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Kallistatin, a new and reliable biomarker for the diagnosis of liver cirrhosis



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KEY WORDS

Kallistatin; Liver cirrhosis; Biomarker: Liver fibrosis; Hepatocellular carcinoma Abstract Kallistatin, which protects organs and cells against inflammation, fibrosis and oxidative stress, is mainly synthesized and secreted in liver. However, its relationship to human liver disease remains unclear. The purpose of this study was to explore the relationship between serum kallistatin and clinical evidence of both cirrhosis and hepatocellular carcinoma (HCC), and to determine if serum kallistatin levels could be used as a diagnostic indicator of hepatic health status, especially human liver cirrhosis (LC). Our cohort consisted of 115 patients with clinically proven liver fibrosis (LF), LC, or HCC by liver biopsies, and 31 healthy controls (CON). Serum kallistatin levels were quantified by ELISA. Results of the present study demonstrated that irrespective of the underlying etiology, serum kallistatin levels were significantly lower in the LF/LC group when compared with the CON group. A decrease in serum kallistatin levels appeared to reflect the extent of cirrhosis, with the lowest levels associated with higher grades of cirrhosis. Patients with LC had a noticeable correlation between serum kallistatin levels and other serum biochemical indicators. The area under the curve (AUC) for LC, viral liver cirrhosis (VLC) and alcoholic liver cirrhosis (ALC) was 0.845, 0.757 and 0.931, respectively. In conclusion, our findings

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Abbreviations: ALT, alanine transaminase; ALB, albumin; ALC, alcoholic liver cirrhosis; ALP, alkaline phosphatase; AUC, area under the curve; AST, aspartate aminotransferase; CE, choline esterase; CAP, community-acquired pneumonia; CON, controls; DBIL, direct bilirubin; GGT, gamma-glutamyl transpeptidase; GLB, globulin; HCC, hepatocellular carcinoma; IBIL, indirect bilirubin; KBP, kallikrein-binding protein; LC, liver cirrhosis; LF, liver fibrosis; NASH, non-alcoholic steatohepatitis; PA, prealbumin; STP, serum total protein; TBA, total bile acid; TBIL, total bilirubin; VLC, viral liver cirrhosis

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demonstrated that kallistatin, a plasma protein produced by the liver, can be a useful and reliable diagnostic indicator of hepatic health status, especially for LC.

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1. Introduction

Liver cirrhosis (LC), characterized by the histological development of regenerative nodules surrounded by fibrous bands, is the most frequent long term consequence of all chronic liver diseases. Cirrhosis is often asymptomatic and unsuspected until complications emerge (*e.g.* liver failure, portal hypertension, ascites, hepatic encephalopathy, variceal hemorrhage, hepatocellular carcinoma (HCC)). The annual incidence rates of liver failure (decompensation), HCC, and death in patients with cirrhosis associated with chronic hepatitis B or C infection are approximately 4%, 3% and 3%, respectively¹. The majority of patients with LC die from lifethreatening complications, which occurred relatively early in the course of the disease. Thus, the early diagnosis of LC is important for patients with chronic liver disease.

The definitive diagnosis of cirrhosis relies on the histological examination of liver tissue. The use of liver biopsies in clinical practice, however, has several limitations. In particular, sample errors are a significant problem, with an estimated mean of 24% of false negatives being reported in series of blind liver biopsies². Therefore routine clinical investigation of liver function employs serum analysis of indicators of liver dysfunction, such as increases in alanine transaminase (ALT) and aspartate aminotransferase (AST) amongst others. The complexity of LC and the influence of a multitude of genetic and environmental factors on the course of disease progression and response to therapy suggest the need to extend the battery of serum based functional assays that could add to the sensitivity and accuracy of currently employed diagnostic and prognostic markers.

Kallistatin, a tissue-kallikrein, has vasodilatory, anti-angiogenic, anti-inflammatory, anti-tumor and anti-oxidant effects^{3–6}. It is present in many human tissues, including the eye, kidney, liver, heart, arteries and veins, atheroma, blood cells and body fluids^{7.8}. As several studies have shown that the liver represents the major site of synthesis and secretion of kallistatin^{7.9}, we therefore hypothesized that serum kallistatin levels could be a potential biomarker for liver cirrhosis. We have explored the relationship between serum kallistatin and clinical evidence of both human cirrhosis and HCC in this study.

2. Participants and methods

2.1. Participants

Serum samples were collected at the Fujian Strait Hospital from 12 patients with LF, 93 with LC and 10 with HCC (without LC) proven by liver biopsies (Figs. S1 and S2 in Supporting information). LC patients had diverse etiologies: 41 patients were diagnosed with hepatitis B or C virus infection, 7 with alcoholic liver cirrhosis (ALC), and 45 had liver cirrhosis of uncertain cause. Eighteen of the patients with viral liver cirrhosis (VLC) (the cause of LC came from HBV infection) received different anti-cirrhotic drugs (Lamivudine, Sebivo or Adefovir dipivoxil, respectively)

and had their serum samples re-collected 30 days after the treatment. Thirty healthy volunteers served as controls. The study was approved by the local ethics committee (Quanzhou, No. 201001) and written informed consent was obtained from all patients involved.

2.2. Serum separation

Serum was collected from clotted blood using serum separator tubes centrifuged at 4000 rpm for 10 min at 4 °C. The serum was snap-frozen and stored at -80 °C until required. All serum samples were thawed only once.

2.3. Determination of serum kallistatin

Kallistatin levels were quantified by ELISA (R&D Systems, Inc. Minneapolis, USA) as previously described¹⁰. Briefly, standards and clinical samples were added in duplicate at appropriate dilutions into 96-well microtiter plate coated with capture antibodies.

2.4. Determination of other serum biochemical indicators

The clinical serum samples were also used to measure: serum prealbumin (PA), serum total protein (STP), albumin (ALB), albumin/globulin ratio (ALB/GLB), choline esterase (CE), globulin (GLB), total bilirubin (TBIL), direct bilirubin (DBIL), indirect bilirubin (IBIL), ALT, AST, gamma-glutamyl transpeptidase (GGT), alkaline phosphatase (ALP) and total bile acid (TBA). All of the parameters were measured using a conventional automated analyzer (Beckman Au680).

2.5. Statistical analysis

Data were expressed as mean \pm SD. All statistical analyses were performed using SPSS 16.0 software. The *t*-test was used for univariate comparisons between patients and the CON group with P < 0.05 considered significant. The Pearson correlation test, Spearman's rank correlation test and Logistic regression analysis were used to determine associations between variables. Receiver operating characteristic (ROC) curve analyses were performed to determine the diagnostic sensitivity and specificity of parameters. The area under the ROC curve (AUC) was calculated for prediction of patients with LC and the best cut-off values were established for the serum kallistatin levels.

3. Results

3.1. The main features and laboratory characteristics of all subjects

The serum levels of liver function tests, including PA, STP, ALB, GLB, ALT, AST, TBLI, DBIL, IBIL, GGT, ALP and TBA in the

LC groups were significantly different from those of the CON groups (Table 1).

3.2. Serum kallistatin levels in patients with LF/LC

Serum kallistatin levels were significantly lower in the 12 LF patients and 93 LC patients as compared to the CON groups (LF 22.77 \pm 5.02 & LC 16.78 \pm 8.65 *vs*. CON 29.03 \pm 8.07 µg/mL; *P*<0.05, *P*<0.01) (Fig. 1). This trend did not appear to be influenced by etiology: whether the cause of LC was viral infection or excessive alcohol consumption, the serum levels of kallistatin were significantly lower than the CON (VLC 19.84 \pm 9.71 & ALC 16.38 \pm 3.82 *vs*. CON 29.03 \pm 8.07 µg/mL; *P*<0.01). Kallistatin levels were similar for the VLC and ALC groups. In addition, various age or sex groups had no significant differences in serum kallistatin levels (data not shown).

Comparison of the LF and LC groups showed that serum kallistatin levels were further decreased (P < 0.05) (Fig. 1). Analysis of kallistatin levels in patients with different severities of LC revealed higher serum kallistatin concentrations in patients with compensated LC compared to patients with decompensated LC (19.98 ± 10.14 vs. 12.25 ± 6.22 µg/mL; P < 0.05), suggesting that serum levels of kallistatin may reflect the degree of liver dysfunction.

A follow-up study of 18 patients with VLC who received the anti-cirrhotic treatment (Lamivudine, Sebivo or Adefovir dipivoxil respectively) for 30 days showed elevated kallistatin levels after the treatment $(13.67 \pm 7.81 \text{ prior to } vs. 17.09 \pm 6.89 \,\mu\text{g/mL}$ following the treatment).

3.3. Serum kallistatin levels in patients with HCC

HCC groups had lower serum kallistatin levels than CON groups $(25.05\pm7.20 \text{ vs. } 29.03\pm8.06 \,\mu\text{g/mL})$, but higher than the LC groups (P < 0.05) (Fig. 1). These results suggest that decreases in

serum kallistatin levels are not unique to patients with LC, but may reflect different etiologies of liver disease.

3.4. Relationship between kallistatin serum levels and liver function

Patients with LC had a noticeable positive correlation between serum kallistatin levels and serum PA, STP, ALB, ALB/GLB and CE. Also, they had a significantly negative correlation between serum kallistatin levels and TBIL, ALT and AST levels, as determined by Pearson and Spearman rank correlation tests (Table 2) and linear regression analysis (Fig. 2).

Table 3 presents the association of each characteristic (serum kallistatin levels and other biochemical indicators) with LC (unadjusted odds ratios). It seemed that all the variables except



Figure 1 Mean concentration of serum kallistatin in the patients with LF, LC and HCC. Kallistatin levels were significantly different between LF, LC, HCC and controls (*P < 0.05, **P < 0.01).

Table 1	Demographic and	d clinical	characteristics	of the	study	population	with LF,	LC and	HCC.
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Parameter	CON (<i>n</i> =31)	LF (<i>n</i> =12)	LC (<i>n</i> =93)	HCC (<i>n</i> =10)
Age (years)	35.10 ± 10.40	41.58 ± 12.75	52.29 ± 11.98	59.40 ± 10.84
Gender (male; female)	8;23	9;3	77;16	7;3
Kallistatin (µg/mL)	29.03 ± 8.07	22.77 ± 5.02	16.78 ± 8.65	25.05 ± 7.20
Prealbumin (g/L)	74.29 ± 3.18	24.89 ± 5.89	11.67 ± 5.49	14.58 ± 5.61
STP (g/L)	43.67 ± 2.24	71.71 ± 6.28	67.33 ± 9.92	70.00 ± 6.74
Albumin (g/L)	30.05 ± 3.13	44.46 ± 3.37	33.52 ± 7.18	37.75 ± 5.40
Globulin (g/L)	21.54 ± 2.82	27.25 ± 4.96	33.80 ± 8.02	32.25 ± 4.81
Albumin/Globulin	1.45 ± 0.12	1.67 ± 0.27	1.05 ± 0.33	1.20 ± 0.29
ALT (U/L)	24.81 ± 14.21	99.89 ± 146.17	48.97 ± 69.74	98.76 ± 156.35
AST (U/L)	18.87 ± 5.49	48.76 ± 44.62	64.27 ± 120.38	76.76 ± 94.59
AST/ALT	0.93 ± 0.43	0.77 ± 0.34	1.34 ± 0.52	1.08 ± 0.57
Choline esterase (KU/L)	8.38 ± 2.2	8.19 ± 1.94	6.38 ± 3.22	8.14 ± 4.03
Total bilirubin (µmol/L)	13.86 ± 5.45	15.90 ± 5.65	48.64 ± 63.13	32.76 ± 48.84
Direct bilirubin (µmol/L)	5.03 ± 2.17	4.09 ± 2.07	20.95 ± 34.73	16.29 ± 26.34
Indirect bilirubin (µmol/L)	10.34 ± 6.82	11.81 ± 3.99	27.36 ± 31.37	21.01 ± 21.58
GGT (U/L)	25.13 ± 13.84	160 ± 424.66	71.44 ± 83.45	169.33 ± 284.42
ALP (U/L)	90.42 ± 32.71	121.50 ± 169.57	135.10 ± 70.82	307.90 ± 355.98
TBA (µmol/L)	6.13 ± 3.85	7.96 ± 6.6	57.85 ± 73.64	34.63 ± 49.82

PA and STP were significantly associated with liver cirrhosis. But only ALB and GLB were included in the final model, which was determined by performing multivariate logistic regression with stepwise selection of LC, and significant variables at level of 0.05. It was indicated from the final model that for 1 µg/mL increase in serum kallistatin concentration, the odds of liver cirrhosis decreases by $(1-0.682) \times 100\% = 31.8\%$ after controlling the effect of all the other covariates.

3.5. Reliability of serum kallistatin level as predictor of LC

The serum kallistatin level predicted the presence of LC estimated with AUC of 0.845 in 93 patients (Fig. 3A). Based on the ROC curve, an optimal cut-off value for LC diagnosis was set $<21.64 \mu g/mL$ of kallistatin. Sensitivity (Se), specificity (Sp), positive predictive value (PPV) and negative predictive value (NPV) values were 87.1%, 74.2%, 91% and 62.3%, respectively (Table S1 in supporting information).

In the cases of discriminating VLC and ALC from the CON patients based on the kallistatin level, the AUCs were 0.757 (P=0.001) and 0.931 (P<0.01) respectively. The results indicate that the predictive value is significantly better for patients with ALC than for the ones with VLC (Fig. 3B, C and Table S1 in Supporting information).

3.7. Disease specificity for LC

Using similar ROC analyses, we found that kallistatin level was not a reliable predictor of HCC, even though the levels in those patients were lower than in the CON groups. Serum kallistatin is therefore a good biomarker for different etiologies of LC with exception of HCC.

Table 2 Pearson correlation (*r*-value) and spearman rank correlation (ρ) between biochemical indices of liver sufficiency and kallistatin concentration in all cirrhotic patients without HCC.

	Pearson correlation		Spearman rank correlation		
Parameter	r	Р	ρ	Р	
Prealbumin (g/L)	0.699	< 0.001	0.671	< 0.001	
STP (g/L)	0.523	< 0.001	0.536	0.005	
Albumin (g/L)	0.313	0.002	0.326	< 0.001	
ALB/GLB	0.372	< 0.001	0.392	< 0.001	
Choline esterase (U/L)	0.699	< 0.001	0.645	< 0.001	
ALT (U/L)			-0.224	0.031	
AST (U/L)			-0.348	0.001	
Total bilirubin (µmol/L)			-0.313	0.002	
Direct bilirubin (µmol/L)			-0.329	0.001	
Indirect bilirubin (µmol/L)	irubin (µmol/L)		-0.288	0.005	
TBA (µmol/L)			-0.257	0.013	



Figure 2 Correlation between serum kallistatin levels and other biomarkers in 93 patients with LC. The vertical axis represents serum kallistatin levels in μ g/mL and the horizontal axis represents the expression levels of serum PA, STP, ALB, ALB/GLB and CE.

Variable	Unadjusted	Unadjusted OR	Final model			
		95% CI	<i>P</i> -value	Unadjusted OR	95% CI	P-value
PA	0.72	0.325-1.594	0.4172			
STP	1.00	0.993-1.007	0.9254			
ALB	1.09	1.019-1.174	0.0133	1.36	1.072-1.716	0.01
GLB	1.73	1.387-2.159	< 0.0001	1.95	1.272-2.990	0.00
ALB/GLB	0.00	< 0.001-0.033	< 0.0001			
CE	1.00	1.0001-1.0008	0.0027			
ALT	1.04	1.008-1.071	0.0122			
AST	1.22	1.121-1.328	< 0.0001			
AST/ALT	9.82	2.777-34.723	0.0004			
TBIL	1.22	1.114-1.334	< 0.0001			
DBIL	1.46	1.178-1.815	0.0006			
IBIL	1.19	1.092-1.298	< 0.0001			
GGT	1.04	1.012-1.063	0.0037			
ALP	1.02	1.008-1.035	0.0015			
TBA	1.19	1.080-1.319	0.0005			

Table 3	Logistic	regression	analysis	of liver	cirrhosis



Figure 3 Serum kallstatin levels can accurately diagnose liver disease. A: LC; B: VLC; C: ALC. The AUC is shown for the performance of the serum kallistatin levels for discriminating LC, VLC and ALC from the healthy control groups. The vertical axis represents the sensitivity and the horizontal axis represents the 1-specificity.

4. Discussion

To our knowledge, the present study is the first to demonstrate kallistatin to be a useful potential biomarker for the detection of human LC. The data presented in this study showed that serum kallistatin levels in patients with LC were significantly lower than those in healthy controls, demonstrating a close correlation between the reduction in serum kallistatin levels and severity of early hepatic disease. Serum kallistatin levels could therefore provide an additional biomarker for the detection of LC and progressive loss of liver function along in response to the therapy.

The biology of LC is characterized by a constant stimulus for hepatocellular regeneration by a microenvironment associated with chronic inflammation and tissue fibrosis. Cirrhosis represents the final common pathological outcome for the majority of chronic liver diseases. Previous studies have shown that 30%-40% of non-alcoholic steatohepatitis (NASH) patients have advancing bridging fibrosis or cirrhosis^{11–13}. Most patients with cirrhosis die from one or more clinical complications including ascites, hepatic encephalopathy and variceal hemorrhage¹⁴. Among the 1.4 million liver-disease-related deaths that occur each year worldwide, over 55%, or 796,000, are directly attributable to cirrhosis¹⁵.

Liver transplantation has been the most effective therapy for patients with many types of advanced liver disease. Unfortunately, many patients cannot obtain transplantation due to the limited availability of donor livers, with over 10% of patients dying whilst on the waiting list. Of those patients who receive liver transplants, 94% survive after three months, 88% after one year and 79% after three years¹⁶. Furthermore, evidence of either fibrotic or cirrhotic regression has now been reported in chronic liver diseases of different etiologies, including viral hepatitis¹⁷⁻²⁴, autoimmune hepatitis²⁵, alcoholic and NASH^{26,27}. While traditionally considered irreversible, recent evidence from animal studies and human clinical observations indicated that even advanced fibrosis can be reversed^{17,28}. This makes it important to identify reliable biomarkers for the early detection of liver disease and subsequent evaluation of response to therapeutic intervention. The ability of serum biomarkers of liver function to further discriminate between LC and HCC would add a major advantage.

Since first identified as a tissue kallikrein-binding protein (KBP), kallistatin mRNA was found in varying amounts in human liver, stomach, pancreas, kidney, aorta, testes, prostate, artery, atrium, ventricle and lung²⁹. By far, the most important site of synthesis of kallistatin is the liver³⁰, explaining the

significantly reduced levels of serum kallistatin in patients with LC.

Kallistatin can inhibit tumor growth⁵, reduce blood pressure³¹, suppress arthritis^{6,32} and protect organs and cells against inflammation, fibrosis, and oxidative stress^{33–35}. The anti-oxidative effect of kallistatin can also inhibit salt-induced renal injury, inflammation, and fibrosis³⁴.

The patients with HCC had lower serum kallistatin levels than normal controls, but significantly higher than in patients with LC. Thus, an inverse relationship between oxidative stress and kallistatin level might exist. This would be consistent with data demonstrating that male Sprague-Dawley (SD) rats exposed to either episodic or sustained hypoxia have reduced renal expression of kallistatin³⁶. Moreover, chronic oxidative organ damage can markedly reduce circulating kallistatin levels in rats, while hydrogen peroxide (H₂O₂) lowers kallistatin mRNA and protein levels in dose-dependent fashion *in vitro*³³. Kallistatin is known to play an important role in prevention of cancer, probably through its anti-angiogenic effects⁵. The human body can therefore upregulate kallistatin expression when cancers occur in a selfprotective mechanism. In agreement with data in our present study, previous examination of plasma kallistatin concentrations detected $22.1\pm3.5 \,\mu\text{g/mL}$ in 30 normal subjects and $21.1 \pm 3.8 \,\mu$ g/mL in five patients with hereditary angioedema⁷. A significant decrease in kallistatin levels $(7.2 \pm 2.5 \,\mu\text{g/mL},$ P < 0.001) was also found in plasma samples from nine patients with liver disease7. Such a discrepancy may be related to differences in the populations studied or may reflect the relatively small sample size. We have previously demonstrated that transgenic expression of kallistatin can significantly attenuate carbon tetrachloride-induced liver damage³⁷. This anti-inflammatory, antifibrotic potential of kallistatin warrants clinical evaluation, either with gene-therapy-based or recombinant-protein-administrationbased evaluation of its therapeutic potential in human LC.

Previous studies showed that other clinical situations can also affect serum kallistatin levels, such as adiposity, communityacquired pneumonia (CAP) and type 1 diabetes38,39. Serum kallistatin levels were significantly increased in diabetic vs. nondiabetic control subjects (12.6+4.2 vs. 10.3+2.8 µg/mL), and showing that levels were higher in diabetic patients with complications but did not differ for complication-free patients³⁸. A study of CAP assessing the correlations of kallistatin with other biomarkers found that kallistatin was significantly consumed in CAP patients (median: 8.3, range 1.3-17.3 µg/mL) compared with healthy subjects (median: 17.2, range 5.3-82.7 μ g/mL)³⁹. These studies emphasize the diagnostic value of serum kallistatin levels in human LC, even though CAP patients have lower serum kallistatin levels, and the downgrading is fairly modest. However, the levels were decreased in liver disease but increased in diabetic patients. Therefore, comprehensive evaluation of kallistatin level and patients with obesity and diabetes history may determine their liver health more accurately.

In conclusion, the present study demonstrated the levels of serum kallistatin reduced significantly in patients with LC of different etiologies, but not with HCC. The magnitude of this decrease appeared to be correlated with the degree of LC and disruption of normal liver function. Kallistatin therefore has a potential as a new biomarker for the diagnosis of, and evaluation of the extent of human LC, along with its response to therapeutic intervention. Furthermore, kallistatin may provide a new therapeutic strategy for the management and treatment of cirrhotic liver damage.

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Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at http://dx.doi.org/10.1016/j.apsb.2015.02.003.

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