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RESEARCH ARTICLE

Immune status changing helps diagnose osteoarticular tuberculosis

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Abstract

Objective

This study is aimed to develop a new nomogram for the clinical diagnosis of osteoarticular tuberculosis (TB).

Methods

xCell score estimation to obtained the immune cell type abundance scores. We downloaded the expression profile of GSE83456 from GEO and proceed xCell score estimation. The routine blood examinations of 326 patients were collected for further validation. We analyzed univariate and multivariate logistic regression to identified independent predicted factor for developing the nomogram. The performance of the nomogram was assessed using the receiver operating characteristic (ROC) curves. The correlation of ESR with lymphocytes, monocytes, and ML ratio was performed and visualized in osteoarticular TB patients.

Results

Compared with the healthy control group in the dataset GSE83456, the xCell score of basophils, monocytes, neutrophils, and platelets was higher, while lymphoid was lower in the EPTB group. The clinical data showed that the cell count of monocytes were much higher, while the cell counts of lymphocytes were lower in the osteoarticular TB group. AUCs of the nomogram was 0.798 for the dataset GSE83456, and 0.737 for the clinical data. We identified the ML ratio, BMI, and ESR as the independent predictive factors for osteoarticular TB diagnosis and constructed a nomogram for the clinical diagnosis of osteoarticular TB. AUCs of this nomogram was 0.843.

Conclusions

We demonstrated a significant change between the ML ratio of the EPTB and non-TB patients. Moreover, we constructed a nomogram for the clinical diagnosis of the osteoarticular TB diagnosis, which works satisfactorily.

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Abbreviations: TB, Tuberculosis; EPTB, Extrapulmonary tuberculosis; TME, Tumor microenvironment; TIICs, Tumor-infiltrating immune cell; GSEA, Gene Set Enrichment Analysis; GO, gene ontology; KEGG, Kyoto Encyclopedia of Genes and Genomes; DEGs, differentially expressed genes; ROC, receiver operating characteristic; ESR, erythrocyte sedimentation rate; BMI, body mass index.

Introduction

World health organization (WHO) estimates that 1.8 billion people, about one-fourth of the global population, are *Mycobacterium tuberculosis* (*M.tb*) infected, out of which roughly 10 million contracted tuberculosis (TB) and 1.6 million died of the disease [1-3]. Therefore, TB remains a leading global public health problem [4, 5]. Till now, TB has been described in all virtual tissues or organs, including the spine, lymph nodes, abdomen, genitourinary tract, skin, joints, and meningeal [6, 7]. Extrapulmonary tuberculosis (EPTB) incidents are higher in immune-compromised individuals and represents 15% of the global TB cases [8]. However, an individual's immunity status decides whether patients with *M.tb* would develop active TB or not [9].

Despite the existence of guidelines for the diagnosis and treatment of TB [10], there tend to be cases of EPTB that present with atypical manifestations such as local pain, weight loss, night sweat and fever, which make the diagnosis difficult [11]. In addition, the anatomical sites frequently involves in EPTB are not easily accessible and require invasive procedures for diagnostic confirmation. The availability of little information is the reason for the difficulty and delay in the diagnosis of EPTB. Therefore, an urgent need arises to invent new methods for EPTB diagnosis.

The absolute number of monocytes or lymphocytes in peripheral blood or yet the ratio of monocytes to lymphocytes (ML ratio) has prognostic value in diseases such as hematopathy and tumors [12]. Previous studies have demonstrated that the ML ratio shows a predictive value for active TB [13]. The development of osteoarticular TB is the result of an immune system disorder. However, spine TB is the most common kind of osteoarticular TB, which influences the proportion of immune cells. Therefore, the discovery of immune status associated with osteoarticular TB can advance osteoarticular TB diagnosis.

In this study, the expression profile of dataset GSE83456 was downloaded to obtain the xCell score of the cell type, including lymphocytes, monocytes, neutrophils, eosinophils, basophils, erythrocyte, and platelet that belongs to the blood routine examination items. We found that the immune status of individuals changes during TB infection. ML ratio was found significantly up-regulated in the EPTB group than that in the non-TB group. Based on this result, we constructed a novel nomogram, which incorporated easy access to clinical characteristics like ML raito, erythrocyte sedimentation rate (ESR) and body mass index (BMI), for osteoarticular TB diagnosis.

Patients and methods

Microarray data and xCell score estimation

Gene Expression Omnibus (GEO, https://www.ncbi.nlm.nih.gov/geo/) is an open database from where we downloaded the gene expression profiles. The GSE83456 with platform GPL10558 (Illumina HumanHT-12 V4.0 expression beadchip) was downloaded and used to extracted blood sample profiles of 47 EPTB and 61 healthy controls (HCs) from the total 202 samples for further analysis. We analyzed the xCell score using the R package 'xCell' (https://github.com/dviraran/xCell), which allowed us to obtained 64 immune cell type abundance scores [14]. Next, the xCell score of lymphocytes, monocytes, neutrophils, eosinophils, basophils, erythrocyte, and platelet belonging to the blood routine examination items was picked out. The xCell score of lymphocytes was a constitution of 21 subtypes (S1 Fig) of lymphoid cells (B cells, T cells and NK cells). This score was consistent with the method of lymphocyte count in the routine blood examination.

Patients

Subjects volunteering for the study had signed informed consent forms. In addition, the Ethics Committee of The First Affiliated Hospital of Guangxi Medical University approved this study.

From 2012 to 2018, we consecutively screened out 173 osteoarticular TB patients from The First Affiliated Hospital of Guangxi Medical University. Following were the included diagnostic criteria for osteoarticular TB: (1) patients with typical symptoms of tuberculous infection, including low fever, night sweats, weight loss, and fatigue; (2) patients with positive *M.tb* antibody; (3) patients with spinal cord compression symptoms, such as pain, myodynamia, muscle tension, tendon reflexes, limited activity, and spinal deformities; (4) patients with the typical features of spinal TB on imageology, such as bone marrow edema, endplate erosion, vertebral destruction, and spinal compression; and (5) patients with tuberculous granuloma [15]. Exclusion criteria were set for osteoarticular TB patients: (1) co-infection with other types of bacteria, virus, and co-morbidities; (2) co-occurrence with tumor (3) already receiving anti-tuberculosis drug treatment; and (4) recurrent tuberculosis.

In The First Affiliated Hospital of Guangxi Medical University, from 2012 and 2018, we randomly screened out the non-TB patients from all the inpatients diagnosed with lumbar disc herniation or lumbar spinal stenosis. After identifying patients, 326 eligible patients were enrolled, including 164 osteoarticular TB patients and 162 non-TB patients.

Statistical analysis

Using the Student's t-test, we analyzed continuous variables such as xCell score of monocytes, peripheral blood of monocytes counts, or lymphocytes. To identify the independent risk factors in the dataset GSE83456 and the clinical data, we performed the univariate and multivariate logistics regression analyses using the R package 'rms' (https://cran.rstudio.com/bin/windows/contrib/4.0/rms_6.1-0.zip). Also, we constructed and compared the nomograms based on the logistics regression result obtained. The performance of the nomogram were assessed using the receiver operating characteristic (ROC) curves. Furthermore, factors in the clinical cohort sample were evaluated via univariate and multivariate logistic regression. The correlation of ESR with lymphocytes, monocytes, and ML ratio in osteoarticular TB patients was performed and visualized using the R package 'corrplot' (https://cran.rstudio.com/bin/windows/contrib/4.0/corrplot_0.84.zip). The value of P < 0.05 was considered a significant difference.

Gene Set Enrichment Analysis (GSEA)

Using GSEA (4.0.3), we identified the potential biological mechanisms of ML ratio, which were involved in the impact of osteoarticular TB [16]. The gene set permutations with 1000-times were conducted to acquire the normalized enrichment score (NES). The normal P-value < 0.05 and false discovery rate (FDR) < 0.25 was used to quantify statistically significant enrichment.

Results

Patient baseline characteristics

Table 1 illustrates the baseline characteristics collected for the 326 patients, such as age, sex, profession, nationality, BMI, ESR. In this study, spine TB accounted for 93.9% and constituted the majority of osteoarticular TB. BMI and hemoglobin were much lower in the osteoarticular TB group than in the non-TB group. However, ESR was much higher in the osteoarticular TB group (Table 1).

Table 1. Baseline characteristics of patients.

Characteristics	Osteoarticular TB (N = 164)	HC (N = 162)	Overall (N = 326)	P value
Mean (SD)	45.2 (17.0)	48.0 (17.2)	46.6 (17.2)	0.137
Median [Min, Max]	46.5 [3.00, 78.0]	51.0 [5.00, 86.0]	49.0 [3.00, 86.0]	
Sex				
Female	69 (42.1%)	69 (42.6%)	138 (42.3%)	1
Male	95 (57.9%)	93 (57.4%)	188 (57.7%)	
Nationality				
Han	80 (48.8%)	97 (59.9%)	177 (54.3%)	0.158
Zhuang	75 (45.7%)	59 (36.4%)	134 (41.1%)	
Yao	8 (4.9%)	4 (2.5%)	12 (3.7%)	
Others	1 (0.6%)	2 (1.2%)	3 (0.9%)	
Profession				
Farmer/Worker	100 (61.0%)	86 (53.1%)	186 (57.1%)	0.153
Office clerk	9 (5.5%)	10 (6.2%)	19 (5.8%)	
Student	7 (4.3%)	17 (10.5%)	24 (7.4%)	
Others	48 (29.3%)	49 (30.2%)	97 (29.8%)	
BMI				
Mean (SD)	20.1 (2.79)	22.5 (4.17)	21.3 (3.76)	< 0.001
Median [Min, Max]	19.8 [13.2, 29.4]	22.7 [13.5, 36.9]	21.0 [13.2, 36.9]	
Missing	20 (12.2%)	10 (6.2%)	30 (9.2%)	
Location				
-	0 (0%)	162 (100%)	162 (49.7%)	< 0.001
joint tuberculosis	10 (6.1%)	0 (0%)	10 (3.1%)	
spine tuberculosis	154 (93.9%)	0 (0%)	154 (47.2%)	
Hemoglobin				
Mean (SD)	119 (17.6)	127 (16.7)	123 (17.6)	< 0.001
Median [Min, Max]	121 [76.4, 153]	128 [75.8, 167]	124 [75.8, 167]	
Erythrocyte				
Mean (SD)	4.71 (3.01)	4.60 (0.663)	4.65 (2.19)	0.646
Median [Min, Max]	4.46 [2.83, 42.0]	4.56 [3.14, 7.07]	4.51 [2.83, 42.0]	
White blood cell				
Mean (SD)	7.21 (2.71)	7.62 (2.57)	7.41 (2.65)	0.163
Median [Min, Max]	6.78 [2.55, 17.2]	7.08 [3.10, 21.2]	6.90 [2.55, 21.2]	
Lymphocytes				
Mean (SD)	1.52 (0.798)	2.06 (0.780)	1.79 (0.834)	<0.001
Median [Min, Max]	1.38 [0.268, 7.21]	1.95 [0.382, 5.55]	1.67 [0.268, 7.21]	
Monocytes				
Mean (SD)	0.645 (0.269)	0.582 (0.205)	0.614 (0.241)	0.017
Median [Min, Max]	0.589 [0.20, 1.72]	0.553 [0.04, 1.52]	0.568 [0.04, 1.72]	
Neutrophil				
Mean (SD)	4 73 (2,25)	4 72 (2,49)	4 72 (2, 37)	0.964
Median [Min, Max]	4.24 [1 16, 14 7]	4.07 [1.55, 19.5]	4.14 [1.16.19.5]	0.001
Eosinophils		107 [100, 170]		
Mean (SD)	0.278 (0.474)	0 229 (0 172)	0 254 (0 357)	0.219
Median [Min. Max]	0 189 [0 5 73]	0.201 [0.0.976]	0.198 [0.573]	0.219
Basophils	0.107 [0, 5.75]	0.201 [0, 0.970]	0.170 [0, 0.75]	
r				

(Continued)

Characteristics	Osteoarticular TB	НС	Overall	P value
	(N = 164)	(N = 162)	(N = 326)	
Mean (SD)	0.034 (0.021)	0.036 (0.019)	0.035 (0.020)	0.225
Median [Min, Max]	0.031 [0, 0.162]	0.033 [0, 0.133]	0.032 [0, 0.162]	
Platelet				
Mean (SD)	290 (85.3)	250 (80.1)	270 (85.1)	< 0.001
Median [Min, Max]	274 [78.5, 562]	239 [98.0, 630]	256 [78.5, 630]	
ESR				
Mean (SD)	38.6 (23.6)	19.5 (19.6)	30.0 (23.8)	< 0.001
Median [Min, Max]	34.0 [1.00, 109]	11.0 [2.00, 94.0]	24.0 [1.00, 109]	
Missing	6 (3.7%)	34 (21.0%)	40 (12.3%)	

Table 1. (Continued)

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Osteoarticular tuberculosis group vs. non-TB group

The xCell score of 64 subtypes for each sample was calculated based on the expression file (S1 File). The xCell score of lymphocytes was sum of all xCell score of lymphoid cells (B cells, T cells and NK cell) in the dataset GSE83456 (S1 Fig). Compared with the HC group in the dataset GSE83456, the EPTB group has a higher xCell score of basophils, monocytes, neutrophils, and platelets, but lower xCell score of lymphocytes (Fig 1A). The cell counts of monocytes and platelets were much higher while that of lymphocytes were lower in the osteoarticular TB group of the clinical data (Fig 1B). Monocytes, lymphocytes and platelet were significantly different between the osteoarticular TB and non-TB group.

A high ML ratio is associated with osteoarticular TB

The ML ratio in the dataset GSE83456 represents the ratio of xCell score of monocytes to lymphocytes. The ML ratio in the clinical data represents the ratio of cell counts of monocytes to lymphocytes. In the EPTB group, the ML ratio was significantly higher in the dataset GSE83456 (Fig 2A) and clinical data (Fig 2B). We identified the ML ratio as an independent risk factor for the diagnosis of EPTB; therefore, nomograms (S2 Fig) based on the ML ratio were constructed for both dataset GSE83456 and clinical data. ROC analysis showed the AUC of the nomogram was 0.798 for the dataset GSE83456, and 0.737 for the clinical dataset (Fig 2C).

GSEA analysis showed that 22 hallmarks pathways were significantly enriched in the EPTB group, while none enriched in the HC group. Fig 2D shows 12 representative pathways, including apoptosis, reactive oxygen species (ROS), inflammatory response, interferon- α/γ response, complement, IL6/JAK/STAT3, and TNF α via NF- κ B. Moreover, GSEA analysis also showed that nine significant enriched pathways in the high ML ratio phenotype were consistent with those of the EPTB group (Fig 2E).

Nomogram for osteoarticular TB diagnosis

Fig 3A shows a nomogram based on three independent risk factors (ML ratio, ESR, and BMI), which we constructed for advancing clinical diagnosis of osteoarticular TB (Table 2). AUC of this nomogram was 0.843 (Fig 3B); also, the calibration curves indicated a satisfactory agreement between nomogram prediction and actual probabilities (Fig 3C). We further evaluated a correlation between monocytes, lymphocytes, and ESR. Monocytes and ML ratio was positively correlated with ESR while lymphocytes was negatively correlated with ESR. Fig 3D illustrates ML ratio was positively correlated with ESR (cor = 0.439, P < 0.0001).



Fig 1. Different cell type between EPTB and healthy controls. (A) Violin plot showed the xCell score of 7 kinds of blood cells in dataset GSE83456. (B) Violin plot showed the cell counting of 7 kinds of blood cells in clinical data. (C) Violin plot showed the cell rate of 5 kinds of blood cells in clinical data.

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Discussion

At present, TB poses a global threat for both developing and developed countries [17, 18], the innate immune response representing one of the most critical determinants associated with the outcome of EPTB infection [19]. In this study, the xCell score of monocytes and platelets is significantly higher, while that of lymphocytes is lower in the EPTB group in dataset GSE83456. Moreover, the ML ratio of the xCell score is significantly higher in the EPTB group. GSEA results showed pathways closely related to the inflammation, such as apoptosis, reactive oxygen species (ROS), inflammatory response, and interferon- α/γ response, were significantly enriched. Previous studies suggest that ROS and interferon- α/γ might be associated with *M.tb* immune escape and disease progression in infected humans [20, 21]. Micheliolide and nitric oxide play an anti-inflammatory role in *M.tb* infection by inhibiting NF- κ B that is a ubiquitously existed transcription factor family [22, 23]. These results demonstrated *M.tb* infection activates these inflammation-related pathways and plays an significant role in the pathogenesis of EPTB. The pathways significantly enriched in the EPTB group were also enriched in the high ML ratio phenotype, which indicates that a high ML ratio is a crucial characteristic of EPTB and is consistent with a previous study [13].

Understanding the immune responses protecting from infection or progression to disease is crucial to allow the development of diagnostic tools for the efficient prevention and



Fig 2. T test of ML ratio between EPTB and healthy controls and GSEA analysis. (A) Violin plot showed the ML ratio between EPTB and healthy controls in dataset GSE83456. (B) Violin plot showed the ML ratio between EPTB and healthy controls in clinical data. (C) AUCs of the nomogram based on ML ratio in GSE83456 and clinical data. (D) The enriched gene sets in HALLMARK collection by the EPTB samples. (E) The enriched gene sets in HALLMARK collection by the high ML ratio samples.

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management of EPTB. The assumption is further confirmed by the analysis of the clinical data. It had been proved that the cell counts and frequency of monocytes and lymphocytes were significantly different between TB disease and HCs [24], which were consistent with our results. Lymphocytes are thought to be the primary effector cells in TB immunity, while myeloid cells as the primary host cell for infection. Therefore, the relative abundance may reflect a balance between effector and target cells. An alternative explanation is that the relative abundance of these cell types could be a marker of hematopoietic parameters associated with TB. The ratio of myeloid transcripts to lymphoid transcripts were altered in inflammation-associated disease. Altogether, these data support the hypothesis that a high or a low ML ratio may be a readily available tool to identify the risk of TB during HIV infection of patients with acceptable combination of antiretroviral therapy [25, 26].

So far, the osteoarticular TB diagnosis relies on the comprehensive analysis of the history, imaging, and blood test. However, the challenges in the clinical identification of osteoarticular TB are compounded by a diagnostic armamentarium with significant limitations. Based on the



Fig 3. Nomogram and correlation analysis. (A) Nomogram for predicting osteoarticular TB probability for clinical data; The red line represents an osteoarticular TB patient while the green line represents an non-TB patient. (B) AUCs of the nomogram for clinical data. (C) Calibration curves for predicting osteoarticular TB probability for clinical data. (D) The correlation of ML ratio with ESR.

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Characteristics	Univariate logistic		Multiv	Multivariate logistic	
	HR	Р	HR	Р	
Age	0.99 [0.98, 1.00]	0.137			
Sex	1.02 [0.66, 1.59]	0.924			
BMI	0.82 [0.75, 0.88]	p<0.0001	0.80 [0.72, 0.88]	p<0.0001	
White blood cell	0.94 [0.87, 1.02]	0.165			
Erythrocyte	1.03 [0.92, 1.21]	0.656			
ESR	1.05 [1.03, 1.06]	p<0.0001	1.03 [1.01, 1.05]	0.001	
Hemoglobin	0.97 [0.96, 0.99]	p<0.0001	1.00 [0.98, 1.03]	0.663	
ML ratio	45.39 [13.37, 173.58]	p<0.0001	21.41 [2.42,222.93]	0.008	
PL ratio	1.01 [1.01, 1.01]	p<0.0001	1.00 [1.00, 1.01]	0.1	
PM ratio	1.00 [1.00, 1.00]	0.384			

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just mentioned facts, we try to benefit from ML ratio, which can help us with osteoarticular TB diagnosis since it is difficult to obtain pus or tissue for *M.tb* culture [27]. Besides, a bacterial culture is frequently culture-negative. In this study, the ML ratio was an independent predictive factor for osteoarticular TB diagnosis in both the dataset GSE83456 and clinical datasets. Higher ML ratio was associated with osteoarticular TB through bioinformatic analysis and clinical validation. ESR is one of the blood markers monitor TB and infection [28]. Moreover, ESR is positively related to the ML ratio. As the results are shown in this studies, osteoarticular TB will lead to anemia, low albumin, and low BMI [29, 30]. Moreover, nutritional deficiencies predispose to a worse osteoarticular TB infection [31]. Results of the study indicate that the ML ratio, BMI, and ESR are independent predictive factors for osteoarticular TB diagnosis. Therefore, we constructed a nomogram to improve the diagnosis accuracy of osteoarticular TB. The AUC of the nomogram, which demonstrated more accurate and practical performance, is 0.843. Our study provides clear indicators, easily acquired in clinical work, for improvement of osteoarticular TB diagnosis. Nonetheless, the osteoarticular TB diagnosis also depends on the imaging. Combine our nomogram with imaging may significantly improve the diagnosis of osteoarticular TB. Therefore, our future research will focus on this combination.

There are three important potential limitations of this study: 1) since this is a retrospective study of osteoarticular TB and non-TB group, there is a possibility of selection bias; 2) The participants of this study did not have TSTs or interferon γ -release assays performed; 3) Most of the diagnosed patients were typically confirmed to be pathologically positive but microbiologically negative.

Conclusion

We can conclude that the immune status of individuals changes during TB infection. Nonetheless, the ML ratio was identified as an independent predictive factor for EPTB diagnosis in the dataset GSE83456 and was considered a significant characteristic of osteoarticular TB. Lastly, the nomogram we constructed showed satisfactory ability to diagnose osteoarticular TB.

Supporting information

S1 Fig. Cell types in the microenvironment. (A, B, C, D, E) xCell score of sixty-four cell types in GSE83456 were grouped into five groups: lymphocytes, myeloids, stem, stromal, and other cells.

(TIF)

S2 Fig. Nomograms. (A) Nomogram for predicting EPTB probability base on ML ratio in dataset GSE83456. (B) Nomogram for predicting osteoarticular TB probability base on ML ratio in clinical data.

(TIF)

S1 File. xCell score of 64 subtypes immune cells of each sample. (XLSX)

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References

- Pai M, Behr MA, Dowdy D, Dheda K, Divangahi M, Boehme CC, et al. Tuberculosis. Nature reviews Disease primers. 2016; 2:16076. Epub 2016/10/28. <u>https://doi.org/10.1038/nrdp.2016.76</u> PMID: 27784885.
- Suarez I, Funger SM, Kroger S, Rademacher J, Fatkenheuer G, Rybniker J. The Diagnosis and Treatment of Tuberculosis. Deutsches Arzteblatt international. 2019; 116(43):729–35. Epub 2019/11/23. https://doi.org/10.3238/arztebl.2019.0729 PMID: 31755407.
- Furin J, Cox H, Pai M. Tuberculosis. Lancet (London, England). 2019; 393(10181):1642–56. Epub 2019/03/25. https://doi.org/10.1016/s0140-6736(19)30308-3 PMID: 30904262.
- Qian X, Nguyen DT, Lyu J, Albers AE, Bi X, Graviss EA. Risk factors for extrapulmonary dissemination of tuberculosis and associated mortality during treatment for extrapulmonary tuberculosis. Emerging microbes & infections. 2018; 7(1):102. Epub 2018/06/07. https://doi.org/10.1038/s41426-018-0106-1 PMID: 29872046; PubMed Central PMCID: PMC5988830.
- Nasiri MJ, Pormohammad A, Goudarzi H, Mardani M, Zamani S, Migliori GB, et al. Latent tuberculosis infection in transplant candidates: a systematic review and meta-analysis on TST and IGRA. Infection. 2019; 47(3):353–61. Epub 2019/02/26. https://doi.org/10.1007/s15010-019-01285-7 PMID: 30805899.
- Mohammed H, Assefa N, Mengistie B. Prevalence of extrapulmonary tuberculosis among people living with HIV/AIDS in sub-Saharan Africa: a systemic review and meta-analysis. HIV/AIDS (Auckland, NZ). 2018; 10:225–37. Epub 2018/11/23. https://doi.org/10.2147/HIV.S176587 PMID: 30464643; PubMed Central PMCID: PMC6225852.
- Gao W, Zeng Y, Chen W. Multiple subcutaneous tuberculous abscesses in a dermatomyositis patient without pulmonary tuberculosis: a case report and literature review. BMC infectious diseases. 2020; 20 (1):409. Epub 2020/06/14. https://doi.org/10.1186/s12879-020-05137-w PMID: 32532200; PubMed Central PMCID: PMC7291664.
- Tahseen S, Khanzada FM, Baloch AQ, Abbas Q, Bhutto MM, Alizai AW, et al. Extrapulmonary tuberculosis in Pakistan- A nation-wide multicenter retrospective study. PloS one. 2020; 15(4):e0232134. Epub 2020/04/29. https://doi.org/10.1371/journal.pone.0232134 PMID: 32343714; PubMed Central PMCID: PMC7188211.
- Cliff JM, Kaufmann SH, McShane H, van Helden P, O'Garra A. The human immune response to tuberculosis and its treatment: a view from the blood. Immunological reviews. 2015; 264(1):88–102. Epub 2015/02/24. https://doi.org/10.1111/imr.12269 PMID: 25703554; PubMed Central PMCID: PMC4368415.
- Pollett S, Banner P, O'Sullivan MV, Ralph AP. Epidemiology, Diagnosis and Management of Extra-Pulmonary Tuberculosis in a Low-Prevalence Country: A Four Year Retrospective Study in an Australian Tertiary Infectious Diseases Unit. PloS one. 2016; 11(3):e0149372. Epub 2016/03/11. https://doi.org/ 10.1371/journal.pone.0149372 PMID: 26963244; PubMed Central PMCID: PMC4786131.
- Kim JH, Kim ES, Jun KI, Jung HG, Bang JH, Choe PG, et al. Delayed diagnosis of extrapulmonary tuberculosis presenting as fever of unknown origin in an intermediate-burden country. BMC infectious diseases. 2018; 18(1):426. Epub 2018/08/30. https://doi.org/10.1186/s12879-018-3349-5 PMID: 30153813; PubMed Central PMCID: PMC6114835.
- Soyano AE, Dholaria B, Marin-Acevedo JA, Diehl N, Hodge D, Luo Y, et al. Peripheral blood biomarkers correlate with outcomes in advanced non-small cell lung Cancer patients treated with anti-PD-1

antibodies. Journal for immunotherapy of cancer. 2018; 6(1):129. Epub 2018/11/25. https://doi.org/10. 1186/s40425-018-0447-2 PMID: 30470260; PubMed Central PMCID: PMC6251165.

- La Manna MP, Orlando V, Dieli F, Di Carlo P, Cascio A, Cuzzi G, et al. Quantitative and qualitative profiles of circulating monocytes may help identifying tuberculosis infection and disease stages. PloS one. 2017; 12(2):e0171358. Epub 2017/02/17. https://doi.org/10.1371/journal.pone.0171358 PMID: 28208160; PubMed Central PMCID: PMC5313257.
- Aran D. Cell-Type Enrichment Analysis of Bulk Transcriptomes Using xCell. Methods in molecular biology (Clifton, NJ). 2020; 2120:263–76. Epub 2020/03/04. <u>https://doi.org/10.1007/978-1-0716-0327-7_19 PMID: 32124326</u>.
- Wang L, Shang X, Qi X, Ba D, Lv J, Zhou X, et al. Clinical Significance of M1/M2 Macrophages and Related Cytokines in Patients with Spinal Tuberculosis. Disease markers. 2020; 2020:2509454. Epub 2020/06/23. https://doi.org/10.1155/2020/2509454 PMID: 32566036; PubMed Central PMCID: PMC7267866.
- Tang Y, Gu Z, Fu Y, Wang J. CXCR3 from chemokine receptor family correlates with immune infiltration and predicts poor survival in osteosarcoma. Bioscience reports. 2019; 39(11). Epub 2019/11/07. https://doi.org/10.1042/BSR20192134 PMID: 31696204; PubMed Central PMCID: PMC6851512.
- Gaensbauer J, Broadhurst R. Recent Innovations in Diagnosis and Treatment of Pediatric Tuberculosis. Current infectious disease reports. 2019; 21(1):4. Epub 2019/02/16. https://doi.org/10.1007/s11908-019-0662-0 PMID: 30767077.
- Meier NR, Jacobsen M, Ottenhoff THM, Ritz N. A Systematic Review on Novel Mycobacterium tuberculosis Antigens and Their Discriminatory Potential for the Diagnosis of Latent and Active Tuberculosis. Front Immunol. 2018; 9:2476. Epub 2018/11/27. https://doi.org/10.3389/fimmu.2018.02476 PMID: 30473692; PubMed Central PMCID: PMC6237970.
- Wassermann R, Gulen MF, Sala C, Perin SG, Lou Y, Rybniker J, et al. Mycobacterium tuberculosis Differentially Activates cGAS- and Inflammasome-Dependent Intracellular Immune Responses through ESX-1. Cell Host Microbe. 2015; 17(6):799–810. Epub 2015/06/07. https://doi.org/10.1016/j.chom. 2015.05.003 PMID: 26048138.
- Krakauer T. Inflammasomes, Autophagy, and Cell Death: The Trinity of Innate Host Defense against Intracellular Bacteria. Mediators of inflammation. 2019; 2019:2471215. Epub 2019/02/08. https://doi. org/10.1155/2019/2471215 PMID: 30728749; PubMed Central PMCID: PMC6341260.
- 21. Ma J, Yang B, Yu S, Zhang Y, Zhang X, Lao S, et al. Tuberculosis antigen-induced expression of IFN-α in tuberculosis patients inhibits production of IL-1β. FASEB journal: official publication of the Federation of American Societies for Experimental Biology. 2014; 28(7):3238–48. Epub 2014/03/29. <u>https://doi.org/10.1096/fj.13-247056</u> PMID: 24675363.
- Braverman J, Stanley SA. Nitric Oxide Modulates Macrophage Responses to Mycobacterium tuberculosis Infection through Activation of HIF-1alpha and Repression of NF-kappaB. Journal of immunology (Baltimore, Md: 1950). 2017; 199(5):1805–16. Epub 2017/07/30. https://doi.org/10.4049/jimmunol. 1700515 PMID: 28754681; PubMed Central PMCID: PMC5568107.
- Zhang Q, Jiang X, He W, Wei K, Sun J, Qin X, et al. MCL Plays an Anti-Inflammatory Role in Mycobacterium tuberculosis-Induced Immune Response by Inhibiting NF-kappaB and NLRP3 Inflammasome Activation. Mediators of inflammation. 2017; 2017:2432904. Epub 2017/06/24. https://doi.org/10.1155/2017/2432904 PMID: 28642632; PubMed Central PMCID: PMC5470027.
- Habtamu M, Abrahamsen G, Aseffa A, Andargie E, Ayalew S, Abebe M, et al. High-throughput analysis of T cell-monocyte interaction in human tuberculosis. Clinical and experimental immunology. 2020; 201 (2):187–99. Epub 2020/04/30. <u>https://doi.org/10.1111/cei.13447</u> PMID: <u>32348546</u>; PubMed Central PMCID: PMC7366737.
- 25. Naranbhai V, Kim S, Fletcher H, Cotton MF, Madhi SAJBM. The association between the ratio of monocytes:lymphocytes at age 3 months and risk of tuberculosis (TB) in the first two years of life. 2014; 12 (1):120.
- 26. Naranbhai V, Hill AV, Abdool Karim SS, Naidoo K, Abdool Karim Q, Warimwe GM, et al. Ratio of monocytes to lymphocytes in peripheral blood identifies adults at risk of incident tuberculosis among HIV-infected adults initiating antiretroviral therapy. The Journal of infectious diseases. 2014; 209(4):500–9. Epub 2013/09/18. https://doi.org/10.1093/infdis/jit494 PMID: 24041796; PubMed Central PMCID: PMC3903371.
- Ma Y, Li R, Shen J, He L, Li Y, Zhang N, et al. Clinical effect of T-SPOT.TB test for the diagnosis of tuberculosis. BMC infectious diseases. 2019; 19(1):993. Epub 2019/11/23. https://doi.org/10.1186/ s12879-019-4597-8 PMID: 31752713; PubMed Central PMCID: PMC6873440.
- Ding RD, Zhang HJ. Effect of linezolid on serum PCT, ESR, and CRP in patients with pulmonary tuberculosis and pneumonia. Medicine. 2018; 97(37):e12177. Epub 2018/09/15. https://doi.org/10.1097/MD. 000000000012177 PMID: 30212947; PubMed Central PMCID: PMC6155971.

- Yadav J, Verma S, Chaudhary D, Jaiwal PK, Jaiwal R. Tuberculosis: Current Status, Diagnosis, Treatment and Development of Novel Vaccines. Current pharmaceutical biotechnology. 2019; 20(6):446–58. Epub 2019/06/19. https://doi.org/10.2174/1389201020666190430114121 PMID: 31208308.
- Tang J, Yam WC, Chen Z. Mycobacterium tuberculosis infection and vaccine development. Tuberculosis (Edinburgh, Scotland). 2016; 98:30–41. Epub 2016/05/10. https://doi.org/10.1016/j.tube.2016.02. 005 PMID: 27156616.
- Moreira-Teixeira L, Mayer-Barber K, Sher A, O'Garra A. Type I interferons in tuberculosis: Foe and occasionally friend. The Journal of experimental medicine. 2018; 215(5):1273–85. Epub 2018/04/19. https://doi.org/10.1084/jem.20180325 PMID: 29666166; PubMed Central PMCID: PMC5940272.