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Enhanced Chitosan Photoluminescence by Incorporation of Lithium Perchlorate

Marlon Gurumendi, Floralba López, Luis J. Borrero-González, Thibault Terencio, Manuel Caetano, Carlos Reinoso, and Gema González*



1. INTRODUCTION

Chitosan (CS) is a semicrystalline polysaccharide derived from chitin, the second most important natural polymer that can be extracted from the shells of crustaceans. Due to its physical and chemical properties, it has been an attractive alternative for several technological applications, including biomedical, pharmaceutical, and environmental applications.¹⁻⁹ CS has also been used extensively as a membrane matrix owing to its excellent film forming with good mechanical properties.^{10,11} The structure comprises repeating units of N-acetyl D-glucosamine and D-glucosamine,¹² shown in Figure S1 in the Supporting Information.

Many hydroxyls and amine groups in polymers can act as electron donors,² facilitating the reactions with many organic and inorganic salts to form polyelectrolytes.^{1,5,13-17} Adding salt to polymers causes changes in the structure, and in optical, electrical, and thermal properties.^{1,14,18-20} In the case of chitosan, it provides ions as charge carriers that can improve the conductivity of the polymer, as has been reported in different works.^{1,21,22} The most used dopants in the preparation of many polymer electrolytes are lithium salts like lithium perchlorate,^{19,23,24} lithium tetraborate,²⁵ lithium hexafluorophosphate, and lithium trifluoromethanesulfonate.^{26,27} Lithium salts are preferred due to their low interfacial strength and ability to coordinate and solvate easily, exhibiting their ionic conductivity character. The advantage of incorporating these salts is that a complexation can be obtained when the polymer electrolytes are prepared due to their high dissociation energy and good solubility in most solvents.¹

The possibility of obtaining a biocompatible organic photoluminescent system is very attractive for bioimaging applications.^{28–31} The photoluminescence properties of chitosan have attracted much attention due to its low toxicity and good biocompatibility, which offers applications as imaging probes. The photoluminescence of chitosan has been reported by some authors,^{9,31–38} taking advantage of photoluminescence spectroscopy nondestructive nature.^{39–41} Although, in general, fluorescence in macromolecules is caused by molecules with delocalized electrons at conjugated double bonds,^{38,42} the CS oligomer featured a molecular structure that has no delocalized electrons. However, they have shown fluorescent properties.^{32,38}

Some amino-containing polymers, such as chitosan, have been reported to present a photoluminescent effect.^{9,32,36,43–46} CS has an emission band in the visible spectrum at 465 nm (with a laser excitation of 369 nm), attributed to its amine groups.⁴⁵ Furthermore, CS can be either stabilized or doped with luminescent agents,^{32–34,38,46,47} adding optically active agents luminescent nanoparticles, such as quantum dots,^{39,48,49} plasmonic nanoparticles (gold or silver),^{6,50} organic complexes,^{51–53} or polymeric nanoparticles.⁵⁴ CS's most common

Received:December 19, 2022Accepted:March 16, 2023Published:April 5, 2023



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Article



© 2023 The Authors. Published by American Chemical Society luminescent systems use lanthanide complexes, such as europium or terbium complexes, to get good luminescence properties for possible applications in electrochemical devices.^{33,46,55}

The addition of lithium salts to CS has been studied mainly to improve ionic conductivity,^{14,19,56,57} however, as far as we know, the photoluminescent effect of lithium salts in CS matrix has been scarcely reported. Therefore, looking for new hybrid biocompatible and environmentally friendly compounds with good film forming capability and high luminescence, chitosan with different amounts of LiClO₄ has been studied, which complement the well-known high ionic conductivity studies. In this work, experimental evidence is complemented for the first time with theoretical studies, and mechanisms that satisfy both observations are proposed to get new insight into the problem.

1.1. Methodology. *1.1.1. Materials.* CS flakes from shrimp shells (>75% deacetylation degree) were purchased from Sigma-Aldrich. Lithium perchlorate was received as a white crystalline salt from Sigma-Aldrich and acetic acid (glacial) 100% from Merck. Distilled water was used throughout the experiment, and all samples were prepared using the solution-casting technique.

1.1.2. Films Preparation. Chitosan solution 1% in acetic acid was prepared under magnetic stirring until the chitosan powder was completely dissolved. For pure chitosan film, the chitosan solution was stored in a plastic Petri dish and left to dry at room temperature. For hybrid films, lithium perchlorate salt was added to the chitosan solution to have 10, 25, 30, and 40% w/w in weight of lithium perchlorate salt concerning chitosan weight in the solution. After 15 min of stirring, the solution was poured into a plastic Petri dish and dried at room temperature for 3 days. The film thickness was measured using a digital micrometer with a precision 0.001 mm. The values obtained for each sample at different locations were averaged. Table S1 in Supporting Information summarizes the prepared hybrid compounds. For clarity and simplicity, the term XLiClO₄/CS will be used, where X refers to the composite percentage ratio of LiClO₄ weight relative to chitosan weight.

1.2. Structural and Morphological Characterization. *1.2.1. Scanning and Transmission Electron Microscopies.* The morphology of the samples was observed using a scanning electron microscope (SEM) FEI, INSPECT F50, and by transmission electron microscopy (TEM) with a Tecnai F20, operating at 200 kV. The samples were prepared by ultramicrotomy, using a Reichert FCS ultramicrotome with a cryo-sectioning accessory.

1.2.2. Fourier Transform Infrared Spectroscopy. The functional groups of chitosan hybrid films were characterized by Fourier transform infrared spectroscopy (FTIR). The spectrophotometer was a PerkinElmer model Spectrum 100, and the data of the samples were acquired in the absorbance mode at room temperature, in the range from 450 to 4500 cm⁻¹.

1.2.3. X-ray Diffractometry. The diffraction patterns of the samples were obtained with X-ray diffractometer PANalytical X'Pert Pro by using X'Pert High Source Plus and X'Pert Data Viewer programs. The equipment was operated using CuK α radiation source $\lambda = 1.542$ Å at 40 kV and 30 mA. The 2 θ angles were scanned between 10° and 70°, with a step size of 0.1°. The degree of crystallinity (χ_c) of the polyelectrolyte was determined by using eq 1:^{13,58}

$$\chi_C = \left[\frac{A_C}{(A_C + A_A)}\right] \times 100\% \tag{1}$$

where A_C and A_A are the areas of crystalline and amorphous peaks, respectively. The determination of amorphous and crystalline peaks was done by deconvolution of the peaks method. Baseline correction has been done before fitting multipeaks using the Gaussian function.

1.2.4. X-ray Photoelectron Spectroscopy. X-ray photoelectron spectroscopy surface analysis was performed on the pristine chitosan and also on each of the final materials having different percentages of lithium incorporated. For these measurements, a PHI VersaProbe III from Physical Scientific was used, equipped with a 180 hemispherical electron energy analyzer; using a monochromatized Al K α source with energy 1486.6 eV operated with an energy band-pass 255 kV for the survey acquisition and 55 kV for high-resolution acquisition. Spot analysis had a diameter of 100 μ m and the detection angle relative to the substrate surface was 45°. Analyses and fitting were performed using peak fit version 4.7 and Qtiplot version 0.9.8.9.

1.3. DFT Calculation. Density functional theory (DFT) calculations were conducted using the software Orca^{59} on a representative glucosamine unit of the material alone and with Li^+ ion in different positions. The functional B3LYP^{60,61} in conjunction with a Def2-TVZP (triple ζ) basis-set were used.^{62,63} Dispersion interactions were included through Grimme approach DFT-D3 with Becke–Johnson damping (D3BJ).^{64,65} All optimized structures were ensured to be minimal through the calculation of their frequencies. Calculations, including solvation, were done using an implicit CPCM model of water.⁶⁶ The simulated UV–vis spectra were generated using the TD-DFT method.

1.4. Optical Properties. *1.4.1. UV–Vis Spectroscopy.* UV–vis spectra of the composite films were collected in total transmittance mode using a UV/vis/NIR double beam spectrometer from PerkinElmer, model Lambda 1050, equipped with a 150 mm integration sphere accessory, in the wavelength range from 200 to 600 nm. Data analysis and visualization were performed using an application developed in R statistical programming language (https://github.com/mcaetanove/chemometrics.git). UV–vis spectra were smoothed by an adaptive-degree polynomial filter, using a maximum polynomial order of 4 and a data window of 11.⁶⁷

The optical absorption for noncrystalline materials is commonly modeled by the Tauc and Davis-Mott equation, $^{68-70}$ given by

$$(\alpha h\nu) = \beta (h\nu - E_g)^{\gamma} \tag{2}$$

where $h\nu$ is the photon energy, h is Planck's constant, ν is the photon frequency, and α is the absorption coefficient. E_g is the energy bandgap (Tauc's bandgap), β is a constant that depends on the probability of transition, and γ is an index that describes the type of electronic transition which could be assumed to be 1/2 for allowed direct optical band gap or 2 for the allowed indirect optical band gap.

The optical absorption coefficient α was calculated according to Lambert's Absorption Law:

$$\alpha(t) = \left(\frac{2.306}{t}\right) \times \text{Abs}$$
(3)

where Abs is the absorbance and t is the thickness of the sample.

1.4.2. Photoluminescence Spectroscopy. The photoluminescence (PL) spectra were recorded on a High-Resolution spectrometer HR4000CG-UV-NIR using the program Ocean Optics SpectraSuite. An excitation wavelength of 400 nm from a diode laser and an integration time of 5 s was used.

2. RESULTS AND DISCUSSION

2.1. Structural and Morphological Analysis. *2.1.1. XRD.* The XRD patterns of the films of pure chitosan and chitosan loaded with different concentrations of LiClO_4 are presented in Figure 1. For pure chitosan, the diffractogram presents the characteristic diffraction peaks at 2θ 11.94° and 18.70°, assigned to intramolecular and intermolecular hydrogen bonds.⁷¹ The films with different concentrations of LiClO₄, as can seen in Figure 1, show slight shifts in 2θ to lower angles from 11.94° to 11.87° and from 18.70° to 18.46°,



Figure 1. Diffractogram of different prepared films among the respective deconvolution to evaluate the crystallinity of different prepared films.

respectively. Additionally, a decrease in the intensity of these peaks is observed as the LiClO₄ concentration increases. The halo in the 2θ region between 15° and 30° becomes broader, especially for the case of the sample with 40LiClO₄/CS, suggesting an increase in the amorphous structure of chitosan.^{13,72} The broadening, shifting, and lowering of the relative intensity of chitosan diffraction peaks with the incorporation of LiClO₄ can be attributed to the complexation of LiClO₄ in the polymer chains, occurring between the functional groups of chitosan and the ions of the dopant salt,^{27,73,74} resulting in an increase amorphization with the increasing addition of LiClO₄. The above agrees with previous studies on chitosan doped with different salts that reported an increase in amorphization with the addition of salts.^{13,19,27,74–76}

The degree of crystallinity, denoted by χ_{c} of the prepared films was determined by eq 1 are indicated in each diffractogram in Figure 1, suggesting that the amorphousness of the poly electrolyte films increased with the salt concentration. The initial chitosan sample (without lithium) had a higher crystallinity percentage, ~28%, progressively decreasing to a value of ~2% for the sample with the highest salt concentration.

2.1.2. SEM and TEM. The morphology of chitosan thin film with different concentrations of LiClO_4 is presented in the SEM images, taken in electron backscattered mode (Figure 2(a-f)). The image of the pure chitosan film, presented in Figure 2(a), shows a smooth and homogeneous nonporous structure. As the LiClO_4 concentration increases, the matrix shows the formation of a second phase.



Figure 2. SEM micrographs of different prepared films. Samples (a) pure chitosan, (b) $10LiClO_4/CS$, (c) and (d) $25LiClO_4/CS$, (e) $30LiClO_4/CS$, (f) $40LiClO_4/CS$. TEM micrographs of different prepared films (g) $25LiClO_4/CS$ and (h) $40LiClO_4/CS$.

For $10LiClO_4/CS$ film, small clusters (<10 nm) with a very uniform distribution are presented in Figure 2(b). For 25LiClO₄/CS the formation of some spherulites⁷⁷ can be seen at low magnification (Figure 2(c)), while at high magnification (Figure 2(d)) a higher density of the small clusters between 50 and 100 nm can be identified, clearly seen by TEM micrographs presented in Figure 2(g). For $30LiClO_4/$ CS film, similar-sized aggregates are present (Figure 2(e)). For 40LiClO₄/CS, a higher density of clusters whose sizes are in the range of 200-400 nm can be identified in Figure 2(f), which is also seen by TEM (Figure 2(h)). It seems that for low LiClO₄ concentration, the acid polysaccharide molecules from CS react with the ionic species forming small aggregates of less than 10 nm (Figure 2(b)) as the $LiClO_4$ concentration increases the aggregate size and density. The resulting structures might be stabilized as the local supersaturation increases, forming nanoclusters with size in the range of 50-400 nm, as can be seen in Figure 2(g,h). Similar cluster sizes have been reported for different authors.53,78

2.1.3. FT-IR. FT-IR studies were conducted to follow the chemical modifications in chitosan structure upon interaction with $LiClO_4$ (Figure 3). The chitosan film spectrum (Figure



Figure 3. FTIR spectra of (a) pure chitosan, (b) $10LiClO_4/CS$, (c) $25LiClO_4/CS$, (d) $30LiClO_4/CS$, and (e) $40LiClO_4/CS$.

3(a)) shows a broad band centered at 3475 cm⁻¹ assigned to the stretching vibration of the hydroxyl group; the presence of the broadband at 3244 cm⁻¹ is attributed to the N–H stretching.⁷⁹ The absorbance band at 2907 cm⁻¹ is associated with C–H symmetric and asymmetric stretching. The peak at 1652 cm⁻¹ is assigned to the C=O stretching in NH–CO– CH₃ group (amide I).⁷⁹ The pH of the precursor suspension of the film constrains the protonation degree of NH group, for pH < 6.5 protonation is favored with the corresponding bending vibrations of the protonated amine group appearing at 1550 cm⁻¹, while the N–H band in the amide group (amide II) appears at a higher frequency around 1560 cm^{-1.79}

The stretching vibration of C–H, CH₂, and CH₃ groups are observed at 1412 cm⁻¹ and around 1375 cm⁻¹. The peak at 1151 cm⁻¹ corresponds to the C–O–C bridge asymmetric stretching mode.^{79,80} The signals in the region around 1080– 1025 cm⁻¹ correspond to the stretching vibration of C–O–H, C–O–C, and CH₂–O–H present in the pyranose ring. The peak observed at 665 cm⁻¹ is ascribed to the N–H twist vibration. Table S2 shown in Supporting Information presents the FTIR peak assignments of chitosan. The different XLiClO₄/CS FTIR spectra (Figure 3(b–e)) show all the main signals of pure chitosan, although some shifts with respect to pure chitosan can be observed. In the chitosan spectrum, the O–H stretching resonance band is significantly broad and centered at 3500 cm^{-1} , clearly signaling the presence of intramolecular hydrogen bonds.⁸¹

To gain more insight into the structural features, the second spectra derivative was taken applying the Norris-Williams Derivative Filter (5 smoothing points and five differential gaps).⁸² This procedure enriches the qualitative interpretation of the FTIR results and provides a basis to discuss the effect of LiClO₄ interaction with chitosan moieties. Figure 4(a,b) shows the second derivative of the FTIR spectra at 3700–3500 cm⁻¹ and 1590–1500 cm⁻¹, respectively, for CS and XLiClO₄/CS.



Figure 4. (a) Second derivative of $3900-3500 \text{ cm}^{-1}$ and (b) $1590-1500 \text{ cm}^{-1}$ spectral regions for different concentrations of LiClO₄.

In the presence of LiClO₄ a new absorption band appears around 3640 cm⁻¹ in the O–H stretching region. The blueshifting of this band was accentuated as the LiClO₄ concentration increased, which seems to suggest an interaction of Li⁺ ions with the OH⁻ of chitosan. This interaction may lead to hydroxyl groups being released from the hydrogen-bond network, the O–H bond length decreases, the constant force increases, and the frequency suffers a blue-shift.⁸¹ In addition, the presence of LiClO₄ causes a shift of the band associated with the N–H bending of amine to lower wavenumbers to the pure chitosan, as can be seen in Figure 4(b). This may be associated with Li⁺ and ClO₄⁻ ions binding to the amine groups.^{19,57,76}

2.1.4. XPS. The survey spectra of $40\text{LiClO}_4/\text{CS}$ and chitosan films are presented in Figure 5(a) and (b), respectively, exhibiting clear features at 56 eV (Li 1s) and 199 eV (Cl 2p) confirming the presence of lithium and chlorine, respectively.^{35,83–88} A magnification of this eV region is presented in the inset of Figure 5(a), so the Li 1s peak at 56 eV can be seen. In contrast, in the inset in Figure 5(b), showing the same eV region but for pure chitosan, this peak is not present, as expected.



Figure 5. XPS survey spectra of prepared films, (a) $40\text{LiClO}_4/\text{CS}$ and (b) pure chitosan. XPS spectra of Li 1s core levels: (c) 40%, (d) 30%, (e) 25%, and (f) 10% of w/w LiClO} $_4/\text{CS}$ films; and N 1s core levels: (g) CS, (h) 10%, (i) 30%, and (j) 40% of w/w LiClO} $_4/\text{CS}$ films. A decrease of counts of NH₂ at 397.9 eV is shown while an increasing signal at 400.2 eV by the incorporation of lithium in the sample is seen.

Different concentrations for the lithium chemical environment are revealed with an improved resolution and displayed in Figure 5(c-f), where a deconvolution process of the four samples is presented. The fitting was performed without background subtraction, using a Voigt adjustment. An error of ± 0.2 eV is due to the data dispersion and to XPS resolution ~0.5 eV. The high resolution spectra recordings lead to finding LiOH, Li₂O, and LiClO₄ at 54.9, 55.6, and 56.4 eV (± 0.2 eV), respectively, indicated in Figure 5(c).^{85–87} The XPS cross-section for lithium is quite small, the reason the counts are very low; however, the XPS technique is able to detect low contributions of lithium.³⁵ The scale of intensity of spectrum for sample 10LiClO₄/CS (Figure 5(f)), has been enlarged twice (2×) for a better visualization. The presence of lithium compounds formed by combination with oxygen and chlorine confirms the existence of lithium in the different samples in

different concentrations. Table 1 presents the comparative contributions of each lithium compound in each sample. This suggests an ionic exchange between chitosan electron charge and lithium perchlorate.

Table 1. Chemical Environment Contr	ribution of Lithium
Dispersed in Chitosan from Fitting XI	PS Spectra

	Lithium species detected		
Sample	LiOH (at.%)	Li ₂ O (at.%)	LiClO ₄ (at.%)
CS	-	-	-
10LiClO ₄ /CS	39.5	60.45	<0.1
25LiClO ₄ /CS	23.15	53.69	<0.1
30LiClO ₄ /CS	43.1	48.39	8.28
40LiClO ₄ /CS	38.4	52.60	8.98

The N 1s core level (Figure 5(g-j)) exhibits a clear interaction between the NH₂ and the lithium, whose presence shifts the binding energy from 397.9 to 400.2 eV that appears in correspondence with the concentration raised of lithium in the sample. This suggests the formation of species $-NH_2-Li^+$ with higher energy bonding. At the same time the binding energy assigned to the -N-C- interaction at 398.8 eV remains almost constant in agreement with the molecular stability of chitosan.^{89,90}

2.2. DFT Calculations. Chitosan is composed of alternating units of β -(1-4)-D-glucosamine and N-acetyl-Dglucosamine. The ratio between one and the other depends on the degree of acetylation, which can be tuned through the preparation medium used. With this consideration in mind, theoretical calculations were conducted on one glucosamine unit⁹¹ considering three different models possible: (i) the protonated form NH_{3} (ii) the normal glucosamine form NH_{2} and (iii) the acetylated form. Using the IR spectrum for each form, the most stable chair conformer and boat conformer were calculated, and they are shown in Figure S2 in the Supporting Information. The comparison with experimental FTIR spectra allows concluding which form is present in the evaluated material. It should be noted that the experimental spectra are much more complex than the theoretical ones as each unit constituting the chitosan could be in a different form, different conformation, and linked to another unit in a different manner.

As expected, the spectra of the three different forms display similar features: a very intense band at 1050 cm⁻¹, two around 3000 cm⁻¹, and two bands at 1450 and 1520 cm⁻¹. However, the acetylated form presents an intense band at 1700 cm⁻¹ characteristic of C=O stretching. This particular feature does not seem to appear in the experimental spectrum suggesting few units are present in the acetylated form. For NH₂ and NH₃ forms, it is difficult to judge which one is more abundant only from the IR spectrum as both present very similar features. However, by careful inspection, the NH₂ form seems more abundant, most notably by the presence of the N-H twist band at 650 cm⁻¹. While the calculations do not take into account solvent effects such as hydrogen bonding with solvent molecules, and hence the calculated N-H and O-H are considerably shifted, it seems that the broadband at higher wavenumber in the experimental spectrum does not correspond only to the different glucosamine forms. This discrepancy could be explained due to the experimental procedure where some water can also be present in the final material. In order to probe this hypothesis, a lone water



Figure 6. Structures of glucosamine with different location sites for lithium and their associated simulated IR-spectra. Atom colors — O: red, N: blue, C: gray, H: white, Li: pink.

molecule, a glucosamine unit with one or two water molecules with explicit and implicit solvation mimicking water was simulated.

Water molecules normally possess 3 normal vibrations modes: O–H stretching symmetric, O–H stretching asymmetric, and H–O–H bending. The water molecules were placed close to the oxygen and nitrogen of the glucosamine unit allowing H-bonding. Spectra calculated for NH_2 forming glucosamine units with these water molecules in different locations are shown in Figure S3 in Supporting Information, displaying a broad band in the higher region, confirming the aforementioned hypothesis. The shift observed between the experimental and calculated frequency for O–H stretching of water is due to the model being limited to only one or two water molecules and the calculation method, but it is wellknown that the frequency corresponding to this vibration should be at lower wavenumbers.

Five different structures, varying the location sites of lithium adsorption on CS, and considering glucosamine units in the NH₂ form, are depicted in Figure 6. In position 1, lithium interacts with three oxygen atoms. In positions 2 and 4, the lithium ion only interacts with one hydroxyl group. Position 3 involves the interaction with both hydroxyl and methyl groups, while in position 5 the lithium ion is positioned between nitrogen and oxygen atoms. Each different structure was optimized, and the frequencies calculated all positives, to ensure minimum energy structures were obtained. One remark is that all optimizations lead to positions close to groups that are more electronegative, e.g., N and O atoms. Logically, this can be explained by considering that Li⁺ establish electrostatic interactions with the partial negative charge in the chitosan.

In the five location sites of lithium considered, there are three different kinds of groups interacting with Li ion: C–OH, NH₂, and C–O–C. Two regions are affected by the change of position of lithium ion, one low region between 400 and 800 cm⁻¹ and the upper region between 3000 and 4000 cm⁻¹. This observation is corroborated by the experimental results where most changes occur also in these regions. In fact, the higher region seems to be more affected by the lithium presence. The positions where the most changes are observed in this region are positions 3 and 4; a very intense band appears at 3460 and 3000 cm^{-1} , respectively. Interestingly, this band corresponds to the O–H stretching in both cases where the lithium is close to this group. However, lithium is also close to an OH group in position 1, but this very intense band is not observed. The main difference between position 1 and position 4 is that the group OH is involved in an intramolecular H-bond in the latter. By the comparison between theoretical and experimental results, we can affirm that lithium is close to groups OH establishing H-bonds and particularly affecting it.

There is a clear interplay between lithium position and Hbonds. In our calculations, only one unit of glucosamine was considered and we observe only intramolecular H-bond, but this effect would probably be the same in the case of intermolecular H-bonds. Lithium ions could play a role in forming the aggregates with chitosan, and probably in the luminescence, not only through electrostatic interaction but also by affecting the H-bonds. Figure 6 shows simulated IR spectra of species with lithium in different positions on the boat form of acetylated species.

Energetically, positions 2 and 5 were found to be the most stable; with a difference of only 4 kJ/mol in favor of position 5. Positions 3 and 1 are less stable than positions 5 by 14 and 17 kJ/mol, respectively. Position 4 is clearly less stable with a difference of 115 kJ/mol higher, highlighting a clear preference of the Li⁺ for more than one neighboring electronegative atom. The exact same tendency is observed when considering Gibbs energies shown in section 3 of Supporting Information (Tables S3 and S4). However, the experimental spectrum presents very broad bands in this region and renders it impossible to conclude the exact location of lithium ion and is probably a combination of the most stable positions, namely, 5 and 2. When considering an implicit solvent model position 5 is clearly more stable than all the other positions (by at least 20 kJ/mol).

Finally, we simulated the different lithium species which could be formed: $LiClO_4$, LiOH, Li_2O , and Li_2CO_3 . Although the comparison with the experimental spectra does not permit us to conclude unequivocally on the presence of one or several of these species, the XPS experimental results showed mainly the presence of LiOH, Li_2O associated with the groups N and OH from chitosan. Section 4 of the Supporting Information shows the calculated IR spectra for these lithium species (Figure S4).

2.3. Optical Properties. 2.3.1. UV-vis Spectroscopy. The UV-vis absorbance spectra of chitosan films containing 0, 10, 25, 30, and 40% w/w LiClO₄/CS are shown in Figure 7(a).



Figure 7. (a) UV–vis absorbance spectra of $XLiClO_4/CS$ and (b) optical absorption coefficients of pure and loaded CS samples as a function of photon energy.

The spectra show two relatively broad absorption bands centered at ca. 250 and 310 nm, which are associated with the $n - \pi^*$ and $\pi - \pi^*$ transitions of the conjugated ring system.^{49,92-94} A positive correlation between optical absorption intensity and incorporation of LiClO₄, as well as a minimal bathochromic shift (redshift) ≈ 4 nm, is observed, related to light scattering from Li⁺/CS aggregates; this is in agreement with SEM and TEM observations. Figure 7(b) represents the optical absorption coefficients of pure and loaded chitosan samples as a function of photon energy.

Table 2 shows the absorption edge values determined by extrapolating the linear portion of the spectra to a zero

Table 2. Estimated Absorption Edges and Optical Bandgaps from XLiClO₄/CS Composite Films

		$E_{g,d} [eV]^a$	$E_{g,i} [eV]^a$
Sample	Absorption Edge [eV] ^a	$\gamma = 1/2$	$\gamma = 2$
CS	5.46	5.66	5.19
10LiClO ₄ /CS	5.41	5.62	4.99
30LiClO ₄ /CS	5.31	5.60	4.85
40LiClO ₄ /CS	5.20	5.51	4.75
^a Standard Deviatio	n = 0.01.		

absorption value from the spectra shown in Figure 7(b). The absorption edge of the chitosan films slightly shifts toward the lower photon energy as the $LiClO_4$ concentration in the chitosan matrix increases.

The optical absorption for noncrystalline materials is commonly modeled by the Tauc and Davis-Mott equation.

Although polymeric materials have highly complicated electronic structures Tauc models could be applicable.^{95,96} The fluctuation of the polymeric lattice configuration blurs the fundamental edge and causes the presence of an Urbach tail. Also, the formation of solutions produces discrete energy levels in the bandgap, which will be reflected in the absorption coefficient.

Allowed direct and indirect transition energies, $E_{g,d}$ and $E_{g,\nu}$ were obtained from both $(\alpha h\nu)^2$ vs $h\nu$ (photon energy) and $(\alpha h\nu)^{1/2}$ vs $h\nu$ plots, respectively, and by extrapolating the linear parts of the curves until they intersect on the *x*-axis (zero absorption), as shown in Figure 8. For pure chitosan, direct and indirect bands lie at 5.66 and 5.19 eV, respectively, while for XLiClO₄/CS doped films, the values are given in Table 2.

From Table 2, it can be seen that the band edge and both direct and indirect band gaps decrease with increasing LiClO_4 concentration. It would be expected that the incorporation of LiClO_4 does not alter the chitosan band gap drastically, only some shifts in the bands attributable to electron-hole interaction between valence and conduction band were observed.

2.3.2. PL Spectroscopy. The photoluminescence (PL) spectra recorded at room temperature of chitosan and chitosan with different LiClO_4/CS percentages (10, 25, 30 and 40%w/w) are shown in Figure 9(A). All spectra show a broad emission band in the visible range. With the incorporation of the salt, the integrated area increases linearly with the subsequent loading (see inset in Figure 9(A)), where the sample loaded with 40% has an integrated area of about 3 times the integrated area of the sample of pure chitosan.

The increase in emission intensity with the incorporation of LiClO_4 can be attributed to the ability of chitosan to chelate metal ions due to its amino and hydroxyl groups. The complexation between chitosan and LiClO_4 has been demonstrated by FTIR, XPS, and DFT calculations, where a shift of the N–H vibrations was observed, and the enhancement of the vibration band located at 1144 cm⁻¹ and of the band at 1067 cm⁻¹, suggests complexation with $\text{Li}^+\text{ClO}_4^-$ and the interaction of the amino groups of chitosan with these species.

A deconvolution was performed to the PL spectra (Figure S5 in the Supporting Information), and based on the peak positions of the fits, an energy band diagram was constructed for the evaluated system; see Figure 9(B). After optical excitation at 400 nm (3.10 eV) within tail states^{70,97,98} below the conduction band, the excitation relaxes via nonradiative processes until finding luminescent centers with a subsequent relaxation to tail states above the valence band. These luminescent centers are related to defect states and are located in the intervals 436–444 nm (2.84–2.79 eV), 491–494 nm (2.52–2.50 eV), and 528–536 nm (2.35–2.31 eV).

The luminescence mechanism could be explained by aggregation-induced emission (AIE) phenomena, where aggregate formation can restrict the intramolecular motions turning on luminescence. The latter can effectively dissipate exciton energy and the radiative transition is activated. TEM analysis (Figure 2) shows the formation of dark clusters in the polymer matrix, with an average size of 50 nm for the composite film $25\text{LiClO}_4/\text{CS}$ and in the range of 120-300 nm for $40\text{LiClO}_4/\text{CS}$. Therefore, the larger the cluster the higher the luminescence intensity (Figure 9(A)). On aggregation, the molecules become rigidified, this kind of luminescence is



Figure 8. Tauc plot for allowed direct—(a) pure chitosan and (b) composite films (10, 30, 40)LiClO₄/CS—and indirect—(c) pure chitosan and (d) composite films $XLiClO_4/CS$ —transitions.



Figure 9. (A) Photoluminescence spectra for different samples of the evaluated films. On the upper left side, a photograph of one of the tested samples when it is being irradiated is shown. On the upper right side, the increasing linear trend of the integrated area can be observed with different percentages of $LiClO_4$. (B) Band diagram.

known as rigidification-induced emission (RIE) and has been observed in different natural polymers. 99,100

On the other hand, the increase in the amorphization character observed by XRD (Figure 1) with the addition of $LiClO_4$ is associated with the increase of inter- and intramolecular charge transfer transitions between lithium ions and the amine and OH groups of chitosan via stretching vibration.¹ Amorphization and cluster formation cause suppression of molecular motions in the glassy matrix, and a rigidification of luminogenic molecules occurs. A similar photoluminescence mechanism has been reported in cotton fibers.¹⁰¹ With the addition of LiClO₄ to chitosan, there are numerous electron-rich heteroatoms with lone-pair electrons in

their molecular structures. The rigidified conformation during cluster formation can become highly luminescent by forming clustoluminogens since the aggregation promotes intra- and intermolecular interaction with the Li^+ and ClO_4^- ions.

Figure 9(B) shows the band diagram associated with the transitions observed during the PL of the different samples. The distributions of conduction, valence, and electronic states in chitosan do not end abruptly at the band edges. Instead, due to defects (amorphous material), some localized states, called tail states, appear deep in the gap region. Defects in chitosan come from dangling bonds, holes, pores, ruptures, and other macroscopic defects. The increase in PL emission with the LiClO₄ concentration could be attributed to the weakening of

hydrogen bonds, increasing the amorphization (defects), and, consequently, the density of tail states. IR spectra, XRD, and theoretical calculations support the above.

Furthermore, it can be seen that the wavelength of the emission peak for the chitosan sample is 493.58 nm, and with the incorporation of the LiClO_4 the peak wavelength increases to 496.16 nm, which corresponds to a peak wavelength shift on the order of 2.6 nm. This shift can be due to a compression of the emitting energy levels of chitosan due to the formation of aggregates.

3. CONCLUSIONS

The incorporation of LiClO₄ affects the structure and, consequently, the optical properties of chitosan, as there is a strong interplay between both phenomena. The complexation between chitosan and Li⁺ has been demonstrated by FTIR, XPS, and DFT calculations, where the specificity of Li⁺ for more electronegative atoms (O and N) was deduced. Moreover, DFT calculations and IR results show that the presence of Li⁺ in the close vicinity of an H-bond would influence the O-H and N-H stretching and, subsequently, the strength of the H-bonds. Since the crystalline structure of chitosan strongly depends on the inter- and intramolecular hydrogen bonds between the OH and NH₂ groups, introducing Li⁺ would logically affect its crystallinity. Indeed, an increase in the amorphization character of the hybrid material, along with the addition of $LiClO_4$, is observed from XRD results.

Concerning the optical properties, the absorption and emission of light have been observed to increase with the incorporation of lithium. While the optical band gap is large for chitosan, small transitions can be observed at energy differences inferior to the band gap due to the presence of tail states. Incorporating lithium, the decrease in crystallinity generates structural defects, which would increase the number of these tail states. Apart from the tail states, the intensity corresponding to each absorption band in UV-vis increase with LiClO₄ incorporation. The increase can be explained by the aggregation of polymeric parts around Li⁺, increasing the local concentration of absorbing groups and allowing the polymeric material to absorb more photons, which permits, in turn, display of higher luminescence. The promising results of optical properties open new possibilities in tailoring the design of molecules. The route used in this work could be generalized to different polymers bearing electronegative atoms such as N or O: introducing lithium to form local aggregates resulting in improved UV-vis photon absorption and, thus, its luminescence.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsomega.2c08072.

Additonal experimental details (PDF)

AUTHOR INFORMATION

Corresponding Author

Gema González – Yachay Tech University, School of Physical Sciences and Nanotechnology, Urcuquí 100119, Ecuador; Centro de Ingenieria de Materiales y Nanotecnologia, Instituto Venezolano de Investigaciones Científicas, Caracas *1020A, Venezuela;* orcid.org/0000-0003-4526-2429; Email: ggonzalez@yachaytech.edu.ec

Authors

- Marlon Gurumendi Yachay Tech University, School of Physical Sciences and Nanotechnology, Urcuquí 100119, Ecuador
- Floralba López Yachay Tech University, School of Chemical Sciences and Engineering, Urcuquí 100119, Ecuador
- Luis J. Borrero-González Pontificia Universidad Católica del Ecuador, Facultad de Ciencias Exactas y Naturales, Escuela de Ciencias Físicas y Matemática, Laboratorio de Óptica Aplicada, Quito 17-01-2184, Ecuador; © orcid.org/ 0000-0003-3853-5433
- Thibault Terencio Yachay Tech University, School of Chemical Sciences and Engineering, Urcuquí 100119, Ecuador
- Manuel Caetano Yachay Tech University, School of Chemical Sciences and Engineering, Urcuquí 100119, Ecuador; Universidad Central de Venezuela, Facultad de Ciencias, Escuela de Química, Caracas 1020A, Venezuela; orcid.org/0000-0002-8891-3140
- Carlos Reinoso Yachay Tech University, School of Physical Sciences and Nanotechnology, Urcuquí 100119, Ecuador

Complete contact information is available at: https://pubs.acs.org/10.1021/acsomega.2c08072

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work employed the Imbabura cluster of Yachay Tech University, which was purchased under contract No. 2017-024 (SIE-UITEY-007-2017). The authors would like to acknowledge the internal grants CHEM19-15 and Proyecto 6: "Estudio teórico de los efectos geométricos del ámbito cercano de los enlaces hidrógenos y su efecto en la interacción con la luz" of Yachay Tech University for supporting this work and providing the facilities required to carry out this work, as well as to Professor César Costa (Escuela Politécnica Nacional in Ecuador), for his collaboration with the photoluminescence measurements.

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