bioRxiv preprint doi: https://doi.org/10.1101/2020.04.02.022764. this version posted April 5, 2020. The copyright holder for this preprint (which was not certified by peer review) is the author/funder. It is made available under a CC-BY-NC-ND 4.0 International license. Potent Antiviral Activities of Type I Interferons to SARS-CoV-2 Infection Emily Mantlo¹, Natalya Bukreyeva¹, Junki Maruyama¹, Slobodan Paessler^{1, 2} and Cheng Huang^{1, 2}* ¹ Department of Pathology and ² Institute for Human Infections and Immunity, University of Texas Medical Branch, Galveston, Texas, United States of America * Correspondence: chhuang@utmb.edu Keywords: SARS-CoV-2; COVID-19; Interferon; antivirals; innate immune

29 Abstract:

30 The historical outbreak of COVID-19 disease not only constitutes a global public health crisis, but also has a devastating social and economic impact. The disease is caused by a newly identified coronavirus. Severe 31 Acute Respiratory Syndrome coronavirus 2 (SARS-CoV-2). There is an urgent need to identify antivirals to 32 curtail the COVID-19 pandemic. Herein, we report the remarkable sensitivity of SARS-CoV-2 to recombinant 33 human interferons α and β (IFN α/β). Treatment with IFN- α or IFN- β at a concentration of 50 international units 34 (IU) per milliliter drastically reduce viral titers by 3.4 log or 4.5 log, respectively in Vero cells. The EC₅₀ of IFN- α 35 36 and IFN-β treatment is 1.35 IU/ml and 0.76 IU/ml, respectively, in Vero cells. These results suggested that SARS-CoV-2 is more sensitive to many other human pathogenic viruses, including the SARS-CoV. Overall. 37 38 our results demonstrate the potent efficacy of human Type I IFN in suppressing SARS-CoV-2 replication, a 39 finding which could inform future treatment options for COVID-19.

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41 Introduction

The COVID-19 outbreak has started from the Wuhan Province, China since December 2019 and rapidly 42 spread globally, causing over 752,000 confirmed cases and 36,000 deaths as of April 1, 2020. The causative 43 agent for the COVID-19 disease is a newly identified Severe Acute Respiratory Syndrome coronavirus 2 44 45 (SARS-CoV-2) (1), which is transmitted through aerosol/airdrop inhalation or contact. The historical outbreak causes a public health crisis much severe than the SARS outbreak, which caused 8,098 infections and 774 46 deaths between November 2002 and July 2003. The COVID-19 disease also has a devastating social and 47 48 economic impact worldwide. WHO has declared COVID-19 disease a pandemic. In USA, there is over 200,000 49 confirmed cases and 4,300 deaths as of April 1, 2020. It is estimated by CDC that the COVID-19 pandemic may claim 200,000 to 1.7 million lives in USA. Treatments are urgently needed. Due to the high rate of new 50 51 infections reported each day, drugs already approved for clinical use for other disease may offer the most expedient option for treating COVID-19, and several such drugs are already being tested in clinical trials. 52

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54 Type I interferons (IFN- α/β) are cytokines that play a pivotal role in inducing an antiviral response across a 55 wide range of cell types. Humans produce 13 types of IFN- α and a singular IFN- β (2). Each Type I IFN 56 ultimately induces a number of interferon-stimulated genes (ISGs) which encode for a variety of antiviral

effectors. Notably, IFN-β production leads to a positive feedback loop that further stimulates the expression of many of the IFN- α genes (3). Clinically, Type I IFNs have already approved for use in the treatment of certain cancers, autoimmune disorders, and viral infections. We assessed the sensitivity of SARS-CoV-2 to both IFN- α and IFN- β . Herein, we report that type I IFNs exhibited potent anti-SARS-CoV-2 activities in cultured cells, demonstrating the therapeutic potency of type I IFNs for COVID-19.

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63 MATERIALS AND METHODS

Virus and Cells. The SARS-CoV-2 (USA-WA1/2020) were obtained from The World Reference Center for
 Emerging Viruses and Arboviruses (WRCEVA), University of Texas Medical Branch, Galveston, TX). Stock

virus were propagated by infecting Vero cells (ATCC CCL-81) at a low multiplicity of infection (MOI) 0.0025.

67 Three days after infection, supernatants were harvested and centrifuged at 2000 rpm for 5 min to remove cell

debris. Stock virus was titrated with a 50% tissue culture infectious dose assay (TCID₅₀) (4). All experiments

69 involving infectious virus were conducted at the University of Texas Medical Branch (Galveston, TX) in

approved biosafety level 3 laboratories in accordance with institutional health and safety guidelines and federal
 regulations.

Virus growth curve. Vero cells were infected by SARS-CoV-2 at MOI 1 or 0.01 for 1 hr. Then inoculum was 72 removed, replaced with media (DMEM+5%FBS) and incubated at 37°C and 5% CO₂. At different time points 73 after infection, supernatants were harvested and virus titers were determined by a TCID₅₀ assay on Vero cells. 74 75 Virus sensitivity to IFN treatment. Vero cells (2x10⁴/well) were seeded into 48-well plates for 24 h and treated with human IFN-β1a (mammalian, cat# 11415, PBL) and IFN-α (Universal Type I alpha A/D (Bg III), 76 PBL, cat# 11200-1) at different concentrations for 16 h. Cells were then infected with SARS-CoV-2 at an MOI 77 of 0.01 TCID₅₀/cell. IFNs were supplemented after virus infection. Supernatants were collected at 22 hr post 78 79 infection and assayed for virus titers.

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81 Results

The growth kinetics of the newly identified SARS-CoV-2 in cultured cells remained to be characterized. Thus, we first examined the growth kinetics of SARS-CoV-2 in Vero cells. Vero cells were infected at either a low MOI (MOI=0.01) or high MOI (MOI=1). Supernatant was collected every 8-16 hours. At both conditions, viral

titers peaked at around 24 hours post-infection (hpi) and remained stable until 40 hours post-infection before declining (Fig. 1). The peak virus titer was $5.5 \times 10^6 \,\text{TCID}_{50}$ /ml at MOI 0.01 and $3.75 \times 10^5 \,\text{TCID}_{50}$ /ml at MOI, indicating that viral replication was more efficient at low MOI (MOI=0.01) than high MOI (MOI=1). Virus infection also caused strong cytotoxic effect (CPE), which was more evident at 48 hpi and later than the peak

89 of virus production (at 40 hpi).

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Next, we examined the effect of recombinant human IFN- α and IFN- β treatment on viral infection. Vero cells 91 were pre-treated with different concentrations of IFN- α or IFN- β ranging from 50-1000 international units (IU) 92 per milliliter for 16 hours prior to infection. After 1 hour of infection with SARS-CoV-2 (MOI 0.01), media 93 94 containing IFN was returned, and cells were incubated for a further 22 hours. Supernatants were then collected, and viral titers were determined via TCID₅₀ assay. The result indicated a potent inhibition of SARS-95 CoV-2 infection by IFN-a treatment. Virus titers were not detectable except at the lowest concentration tested 96 97 (50 IU/ml), at which IFN- α drastically reduced viral titers by 4-logs of magnitude (Fig. 2). IFN- β exhibited more 98 potent anti-SARS-CoV-2 activity than IFN- α ; the virus titer was below the detection limit at all concentrations 99 tested (50 u/ml-1000u/ml). Consistently, no CPE was observable under microscope examination in all IFNtreated samples. 100

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We next tested the antiviral efficacy of IFN- α and IFN- β at lower concentrations ranging from 1-50 IU/ml. For 102 both IFNs, a dose-dependent effect was clearly observed using these lower concentrations (Fig. 3). For IFN- α . 103 the anti-SARS-CoV-2 activity could be noticed as low as 5 IU/ml, at which the virus titer was significantly 104 reduced by over 1 log (P<0.01). With increasing IFN- α concentrations, the virus titers steadily decreased. For 105 IFN-β, treatment with 1 IU/ml of IFN-β resulted in a moderate (approximately 70%) but significant decrease in 106 virus titer (P<0.05, Student t test). Virus levels were nearly undetectable upon treatment with 10, 25, and 50 107 IU/ml of IFN- β . The EC₅₀ of IFN- α and IFN- β treatment is 1.35 IU/ml and 0.76 IU/ml, respectively. Taken 108 together, these results indicated that treatment with low concentrations of both IFN- α and IFN- β significantly 109 inhibited viral replication, with IFN- β being slightly more effective than IFN- α . 110

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- 112 Discussion

Our data clearly demonstrated that SARS-CoV-2 is highly sensitive to both IFN- α and IFN- β treatment in 113 cultured cells. The experiment was performed in the IFN- α/β gene-defective Vero cells. It is plausible that the 114 anti-SARS-CoV-2 efficacy of exogenous IFN treatment is more potent in IFN-competent cells, as IFN- β is 115 known to mediate the expression of other subtypes of Type I IFNs. Our data may also explain, at least in part, 116 that approximately 80% of patients actually only developed mild symptoms and recovered (5). It is possible 117 that many infected people developed IFN- α and IFN- β -mediated innate immune response upon SARS-CoV-2 118 infection, which limits virus infection during initial stage. Later on, adaptive immune response, such as antibody 119 production, may eventually help the recovery from the COVID-19 disease. 120

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Compared to SARS-CoV-2, it seems SARS-CoV is not relatively less sensitive to IFN treatment in vitro (6, 7). 122 One study reported that the EC₅₀ of IFN- β for SARS-CoV is 95 or 105 IU/ml depending on virus strain (8). 123 Many other highly pathogenic viruses are also resistant to exogenous IFN treatment. Treatment of Ebola-124 125 infected cells with exogenous IFN- α does not lead to any reduction in viral replication (9), which is hypothesized to be a result of viral protein antagonism of the IFN response. Also, arenavirus Junín virus, which 126 127 causes Argentine Hemorrhagic Fever, is also found to be insensitive to IFN treatment. In Vero cells, the titers of JUNV were reduced by less than 1-log when treated with a high concentration of human IFN-α. β or v (1000 128 U/ml) (10). By contrast, the remarkable sensitivity of SARS-CoV-2 to IFN treatment reveals a weakness of this 129 virus, which may be informative to antiviral development. Further work is needed to characterize the IFN 130 response during SARS-CoV-2 infection and any possible viral interference. 131

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In vitro, we have demonstrated that SARS-CoV-2 replication is inhibited by clinically-achievable doses of both IFN- α and IFN- β . Thus, the use of IFN may be a viable treatment option for COVID-19. Serum levels of Roferon-A and Intron-A, two recombinant IFN- α drugs already approved for the treatment of hepatitis B and C, can reach up to 330 IU/ml and 204 IU/ml, respectively (11). Betaferon and Rebif, two recombinant IFN- β drugs approved for use in the treatment of multiple sclerosis, can reach respective concentrations of 40 IU/ml and 4.1 IU/ml in the serum (11). Some of these drugs may therefore have the potential to be repurposed for the treatment of COVID-19 either alone or in combination with other antiviral therapies.

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148	
149	Figure legend:
150	Figure 1: Vero cells were infected by SARS-CoV-2 at MOI 1 or 0.01 for 1 hr. At different time points after
151	infection, supernatants were harvested and virus titers were determined by a $TCID_{50}$ assay on Vero cells. The
152	average of triplicates and Standard deviation are shown. Dotted line indicates the detection limit.
153	
154	Figure 2: Vero cells were pretreated with human IFN- α or IFN- β (0, 50, 125, 250, 500, 1000 IU/ml) for 16
155	hours. Cells were then infected with SARS-CoV2 for 1 hour at an MOI of 0.01. Viral inoculums were removed
156	and replaced with fresh media containing listed concentrations of IFN- α or IFN- β . Media was collected at 22
157	hpi and titers were determined via TCID50 assay on Vero cells. The average of triplicates and Standard
158	deviation are shown. Dotted line indicates the detection limit.

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Figure 3: Vero cells were pretreated with human IFN- α or IFN- β (0, 1, 5, 10, 25, 50 U/ml) for 16 hours and then infected with SARS-CoV2 at an MOI of 0.01. Viral inoculums were removed and replaced with fresh media containing listed concentrations of IFN- α or IFN- β . Media was collected at 22 hpi and virus titers were determined via TCID50 assay. The average of triplicates and Standard deviation are shown. Dotted line indicates the detection limit. (*, P<0.05; **, P<0.01; n.s. not significant, one tail Student T test)

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