

# Draft Genome Sequence of *Streptomyces viridochromogenes* Strain Tü57, Producer of Avilamycin

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**Here we present the draft genome sequence of *Streptomyces viridochromogenes* Tü57. This strain is a producer of avilamycin A, an oligosaccharide antibiotic from the orthosomycin group, which is active against Gram-positive bacteria.**

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*Streptomyces viridochromogenes* strain Tü57 is a producer of avilamycin A, a potent antibiotic against Gram-positive pathogenic bacteria. Avilamycin is an oligosaccharide antibiotic which belongs to the orthosomycin group (1). Members of this group show high antibacterial activity against glycopeptide-resistant enterococci, penicillin-resistant streptococci, and methicillin-resistant staphylococci (2–4). Avilamycin A consists of a hepta-saccharide side chain and a polyketide-derived dichloroisoeverninic acid as aglycone. Several genes from the secondary metabolite gene cluster which encodes the biosynthesis of avilamycin A have been characterized previously (5–7).

The whole-genome shotgun sequence was determined by Roche/454 GS FLX titanium pyrosequencing technology (8). Generation of a paired-end library consisting of 855,621 reads was performed, which corresponds to 17-fold coverage. All reads were assembled using Newbler 2.5.3, and 379 contigs were obtained. The genome of *Streptomyces viridochromogenes* Tü57 is composed of 9.7 Mbp, with an average G+C content of 71%.

For the analysis of the *S. viridochromogenes* Tü57 genome, open reading frames were predicted by Glimmer3 using a specific *Streptomyces* training set (9). Genome annotation was performed by applying an in-house-developed Galaxy-based automatic genome annotation pipeline for *Streptomyces*. In order to determine putative secondary metabolites, prediction of gene clusters was performed using antiSMASH (10). The analysis revealed that 846 genes belong to 33 secondary metabolite gene clusters. From those, we identified two polyketide synthase type I (PKSI) gene clusters, one PKSII, two PKSIII, 9 nonribosomal peptide synthetase (NRPS), two hybrid NRPS-PKSI, three terpene, and 14 other gene clusters. Furthermore, the analysis of nontranslating genes predicted at least 68 tRNAs and 1 tmRNA on the genome (11, 12).

**Nucleotide sequence accession number.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/

GenBank under the accession number [AML000000000](https://www.ncbi.nlm.nih.gov/nuccore/AML000000000). The version described in this paper is the first version.

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## REFERENCES

1. Buzzetti F, Eisenberg F, Grant HN, Keller-Schierlein W, Voser W, Zähler H. 1968. Avilamycin. *Experientia* 24:320–324.
2. Champney WS, Tober CL. 2000. Evernonimycin (SCH27899) inhibits both translation and 50S ribosomal subunit formation in *Staphylococcus aureus* cells. *Antimicrob. Agents Chemother.* 44:1413–1417.
3. Fuchs PC, Barry AL, Brown SD. 1999. In vitro activities of SCH27899 alone and in combination with 17 other antimicrobial agents. *Antimicrob. Agents Chemother.* 43:2996–2997.
4. Foster DR, Rybak MJ. 1999. Pharmacologic and bacteriologic properties of SCH27899 (Ziracin), an investigational antibiotic from evernonimycin family. *Pharmacotherapy* 19:1111–1117.
5. Gaisser S, Trefzer A, Stockert S, Kirschning A, Bechthold A. 1997. Cloning of an avilamycin biosynthetic gene cluster from *Streptomyces viridochromogenes* Tü57. *J. Bacteriol.* 179:6271–6278.
6. Weitnauer G, Mühlenweg A, Trefzer A, Hoffmeister D, Süßmuth RD, Jung G, Welzel K, Vente A, Girreser U, Bechthold A. 2001. Biosynthesis of the orthosomycin antibiotic avilamycin A: deductions from the molecular analysis of the avi biosynthetic gene cluster of *Streptomyces viridochromogenes* Tü57 and production of new antibiotics. *Chem. Biol.* 8:569–581.
7. Weitnauer G, Gaisser S, Kellenberger L, Leadlay PF, Bechthold A. 2002. Analysis of a C-methyltransferase gene (aviG1) involved in avilamycin biosynthesis in *Streptomyces viridochromogenes* Tü57 and complementation of a *Saccharopolyspora erythraea* eryBIII mutant by aviG1. *Microbiology* 148:373–379.
8. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen YJ, Chen Z, Dewell SB, Du L, Fierro JM, Gomes XV, Godwin BC, He W, Helgesen S, Ho CH, Irzyk GP, Jando SC, Alenquer ML, Jarvie TP, Jirage KB, Kim JB, Knight JR, Lanza JR, Leamon JH, Lefkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhijani VB, McDade KE, McKenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JF, Simpson JW, Srinivasan M, Tartaro KR, Tomasz A, Vogt

- KA, Volkmer GA, Wang SH, Wang Y, Weiner MP, Yu P, Begley RF, Rothberg JM. 2005. Genome sequencing in microfabricated high-density picolitre reactors. *Nature* 437:376–380.
9. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. *Bioinformatics* 23:673–679.
  10. Medema MH, Blin K, Cimermancic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. 2011. antiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene clusters in bacterial and fungal genome sequences. *Nucleic Acids Res.* 39:W339–W346.
  11. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res.* 32:11–16.
  12. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res.* 25: 955–964.