




Review

Intestinal Chelators, Sorbants, and Gut-Derived Uremic Toxins

Solène M. Laville ¹, Ziad A. Massy ^{1,2}, Said Kamel ^{3,4}, Jean Marc Chillon ^{3,5}, Gabriel Choukroun ^{3,6} and Sophie Liabeuf ^{3,5,7,*}

- ¹ Centre for Research in Epidemiology and Population Health (CESP), INSERM UMRS 1018, Université Paris-Saclay, F-94807 Villejuif, France; solene.laville@inserm.fr (S.M.L.); ziad.massy@aphp.fr (Z.A.M.)
- ² Department of Nephrology, Ambroise Paré University Hospital, APHP, Boulogne Billancourt, F-92100 Paris, France
- ³ MP3CV Laboratory, EA7517, Jules Verne University of Picardie, F-80000 Amiens, France; said.kamel@u-picardie.fr (S.K.); jean-marc.chillon@u-picardie.fr (J.M.C.); choukroun.gabriel@chu-amiens.fr (G.C.)
- ⁴ Department of Biochemistry, Amiens University Medical Center, F-80000 Amiens, France
- ⁵ Direction of Clinical Research, Amiens University Medical Center, F-80000 Amiens, France
- ⁶ Department of Nephrology Dialysis Transplantation, Amiens University Medical Center, F-80000 Amiens, France
- ⁷ Department of Clinical Pharmacology, Amiens University Medical Center, F-80000 Amiens, France
- * Correspondence: liabeuf.sophie@chu-amiens.fr; Tel.: +33-322-087-995

Abstract: Chronic kidney disease (CKD) is a highly prevalent condition and is associated with a high comorbidity burden, polymedication, and a high mortality rate. A number of conventional and nonconventional risk factors for comorbidities and mortality in CKD have been identified. Among the nonconventional risk factors, uremic toxins are valuable therapeutic targets. The fact that some uremic toxins are gut-derived suggests that intestinal chelators might have a therapeutic effect. The phosphate binders used to prevent hyperphosphatemia in hemodialysis patients act by complexing inorganic phosphate in the gastrointestinal tract but might conceivably have a nonspecific action on gut-derived uremic toxins. Since phosphorous is a major nutrient for the survival and reproduction of bacteria, changes in its intestinal concentration may impact the gut microbiota's activity and composition. Furthermore, AST-120 is an orally administered activated charcoal adsorbent that is widely used in Asian countries to specifically decrease uremic toxin levels. In this narrative review, we examine the latest data on the use of oral nonspecific and specific intestinal chelators to reduce levels of gut-derived uremic toxins.

Keywords: uremic toxins; phosphate binders; chronic kidney disease

Key Contribution: Even though gut-derived uremic toxins appear to be valuable therapeutic targets, repositioned phosphate binders do not appear to effectively decrease circulating levels of these toxic compounds. Specific uremic toxins sorbants such as AST-120 need to demonstrate putative effectiveness on clinical outcomes and mortality in further trials.



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1. Introduction

Chronic kidney disease (CKD) is a highly prevalent condition that generates a substantial disease burden worldwide. Over the last few decades, the burden of CKD has not declined to the same extent as it has for many other important noncommunicable diseases [1]. In 2017, an estimated 700 million individuals had CKD (regardless of the stage), more than those affected by diabetes, asthma, chronic obstructive pulmonary disease, or depressive disorders [1]. It was recently shown that patients seen by nephrologists are more difficult to treat than patients seen by other subspecialists, due to the large number of comorbidities, polymedication, and the high mortality risk [2].

A number of conventional and nonconventional risk factors for comorbidities and mortality in CKD have been identified. The nonconventional risk factors include uremic toxins, various harmful compounds that accumulate as renal function declines and that are potential therapeutic targets [3,4]. The high comorbidity burden in CKD means that polymedication is common [5,6]. Specific treatment approaches are implemented for complications that can be readily identified and quantified, such as cardiovascular disease, hypertension, anemia, mineral bone disorder, volume overload, electrolyte disorders, and acid-base disorders. At present, there are few specific or nonspecific pharmacological strategies for decreasing uremic toxin levels. The objective of the present narrative review was to assess the efficacy of intestinal chelators (e.g., phosphate binders and the orally administered activated charcoal sorbent AST-120 (Kremezin®, Kureha Corporation, Tokyo, Japan)) for the adsorption of gut-derived uremic toxins. We will describe gut-derived uremic toxins, phosphate binders, and then the effects of phosphate binders and AST-120 on uremic toxin levels.

2. Gut-Derived Uremic Toxins

According to the European Society of Artificial Organs' European Uremic Toxins Work Group [7], uremic toxins are harmful compounds that accumulate in the body during periods of renal function decline. Uremic toxins can be classified according to their molecular weight, water solubility, and protein-binding status. Alternatively, uremic toxins can be classified by their origin: gut-derived or not gut-derived. The present review focused on gut-derived uremic toxins.

2.1. Trimethylamine-N-Oxide

Trimethylamine-N-oxide (TMAO) is a small, water-soluble, gut-derived uremic toxin produced by the oxidation of trimethylamine (TMA). TMAO is derived primarily from dietary choline and carnitine through the action of the gut microbiota, which metabolizes these constituents to TMA. Red meat, eggs, dairy products, and saltwater fish are rich in choline, lecithin, and L-carnitine, and thus constitute potential sources of TMAO [8]. Many bacterial species are involved in producing TMA, including *Clostridia*, *Proteus*, *Shigella*, and *Aerobacter*. Over the last decade, a growing body of preclinical and clinical evidence has identified TMAO as an important contributor to the pathogenesis of cardiovascular disease [9]. Indeed, elevated TMAO levels are associated with a greater incident risk of major cardiovascular adverse events [10–12].

2.2. Indoxyl Sulfate

Indoxyl sulfate (IS, a protein-binding uremic toxin) is generated after tryptophan present in food is metabolized to indole by intestinal bacteria (mainly *Escherichia coli*). The indole is absorbed by the intestine and then circulates in the blood to the liver. After hydroxylation and sulfation in the liver, indole becomes IS and re-enters the circulation. When the kidney is operating normally, IS in serum enters renal tubular cells via the organic anion transporters (OATs) 1 and 3, located in the proximal basolateral membrane, and is subsequently drained into the renal tubules via OAT4 in the renal tubular cells' apical membrane [3]. Various studies have suggested that IS is an agonist of the transcription factor aryl hydrocarbon receptor (AhR) and increases oxidative stress and inhibits nitric oxide production in endothelial cells, inhibits neovascularization, and enhances coagulation [3]. In cohort studies of predialysis and dialysis patients, high levels of IS were associated independently with cardiovascular events, renal function decline, and mortality [13,14].

2.3. Indole Acetic Acid

Indole-3 acetic acid (IAA) is a protein-bound uremic toxin generated by the metabolism of tryptophan and that belongs to the indole family of uremic solutes (like IS). Serum IAA levels are elevated in patients with CKD. IAA is an agonist of the transcription factor AhR but has been less extensively studied than IS as a uremic toxin. Nevertheless, IAA has been linked to cardiovascular disease via the induction of inflammation and the production of

pro-coagulant tissue factors [15]. As with IS, these effects are mainly linked to activation of the AhR, which appears to have a key role in the biological action of several indoles [3,16]. IAA has been linked to elevated mortality and the incidence of cardiovascular events in CKD patients [15].

2.4. P-Cresyl Sulfate

P-cresyl sulfate (pCS) is a protein-bound, gut-derived uremic toxin. P-cresol is produced by the bacterial metabolism of tyrosine and phenylalanine in the intestine. Tyrosine and phenylalanine are essential amino acids for human beings and are found in protein-rich foods like meat, dairy products, eggs, and nuts. Conjugation of p-cresol creates pCS and, to a lesser extent, p-cresylglucuronide. The pCS is normally excreted by the kidneys. Data from animal studies suggest that pCS is harmful in several respects. Firstly, pCS contributes to cardiovascular and renal damage. An elevated pCS plasma concentration alters vascular function and in vitro remodeling [4] and is toxic for renal tubular cells and kidney fibrosis [17]. Furthermore, free pCS was found to be a predictor of survival in both dialyzed and nondialyzed CKD patients [14,18,19].

2.5. Nonpharmacological Interventions, Dietary Changes, and Uremic Toxin Concentrations

Although this review focused on pharmacological interventions (intestinal chelators) that modulate the concentrations of uremic toxins and prevent the development of harmful effect in CKD patients, diet may also have a role in the therapeutic strategy. Indeed, plant nutrients and plant-based diets can have beneficial effects in patients with CKD [20]. In CKD, there is a direct relationship between the protein/dietary fiber ratio and the IS and pCS levels. Di Iorio et al. demonstrated that nutritional therapy such as Mediterranean diet and very low protein diet (which is a vegetarian diet) are effective in lowering IS and pCS serum levels and ameliorated the intestinal permeability in CKD patients [20]. In patients on maintenance hemodiafiltration, plasma levels of IS and pCS were lower in vegetarian individuals than in nonvegetarian individuals [21]. This finding was confirmed in a randomized controlled trial of synbiotic therapy in nondialysis patients [22]. Likewise, a study of people with normal renal function found that pCS and IS production rates were markedly lower in vegetarians than in individuals consuming an unrestricted diet [23]. Furthermore, vegetarian diets contain less lecithin, choline, and l-carnitine; this might result in less TMAO production [24].

3. Phosphate Binders

Several lines of evidence link high phosphate concentrations to adverse health effects in patients with CKD [25–27]. Intestinal phosphate binders have been authorized for lowering phosphate concentrations in dialysis patients and for sevelamer carbonate in nondialyzed patients [28]. The phosphate binders act by decreasing the absorption of ingested phosphate and converting it into an insoluble form that is excreted in the stools. This mechanism of action explains why the binders must be taken at the same time as phosphate-containing foods.

There are two categories of phosphate binder: (1) calcium-based binders (calcium carbonate, calcium acetate, and calcium acetate/magnesium carbonate) and (2) noncalcium-based binders (sevelamer, lanthanum, and, more recently, iron-based binders). Although the two categories are equally effective in lowering serum phosphate concentrations when well titrated [29], each has various drawbacks, e.g., a positive calcium balance and potentially harmful cardiovascular outcomes for calcium-based binders [30] and the higher cost of noncalcium-based binders. Furthermore, the pill burden and gastrointestinal side effects associated with phosphate binders are major causes of poor treatment compliance [31,32] (Table 1).

Table 1. Advantages and disadvantages of various phosphate binders.

	Calcium-Based Phosphate Binders	Magnesium/Calcium-Based Phosphate Binders	Calcium-Free Phosphate Binders			
	<i>Calcium carbonate</i> <i>Calcium acetate</i>	<i>Magnesium carbonate</i>	<i>Sevelamer</i>	<i>Lanthanum carbonate</i>	<i>Iron-Based Phosphate Binders</i>	
					<i>Ferric citrate</i>	<i>Sucroferric oxyhydroxide</i>
Advantages	<ul style="list-style-type: none"> - Correction of hypocalcemia - Low cost 	<ul style="list-style-type: none"> - Low cost - Potential benefit on vascular calcification [33] 	<ul style="list-style-type: none"> - No systemic absorption - No hypercalcemia [29] - Reduction in vascular calcification vs. calcium-based phosphate binders [34] - Reduction in LDL-cholesterol [35] 	<ul style="list-style-type: none"> - Low pill burden - Reduction in vascular calcification vs. calcium-based phosphate binders [36,37] 	<ul style="list-style-type: none"> - Beneficial effect on iron-deficiency anemia [38,39] 	<ul style="list-style-type: none"> - Low pill burden [40–42] - Low systemic absorption [43]
Disadvantages	<ul style="list-style-type: none"> - Moderate pill burden - Systemic absorption - Positive calcium balance (hypercalcemia) - Progression of extraskelatal calcification (vascular calcification [44]) 	<ul style="list-style-type: none"> - High pill burden 	<ul style="list-style-type: none"> - High pill burden - High cost - Reduction in intestinal absorption of certain drugs and vitamins [45] - Gastrointestinal adverse events (constipation) - Gastropathy (sevelamer crystals and mucosa injury) [46–48] 	<ul style="list-style-type: none"> - High cost - Difficult to chew - Gastrointestinal adverse events - Lanthanum gastropathy (gastrointestinal deposits) [49–52] 	<ul style="list-style-type: none"> - High pill burden - High cost - Potential iron overload - Gastrointestinal adverse events (diarrhea) 	<ul style="list-style-type: none"> - High cost - Gastrointestinal adverse events

3.1. Calcium-Based Phosphate Binders

For many years, calcium-based phosphate binders constituted the cornerstone of hyperphosphatemia treatment. Although these binders are cheap and effective in lowering serum phosphate in dialysis patients and have a satisfactory safety profile, they have been linked to hypercalcemia (more frequently with calcium acetate than with calcium carbonate [53]) and the progression of vascular calcification [54]. Patients with high calcemia values should not, therefore, be treated with calcium-based phosphate binders [55].

3.2. Magnesium-Based Phosphate Binders.

A magnesium–calcium combination is a well-tolerated alternative to calcium acetate alone; it reduces the calcium load in hemodialysis patients and is not inferior to sevelamer hydrochloride [56]. Furthermore, magnesium may be of additional value in CKD patients because it appears to inhibit calcification [57–59].

3.3. Calcium-Free/Noncalcium-Based Phosphate Binders

Sevelamer was the first metal-free, calcium-free phosphate binder (first as the hydrochloride and then sevelamer carbonate) to become available for the treatment of hyperphosphatemia in patients on dialysis. Several randomized trials have compared sevelamer with calcium-based phosphate binders, particularly with regard to the progression of cardiovascular disease at different CKD stages. The majority of these trials showed that treatment with sevelamer was associated with less vascular calcification and that calcium-based binders were associated with more vascular calcification [34,44,60,61]. Sevelamer appears to induce constipation more frequently than calcium-based lanthanum and iron-based phosphate binders do [29].

Lanthanum carbonate is a powerful metal-based but calcium-free phosphate chelator [62]. Concerns have been raised about its safety, especially with regard to possible accumulation in the liver (like aluminum). However, this has not been confirmed in studies in humans [63]. Like most noncalcium binders, lanthanum carbonate is associated with less vascular calcification when compared with calcium-based binders [36]. Although the pill burden is lower than for sevelamer, gastrointestinal effects (such as nausea, vomiting, and abdominal cramps) are common [29,64].

3.4. Iron-Based Phosphate Binders

Two iron-based phosphate binders are currently on the market: ferric citrate and sucroferric oxyhydroxide. Both are effective phosphate binders and are not inferior to other compounds in this respect [65,66].

Ferric citrate binds phosphate in exchange for citrate to form ferric phosphate, which is insoluble and is excreted in the feces. In one trial, ferric citrate was non-inferior to the comparator arm (sevelamer or calcium-containing) in controlling serum phosphate levels [39]. Furthermore, ferric citrate was associated with an elevation in serum ferritin, which reduced the need for intravenous iron and erythropoietin-stimulating agents and increased hemoglobin levels [39,67,68].

Sucroferric oxyhydroxide is a novel, polynuclear, chewable, iron-based phosphate binder that can produce an insoluble complex in the gastrointestinal tract. In a Phase III randomized trial, sucroferric oxyhydroxide was as effective as sevelamer in lowering serum phosphate in dialysis patients, and the pill burden was 75% lower [42]. Furthermore, use of sucroferric oxyhydroxide led to an elevation in serum iron levels, albeit to a much lesser extent than for ferric citrate [43].

Both of these iron-based binders are associated with mild to moderate diarrhea and stool discoloration [29,64].

The 2017 Kidney Disease: Improving Global Outcomes (KDIGO) guidelines recommend “lowering elevated phosphate levels toward the normal range” but do not specify which types of phosphate binder should be used. However, the KDIGO guidelines also suggest avoiding hypercalcemia [55].

Observational studies have shown that the risk of all-cause mortality and cardiovascular disease among dialysis patients is lower among those treated with phosphate binders [69]. A meta-analysis showed that noncalcium chelators (sevelamer and lanthanum) were associated with a lower risk of mortality than calcium chelators (carbonate and calcium acetate) [70]. However, placebo-controlled, randomized trials have not found a beneficial impact of phosphate binders on mortality [29]. However, these trials included small numbers of patients and did not have long-term follow-up. Sevelamer could lead to a reduction in vascular calcification vs. calcium-based phosphate binders [71]. The advantages and disadvantages of phosphate chelators are summarized in Table 1.

4. Phosphate Binders and Gut-Derived Uremic Toxins

4.1. Phosphate Binders and the Gut Microflora

Due to the diversity and adaptability of the gut microbiome, changes in diet can quickly and dramatically influence the species of bacteria present. Thus, altering the concentrations and effects of bacterial metabolites might influence the host's metabolism [72,73]. CKD patients (especially those on dialysis) have to follow a low-phosphorus, low-potassium diet due to the association between high serum levels of these metabolites and the mortality risk [74]. Since phosphorous is a major nutrient for the survival and reproduction of bacteria, changes in its intestinal concentration may impact the gut microbiota's activity and composition [73]. Therefore, these dietary restrictions alter the composition of the gut microbiome. Hence, the gut microbiome is influenced not only by CKD but also by phosphate binders [75,76].

4.2. Phosphate Binders and Gut-Derived Uremic Toxins

4.2.1. Experimental Data

The phosphate binders' impact on the gut microflora can potentially modify the generation of gut-derived uremic toxins. Phosphate binders might prevent the absorption of both phosphate and gut-derived uremic toxins. There are few published in vitro studies of the impact of phosphate binders on precursors or uremic toxins present in the intestine. A recent report (published as a poster only) suggested that sevelamer hydrochloride induced a decrease in levels of indole and p-cresol (the gut precursors of pCS and IS) [77]. Another recent report indicated that sevelamer carbonate was able to adsorb IAA but not indole and p-cresol. The results can, perhaps, be explained by the compounds' respective chemical structures: IAA has a carboxylic acid group (like bile acids, which are also chelated by sevelamer) but none of the precursors have functional groups that would enable chelation by sevelamer [78] (Table 2).

In a study of uremic apolipoprotein-E-deficient mice, the serum concentrations of IS and IAA did not fall significantly, in contrast to the serum phosphorus concentration, after an eight-week sevelamer-containing diet [79].

Clinical studies of the impact of phosphate binders on gut-derived uremic toxin concentrations are summarized in Table 2.

Table 2. Summary of preclinical and clinical studies of the impact of phosphate binders on concentrations of gut-derived uremic toxins

In Vitro Studies					
First Author Year	Type of Binder	Type of Gut-Derived Uremic Toxin	Results		
De Smet R. 2016 (abstract) [77]	Sevelamer hydrochloride	IAA, p-cresol, indole	Sevelamer hydrochloride was able to adsorb indole, IAA and p-cresol		
Bennis Y. 2019 [78]	Sevelamer carbonate	IAA, p-cresol, indole	Sevelamer carbonate was able to adsorb IAA but not indole or p-cresol		
Animal Studies					
First Author Year	Type of Models	Type of Binder	Type of Gut-Derived Uremic Toxin	Results	
Phan O. 2005 [79]	apolipoprotein E-deficient mice	8 weeks of a sevelamer-containing diet	ISI AA	Levels of IS and IAA had not decreased significantly after 8 weeks of a sevelamer-containing diet	
Clinical Studies					
First Author Year	Type of Study	Patient Type and Numbers	Type of Phosphate Binder	Type of Gut-Derived Uremic Toxin	Results
Observational Studies					
Guida B. 2013 [80]	Cross-sectional observational study	57 patients on peritoneal dialysis	Sevelamer (<i>n</i> = 29) Lanthanum (<i>n</i> = 16) No binders (<i>n</i> = 12)	P-cresol	Patients on sevelamer had p-cresol levels significantly lower than those receiving lanthanum or no drug
Lin C. 2017 [81]	Observational Noncomparative	5 hemodialysis patients	Sevelamer for 3 months	IS pCS	Significant Reduction in pCS but not IS
Iguchi A. 2020 [76]	Observational cohort	18 hemodialysis patients	Sucroferric oxyhydroxide for 3 months	IS pCS	Increase levels of IS and pCS
Dai L. 2020 [82]	Cross-sectional observational cohort	423 ESKD patients	Calcium-containing phosphate binders Sevelamer	IS pCS TMAO	Increased levels of IS and TMAO, no change in pCS in sevelamer users vs. sevelamer non-users
Interventional studies					
Brandenburg V.M. 2010 [83]	Clinical trial (controlled crossover study)	57 hemodialysis patients	3-phase trial (A-B-A design; 8 weeks per phase). Sevelamer was only administered in the middle phase of the study.	ISI AA p-cresol	No impact on IS and IAA levels and a significant rise in p-cresol during the sevelamer period
Riccio E. 2018 [84]	Clinical trial (single-blind, placebo-controlled randomized trial)	69 CKD patients (grade 3–5, not on dialysis)	Sevelamer vs. placebo for 3 months	p-cresol	Significant Reduction in p-cresol after 3 months of treatment by sevelamer but not placebo
Bennis Y. 2019 [78]	Clinical trial (multicenter, double-blind, placebo-controlled, randomized)	78 CKD patients (grade 3–4)	Sevelamer vs. placebo for 3 months	IS pCS IAA	No significant changes in IS, pCS and IAA levels in the sevelamer arm
Lenglet A. 2019 [85]	Clinical trial (multicenter, open-label, randomized controlled trial)	100 hemodialysis patients	Sevelamer vs. nicotinamide	IS pCS TMAO	No impact on IS, pCS or TMAO levels in either arm

ESKD: end-stage kidney disease.

4.2.2. Observational Studies

In an observational, noncomparative study of five hemodialysis patients, Lin et al. evaluated serum levels of IS and pCS after 12 weeks of sevelamer hydrochloride treatment. A significant decrease was seen for levels of pCS, when a nonsignificant trend was observed for IS levels [81]. In a cross-sectional, noncomparative, observational study, Guida et al. assessed the serum pCS levels in a total of 57 patients on peritoneal dialysis. Of the 45 patients with hyperphosphatemia, 29 received sevelamer hydrochloride and 16 were treated with another phosphate binder (lanthanum). Patients with normal phosphatemia were not treated ($n = 12$). Plasma p-cresol concentrations were significantly lower in patients on sevelamer than in patients receiving lanthanum or those not treated. [80]. To note, the circulating toxin is pCS and not p-cresol; the latter is essentially generated *ex vivo* during the processing of phenol-containing blood samples [86].

Recently, a study in 18 hemodialysis patients found that 12 weeks of treatment with sucroferic oxyhydroxide was associated with higher IS and pCS concentrations [76]. On the same lines, the results of Dai et al.'s observational study of 423 patients with end-stage renal disease showed that serum levels of IS and TMAO (but not pCS) were significantly higher in sevelamer users than in non-users [82]. However, the observational design of these two recent studies constitutes an important limitation on the interpretation of the results; indication bias might account for the elevated levels of gut-derived uremic toxins observed in the sevelamer groups. The data from the interventional studies described below appear to be more robust.

4.2.3. Interventional Studies

An interventional, controlled, crossover study included 57 hemodialysis patients, during which they received an eight-week treatment of sevelamer hydrochloride. No changes in serum levels were observed for IS and IAA; unexpectedly, the serum concentration of pCS increased significantly [83].

In nondialyzed patients, in a single-blind, placebo-controlled, randomized clinical trial, plasma levels of p-cresol decreased significantly in the sevelamer group only [84]. In a recent, multicenter, comparative, randomized, clinical trial in dialysis patients, neither sevelamer nor nicotinamide induced a change in gut-derived uremic toxin levels [85]. Likewise, in a multicenter, double-blind, placebo-controlled, randomized, clinical trial in grades 3 and 4 CKD patients, a three-month course of sevelamer carbonate was not associated with significant changes in the serum concentrations of pCS, IS, or IAA [78].

Hence, in view of the literature data from interventional studies, sevelamer has no real effect on gut-derived uremic toxins.

5. Sorbents and Uremic Toxins

AST-120 is an oral-activated, charcoal adsorbent consisting of water-insoluble porous carbon particles (diameter: 0.2–0.4 mm). The adsorbent has been commercialized since 1991 in Japan, since 2004 in Korea, since 2007 in Taiwan, and since 2010 in the Philippines. It is indicated for the treatment of symptoms of uremia and to prolong the time to dialysis initiation in patients with progressive CKD [87].

In mechanistic terms, AST-120 might act by adsorbing uremic toxins and their precursors in the gastrointestinal tract, allowing them to be excreted in the feces before they can be absorbed into the bloodstream. AST-120 can adsorb the IS and pCS precursors generated by amino acid metabolism in the intestine. Most of the data on potential decreases in gut-derived uremic toxin levels by AST-120 concern IS. Administration of IS increases oxidative stress in the rat kidney, whereas co-administration of AST-120 reduced oxidative stress in CKD rats [88]. In an experimental model of apoE^{-/-} mice with CKD, IS appears to be an important contributor to the vascular dysfunction, and AST-120 treatment ameliorates this dysfunction, possibly via a decrease in serum IS concentration [89]. In a dose-ranging study in 164 CKD patients in the USA, AST-120 decreased serum IS levels in a dose-dependent fashion [90]. In the Kremezin Study against Renal Disease Progression

performed in 579 grades 3 and 4 CKD patients in Korea, the AST-120-induced decrease in the serum IS concentration was inversely correlated with the occurrence of the composite primary outcome (progression of renal disease) [91].

From 2007 to 2012, the multinational, randomized, double-blind, placebo-controlled Evaluating Prevention of Progression in CKD 1 and 2 trials were conducted in North America, Latin America, and Europe. The objective was to evaluate the effects of add-on AST-120 therapy (vs. placebo) on CKD progression [92]. The primary endpoint was a composite of serum creatinine doubling and number of dialysis initiation and kidney transplantation. The time to the primary endpoint was similar in the AST-120 and placebo groups in each trial individually and in a pooled analysis of the two trials. However, the serum levels of IS (a guide to the patients' treatment compliance) were not evaluated. Indeed, in a post hoc analysis of the patients in the USA, there was a significant intergroup difference in the time to achieve the primary endpoint (hazard ratio (95% confidence interval) = 0.74 (0.56–0.97) in the per-protocol population, with compliance rates of $\geq 67\%$) [93].

The majority of clinical trials on AST-120 have been performed in Asia and essentially focused on kidney disease progression. As this compound seems to effectively reduce gut-derived uremic toxin levels and given the organ toxicity of these toxins, there is a need for new studies of AST-120's effects on other outcomes (e.g., patient-reported outcomes, such as symptoms potentially due to the accumulation of uremic toxins).

6. Conclusions

Even though gut-derived uremic toxins appear to be valuable therapeutic targets, repositioned phosphate binders do not appear to effectively decrease circulating levels of these toxic compounds. AST-120, a specific uremic toxin sorbent only commercialized in Asia, is effective in reducing uremic toxin levels, mostly IS. However, the sorbent's putative effectiveness on clinical outcomes and mortality must now be assessed in further trials.

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References

1. Levin, A.; Tonelli, M.; Bonventre, J.; Coresh, J.; Donner, J.-A.; Fogo, A.B.; Fox, C.S.; Gansevoort, R.T.; Heerspink, H.J.L.; Jardine, M.; et al. Global Kidney Health 2017 and beyond: A Roadmap for Closing Gaps in Care, Research, and Policy. *Lancet* **2017**, *390*, 1888–1917. [[CrossRef](#)]
2. Tonelli, M.; Wiebe, N.; Manns, B.J.; Klarenbach, S.W.; James, M.T.; Ravani, P.; Pannu, N.; Himmelfarb, J.; Hemmelgarn, B.R. Comparison of the Complexity of Patients Seen by Different Medical Subspecialists in a Universal Health Care System. *JAMA Netw. Open* **2018**, *1*, e184852. [[CrossRef](#)] [[PubMed](#)]
3. Vanholder, R.; Pletinck, A.; Schepers, E.; Glorieux, G. Biochemical and Clinical Impact of Organic Uremic Retention Solutes: A Comprehensive Update. *Toxins* **2018**, *10*, 33. [[CrossRef](#)]
4. Six, I.; Flissi, N.; Lenglet, G.; Louvet, L.; Kamel, S.; Gallet, M.; Massy, Z.A.; Liabeuf, S. Uremic Toxins and Vascular Dysfunction. *Toxins* **2020**, *12*, 404. [[CrossRef](#)] [[PubMed](#)]
5. Hayward, S.; Hole, B.; Denholm, R.; Duncan, P.; Morris, J.E.; Fraser, S.D.S.; Payne, R.A.; Roderick, P.; Chesnaye, N.C.; Wanner, C.; et al. International Prescribing Patterns and Polypharmacy in Older People with Advanced Chronic Kidney Disease: Results from the European Quality Study. *Nephrol. Dial. Transpl.* **2020**. [[CrossRef](#)]
6. Liabeuf, S.; Laville, M. Drug Prescription in Patients with Chronic Kidney Disease: A True Challenge. *Nephrol. Dial. Transpl.* **2020**. [[CrossRef](#)]

7. Vanholder, R.; De Smet, R.; Glorieux, G.; Argilés, A.; Baurmeister, U.; Brunet, P.; Clark, W.; Cohen, G.; De Deyn, P.P.; Deppisch, R.; et al. Review on Uremic Toxins: Classification, Concentration, and Interindividual Variability. *Kidney Int.* **2003**, *63*, 1934–1943. [[CrossRef](#)]
8. Cho, C.E.; Caudill, M.A. Trimethylamine-N-Oxide: Friend, Foe, or Simply Caught in the Cross-Fire? *Trends Endocrinol. Metab.* **2017**, *28*, 121–130. [[CrossRef](#)]
9. Subramaniam, S.; Fletcher, C. Trimethylamine N-Oxide: Breathe New Life. *Br. J. Pharm.* **2018**, *175*, 1344–1353. [[CrossRef](#)]
10. Tang, W.H.W.; Wang, Z.; Levison, B.S.; Koeth, R.A.; Britt, E.B.; Fu, X.; Wu, Y.; Hazen, S.L. Intestinal Microbial Metabolism of Phosphatidylcholine and Cardiovascular Risk. *N. Engl. J. Med.* **2013**, *368*, 1575–1584. [[CrossRef](#)]
11. Tomlinson, J.A.P.; Wheeler, D.C. The Role of Trimethylamine N-Oxide as a Mediator of Cardiovascular Complications in Chronic Kidney Disease. *Kidney Int.* **2017**, *92*, 809–815. [[CrossRef](#)] [[PubMed](#)]
12. Kim, R.B.; Morse, B.L.; Djurdjev, O.; Tang, M.; Muirhead, N.; Barrett, B.; Holmes, D.T.; Madore, F.; Clase, C.M.; Rigatto, C.; et al. Advanced Chronic Kidney Disease Populations Have Elevated Trimethylamine N-Oxide Levels Associated with Increased Cardiovascular Events. *Kidney Int.* **2016**, *89*, 1144–1152. [[CrossRef](#)] [[PubMed](#)]
13. Barreto, F.C.; Barreto, D.V.; Liabeuf, S.; Meert, N.; Glorieux, G.; Temmar, M.; Choukroun, G.; Vanholder, R.; Massy, Z.A.; European Uremic Toxin Work Group (EUTox). Serum Indoxyl Sulfate Is Associated with Vascular Disease and Mortality in Chronic Kidney Disease Patients. *Clin. J. Am. Soc. Nephrol.* **2009**, *4*, 1551–1558. [[CrossRef](#)] [[PubMed](#)]
14. Liabeuf, S.; Drüeke, T.B.; Massy, Z.A. Protein-Bound Uremic Toxins: New Insight from Clinical Studies. *Toxins* **2011**, *3*, 911–919. [[CrossRef](#)] [[PubMed](#)]
15. Dou, L.; Sallée, M.; Cerini, C.; Poitevin, S.; Gondouin, B.; Jourde-Chiche, N.; Fallague, K.; Brunet, P.; Calaf, R.; Dussol, B.; et al. The Cardiovascular Effect of the Uremic Solute Indole-3 Acetic Acid. *J. Am. Soc. Nephrol.* **2015**, *26*, 876–887. [[CrossRef](#)]
16. Sallée, M.; Dou, L.; Cerini, C.; Poitevin, S.; Brunet, P.; Burtsey, S. The Aryl Hydrocarbon Receptor-Activating Effect of Uremic Toxins from Tryptophan Metabolism: A New Concept to Understand Cardiovascular Complications of Chronic Kidney Disease. *Toxins* **2014**, *6*, 934–949. [[CrossRef](#)]
17. Gryp, T.; Vanholder, R.; Vanechoutte, M.; Glorieux, G. P-Cresyl Sulfate. *Toxins* **2017**, *9*, 52. [[CrossRef](#)]
18. Liabeuf, S.; Barreto, D.V.; Barreto, F.C.; Meert, N.; Glorieux, G.; Schepers, E.; Temmar, M.; Choukroun, G.; Vanholder, R.; Massy, Z.A.; et al. Free P-Cresylsulphate Is a Predictor of Mortality in Patients at Different Stages of Chronic Kidney Disease. *Nephrol. Dial. Transpl.* **2010**, *25*, 1183–1191. [[CrossRef](#)]
19. Meijers, B.K.I.; De Preter, V.; Verbeke, K.; Vanrenterghem, Y.; Evenepoel, P. P-Cresyl Sulfate Serum Concentrations in Haemodialysis Patients Are Reduced by the Prebiotic Oligofructose-Enriched Inulin. *Nephrol. Dial. Transpl.* **2010**, *25*, 219–224. [[CrossRef](#)]
20. Di Iorio, B.R.; Rocchetti, M.T.; De Angelis, M.; Cosola, C.; Marzocco, S.; Di Micco, L.; di Bari, I.; Accetturo, M.; Vacca, M.; Gobetti, M.; et al. Nutritional Therapy Modulates Intestinal Microbiota and Reduces Serum Levels of Total and Free Indoxyl Sulfate and P-Cresyl Sulfate in Chronic Kidney Disease (Medika Study). *J. Clin. Med.* **2019**, *8*, 1424. [[CrossRef](#)]
21. Kandouz, S.; Mohamed, A.S.; Zheng, Y.; Sandeman, S.; Davenport, A. Reduced Protein Bound Uraemic Toxins in Vegetarian Kidney Failure Patients Treated by Haemodiafiltration. *Hemodial. Int.* **2016**, *20*, 610–617. [[CrossRef](#)] [[PubMed](#)]
22. Rossi, M.; Johnson, D.W.; Xu, H.; Carrero, J.J.; Pascoe, E.; French, C.; Campbell, K.L. Dietary Protein-Fiber Ratio Associates with Circulating Levels of Indoxyl Sulfate and p-Cresyl Sulfate in Chronic Kidney Disease Patients. *Nutr. Metab. Cardiovasc. Dis.* **2015**, *25*, 860–865. [[CrossRef](#)] [[PubMed](#)]
23. Patel, K.P.; Luo, F.J.-G.; Plummer, N.S.; Hostetter, T.H.; Meyer, T.W. The Production of P-Cresol Sulfate and Indoxyl Sulfate in Vegetarians Versus Omnivores. *Clin. J. Am. Soc. Nephrol.* **2012**, *7*, 982–988. [[CrossRef](#)] [[PubMed](#)]
24. Tomova, A.; Bukovsky, I.; Rembert, E.; Yonas, W.; Alwarith, J.; Barnard, N.D.; Kahleova, H. The Effects of Vegetarian and Vegan Diets on Gut Microbiota. *Front. Nutr.* **2019**, *6*. [[CrossRef](#)] [[PubMed](#)]
25. Block, G.A.; Hulbert-Shearon, T.E.; Levin, N.W.; Port, F.K. Association of Serum Phosphorus and Calcium x Phosphate Product with Mortality Risk in Chronic Hemodialysis Patients: A National Study. *Am. J. Kidney Dis.* **1998**, *31*, 607–617. [[CrossRef](#)]
26. Chartsrisak, K.; Vipattawat, K.; Assanatham, M.; Nongnuch, A.; Ingsathit, A.; Domrongkitchaiporn, S.; Sumethkul, V.; Distha-Banchong, S. Mineral Metabolism and Outcomes in Chronic Kidney Disease Stage 2–4 Patients. *BMC Nephrol.* **2013**, *14*, 14. [[CrossRef](#)]
27. Eddington, H.; Hoefield, R.; Sinha, S.; Chrysochou, C.; Lane, B.; Foley, R.N.; Hegarty, J.; New, J.; O'Donoghue, D.J.; Middleton, R.J.; et al. Serum Phosphate and Mortality in Patients with Chronic Kidney Disease. *Clin. J. Am. Soc. Nephrol.* **2010**, *5*, 2251–2257. [[CrossRef](#)]
28. Ix, J.H. Binder Blunder in CKD. *J. Am. Soc. Nephrol.* **2020**, *31*, 2499–2501. [[CrossRef](#)]
29. Palmer, S.C.; Gardner, S.; Tonelli, M.; Mavridis, D.; Johnson, D.W.; Craig, J.C.; French, R.; Ruospo, M.; Strippoli, G.F.M. Phosphate-Binding Agents in Adults with CKD: A Network Meta-Analysis of Randomized Trials. *Am. J. Kidney Dis.* **2016**, *68*, 691–702. [[CrossRef](#)]
30. Raggi, P.; Bommer, J.; Chertow, G.M. Valvular Calcification in Hemodialysis Patients Randomized to Calcium-Based Phosphorus Binders or Sevelamer. *J. Heart Valve Dis.* **2004**, *13*, 134–141.
31. Chiu, Y.-W.; Teitelbaum, I.; Misra, M.; de Leon, E.M.; Adzize, T.; Mehrotra, R. Pill Burden, Adherence, Hyperphosphatemia, and Quality of Life in Maintenance Dialysis Patients. *Clin. J. Am. Soc. Nephrol.* **2009**, *4*, 1089–1096. [[CrossRef](#)] [[PubMed](#)]

32. Fissell, R.B.; Karaboyas, A.; Bieber, B.A.; Sen, A.; Li, Y.; Lopes, A.A.; Akiba, T.; Bommer, J.; Ethier, J.; Jadoul, M.; et al. Phosphate Binder Pill Burden, Patient-Reported Non-Adherence, and Mineral Bone Disorder Markers: Findings from the DOPPS. *Hemodial. Int.* **2016**, *20*, 38–49. [[CrossRef](#)] [[PubMed](#)]
33. Matias, P.J.; Jorge, C.; Azevedo, A.; Laranjinha, I.; Navarro, D.; Mendes, M.; Amaral, T.; Ferreira, C.; Aires, I.; Gil, C.; et al. Calcium Acetate/Magnesium Carbonate and Cardiovascular Risk Factors in Chronic Hemodialysis Patients. *Nephron* **2016**, *132*, 317–326. [[CrossRef](#)] [[PubMed](#)]
34. Chertow, G.M.; Burke, S.K.; Raggi, P. Treat to Goal Working Group Sevelamer Attenuates the Progression of Coronary and Aortic Calcification in Hemodialysis Patients. *Kidney Int.* **2002**, *62*, 245–252. [[CrossRef](#)]
35. Sadek, T.; Mazouz, H.; Bahloul, H.; Oprisiu, R.; El Esper, N.; El Esper, I.; Boitte, F.; Brazier, M.; Moriniere, P.; Fournier, A. Sevelamer Hydrochloride with or without Alphacalcidol or Higher Dialysate Calcium vs Calcium Carbonate in Dialysis Patients: An Open-Label, Randomized Study. *Nephrol. Dial. Transplant.* **2003**, *18*, 582–588. [[CrossRef](#)]
36. Toussaint, N.D.; Lau, K.K.; Polkinghorne, K.R.; Kerr, P.G. Attenuation of Aortic Calcification with Lanthanum Carbonate versus Calcium-Based Phosphate Binders in Haemodialysis: A Pilot Randomized Controlled Trial. *Nephrology* **2011**, *16*, 290–298. [[CrossRef](#)]
37. Fujii, H.; Kono, K.; Nakai, K.; Goto, S.; Nishii, T.; Kono, A.; Nishi, S. Effects of Lanthanum Carbonate on Coronary Artery Calcification and Cardiac Abnormalities After Initiating Hemodialysis. *Calcif. Tissue Int.* **2018**, *102*, 310–320. [[CrossRef](#)]
38. Yokoyama, K.; Fukagawa, M.; Akiba, T.; Nakayama, M.; Ito, K.; Hanaki, K.; Wolf, M.; Hirakata, H. Randomised Clinical Trial of Ferric Citrate Hydrate on Anaemia Management in Haemodialysis Patients with Hyperphosphataemia: ASTRIO Study. *Sci. Rep.* **2019**, *9*, 8877. [[CrossRef](#)]
39. Lewis, J.B.; Sika, M.; Koury, M.J.; Chuang, P.; Schulman, G.; Smith, M.T.; Whittier, F.C.; Linfert, D.R.; Galphin, C.M.; Athreya, B.P.; et al. Ferric Citrate Controls Phosphorus and Delivers Iron in Patients on Dialysis. *J. Am. Soc. Nephrol.* **2015**, *26*, 493–503. [[CrossRef](#)]
40. Kendrick, J.; Parameswaran, V.; Ficociello, L.H.; Ofsthun, N.J.; Davis, S.; Mullon, C.; Kossmann, R.J.; Kalantar-Zadeh, K. One-Year Historical Cohort Study of the Phosphate Binder Sucroferic Oxyhydroxide in Patients on Maintenance Hemodialysis. *J. Ren. Nutr.* **2019**, *29*, 428–437. [[CrossRef](#)]
41. Coyne, D.W.; Ficociello, L.H.; Parameswaran, V.; Anderson, L.; Vemula, S.; Ofsthun, N.J.; Mullon, C.; Maddux, F.W.; Kossmann, R.J.; Sprague, S.M. Real-World Effectiveness of Sucroferic Oxyhydroxide in Patients on Chronic Hemodialysis: A Retrospective Analysis of Pharmacy Data. *Clin. Nephrol.* **2017**, *88*, 59–67. [[CrossRef](#)] [[PubMed](#)]
42. Floege, J.; Covic, A.C.; Ketteler, M.; Rastogi, A.; Chong, E.M.F.; Gaillard, S.; Lisk, L.J.; Sprague, S.M. PA21 Study Group A Phase III Study of the Efficacy and Safety of a Novel Iron-Based Phosphate Binder in Dialysis Patients. *Kidney Int.* **2014**, *86*, 638–647. [[CrossRef](#)] [[PubMed](#)]
43. Covic, A.C.; Floege, J.; Ketteler, M.; Sprague, S.M.; Lisk, L.; Rakov, V.; Rastogi, A. Iron-Related Parameters in Dialysis Patients Treated with Sucroferic Oxyhydroxide. *Nephrol. Dial. Transplant.* **2017**, *32*, 1330–1338. [[CrossRef](#)] [[PubMed](#)]
44. Block, G.A.; Spiegel, D.M.; Ehrlich, J.; Mehta, R.; Lindbergh, J.; Dreisbach, A.; Raggi, P. Effects of Sevelamer and Calcium on Coronary Artery Calcification in Patients New to Hemodialysis. *Kidney Int.* **2005**, *68*, 1815–1824. [[CrossRef](#)]
45. Sprague, S.M.; Covic, A.C.; Floege, J.; Ketteler, M.; Botha, J.; Chong, E.M.; Rastogi, A. Pharmacodynamic Effects of Sucroferic Oxyhydroxide and Sevelamer Carbonate on Vitamin D Receptor Agonist Bioactivity in Dialysis Patients. *Am. J. Nephrol.* **2016**, *44*, 104–112. [[CrossRef](#)]
46. Kim, J.; Olson, K.; Butani, L. Sevelamer Crystals in the Mucosa of the Gastrointestinal Tract in a Teenager with End-Stage Renal Disease. *Pediatr. Nephrol.* **2016**, *31*, 339–341. [[CrossRef](#)]
47. Nambiar, S.; Pillai, U.K.; Devasahayam, J.; Oliver, T.; Karippot, A. Colonic Mucosal Ulceration and Gastrointestinal Bleeding Associated with Sevelamer Crystal Deposition in a Patient with End Stage Renal Disease. *Case Rep. Nephrol.* **2018**, *2018*, 4708068. [[CrossRef](#)]
48. Tieu, C.; Moreira, R.K.; Song, L.M.W.K.; Majumder, S.; Papadakis, K.A.; Hogan, M.C. A Case Report of Sevelamer-Associated Recto-Sigmoid Ulcers. *BMC Gastroenterol.* **2016**, *16*, 20. [[CrossRef](#)]
49. Yabuki, K.; Haratake, J.; Tsuda, Y.; Shiba, E.; Harada, H.; Yorita, K.; Uchihashi, K.; Matsuyama, A.; Hirata, K.; Hisaoka, M. Lanthanum-Induced Mucosal Alterations in the Stomach (Lanthanum Gastropathy): A Comparative Study Using an Animal Model. *Biol. Trace Elem. Res.* **2018**, *185*, 36–47. [[CrossRef](#)]
50. Ban, S.; Suzuki, S.; Kubota, K.; Ohshima, S.; Satoh, H.; Imada, H.; Ueda, Y. Gastric Mucosal Status Susceptible to Lanthanum Deposition in Patients Treated with Dialysis and Lanthanum Carbonate. *Ann. Diagn. Pathol.* **2017**, *26*, 6–9. [[CrossRef](#)]
51. Hattori, K.; Maeda, T.; Nishida, S.; Imanishi, M.; Sakaguchi, M.; Amari, Y.; Moriya, T.; Hirose, Y. Correlation of Lanthanum Dosage with Lanthanum Deposition in the Gastroduodenal Mucosa of Dialysis Patients. *Pathol. Int.* **2017**, *67*, 447–452. [[CrossRef](#)] [[PubMed](#)]
52. Hoda, R.S.; Sanyal, S.; Abraham, J.L.; Everett, J.M.; Hundemer, G.L.; Yee, E.; Lauwers, G.Y.; Tolckoff-Rubin, N.; Misdraji, J. Lanthanum Deposition from Oral Lanthanum Carbonate in the Upper Gastrointestinal Tract. *Histopathology* **2017**, *70*, 1072–1078. [[CrossRef](#)] [[PubMed](#)]
53. Pflanz, S.; Henderson, I.S.; McElduff, N.; Jones, M.C. Calcium Acetate versus Calcium Carbonate as Phosphate-Binding Agents in Chronic Haemodialysis. *Nephrol. Dial. Transplant.* **1994**, *9*, 1121–1124. [[CrossRef](#)] [[PubMed](#)]

54. Goodman, W.G.; Goldin, J.; Kuizon, B.D.; Yoon, C.; Gales, B.; Sider, D.; Wang, Y.; Chung, J.; Emerick, A.; Greaser, L.; et al. Coronary-Artery Calcification in Young Adults with End-Stage Renal Disease Who Are Undergoing Dialysis. *N. Engl. J. Med.* **2000**, *342*, 1478–1483. [[CrossRef](#)] [[PubMed](#)]
55. Kidney Disease: Improving Global Outcomes (KDIGO) CKD-MBD Update Work Group KDIGO 2017 Clinical Practice Guideline Update for the Diagnosis, Evaluation, Prevention, and Treatment of Chronic Kidney Disease-Mineral and Bone Disorder (CKD-MBD). *Kidney Int. Suppl.* **2017**, *7*, 1–59. [[CrossRef](#)] [[PubMed](#)]
56. de Francisco, A.L.M.; Leidig, M.; Covic, A.C.; Ketteler, M.; Benedyk-Lorens, E.; Mircescu, G.M.; Scholz, C.; Ponce, P.; Passlick-Deetjen, J. Evaluation of Calcium Acetate/Magnesium Carbonate as a Phosphate Binder Compared with Sevelamer Hydrochloride in Haemodialysis Patients: A Controlled Randomized Study (CALMAG Study) Assessing Efficacy and Tolerability. *Nephrol. Dial. Transplant.* **2010**, *25*, 3707–3717. [[CrossRef](#)]
57. Tzanakis, I.P.; Stamataki, E.E.; Papadaki, A.N.; Giannakis, N.; Damianakis, N.E.; Oreopoulos, D.G. Magnesium Retards the Progress of the Arterial Calcifications in Hemodialysis Patients: A Pilot Study. *Int. Urol. Nephrol.* **2014**, *46*, 2199–2205. [[CrossRef](#)]
58. Massy, Z.A.; Drüeke, T.B. Magnesium and Outcomes in Patients with Chronic Kidney Disease: Focus on Vascular Calcification, Atherosclerosis and Survival. *Clin. Kidney J.* **2012**, *5*, i52–i61. [[CrossRef](#)]
59. Massy, Z.A.; Drüeke, T.B. Magnesium and Cardiovascular Complications of Chronic Kidney Disease. *Nat. Rev. Nephrol.* **2015**, *11*, 432–442. [[CrossRef](#)]
60. Patel, L.; Bernard, L.M.; Elder, G.J. Sevelamer Versus Calcium-Based Binders for Treatment of Hyperphosphatemia in CKD: A Meta-Analysis of Randomized Controlled Trials. *Clin. J. Am. Soc. Nephrol.* **2016**, *11*, 232–244. [[CrossRef](#)]
61. Qunibi, W.Y.; Hootkins, R.E.; McDowell, L.L.; Meyer, M.S.; Simon, M.; Garza, R.O.; Pelham, R.W.; Cleveland, M.V.B.; Muenz, L.R.; He, D.Y.; et al. Treatment of Hyperphosphatemia in Hemodialysis Patients: The Calcium Acetate Renagel Evaluation (CARE Study). *Kidney Int.* **2004**, *65*, 1914–1926. [[CrossRef](#)] [[PubMed](#)]
62. Sprague, S.M.; Ross, E.A.; Nath, S.D.; Zhang, P.; Pratt, R.D.; Krause, R. Lanthanum Carbonate vs. Sevelamer Hydrochloride for the Reduction of Serum Phosphorus in Hemodialysis Patients: A Crossover Study. *CN* **2009**, *72*, 252–258. [[CrossRef](#)] [[PubMed](#)]
63. Hutchison, A.J.; Barnett, M.E.; Krause, R.; Siami, G.A. Lanthanum Carbonate Study Group Lanthanum Carbonate Treatment, for up to 6 Years, Is Not Associated with Adverse Effects on the Liver in Patients with Chronic Kidney Disease Stage 5 Receiving Hemodialysis. *Clin. Nephrol.* **2009**, *71*, 286–295. [[PubMed](#)]
64. Ketteler, M.; Liangos, O.; Biggar, P.H. Treating Hyperphosphatemia – Current and Advancing Drugs. *Expert Opin. Pharmacother.* **2016**, *17*, 1873–1879. [[CrossRef](#)] [[PubMed](#)]
65. Negri, A.L.; Ureña Torres, P.A. Iron-Based Phosphate Binders: Do They Offer Advantages over Currently Available Phosphate Binders? *Clin. Kidney J.* **2015**, *8*, 161–167. [[CrossRef](#)]
66. Barreto, F.C.; Barreto, D.V.; Massy, Z.A.; Drüeke, T.B. Strategies for Phosphate Control in Patients With CKD. *Kidney Int. Rep.* **2019**, *4*, 1043–1056. [[CrossRef](#)]
67. Yokoyama, K.; Hirakata, H.; Akiba, T.; Fukagawa, M.; Nakayama, M.; Sawada, K.; Kumagai, Y.; Block, G.A. Ferric Citrate Hydrate for the Treatment of Hyperphosphatemia in Nondialysis-Dependent CKD. *Clin. J. Am. Soc. Nephrol.* **2014**, *9*, 543–552. [[CrossRef](#)]
68. Umanath, K.; Jalal, D.I.; Greco, B.A.; Umeukeje, E.M.; Reisin, E.; Manley, J.; Zeig, S.; Negroi, D.G.; Hiremath, A.N.; Blumenthal, S.S.; et al. Ferric Citrate Reduces Intravenous Iron and Erythropoiesis-Stimulating Agent Use in ESRD. *JASN* **2015**, *26*, 2578–2587. [[CrossRef](#)]
69. Cannata-Andía, J.B.; Fernández-Martín, J.L.; Locatelli, F.; London, G.; Gorriz, J.L.; Floege, J.; Ketteler, M.; Ferreira, A.; Covic, A.; Rutkowski, B.; et al. Use of Phosphate-Binding Agents Is Associated with a Lower Risk of Mortality. *Kidney Int.* **2013**, *84*, 998–1008. [[CrossRef](#)]
70. Jamal, S.A.; Vandermeer, B.; Raggi, P.; Mendelsohn, D.C.; Chatterley, T.; Dorgan, M.; Lok, C.E.; Fitchett, D.; Tsuyuki, R.T. Effect of Calcium-Based versus Non-Calcium-Based Phosphate Binders on Mortality in Patients with Chronic Kidney Disease: An Updated Systematic Review and Meta-Analysis. *Lancet* **2013**, *382*, 1268–1277. [[CrossRef](#)]
71. Toussaint, N.D.; Pedagogos, E.; Lioufas, N.M.; Elder, G.J.; Pascoe, E.M.; Badve, S.V.; Valks, A.; Block, G.A.; Boudville, N.; Cameron, J.D.; et al. A Randomized Trial on the Effect of Phosphate Reduction on Vascular End Points in CKD (IMPROVE-CKD). *J. Am. Soc. Nephrol.* **2020**, *31*, 2653–2666. [[CrossRef](#)] [[PubMed](#)]
72. Arora, T.; Bäckhed, F. The Gut Microbiota and Metabolic Disease: Current Understanding and Future Perspectives. *J. Intern. Med.* **2016**, *280*, 339–349. [[CrossRef](#)] [[PubMed](#)]
73. Rahbar Saadat, Y.; Niknafs, B.; Hosseiniyan Khatibi, S.M.; Ardalan, M.; Majdi, H.; Bahmanpoor, Z.; Abediazar, S.; Zununi Vahed, S. Gut Microbiota; an Overlooked Effect of Phosphate Binders. *Eur. J. Pharmacol.* **2020**, *868*, 172892. [[CrossRef](#)] [[PubMed](#)]
74. Kalantar-Zadeh, K.; Tortorici, A.R.; Chen, J.L.T.; Kamgar, M.; Lau, W.-L.; Moradi, H.; Rhee, C.M.; Streja, E.; Kovesdy, C.P. Dietary Restrictions in Dialysis Patients: Is There Anything Left to Eat? *Semin. Dial.* **2015**, *28*, 159–168. [[CrossRef](#)] [[PubMed](#)]
75. Lau, W.L.; Vaziri, N.D.; Nunes, A.C.F.; Comeau, A.M.; Langille, M.G.I.; England, W.; Khazaeli, M.; Suematsu, Y.; Phan, J.; Whiteson, K. The Phosphate Binder Ferric Citrate Alters the Gut Microbiome in Rats with Chronic Kidney Disease. *J. Pharmacol. Exp. Ther.* **2018**, *367*, 452–460. [[CrossRef](#)]
76. Iguchi, A.; Yamamoto, S.; Oda, A.; Tanaka, K.; Kazama, J.J.; Saeki, T.; Yamazaki, H.; Ishioka, K.; Suzutani, T.; Narita, I. Effect of Sucroferric Oxyhydroxide on Gastrointestinal Microbiome and Uremic Toxins in Patients with Chronic Kidney Disease Undergoing Hemodialysis. *Clin. Exp. Nephrol.* **2020**, *24*, 725–733. [[CrossRef](#)]

77. De Smet, R.; Thermote, F.; Lameyre, N.; Vanholder, R. Sevelamer Hydrochloride (Renagel) Adsorbs Theuremic Compounds Indoxyl Sulfate, Indole and p-Cresol. *J. Am. Soc. Nephrol.* **2004**, *15*, 505.
78. Bennis, Y.; Cluet, Y.; Titeca-Beauport, D.; El Esper, N.; Ureña, P.; Bodeau, S.; Combe, C.; Dussol, B.; Fouque, D.; Choukroun, G.; et al. The Effect of Sevelamer on Serum Levels of Gut-Derived Uremic Toxins: Results from In Vitro Experiments and A Multicenter, Double-Blind, Placebo-Controlled, Randomized Clinical Trial. *Toxins* **2019**, *11*, 279. [[CrossRef](#)]
79. Phan, O.; Ivanovski, O.; Nguyen-Khoa, T.; Mothu, N.; Angulo, J.; Westenfeld, R.; Ketteler, M.; Meert, N.; Maizel, J.; Nikolov, I.G.; et al. Sevelamer Prevents Uremia-Enhanced Atherosclerosis Progression in Apolipoprotein E-Deficient Mice. *Circulation* **2005**, *112*, 2875–2882. [[CrossRef](#)]
80. Guida, B.; Cataldi, M.; Riccio, E.; Grumetto, L.; Pota, A.; Borrelli, S.; Memoli, A.; Barbato, F.; Argentino, G.; Salerno, G.; et al. Plasma P-Cresol Lowering Effect of Sevelamer in Peritoneal Dialysis Patients: Evidence from a Cross-Sectional Observational Study. *PLoS ONE* **2013**, *8*, e73558. [[CrossRef](#)]
81. Lin, C.-J.; Pan, C.-F.; Chuang, C.-K.; Liu, H.-L.; Huang, S.-F.; Chen, H.-H.; Wu, C.-J. Effects of Sevelamer Hydrochloride on Uremic Toxins Serum Indoxyl Sulfate and P-Cresyl Sulfate in Hemodialysis Patients. *J. Clin. Med. Res.* **2017**, *9*, 765–770. [[CrossRef](#)] [[PubMed](#)]
82. Dai, L.; Meijers, B.K.; Bammens, B.; de Loor, H.; Schurgers, L.J.; Qureshi, A.R.; Stenvinkel, P.; Evenepoel, P. Sevelamer Use in End-Stage Kidney Disease (ESKD) Patients Associates with Poor Vitamin K Status and High Levels of Gut-Derived Uremic Toxins: A Drug-Bug Interaction? *Toxins* **2020**, *12*, 351. [[CrossRef](#)] [[PubMed](#)]
83. Brandenburg, V.M.; Schlieper, G.; Heussen, N.; Holzmann, S.; Busch, B.; Evenepoel, P.; Vanholder, R.; Meijers, B.; Meert, N.; Fassbender, W.J.; et al. Serological Cardiovascular and Mortality Risk Predictors in Dialysis Patients Receiving Sevelamer: A Prospective Study. *Nephrol. Dial. Transplant.* **2010**, *25*, 2672–2679. [[CrossRef](#)] [[PubMed](#)]
84. Riccio, E.; Sabbatini, M.; Bruzzese, D.; Grumetto, L.; Marchetiello, C.; Amicone, M.; Andreucci, M.; Guida, B.; Passaretti, D.; Russo, G.; et al. Plasma P-Cresol Lowering Effect of Sevelamer in Non-Dialysis CKD Patients: Evidence from a Randomized Controlled Trial. *Clin. Exp. Nephrol.* **2018**, *22*, 529–538. [[CrossRef](#)] [[PubMed](#)]
85. Lenglet, A.; Fabresse, N.; Taupin, M.; Gomila, C.; Liabeuf, S.; Kamel, S.; Alvarez, J.C.; Druke, T.B.; Massy, Z.A. Does the Administration of Sevelamer or Nicotinamide Modify Uremic Toxins or Endotoxemia in Chronic Hemodialysis Patients? *Drugs* **2019**, *79*, 855–862. [[CrossRef](#)] [[PubMed](#)]
86. Vanholder, R.; Bammens, B.; de Loor, H.; Glorieux, G.; Meijers, B.; Schepers, E.; Massy, Z.; Evenepoel, P. Warning: The Unfortunate End of p-Cresol as a Uraemic Toxin. *Nephrol. Dial. Transplant.* **2011**, *26*, 1464–1467. [[CrossRef](#)]
87. Asai, M.; Kumakura, S.; Kikuchi, M. Review of the Efficacy of AST-120 (KREMEZIN®) on Renal Function in Chronic Kidney Disease Patients. *Ren. Fail.* **2019**, *41*, 47–56. [[CrossRef](#)]
88. Bolati, D.; Shimizu, H.; Yisireyili, M.; Nishijima, F.; Niwa, T. Indoxyl Sulfate, a Uremic Toxin, Downregulates Renal Expression of Nrf2 through Activation of NF- κ B. *BMC Nephrol.* **2013**, *14*, 56. [[CrossRef](#)]
89. Six, I.; Gross, P.; Rémond, M.C.; Chillon, J.M.; Poirot, S.; Druke, T.B.; Massy, Z.A. Deleterious Vascular Effects of Indoxyl Sulfate and Reversal by Oral Adsorbent AST-120. *Atherosclerosis* **2015**, *243*, 248–256. [[CrossRef](#)]
90. Niwa, T.; Nomura, T.; Sugiyama, S.; Miyazaki, T.; Tsukushi, S.; Tsutsui, S. The Protein Metabolite Hypothesis, a Model for the Progression of Renal Failure: An Oral Adsorbent Lowers Indoxyl Sulfate Levels in Undialyzed Uremic Patients. *Kidney Int. Suppl.* **1997**, *62*, S23–S28.
91. Cha, R.-H.; Kang, S.W.; Park, C.W.; Cha, D.R.; Na, K.Y.; Kim, S.G.; Yoon, S.A.; Kim, S.; Han, S.Y.; Park, J.H.; et al. Sustained Uremic Toxin Control Improves Renal and Cardiovascular Outcomes in Patients with Advanced Renal Dysfunction: Post-Hoc Analysis of the Kremezin Study against Renal Disease Progression in Korea. *Kidney Res. Clin. Pract.* **2017**, *36*, 68–78. [[CrossRef](#)] [[PubMed](#)]
92. Schulman, G.; Berl, T.; Beck, G.J.; Remuzzi, G.; Ritz, E.; Arita, K.; Kato, A.; Shimizu, M. Randomized Placebo-Controlled EPPIC Trials of AST-120 in CKD. *J. Am. Soc. Nephrol.* **2015**, *26*, 1732–1746. [[CrossRef](#)] [[PubMed](#)]
93. Schulman, G.; Berl, T.; Beck, G.J.; Remuzzi, G.; Ritz, E.; Shimizu, M.; Shobu, Y.; Kikuchi, M. The Effects of AST-120 on Chronic Kidney Disease Progression in the United States of America: A Post Hoc Subgroup Analysis of Randomized Controlled Trials. *BMC Nephrol.* **2016**, *17*, 141. [[CrossRef](#)] [[PubMed](#)]