971; Bolles & 444 Fanselow, 1980; Fanselow & Lester, 1988), which postulates that as threats shift from distal to 445 proximal, animals respond with distinct, ethologically appropriate behavioral strategies, likely 446 mediated by distinct neural circuits (Deng et al., 2016; Fanselow, 1991, 1994; Gale & Murphy, 447 2014: Shang et al., 2018). For example, a distal aerial predator simply flying overhead may 448 initiate a 'passive coping strategy' such as freezing to avoid detection whereas an approaching 449 aerial predator will elicit more active avoidance strategies, such as flight, to evade capture (De 450 Franceschi et al., 2016; Tafreshiha et al., 2021; Yilmaz & Meister, 2013). It is worth noting that 451 the dichotomy of passive vs. active avoidance strategies is largely one of semantic 452 convenience, as 'passive coping strategies' are intentionally engaged and often involve 453 complex, whole-body motor coordination, and are therefore not truly passive. Despite this 454 qualification, the threat imminence theory is an influential model to describe how animals 455 appropriately regulate or adapt their defensive strategy in response to varied environmental 456 threats.

Innate fear responses have evolved from an evolutionary pressure to avoid predation risk,
resulting in species-specific defensive reactions that are tuned to specific ecological niches. For
example, closely related species of *Peromyscus* mice engage with identical predator threats
using distinct behavioral strategies that are ethologically matched to their evolutionary history
(Baier et al., 2023; Hirsch & Bolles, 1980). In addition to the diversity of behavioral strategies

462 observed, prey species must also show a high degree of behavioral flexibility to select the 463 optimal defensive strategy depending on external conditions such as proximity to a nest or other 464 environmental factors (Campagner et al., 2022; Evans et al., 2018; Lefler et al., 2020; Vale et 465 al., 2017). Although the threat imminence theory provides a powerful general framework to 466 describe how threat proximity influences defensive strategies, understanding how threat 467 imminence is encoded in other behavioral metrics of fear, such as behavioral vigor, has been 468 much more difficult to assess. This difficulty arises, in part, due to the inherent variability in 469 behavioral strategy employed by mice exposed to proximal or distal threats.

470 To facilitate a more direct comparison of behavioral vigor across proximal and distal threats, we 471 designed an innate fear behavioral paradigm in which both looming and sweeping visual stimuli 472 were presented to mice in a chamber lacking a protective shelter. Our results indicate, 473 consistent with previous literature (De Franceschi et al., 2016; Yilmaz & Meister, 2013), that 474 under such conditions mice engage in freezing behaviors in response to both looming and 475 sweeping visual stimuli. Here we have directly compared responses to looming and sweeping 476 visual threats, which is consistent with previous literature in experiments without a shelter (De 477 Franceschi et al., 2016). Our results expand on the previous findings by suggesting that mice 478 still encode threat proximity in the behavioral vigor with which they respond, as more proximal 479 threats elicited more robust (longer bouts of) defensive freezing.

480 It is worth noting that freezing in response to a looming predator may be considered 481 ethologically counter-intuitive as immobility in the face of an approaching predator may increase 482 the risk of predation. However, our data indicates that the strict behavioral hierarchy proposed 483 by the threat imminence theory is incomplete. In the absence of a protective shelter, mice 484 engage in defensive immobility, presumably to reduce the odds of detection while predators 485 make fine-scale adjustments to their attack trajectory (i.e. not all attacks are ballistic). In fact, 486 our data suggest that proximal threats result in increased behavioral vigor, as demonstrated by 487 the increased duration of immobility. In such cases (i.e. the absence of a shelter) the 'optimal' 488 strategy may be prolonged freezing to increase the probability that the predation attempt is 489 unsuccessful. Conversely, more distal threats, by definition, involve less risk, which results in 490 reduced behavioral vigor. Overall, our results reinforce the concept that fear limits the available 491 behavioral repertoire and further reinforce the threat imminence theory, by suggesting that in 492 addition to threat proximity influencing behavioral choice, it is also encoded in response vigor.

493 Behavioral flexibility is critical for appropriate fear responses

494 In addition to ethological pressures to select the optimal behavioral strategy to avoid predation, 495 animals must also be able to assess and adjust their defensive responses. As such, properly 496 regulated fear responses require animals to both respond appropriately to threats in the 497 environment while simultaneously adjusting fear responses to non-threatening stimuli 498 (Tafreshiha et al., 2021). Consistent with this view, our data demonstrate that repeated 499 presentation of looming and/or sweeping visual stimuli resulted in rapid decreases in freezing 500 behavior across repeated trials, which could be engaged with inter-trial intervals as short at 5 501 minutes. Such habituation may be ethologically adaptive, as it allows mice to re-evaluate 502 whether specific sensory inputs reflect acute threats in the environment. The relatively short 503 timescales at which habituation was observed suggests that the neural mechanisms underlying 504 such habituation may be mediated by rapid changes in synaptic integration in central fear 505 circuits. For example, emerging evidence suggests that circuits in the periaqueductal gray, a 506 central hub for generating fear behaviors (D. C. Blanchard & Blanchard, 2008; Koutsikou et al., 507 2015; C. Silva & McNaughton, 2019), may be modulated by numerous upstream circuits 508 including the cerebellum (Vaaga et al., 2020), in a direction predicted to reduce freezing 509 responses.

510 The behavioral habituation observed in response to repeated innate visual threats is reminiscent

511 of extinction learning in instrumental or Pavlovian conditioning paradigms (Maren et al., 2013),

512 although the underlying neural mechanisms may be entirely distinct. In conditioned paradigms,

513 extinction learning occurs when the reinforcing unconditioned stimulus (i.e. foot shock) is no

514 longer presented in conjunction with the conditioned stimulus (i.e. tone) (Bouton et al., 2021;

515 Delamater & Westbrook, 2014; Quirk & Mueller, 2008). There has been great interest in

516 understanding the behavioral and neural mechanisms underlying extinction, as it allows animals

517 to adjust their behavior to novel environments. Our results demonstrate that numerous intrinsic

518 and extrinsic factors (such as inter-trial interval, chamber familiarity and exposure to acute

519 stressors) alter the rate of habituation in an innate fear paradigm across repeated trials.

520 Impact of internal state on innate fear responses

521 Of particular interest is the observed impact of acute stress on innate fear responsivity both at 1

522 hour and 24 hours after stress exposure. Previous work has demonstrated that acute,

523 unpredicted stress significantly increases both associative and non-associative fear responses

in both mice and rats (Hassien et al., 2020; Perusini et al., 2016; Perusini & Fanselow, 2015;

525 Rau et al., 2005; Rau & Fanselow, 2009), which has been proposed as a fundamental model of

526 post-traumatic stress disorder (PTSD). Our work suggests that while acute stress does not 527 significantly impact the fear response on the first trial, it does significantly delay fear habituation. 528 Functionally, the delayed habituation represents an enhanced fear state – in which mice 529 maintain robust freezing responses for a prolonged period. This enhanced fear state is 530 consistent with the observed effects on fear learning, which are enhanced following similar 531 acute stress protocols. The increase in fear state across 24 hours suggests that the effects are 532 not mediated directly by enhanced circulating corticosterone, but rather may involve long-term 533 synaptic remodeling in innate fear circuitry, including the periagueductal gray (Myers et al., 534 2014). Interestingly, NMDA receptor activation is required for acquisition of associative fear 535 memory in the stress context (Rau et al., 2005), whereas circulating corticosterone is required 536 for both increased associative fear memory and stress-enhanced fear learning (Perusini & 537 Fanselow, 2015). Finally, our data support the accumulating evidence in favor of the stressenhanced fear learning paradigm as a model for PTSD (Hassien et al., 2020; Perusini & 538 539 Fanselow, 2015; Rau et al., 2005), as the delayed habituation profile observed at 24 hours post-540 stress exposure likely represents a unique form of delayed fear extinction, a hallmark clinical 541 feature of PTSD. Future work is therefore needed to resolve the neural mechanisms underlying

542 the stress-induced changes in innate fear responsivity and fear habituation.

### 543 **References**:

- Baier, F., Reinhard, K., Tong, V., Murmann, J., Farrow, K., & Hoekstra, H. E. (2023). The neural
  basis of defensive behaviour evolution in Peromyscus mice. *BioRxiv.Org: The Preprint Server for Biology*. https://doi.org/10.1101/2023.07.04.547734
- 547 Bandler, R., Depaulis, A., & Vergnes, M. (1985). Identification of midbrain neurones mediating
   548 defensive behaviour in the rat by microinjections of excitatory amino acids. *Behavioural* 549 *Brain Research*, *15*(2), 107–119.
- Bandler, R., Keay, K. A., Floyd, N., & Price, J. (2000). Central circuits mediating patterned
  autonomic activity during active vs. passive emotional coping. *Brain Research Bulletin*,
  552 53(1), 95–104.
- 553 Bandler, R., & Shipley, M. T. (1994). Columnar organization in the midbrain periaqueductal 554 gray: modules for emotional expression? *Trends in Neurosciences*, *17*(9), 379–389.
- Blanchard, D. C., & Blanchard, R. J. (2008). Chapter 2.4 Defensive behaviors, fear, and anxiety.
  In R. J. Blanchard, D. C. Blanchard, G. Griebel, & D. Nutt (Eds.), *Handbook of Behavioral Neuroscience* (Vol. 17, pp. 63–79). Elsevier.
- Blanchard, R. J., & Blanchard, D. C. (1971). Defensive reactions in the albino rat. *Learning and Motivation*, 2(4), 351–362.
- Bolles, R. C. (1970). Species-specific defense reactions and avoidance learning. *Psychological Review*, 77(1), 32.
- 562Bolles, R. C., & Fanselow, M. S. (1980). A perceptual-defensive-recuperative model of fear and563pain. The Behavioral and Brain Sciences, 3(2), 291–301.
- Bouton, M. E., Maren, S., & McNally, G. P. (2021). Behavioral and neurobiological mechanisms
   of Pavlovian and instrumental extinction learning. *Physiological Reviews*, *101*(2), 611–
   681.
- 567 Campagner, D., Vale, R., Tan, Y. L., Iordanidou, P., Pavón Arocas, O., Claudi, F., Stempel, A.
  568 V., Keshavarzi, S., Petersen, R. S., Margrie, T. W., & Branco, T. (2022). A cortico-
- 569 collicular circuit for orienting to shelter during escape. *Nature*.
- 570 https://doi.org/10.1038/s41586-022-05553-9
- 571 Carrive, P. (1993). The periaqueductal gray and defensive behavior: functional representation 572 and neuronal organization. *Behavioural Brain Research*, *58*(1–2), 27–47.
- 573 Crawford, M., & Masterson, F. A. (1982). Species-specific defense reactions and avoidance
  574 learning. An evaluative review. *The Pavlovian Journal of Biological Science*, *17*(4), 204–
  575 214.

- 576 De Franceschi, G., Vivattanasarn, T., Saleem, A. B., & Solomon, S. G. (2016). Vision Guides
  577 Selection of Freeze or Flight Defense Strategies in Mice. *Current Biology: CB*, *26*(16),
  578 2150–2154.
- 579 Delamater, A. R., & Westbrook, R. F. (2014). Psychological and neural mechanisms of
  580 experimental extinction: a selective review. *Neurobiology of Learning and Memory*, *108*,
  581 38–51.
- 582 Deng, H., Xiao, X., & Wang, Z. (2016). Periaqueductal Gray Neuronal Activities Underlie
   583 Different Aspects of Defensive Behaviors. *The Journal of Neuroscience: The Official* 584 *Journal of the Society for Neuroscience*, *36*(29), 7580–7588.
- Evans, D. A., Stempel, A. V., Vale, R., Ruehle, S., Lefler, Y., & Branco, T. (2018). A synaptic
  threshold mechanism for computing escape decisions. *Nature*.
- 587 https://doi.org/10.1038/s41586-018-0244-6
- 588 Fanselow, M. S. (1991). The Midbrain Periaqueductal Gray as a Coordinator of Action in 589 Response to Fear and Anxiety. In A. Depaulis & R. Bandler (Eds.), *The Midbrain*
- 590 Periaqueductal Gray Matter: Functional, Anatomical, and Neurochemical Organization
  591 (pp. 151–173). Springer US.
- Fanselow, M. S. (1994). Neural organization of the defensive behavior system responsible for
   fear. *Psychonomic Bulletin & Review*, 1(4), 429–438.
- Fanselow, M. S. (2018). The Role of Learning in Threat Imminence and Defensive Behaviors.
   *Current Opinion in Behavioral Sciences*, 24, 44–49.
- Fanselow, M. S., & Lester, L. S. (1988). A functional behavioristic approach to aversively
  motivated behavior: Predatory imminence as a determinant of the topography of
  defensive behavior. In R. C. Bolles (Ed.), *Evolution and learning (pp* (Vol. 263, pp. 185–
  212). Lawrence Erlbaum Associates, Inc, xi.
- Friedman, M. J., Resick, P. A., Bryant, R. A., & Brewin, C. R. (2011). Considering PTSD for
  DSM-5. *Depression and Anxiety*, 28(9), 750–769.
- Gale, S. D., & Murphy, G. J. (2014). Distinct representation and distribution of visual information
   by specific cell types in mouse superficial superior colliculus. *The Journal of*
- Neuroscience: The Official Journal of the Society for Neuroscience, 34(40), 13458–
  13471.
- Gruene, T. M., Flick, K., Stefano, A., Shea, S. D., & Shansky, R. M. (2015). Sexually divergent
  expression of active and passive conditioned fear responses in rats. *ELife*, *4*.
  https://doi.org/10.7554/eLife.11352

609 Hassien, A. M., Shue, F., Bernier, B. E., & Drew, M. R. (2020). A mouse model of stress-610 enhanced fear learning demonstrates extinction-sensitive and extinction-resistant effects 611 of footshock stress. Behavioural Brain Research, 379, 112391. 612 Hirsch, S. M., & Bolles, R. C. (1980). On the ability of prey to recognize predators. Zeitschrift 613 Für Tierpsychologie, 54(1), 71–84. 614 Igbal, J., Huang, G.-D., Xue, Y.-X., Yang, M., & Jia, X.-J. (2023). The neural circuits and 615 molecular mechanisms underlying fear dysregulation in posttraumatic stress disorder. 616 Frontiers in Neuroscience, 17, 1281401. 617 Kavaliers, M., & Choleris, E. (2001). Antipredator responses and defensive behavior: ecological 618 and ethological approaches for the neurosciences. Neuroscience and Biobehavioral 619 *Reviews*, 25(7–8), 577–586. 620 Keay, K. A., & Bandler, R. (2001). Parallel circuits mediating distinct emotional coping reactions 621 to different types of stress. Neuroscience and Biobehavioral Reviews, 25(7-8), 669-678. 622 Kikuchi, D. W., Allen, W. L., Arbuckle, K., Aubier, T. G., Briolat, E. S., Burdfield-Steel, E. R., 623 Cheney, K. L., Daňková, K., Elias, M., Hämäläinen, L., Herberstein, M. E., Hossie, T. J., 624 Joron, M., Kunte, K., Leavell, B. C., Lindstedt, C., Lorioux-Chevalier, U., McClure, M., 625 McLellan, C. F., ... Exnerová, A. (2023). The evolution and ecology of multiple 626 antipredator defences. Journal of Evolutionary Biology, 36(7), 975-991. 627 Koenen, K. C., Ratanatharathorn, A., Ng, L., McLaughlin, K. A., Bromet, E. J., Stein, D. J., 628 Karam, E. G., Meron Ruscio, A., Benjet, C., Scott, K., Atwoli, L., Petukhova, M., Lim, C. 629 C. W., Aguilar-Gaxiola, S., Al-Hamzawi, A., Alonso, J., Bunting, B., Ciutan, M., de 630 Girolamo, G., ... Kessler, R. C. (2017). Posttraumatic stress disorder in the World Mental 631 Health Surveys. Psychological Medicine, 47(13), 2260–2274. 632 Koutsikou, S., Watson, T. C., Crook, J. J., Leith, J. L., Lawrenson, C. L., Apps, R., & Lumb, B. 633 M. (2015). The Periaqueductal Gray Orchestrates Sensory and Motor Circuits at Multiple 634 Levels of the Neuraxis. The Journal of Neuroscience: The Official Journal of the Society 635 for Neuroscience, 35(42), 14132–14147. 636 La-Vu, M. Q., Sethi, E., Maesta-Pereira, S., Schuette, P. J., Tobias, B. C., Reis, F. M. C. V., 637 Wang, W., Torossian, A., Bishop, A., Leonard, S. J., Lin, L., Cahill, C. M., & Adhikari, A. 638 (2022). Sparse genetically defined neurons refine the canonical role of periagueductal 639 gray columnar organization. ELife, 11. https://doi.org/10.7554/eLife.77115 640 LeDoux, J. (2012). Rethinking the emotional brain. Neuron, 73(4), 653-676. 641 Lefler, Y., Campagner, D., & Branco, T. (2020). The role of the periaqueductal gray in escape 642 behavior. Current Opinion in Neurobiology, 60, 115-121.

- Lenzi, S. C., Cossell, L., Grainger, B., Olesen, S. F., Branco, T., & Margrie, T. W. (2022). Threat
  history controls flexible escape behavior in mice. *Current Biology: CB*, *32*(13), 29722979.e3.
- Liu, D., Li, S., Ren, L., Liu, X., Li, X., & Wang, Z. (2022). Different coding characteristics
  between flight and freezing in dorsal periaqueductal gray of mice during exposure to
  innate threats. *Animal Models and Experimental Medicine*, *5*(6), 491–501.
- Maren, S., Phan, K. L., & Liberzon, I. (2013). The contextual brain: implications for fear
  conditioning, extinction and psychopathology. *Nature Reviews. Neuroscience*, *14*(6),
  417–428.
- Myers, B., McKlveen, J. M., & Herman, J. P. (2014). Glucocorticoid actions on synapses,
  circuits, and behavior: implications for the energetics of stress. *Frontiers in Neuroendocrinology*, *35*(2), 180–196.
- Perusini, J. N., & Fanselow, M. S. (2015). Neurobehavioral perspectives on the distinction
  between fear and anxiety. *Learning & Memory*, 22(9), 417–425.
- Perusini, J. N., Meyer, E. M., Long, V. A., Rau, V., Nocera, N., Avershal, J., Maksymetz, J.,
  Spigelman, I., & Fanselow, M. S. (2016). Induction and Expression of Fear Sensitization
  Caused by Acute Traumatic Stress. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, *41*(1), 45–57.
- Quirk, G. J., & Mueller, D. (2008). Neural mechanisms of extinction learning and retrieval.
   *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology*, 33(1), 56–72.
- Rankin, C. H., Abrams, T., Barry, R. J., Bhatnagar, S., Clayton, D. F., Colombo, J., Coppola, G.,
  Geyer, M. A., Glanzman, D. L., Marsland, S., McSweeney, F. K., Wilson, D. A., Wu, C.F., & Thompson, R. F. (2009). Habituation revisited: an updated and revised description
  of the behavioral characteristics of habituation. *Neurobiology of Learning and Memory*,
- *668 92*(2), 135–138.
- Rau, V., DeCola, J. P., & Fanselow, M. S. (2005). Stress-induced enhancement of fear learning:
  an animal model of posttraumatic stress disorder. *Neuroscience and Biobehavioral Reviews*, *29*(8), 1207–1223.
- Rau, V., & Fanselow, M. S. (2009). Exposure to a stressor produces a long lasting
  enhancement of fear learning in rats. *Stress*, *12*(2), 125–133.
- 674 Shang, C., Chen, Z., Liu, A., Li, Y., Zhang, J., Qu, B., Yan, F., Zhang, Y., Liu, W., Liu, Z., Guo,
- K., Li, D., Wang, Y., & Cao, P. (2018). Divergent midbrain circuits orchestrate escape
  and freezing responses to looming stimuli in mice. *Nature Communications*, *9*(1), 1232.

- 677 Silva, B. A., Gross, C. T., & Gräff, J. (2016). The neural circuits of innate fear: detection,
- 678 integration, action, and memorization. *Learning & Memory*, 23(10), 544–555.
- Silva, C., & McNaughton, N. (2019). Are periaqueductal gray and dorsal raphe the foundation of
  appetitive and aversive control? A comprehensive review. *Progress in Neurobiology*,
  177, 33–72.
- Solomon, S. G., Janbon, H., Bimson, A., & Wheatcroft, T. (2023). Visual spatial location
  influences selection of instinctive behaviours in mouse. *Royal Society Open Science*,
  10(4), 230034.
- Tafreshiha, A., van der Burg, S. A., Smits, K., Blömer, L. A., & Heimel, J. A. (2021). Visual
  stimulus-specific habituation of innate defensive behaviour in mice. *The Journal of Experimental Biology*, *224*(Pt 6). https://doi.org/10.1242/jeb.230433
- 688Tovote, P., Esposito, M. S., Botta, P., Chaudun, F., Fadok, J. P., Markovic, M., Wolff, S. B. E.,689Ramakrishnan, C., Fenno, L., Deisseroth, K., Herry, C., Arber, S., & Lüthi, A. (2016).
- 690 Midbrain circuits for defensive behaviour. *Nature*, 534(7606), 206–212.
- Vaaga, C. E., Brown, S. T., & Raman, I. M. (2020). Cerebellar modulation of synaptic input to
   freezing-related neurons in the periaqueductal gray. *ELife*, *9*.
- 693 https://doi.org/10.7554/eLife.54302
- Vale, R., Evans, D. A., & Branco, T. (2017). Rapid Spatial Learning Controls Instinctive
  Defensive Behavior in Mice. *Current Biology: CB*, *27*(9), 1342–1349.
- Yilmaz, M., & Meister, M. (2013). Rapid innate defensive responses of mice to looming visual
  stimuli. *Current Biology: CB*, 23(20), 2011–2015.
- 598 Zhang, S. P., Bandler, R., & Carrive, P. (1990). Flight and immobility evoked by excitatory
- 699 amino acid microinjection within distinct parts of the subtentorial midbrain periaqueductal
- 700 gray of the cat. *Brain Research*, *520*(1–2), 73–82.