



## Draft Genome Sequence of Environmental Bacterium Vibrio vulnificus CladeA-yb158

## Yael Danin-Poleg,<sup>a</sup> Nili Raz,<sup>a</sup> Francisco J. Roig,<sup>b</sup> Carmen Amaro,<sup>b</sup> Yechezkel Kashi<sup>a</sup>

Faculty of Biotechnology and Food Engineering, Technion-Israel Institute of Technology, Haifa, Israel<sup>a</sup>; Estructura de Investigación Interdisciplinar en Biotecnología y Biomedicina ERI BioTecMed, Departamento Microbiología y Ecología, University of Valencia, Valencia, Spain<sup>b</sup>

We report the genome sequence of the environmental *Vibrio vulnificus* biotype 1\_cladeA. This draft genome of the CladeA-yb158 strain, isolated in Israel, represents this newly emerged clonal group that contains both clinical and environmental strains.

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Address correspondence to Yechezkel Kashi, kashi@tx.technion.ac.il

**V***ibrio vulnificus* is an aquatic bacterium and an important human pathogen (1–4). Strains of *V. vulnificus* are biochemically divided into three biotypes. Biotype 1 is a worldwide distributed pathogen and highly varied (5, 6). Recently, we found a new phylogroup, clade\_A, which includes both environmental and clinical isolates and presents biochemical characteristics that differ from those of biotypes 1 and 3 (7, 8). Here, we describe the draft genome sequence of the environmental *V. vulnificus* biotype 1 strain CladeA-yb158, isolated in 2005 from tilapia fish grown at aquaculture ponds in northern Israel (9, 10) and subjected to wholegenome shotgun sequencing.

The 300-bp library (233 to 414 bp) was sequenced using Illumina HiSeq 2000, generating 82,200,000 past-filtered 100-bp paired-end reads, with a coverage of 500×. Reads were de novo assembled with Velvet 1/1/06, SOAPdenovo 0.9.5, and GapCloser 1.10(11-13) (without filtering) generating 448 segments (kmer = 83) using reads with a minimum quality of 30 for each base. The assembly that contains 76 scaffolds of  $\geq$  200 bp, covers 5,294,657 bp, with an  $N_{50}$  of 543,331 bp and a longest segment of 1,472,236 bp and second longest segment of 635,056 bp. Mapping was done using BWA 0.9.5 with a maximum of two differences from the reference sequence per paired-end read and a maximum of one gap not in the 5 bp of the read. A total of 97.96% of the single reads were mapped to the assembly (with 0.37% singletons), and 68% of the single reads were mapped to the V. vulnificus YJ016 genome (14). There is evidence for the presence of a plasmid related to pYJ016 (scaffold\_27 and contig\_C1443).

Various bioinformatics approaches were applied with the aim of reducing the number of contigs without any success, maybe due to the genome complexity of CladeA-yb158(BT1), derived from the high rates of horizontal gene transfer in the *Vibrio* species (5, 6, 15) and the presence of multiple repetitive regions. However, there are three scaffolds that account for more than 50% of the assembly (L50), indicating the high quality of the assembly taking into consideration the genome complexity of this phylogroup.

The draft genome of CladeA-yb158(BT1) consists of 76 segments covering 2 chromosomes and a plasmid (5.29 Mbp; 46.7% G+C content). A total of 4,574 coding sequences (CDS), 82 pseudo genes, 7 rRNAs, 94 tRNAs, and 1 non-coding RNA (ncRNA) were predicted and annotated by the NCBI Prokaryotic Genome Annotation Pipeline (16), similar to the annotation predicted by RAST (17).

Genome comparison of the CladeA-yb158 genome to two published *V. vulnificus* biotype I genomes using the SEED viewer in RAST (17, 18) revealed 88.32% (2,847 I chromosomes, 1,473 II chromosomes, and 34 plasmids) and 91.54% (2,685 I chromosomes and 1,448 II chromosomes) common genes to YJ016 (14) and CMCP6 (5, 19), respectively, suggesting higher resemblance to the CMCP6 strain.

Since clade\_A is highly clonal, the genome of the CladeAyb158 strain provides a representation of this phylogroup, contributing to the understanding of the evolution of this human pathogen in the environment.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. LBNN00000000. The version described in this paper is version LBNN01000000.

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## REFERENCES

- 1. Jones MK, Oliver JD. 2009. *Vibrio vulnificus*: disease and pathogenesis. Infect Immun 77:1723–1733. http://dx.doi.org/10.1128/IAI.01046-08.
- 2. Oliver JD. 2006. *Vibrio vulnificus. In* Thompson FL, Austin B, Swings J (ed), The biology of vibrios. ASM Press, Washington, DC.
- Strom MS, Paranjpye RN. 2000. Epidemiology and pathogenesis of Vibrio vulnificus. Microbes Infect 2:177–188. http://dx.doi.org/10.1016/ S1286-4579(00)00270-7.
- 4. Oliver JD, Warner RA, Cleland DR. 1982. Distribution and ecology of *Vibrio vulnificus* and other lactose-fermenting marine vibrios in coastal waters of the southeastern United States. Appl Environ Microbiol 44: 1404–1414.

- Kim HU, Kim SY, Jeong H, Kim TY, Kim JJ, Choy HE, Yi KY, Rhee JH, Lee SY. 2011. Integrative genome-scale metabolic analysis of *Vibrio vulnificus* for drug targeting and discovery. Mol Syst Biol 7:460. http:// dx.doi.org/10.1038/msb.2010.115.
- Quirke AM, Reen FJ, Claesson MJ, Boyd EF. 2006. Genomic island identification in Vibrio vulnificus reveals significant genome plasticity in this human pathogen. Bioinformatics 22:905–910. http://dx.doi.org/ 10.1093/bioinformatics/btl015.
- Broza YY, Raz N, Lerner L, Danin-Poleg Y, Kashi Y. 2012. Genetic diversity of the human pathogen *Vibrio vulnificus*: a new phylogroup. Int J Food Microbiol 153:436–443. http://dx.doi.org/10.1016/ j.ijfoodmicro.2011.12.011.
- Raz N, Danin-Poleg Y, Hayman RB, Bar-On Y, Linetsky A, Shmoish M, Sanjuán E, Amaro C, Walt DR, Kashi Y. 2014. Genome-wide SNPgenotyping array to study the evolution of the human pathogen *Vibrio vulnificus* biotype 3. PLoS One 9:e114576. http://dx.doi.org/10.1371/ journal.pone.0114576.
- Broza YY, Danin-Poleg Y, Lerner L, Broza M, Kashi Y. 2007. Vibrio vulnificus typing based on simple sequence repeats: insights into the biotype 3 group. J Clin Microbiol 45:2951–2959. http://dx.doi.org/10.1128/ JCM.00236-07.
- Broza YY, Danin-Poleg Y, Lerner L, Valinsky L, Broza M, Kashi Y. 2009. Epidemiologic study of *Vibrio vulnificus* infections by using variable number tandem repeats. Emerg Infect Dis 15:1282–1285. http://dx.doi.org/ 10.3201/eid1508.080839.
- 11. Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Xiaoqian Z, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read *de novo* assembler. GigaScience 1:18. http://dx.doi.org/10.1186/2047-217X-1-18.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res 18:821–829. http:// dx.doi.org/10.1101/gr.074492.107.

- Li R, Zhu H, Ruan J, Qian W, Fang X, Shi Z, Li Y, Li S, Shan G, Kristiansen K, Li S, Yang H, Wang J, Wang J. 2010. *De novo* assembly of human genomes with massively parallel short read sequencing. Genome Res 20:265–272. http://dx.doi.org/10.1101/gr.097261.109.
- Chen CY, Wu KM, Chang YC, Chang CH, Tsai HC, Liao TL, Liu YM, Chen HJ, Shen AB, Li JC, Su TL, Shao CP, Lee CT, Hor LI, Tsai SF. 2003. Comparative genome analysis of *Vibrio vulnificus*, a marine pathogen. Genome Res 13:2577–2587. http://dx.doi.org/10.1101/gr.1295503.
- Urbanczyk H, Ast JC, Kaeding AJ, Oliver JD, Dunlap PV. 2008. Phylogenetic analysis of the incidence of lux gene horizontal transfer in *Vibrionaceae*. J Bacteriol 190:3494–3504. http://dx.doi.org/10.1128/ JB.00101-08.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufo S, Li W. 2013. Prokaryotic genome annotation pipeline. The NCBI handbook [Internet], 2nd ed. National Center for Biotechnology Information, Bethesda, MD. http://www.ncbi.nlm.nih.gov/books/NBK174280/.
- 17. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/ 1471-2164-9-75.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42: D206–D214. http://dx.doi.org/10.1093/nar/gkt1226.
- Kim YR, Lee SE, Kim CM, Kim SY, Shin EK, Shin DH, Chung SS, Choy HE, Progulske-Fox A, Hillman JD, Handfield M, Rhee JH. 2003. Characterization and pathogenic significance of *Vibrio vulnificus* antigens preferentially expressed in septicemic patients. Infect Immun 71:5461–5471. http://dx.doi.org/10.1128/IAI.71.10.5461-5471.2003.