

Thyrotoxic Periodic Paralysis and Polymorphisms of the *ADRB2*, *AR*, and *GABRA3* Genes in Men with Graves Disease

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Background: Thyrotoxic periodic paralysis (TPP) is a rare complication of thyrotoxicosis characterized by acute attacks of muscle weakness and hypokalemia. Recently, variation in several genes was suggested to be associated with TPP. This study evaluated the genetic predisposition to TPP in terms of the β 2-adrenergic receptor (*ADRB2*), androgen receptor (*AR*), and γ -aminobutyric acid receptor α 3 subunit (*GABRA3*) genes.

Methods: This study enrolled 48 men with Graves disease (GD) and TPP, and 48 GD patients without TPP. We compared the frequencies of candidate polymorphisms between the two groups.

Results: The frequency of the Gly16/Gly16 genotype in *ADRB2* was not significantly associated with TPP ($P=0.32$). More CAG repeats (≥ 26) in the *AR* gene were not correlated with TPP (odds ratio [OR], 2.46; 95% confidence interval [CI], 0.81 to 8.09; $P=0.08$). The allele frequency of the TT genotype in the *GABRA3* gene was not associated with TPP (OR, 1.83; 95% CI, 0.54 to 6.74; $P=0.41$).

Conclusion: The polymorphisms in the *ADRB2*, *AR*, and *GABRA3* genes could not explain the genetic susceptibility to TPP in Korean men with GD.

Keywords: Periodic paralysis-hypokalemic; Polymorphism; *ADRB2*; Receptors, androgen; *GABRA3*

INTRODUCTION

Thyrotoxic periodic paralysis (TPP) is a rare complication of thyrotoxicosis characterized by acute, reversible episodes of muscle weakness and low serum potassium levels [1]. TPP is prevalent in Asian populations, particularly among Chinese, Japanese, Vietnamese, Filipino, and Korean nationals [2,3]. Although the pathogenesis of TPP remains unclear, the recurrent

paralytic muscle weakness is caused by hypokalemia followed by an intracellular potassium shift [3]. The clinical features of and factors precipitating TPP are similar to those of familial hypokalemic periodic paralysis (FHPP), which is an autosomal dominant trait that is common in Caucasians, although hyperthyroidism is not present [2].

Several genetic variants have been studied in patients with and without TPP. Mutations in the skeletal muscle calcium

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channel (*CACNA1S*) [4-6], sodium channel (*SCN4A*) [5-7], and voltage-gated potassium channel (*KCNE3*) genes [5,7] were found in patients with FHPP. The similar clinical manifestations of TPP and FHPP, and racial predilection, suggested a strong genetic predisposition in TPP. However, in TPP patients there were no mutations in the *CACNA1S* and *SCN4A* genes and only one patient (a 44-year-old Portuguese) was reported to have a *KCNE3* mutation [8]. Recently, a genome-wide association study (GWAS) of a Thai group suggested that a single nucleotide polymorphism (SNP; rs750841) in intron 3 of the γ -aminobutyric acid receptor $\alpha 3$ subunit (*GABRA3*) gene was associated with TPP [9]. The *GABRA3* gene is one of the chloride channel genes.

Recently, we evaluated the Arg16Gly polymorphism in the $\beta 2$ -adrenergic receptor (*ADRB2*) gene and the number of CAG repeats in the androgen receptor (*AR*) gene in a small number of patients with or without TPP [10,11]. The genetic alterations in these genes were not associated with the genetic susceptibility to TPP in Korean males with Graves disease (GD).

In this study, we evaluated genetic differences in the *ADRB2*, *AR*, and *GABRA3* genes in a larger case-control series of Korean males with GD.

METHODS

Subjects

We enrolled 48 male GD patients with TPP and 48 more without TPP between 2001 and 2006 at Asan Medical Center, Seoul, Korea. All of the subjects had been diagnosed with GD based on clinical and laboratory criteria, including suppressed serum thyroid stimulating hormone (TSH) levels and increased serum free thyroxine (T_4) levels, a diffuse goiter, and positive for at least one antithyroid antibody. In the subjects, TPP was confirmed using acute-onset muscle weakness and biochemical hypokalemia (<3.5 mmol/L). We obtained informed consent from all patients, and the study was approved by the Institutional Review Board of Asan Medical Center.

DNA analysis

Genomic DNA was extracted from peripheral blood samples using the Puregene DNA Isolation Kit (Gentra Systems, Minneapolis, MN, USA). To evaluate the frequencies of the three genetic variants, genes were amplified by polymerase chain reaction (PCR) using the primers 5' GCACCCGACAAGCTGA 3' (F-*ADRB2*), 5' TCCAAACTCGCACCA 3' (R-*ADRB2*; J2960), 5'-ACCGAG-GAGCTTCCAGAAT-3' (F-*AR*), 5'-CTCATCCAGGAC CAG-

GTAGC-3' (R-*AR*; AL049564), 5' CACTGCCTGTTTCCCAAAT 3' (F-*GABRA3*), and 5' TGGAATGCTGAGAGTGATGG 3' (R-*GABRA3*; NT 011726.13).

Each reaction was performed with 100 ng of genomic DNA, 0.5 units of *Taq* polymerase (Takara Shuzo, Otsu, Japan), 2 pmol of each primer, and 4 nmol of each deoxynucleotide in a total reaction volume of 20 μ L using the following cycles: denaturation at 94°C for 5 minutes, followed by 42 cycles of 45 seconds at 94°C, 45 seconds at 56°C, and 45 seconds at 72°C, with a final 10-minute extension at 72°C. The PCR products were subject to agarose gel electrophoresis and DNA was extracted using a gel extraction kit (QIAGEN, Hilden, Germany). The extracted DNA was sequenced using an automated DNA sequence analyzer (PRISM 373 A, Applied Biosystems, Foster City, CA, USA) via a PCR reaction.

Laboratory measurement

As previously described, the serum levels of thyroid hormone (free T_4) and TSH were measured with commercial radioimmunoassay (Abbott, North Chicago, IL, USA) and immunoradiometric assay (SPAC-5 TSH kit, Daiichi, Tokyo, Japan) kits. TBII activity was measured with a radioreceptor assay kit (R. S.R., Cardiff, UK) and anti-thyroglobulin (anti-Tg) and anti-thyroperoxidase (anti-TPO) antibodies were measured with a radioligand assay (Henning test anti-Tg or anti-TPO, Diagnostica, Wiesbaden, Germany) [12]. The serum potassium level was measured using an ion-specific electrode, and all patients with TPP had hypokalemia (<3.5 mmol/L).

Statistical analysis

Associations between variables were analyzed using contingency tables and Fisher exact test. For multiple comparisons of genotype frequencies among the patients with or without TPP, the corrected *P* value was calculated by multiplying the *P* value by the number of comparisons. The Hardy-Weinberg proportion was tested using free software available at <http://www2.biology.ualberta.ca/jbrzusto/hwenj.html>, which was based on a report by Guo and Thompson [13]. The odds ratio (OR) and 95% confidence interval (CI) were calculated using the modified method of Woolf and Haldane. $P < 0.05$ was considered significant.

RESULTS

Baseline clinical characteristics of patients with or without TPP

The mean age of the 96 men with GD was 35.07 ± 10.9 years. Their mean serum TSH and free T_4 levels were 0.023 ± 0.13

$\mu\text{U/mL}$ and $4.03 \pm 1.56 \text{ ng/dL}$, respectively. There was a significant ($P=0.01$) difference in mean age between the patients with and without TPP (Table 1). Although the TPP patients were younger than those without TPP, age does not affect DNA information. There was no significant difference in the serum TSH and free T_4 levels between the two groups ($P=0.25$ and $P=0.32$, respectively). Five patients with TPP had a family history of thyroid disease versus six in the group without TPP. There was no history of TPP in a family member in either group.

Table 1. Baseline Clinical Characteristics of the Men with Graves Disease according to the Presence of TPP

Characteristic	Total (n=96)	Without TPP (n=48)	TPP (n=48)	P value
Age, yr	35.07±10.9	37.54±10.61	32.60±8.28	0.01
TSH, $\mu\text{U/mL}$	0.023±0.13	0.005±0.017	0.038±0.18	0.25
Free T_4 , ng/dL	4.03±1.56	3.83±1.39	4.19±1.67	0.32

Values are expressed as mean±SD.

TPP, thyrotoxic periodic paralysis; TSH, thyroid stimulating hormone; T_4 , thyroxine.

Arg16Gly polymorphism in the *ADRB2* gene

The genotype frequencies of the Arg16Gly polymorphism in the *ADRB2* gene were compared between patients with and without TPP. The Arg16/Arg16 genotype was found in 18 of 48 patients with TPP (38%) and 19 of 48 patients without TPP (40%) (Table 2). The Gly16/Gly16 genotype was identified in 7 of 48 patients with TPP (14%) and 12 of 48 patients without TPP (25%). There were no significant differences in the genotype of the Arg16Gly polymorphism between patients with and without TPP ($P=0.32$). The allele frequency did not differ (OR, 1.18; 95% CI, 0.64 to 2.2; $P=0.65$).

Number of CAG repeats in the *AR* gene

“Long AR” was defined as at least 26 CAG repeats in the promoter region of the *AR* gene, as previously described [11]. Long AR was found in 14 of 47 patients with TPP (30%) and 7 of 48 patients without TPP (15%) (Table 3). The number of CAG repeats in the *AR* gene was not associated with TPP (OR, 2.46; 95% CI, 0.81 to 8.09; $P=0.08$).

Table 2. Polymorphism in the β_2 -Adrenergic Receptor (*ADRB2*) Gene between Graves Disease Patients with and without TPP

	Without TPP (n=48)	TPP (n=48)	OR	95% CI	P value
Genotype frequency					
Arg16/Arg16	19 (0.40)	18 (0.38)			0.32
Arg16/Gly16	17 (0.35)	23 (0.48)			
Gly16/Gly16	12 (0.25)	7 (0.14)			
Allele frequency					
Arg16	55 (0.57)	59 (0.61)	1.18	0.64–2.2	0.65
Gly16	41 (0.43)	37 (0.39)			

Values are expressed as number (%).

TPP, thyrotoxic periodic paralysis; OR, odds ratio; CI, confidence interval.

Table 3. The Number of CAG Repeats (≥ 26 Repeats) in the Androgen Receptor (*AR*) Gene in Graves Disease Patients with and without TPP

	Without TPP (n=48)	TPP (n=47)	OR	95% CI	P value
Long AR ^a	7 (0.15)	14 (0.30)	2.46	0.81–8.0	0.08
Short AR ^b	41 (0.85)	33 (0.70)			

Values are expressed as number (%).

TPP, thyrotoxic periodic paralysis; OR, odds ratio; CI, confidence interval.

^aAt least 26 CAG repeats; ^bFewer than 26 CAG repeats.

Table 4. Allele Frequency of the Single Nucleotide Polymorphism (rs750841) in the γ -Aminobutyric Acid Receptor $\alpha 3$ Subunit (*GABRA3*) Gene in Graves Disease Patients with and without TPP

rs750841	Without TPP (n=48)	TPP (n=48)	OR	95% CI	P value
A	10 (0.21)	6 (0.13)	1.83	0.54–6.74	0.41
T	38 (0.79)	42 (0.87)			

Values are expressed as number (%).

TPP, thyrotoxic periodic paralysis; OR, odds ratio; CI, confidence interval.

SNP in intron 3 of the *GABRA3* gene (rs750841)

The SNP located in intron 3 of *GABRA3* gene was evaluated in the two groups (Table 4). One study reported that allele A was significantly associated with TPP cases [9]. However, 6 of 48 patients (13%) with TPP and 10 of 48 patients without TPP had allele A in rs750841 (Table 3). Therefore, there was no significant association between this polymorphism and TPP (OR, 1.83; 95% CI, 0.54 to 6.74; $P=0.41$).

DISCUSSION

In this study, we evaluated the associations of TPP with genetic variants in the *ADRB2*, *AR*, and *GABRA3* genes in men with GD. Recently, a GWAS of a Thai group suggested that a genetic variant in the *GABRA3* gene was related to TPP [9]. The SNP is rs750841, which is located in intron 3 between exons 3 and 4 of *GABRA3*. *GABRA3* encodes the GABA receptor $\alpha 3$ subunit, which is a member of the chloride channel family. That study found that allele A was significantly associated with TPP and identified putative thyroid hormone response elements. As it is located on the X chromosome, it might explain the sexual predominance in TPP. Therefore, we evaluated this polymorphism in our study, but found no significant association with TPP in Koreans with GD. Moreover, the allele frequency trend was also different. This may reflect ethnic differences.

The *ADRB2* gene encodes a type of *ADRB2* in skeletal muscle, and thyroid hormone sensitizes cells to β -adrenergic stimulation of the sodium pump [14,15]. Many studies have reported that an *ADRB2* gene polymorphism was associated with regulation of adrenergic activity. Several SNPs (-47T/C, Arg16Gly, and Gln27Glu) have been reported to be associated with increased *ADRB2* expression. Although the 47T/C and Gln27Glu polymorphisms are rare in Asian populations [16], the Gly16 *ADRB2* variant plays a role in endogenous agonist-induced downregulation [17]. It has been hypothesized that the Gly16/Gly16 allele in *ADRB2* is associated with TPP development, but that study was limited due to the small number of patients [10]. In this study, we evaluated the *ADRB2* gene allele frequencies in a relatively large cohort and found no significant correlation, consistent with our previous data [10,11].

Patients with TPP tend to experience symptoms after heavy carbohydrate meals [1]. This suggests that hyperinsulinemia enhances the sodium-potassium pump activity [3]. The number of polymorphic CAG repeats in the *AR* gene is independently and positively associated with the serum insulin concentration

[18]. Furthermore, there are ethnic differences in the number of CAG repeats of the *AR* gene. The mean number of CAG repeats is 21 to 22 in Caucasians and 22 to 23 in East Asians [19]. We thought that this ethnic difference could explain the different ethnic prevalence of TPP. However, having more CAG repeats in the *AR* gene was not associated with TPP.

This study had several limitations. The number of subjects limited the ability to provide statistical power to explain the genetic susceptibility of TPP in patients with GD. Moreover, it is difficult to generalize to the general population because this is a single-center study that included only Korean men with GD. However, our study is a relatively large cohort study that evaluated several candidate genes in patients with or without TPP who had confirmed GD. We were also limited because of the candidate gene approach used to evaluate the genetic predisposition to TPP. Further studies that evaluate the entire genomes of patients with or without TPP could provide definite evidence about the pathogenesis of TPP in patients with GD.

In conclusion, we evaluated genetic variants in the *ADRB2*, *AR*, and *GABRA3* genes in patients with or without TPP. These genetic variants could not explain the genetic susceptibility to TPP in Korean men with GD.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

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