RESEARCH



Open Access

Detection of Hepatitis B virus subgenotype A1 in a Quilombo community from Maranhão, Brazil

Mónica V Alvarado-Mora^{1*}, Livia Botelho¹, Michele S Gomes-Gouvêa¹, Vanda F de Souza², Maria C Nascimento², Claudio S Pannuti², Flair J Carrilho¹ and João RR Pinho¹

Abstract

Background: The Brazilian population is mainly descendant from European colonizers, Africans and Native Americans. Some Afro-descendants lived in small isolated communities since the slavery period. The epidemiological status of HBV infection in Quilombos communities from northeast of Brazil remains unknown. The aim of this study was to characterize the HBV genotypes circulating inside a Quilombo isolated community from Maranhão State, Brazil.

Methods: Seventy-two samples from Frechal Quilombo community at Maranhão were collected. All serum samples were screened by enzyme-linked immunosorbent assays for the presence of hepatitis B surface antigen (HBsAg). HBsAg positive samples were submitted to DNA extraction and a fragment of 1306 bp partially comprising HBsAg and polymerase coding regions (S/POL) was amplified by nested PCR and its nucleotide sequence was determined. Viral isolates were genotyped by phylogenetic analysis using reference sequences from each genotype obtained from GenBank (n = 320). Sequences were aligned using Muscle software and edited in the SE-AL software. Bayesian phylogenetic analyses were conducted using Markov Chain Monte Carlo (MCMC) method to obtain the MCC tree using BEAST v.1.5.3.

Results: Of the 72 individuals, 9 (12.5%) were HBsAg-positive and 4 of them were successfully sequenced for the 1306 bp fragment. All these samples were genotype A1 and grouped together with other sequences reported from Brazil.

Conclusions: The present study represents the first report on the HBV genotypes characterization of this community in the Maranhão state in Brazil where a high HBsAg frequency was found. In this study, we reported a high frequency of HBV infection and the exclusive presence of subgenotype A1 in an Afro-descendent community in the Maranhão State, Brazil.

Keywords: Hepatitis B virus, Genotype A1, Quilombo community, Maranhão state, Brazil, Bayesian Analysis

Introduction

It is estimated that two billion people have been infected with Hepatitis B virus (HBV) and that more than 350 million are chronic carriers of this virus [1]. HBV strains have distinct geographical distribution and are classified into nine genotypes, A to I, on basis of genome diversity [2]. Genotype A is found mainly in North America and Africa and has been found in six genetically distinct

* Correspondence: monica.viviana@usp.br

subgenotypes (A1 to A6) [3]. Genotypes B and C are prevalent in Southeast Asia and the Far East. Genotype D has a worldwide distribution and is found predominantly in the Mediterranean region and Central Asia. Genotypes E and F are prevalent in West Africa and in the Native American population, respectively [4]. In addition, genotype G has been reported in the USA, France [5], Colombia [6] and Brasil [7] and genotype H has been found in North and Central America [8]. Recently, by using phylogenetic analyses, a new genotype was characterized in Vietnam and Laos and designated as genotype I [2,9,10].



© 2011 Alvarado-Mora et al; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

¹Laboratory of Tropical Gastroenterology and Hepatology, São Paulo Institute of Tropical Medicine and Department of Gastroenterology, School of Medicine, University of São Paulo, São Paulo, Brazil

Full list of author information is available at the end of the article

In Brazil, a wide variation of HBV infection prevalence was reported, particularly dependent upon the geographical region of this country [11]. Genotype A is the most prevalent and genotypes B, C, D and F are also circulating in the population [12-15]. The presence of these genotypes reflects the mixture of cultures in the Brazil: Native American, European and African ancestral roots, showing this country as an important model for studies of population genetics [16].

During the slavery period in Brazil (from XVI to XIX centuries), some African people managed to escape to refuge areas, living with others in well hidden places in the woods. These places were known as Quilombos and they were regions of large concentration of runaway-slaves, far from urban centers and located in areas with difficult access. Their inhabitants generally stayed in culturally isolated communities without significant additional admixture since their establishment. Quilombos were located in different Brazilian states around the country: Pará (PA), Maranhão (MA), Alagoas (AL), Pernambuco (PE), Bahia (BA), Goiás (GO), Mato Grosso do Sul (MS), Minas Gerais (MG), Rio de Janeiro (RJ) and São Paulo (SP) [17].

The quilombo Frechal is located in the municipality of Mirinzal, at the region of lower western Maranhão State, Brazil (Figure 1). The Frechal community was founded in the late XVIII century and was devoted to sugar production till the XIX century. The Frechal community is one of the oldest and most important Quilombos located in Maranhão State (http://www.cpisp.org.br/comunidades/html/brasil/ma/ma_comunidades_frechal. html, accessed at 11/01/2011).

The aim of this study was to analyze the presence of current HBV infection by HBsAg and HBV DNA detection in the current inhabitants of Frechal community and to determine which HBV genotypes are found among these cases.

Materials and methods

Study Population

Seventy-two samples from inhabitants from Frechal, Maranhão were collected from 2000 to 2001 (Figure 1). Ethical Committee of the University of Sao Paulo, Medical School, Sao Paulo, Brazil, approved this protocol. All patients have signed an informed consent form before the samples were collected for research. The samples were screened for HBsAg using commercially available kits (DiaSorin Ltda, Saluggia, Italy). HBsAg positive samples were submitted to PCR amplification to detect HBV DNA.

HBV DNA extraction

Viral nucleic acids (HBV DNA and HDV RNA) extraction was carried out from 100 μl of sera for each sample

using the acid guanidinium thiocyanate/phenol/chloroform method [18]. To avoid false-positive results, strict procedures for nucleic acid amplification diagnostic techniques were followed [19].

HBV PCR Amplification

Samples were first amplified with the primers described by Sitnik et al. [13] in order to obtain a 416 base pairs (bp) fragment partially covering the HBsAg coding region (S) to confirm the presence of HBV-DNA in the sample.

To characterize HBV genotypes, a fragment of 1306 bp partially comprising HBsAg and the polymerase coding regions (S/POL) of HBV genome was amplified by nested PCR using the primers PS3132F (3132 nt - 3151 nt)/2920R (1417 nt - 1398 nt) and PS3201F (3201 nt -3221 nt)/P1285R (1285 nt - 1266 nt) [6].

HBV Nucleotide Sequencing

Amplified DNA was purified using ChargeSwitch[®] PCR Clean-Up Kit (Invitrogen, São Paulo, Brazil). Sequencing was performed in an ABI Prism[®] 377 Automatic Sequencer (Applied Biosystems, Foster City, CA, USA), based on the protocol described by Sanger et al. [20],



Figure 1 Geographical localization of the Afro-Brazilian community in Maranhão State - Brazil (modified of http://pt. wikipedia.org/wiki/Ficheiro:Maranhao_Municip_Mirinzal.svg).

using dideoxy nucleotide triphosphates (ddNTPs) containing fluorescent markers (*Big Dye*[®] *Terminator v3.1 Cycle Sequencing Ready Reaction kit* - Applied Biosystems, Foster City, CA, USA). The quality of each electropherogram was evaluated using the Phred-Phrap software [21] and consensus sequences were obtained by using an alignment constructed with CAP3 software available at the web page Electropherogram quality analysis (http://asparagin.cenargen.embrapa.br/phph/).

HBV Genotyping Analysis

Sequences were assigned to HBV genotypes after phylogenetic analysis using reference sequences from each HBV genotype obtained from the GenBank (n = 320). These sequences comprised partial HBsAg and polymerase coding regions (S/POL). They were aligned using Muscle Software [22] and edited with the SE-AL software (available at http://tree.bio.ed.ac.uk/software/seal/). To perform the phylogenetic analysis, the missing nucleotides were coded as "missing characters" in nexus block. Bayesian phylogenetic analyses were done applying Markov Chain Monte Carlo simulation using BEAST v.1.5.3 [23], and 10 million generations were sufficient to obtain the convergence of parameters. The relaxed uncorrelated log_{normal} was the best molecular clock for the dataset. The maximum clade credibility (MCC) tree was obtained from summarizing the 10,000 substitution trees and then it was removed 10% of burnin using Tree Annotator v.1.5.3 [23].

Results

Of the 72 samples, 9 (12.5%) were HBsAg-positive. Since we did not have epidemiological data about this population, it was not possible to compare HBsAg results with other demographic information. Of these nine samples, six were positive by nested PCR for the S fragment (416 bp) and among them, 4 also amplified the S/POL region (1306 bp).

To perform the phylogenetic analysis, the longest fragment available from each sample was sequenced and classified as subgenotype A1 (subtype *adw2*). Although the four samples from Frechal were grouped in the same cluster in the tree, it cannot be assumed that there was a founder effect in this community because this group is supported only with a low posterior probability (0.10) (Figure 2). Sequences were deposited in GenBank at accession numbers: HM772994 - HM772997.

Discussion

This is the first study that characterized HBV genotypes present in an isolated community from Maranhão state, Brazil. Since the genotype A1 has been reported as a common genotype in African and Brazilian populations [12,13,24-28], the four samples were compared with others previously reported sequences from Quilombos in Brazil [28] and Venezuela [29] and with sequences from Rio de Janeiro, as in this place there was a constant slave traffic from Africa that was most intense between 1795 to 1811 [30]. The subgenotype A1 sequences from Quilombos and Rio de Janeiro did not assemble in a single group in the tree. These results suggest the presence of the different A1 strains within the Quilombos populations. These variants may have come from Africa before these groups were created and actually the HBV/A1 strains continued to evolve in the Afro-descendent population after they came to Brazil.

A lot of Afro-descendants live in Bahia in Northeast Brazil. The geographic distributions of HBV genotypes have showed that genotype A (subtype *adw2*) is the most frequent in according to the ethnic background of the population [31]. It was reported a high prevalence of genotype A in Bahia but short-length sequences available prevented performing phylogenetic analysis together with the other ones analyzed in this study. Moreover, it is possible that different A1 variants have different African origin, as during the slave trade time, slaves from different countries from Africa were mixed on the boats and then were sold and distributed in different Brazilian regions [32].

We found that the three sequences from Afro-Venezuelan population [29] were classified as subgenotype A2 (Figure 3), which suggest different origin of HBV strains circulating in this population compared with African descendants in Brazil. Moreover, a specific polymorphism was found in HBV S region that distinguish subgenotypes A1 and A2 and it was agreed with HBV genotype A classification previously reported [33]. The synonymous substitutions at nucleotides in the third-positions: 201 nt (TCC \rightarrow TCA) Serine [S]; 222 nt (CCC \rightarrow CCA) Proline [P]; 324 nt (TCA \rightarrow TCG) Serine [S]; 327 nt (TCT \rightarrow TCC) Serine [S] and 462 nt (TAC \rightarrow TAT) Tryptofan [W] were identified and they confirm the results of subgenotype classification (Figure 3).

Finally, we found a high frequency of HBsAg in the Frechal population (12.5%). Kalunga population, the largest Afro-Brazilian isolated community located in Goiás state, showed lower frequency for HBsAg (1.8% - 16/878). HBV subgenotype (A1) was also found in all the positive samples from Goiás [34].

Another study, with 1058 individuals living in12 different isolated Afro-descendant communities, was carried out in Mato Grosso do Sul state and showed that among 1058 individuals, 23 (2.2%) of them were HBsAg positive. The highest prevalence was detected in Furnas dos Dionisios community (42.4% to anti-HBc and 7.4% to HBsAg) and the overall prevalence for HBV infection was 19.8% [35]. Subgenotype A1 was the most frequent



Figure 2 The maximum clade credibility (MCC) tree was estimated by Bayesian analysis of 320 S/POL sequences with 1306 bp of Hepatitis B virus strains. The posterior probabilities of the key nodes (internal nodes) are depicted above the respective nodes. Samples HBV/A1 obtained from Frechal (n = 4, red taxa) were analyzed together with other worldwide strains by Maximum Likelihood (ML) method (n = 134). The clusters containing the strains of others Afro-Brazilian communities previously reported are highlighted (Blue taxes). Genotype B (n = 19), genotype C (n = 20), genotype D (n = 24), genotype E (n = 7), genotype F (n = 25), genotype G (n = 4), genotype H (n = 6) and genotype I (n = 9) branches were collapsed. Also, the subgenotypes A2 (n = 52), A3, A4, and A5 (n = 10), A6 (n = 3) and A7 (n = 7) branches were collapsed.

	[184	4				201	L						222		228]	[313			324	327			33	9]	[457	462		468]	nt
A1 UFE220 Bie de Janeiro	I	CTA				TEC				ACT				TCC			ATC					ст т			I .				
A1_U55221_Rio_de_Janeiro	CCT	CTA	ATT	CCA	GGA	the	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	000	CCC	ATC	CCA						CA .	TGG		TGG	666	
A1 U55222 Rio de Janeiro	CCT	СТА	ATT	CCA	GGA	TIC	ACA	ACA	ACC	AGT	ACG	GGA	cec	TGC	۵۵۵	CCC	ATC	CCA	TAA	1 1	GG G	стт	TC G	CA .	TGG	TAC	TGG	GGG	
A1 AY344100 Rio de Janeiro	CCT	CTA	ATT	CCA	GGA	TEC	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	tat h	GG G	стт	TC G	CA	TGG	TAC	TGG	GGG	
360 Maranhao	CCT	CTA	ATT	CCA	GGA	TEC	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	Tet h	GG G	стт	TC G	CA	TGG	TAC	TGG	GGG	
620 Maranhao	CCT	CTA	ATT	CCA	GGA	TEC	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	Tet h	GG G	стт	TC G	CA	TGG	TAC	TGG	GGG	
650 Maranhao	ССТ	CTA	ATT	CCA	GGA	TCC	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	TET	GG G	ст т	TC G	CA	TGG	TAC	TGG	GGG	
700 Maranhao	ССТ	СТА	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	tet h	GG G	ст т	TC G	CA	TGG	TAC	TGG	GGG	
A1_FJ174795_Mato_Grosso_do_Sul	ССТ	СТА	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	t¢t h	GG G	ст т	TC G	CA	TGG	тлс	TGG	GGG	
A1_FJ174794_Mato_Grosso_do_Sul	ССТ	СТА	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	t¢t 1	GG G	ст т	TC G	CA	TGG	ТАС	TGG	GGG	
A1_FJ174796_Mato_Grosso_do_Sul	ССТ	CTA	ATT	CCA	GGA	TCC	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	T¢A	t¢t 1	GG G	ст т	TC G	CA	TGG	тас	TGG	GGG	
A1_FJ174797_Mato_Grosso_do_Sul	ССТ	CTA	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	T¢A	t¢t 1	GG G	ст т	TC G	CA I	GGG	тас	TGG	GGG	
A1_FJ174798_Mato_Grosso_do_Sul	ССТ	CTA	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	T¢A	T¢T 1	GG G	ст т	TC G	CA	TGG	ТАС	TGG	GGG	
A1_FJ174799_Mato_Grosso_do_Sul	ССТ	CTA	ATT	CCA	GGA	TCC	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	T¢A	t¢t 1	GG G	ст т	TC G	CA	TGG	тас	TGG	GGG	
A1_FJ174800_Mato_Grosso_do_Sul	ССТ	СТА	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	T¢A	t¢t 1	GG G	ст т	TC G	CA	TGG	TAC	TGG	GGG	
A1_EF547829_Mato_Grosso_do_Sul	ССТ	СТА	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	AGA	CCC	TGC	AAA	CCC	ATC	CCA	T¢A	T¢T 1	GG G	ст т	TC G	CA	TGG	TAC	TGG	GGG	
A1_EF547830_Mato_Grosso_do_Sul	ССТ	СТА	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	AGA	CCC	TGC	AAA	CCC	ATC	CCA	TÇA	T¢T 1	GG G	ст т	TC	CA	TGG	TAC	TGG	GGG	
A1_EF547831_Mato_Grosso_do_Sul	ССТ	СТА	ATT	CCA	GGA	тсс	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	T¢T 1	GG G	ст т	TC G	CA	TGG	TAC	TGG	GGG	
A1_EF547832_Mato_Grosso_do_Sul	ССТ	СТА	ATT	СТА	GGA	TCC	ACA	ACA	ACC	AGT	ACG	GGA	CCC	TGC	AAA	CCC	ATC	CCA	TCA	TET	GG G	ст т	TC G	CA	TGG	TAC	TGG	GGG	
A2_AF479492_Afro_Venezuelan	ССТ	СТА	ATT	CCA	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TCG	TCC 1	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A2_AF479491_Afro_Venezue1an	CCT	CTA	ATT	CCA	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	IGC	AAA	CCC	ATC	CCA	TEG		GG G		TC G	CA	TGG		TGG	GGG	
A2_AF479490_Afro_Venezuelan	CCT	CIA	ATT	CCA	GGA	1LA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	IGC	AAA	CCC	ATC	CCA	TEG		GG G		TC G	CA	IGG		IGG	GGG	
A2_V00866_USA	CCT	CTA	ATT	CCA	GGA		ACA	ACA	ALC	AGI	ACG	GGA	CCA	TGC	AAA	ccc	ATC	CCA						CA	TGG		TGG	GGG	
A2_235/1/_POTand	CCT	CTA	ATT	CCA	GGA		ACA	ACA	ACC	AGT	ACG	GGA	CCA	TCC	AAA	ccc	ATC	CCA						CA	TCC	11		666	
A2_A02703_05A	CCT	CTA	ATT	CCA	GGA	114	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	ccc	ATC	CCA	TAG					CA	TGG	-1-	TGG	666	
A2_08//30_SATFICA	CCT	CTA	ATT	CCA	GGA	114	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	Tre					CA	TCC	-1-1	TCC	666	
A2_AF369532 Mexico	CCT	CTA	ATT	CCA	GGA		ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	000	CCC	ATC	CCA	Tra					CA .	TGG	11	TGG	666	
A2_AF965116 Germany	CCT	CTA	ATT	CCA	GGA	TA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC		CCC	ATC	CCA	TEG		GG G	стт	TC G	CA .	TGG	TAT I	TGG	GGG	
A2 AF209395 Germany	ССТ	CTA	ATT	CCA	GGA	TEA	ACA	ACA	ACC	AGT	ACG	GGA	ACA	TGC	AAA	CCC	ATC	CCA	TEG	Tec h	GG G	стт	TC G	CA	TGG	TAT I	TGG	GGG	
A2 AF065111 Germany	CCT	CTA	ATT	CTA	GGA	TEA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TEG	Tec h	GG G	стт	TC G	CA	TGG	TAT	TGG	GGG	
A2 AF129506 UK	CCT	CTA	ATT	CCA	GGA	TEA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TCG	TCC h	GG G	стт	TC G	CA	TGG	TAT	TGG	GGG	
A2 AF132443 UK	ССТ	СТА	ATT	CCA	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TCG	тес п	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A2_AF209399_Germany	ССТ	СТА	ATT	CCA	GGA	ACA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TCG	TCC 1	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A2_AF065113_Germany	ССТ	СТА	ATT	CCA	GGA	ACA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TCG	t¢t h	GG G	ст т	TC G	CA	TGG	ТАТ	TGG	GGG	
A2_AF065115_Germany	ССТ	СТА	ATT	CCA	GGA	ACA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TCG	t¢t 1	GG G	ст т	TC G	CA	TGG	ТАТ	TGG	GGG	
A2_AF222311_USA	ССТ	CTA	ATT	CCA	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	T¢G	t¢t 1	GG G	ст т	TC G	CA	TGG	тат	TGG	GGG	
A2_AF208866_USA	ССТ	ATA	CTT	CCA	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	T¢G	t¢c 1	GG G	ст т	TC G	CA	TGG	TAT	tgg	GGG	
A2_AF209392_Germany	ССТ	CTA	ATT	СТА	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	T¢G	T¢T 1	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A2_AF209400_Germany	ССТ	СТА	ATT	CCA	GGA	ACA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	T¢G	t¢c 1	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A2_U91824_Nicaragua	ССТ	СТА	ATT	CCA	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TCG	TCC 1	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A2_AF209404_Germany	ССТ	СТА	ATT	CCA	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	CCC	ATC	CCA	TCG	TCC 1	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A2_087729_SAfrica	CCT	CTA	ATT	CCA	GGA	TCA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	IGC	AAA	CCC	ATC	CCA	TEG		GG G		TC G	CA	TGG		TGG	GGG	
A2_AF090841_BEL	CCT	CIA	ATT	CCA	GGA	1LA	ACA	ACA	ACC	AGT	ACG	GGA	CCA	IGC	AAA	CCC	ATC	CCA	TEG		GG G		TC G	CA	IGG		IGG	GGG	
A2_AF143299_Germany	CCT	CTA	ATT	CCA	GGA		ACA	ACA	ALC	AGI	ACG	GGA	CCA	TGC	AAA	ccc	ATC	CCA					TC G	CA	TGG		TGG	GGG	
A2_AF090039_DEL	CCT	CTA	ATT	CCA	GGA	114	ACA	ACA	ACC	AGT	ACG	GGA	CCA	TGC	AAA	ccc	ATC	CCA						CA	TGG	-1-	TGG	666	
A2_2/24/8_Germany	CCT	CTA	ATT	CCA	GGA	TIC	ACA	ACC	ACC	AGT	ACG	GGA	CCC	TGC	AGA	000	ATC	CCA	TCA			CT 1	TC 6	CA .	TGG	i li l	TGG	666	
A3 EN545829 Nigeria	ССТ	CTA	ATT	CCA	GGA	TIC	ACA	ACC	ACC	AGT	ACG	GGA	CCC	TGC	AGA	CCC	ATC	CCA	TCA		GG G	стт	TC G	CA .	TGG	TAT I	TGG	GGG	
A3 AM184126 GABON	ССТ	CTA	ATT	CCA	GGA	TEC	ACA	ACC	ACC	AGT	ACG	GGA	CCC	TGC	AGA	CCC	ATC	CCA	TCA	TCC I	GG G	стт	TC G	CA	TGG	TAT	TGG	GGG	
A3 AM184125 GABON	CCT	CTA	ATT	CCA	GGA	TEC	ACA	ACC	ACC	AGT	ACG	GGA	ccc	TGC	AGA	CCC	ATC	CCA	TCA	Tec h	GG G	стт	TC G	CA	TGG	TAT	TGG	GGG	
A4 AM180623 MAL36	CCT	CTA	ATT	CCA	GGA	TEC	ACA	ACA	ACC	AGT	ACG	GGA	ccc	TGC	AGG	CCC	ATC	CCA	TCA	TCA	GG G	стт	TC G	CA	TGG	TAT	TGG	GGG	
A4 AY934764 Gambia	CCT	CTA	ATT	CCA	GGA	тс	ACA	ACA	ACC	AGC	ACG	GGA	ccc	TGC	AGA	CCC	ATC	CCA	TCA	TCA	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A5 FJ692613 Haiti	ССТ	CTA	ATT	CCA	GGA	тс	ACA	ACC	ACC	AGT	ACG	GGA	CCC	TGC	AGA	CCC	ATC	CCA	TCA	TCC h	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A5_FJ692603_Haiti	CCT	CTA	ATT	CCA	GGA	тсс	ACA	ACC	ACC	AGT	ACG	GGA	CCC	TGC	AGA	CCC	ATC	CCA	TCA	TCC 1	GG G	ст т	TC G	CA	TGG	TAT	TGG	GGG	
A6_GQ331046_Belgium	CCT	CTA	ATT	CCA	GGA	ACC	ACA	ACA	ACC	AGT	ACG	GGA	ccc	TGC	AGA	CCC	ATC	CCA	TCA	TET	GG G	ст т	TC G	CA	TGG	TAC	TGG	GGG	
Figure 3 Multiple alignment	t of	ра	rtial	ΗВ	V/S	gei	ne (184	to	468	3 nt)) со	mpr	isir	ng ⊦	IBV/	/A1	seq	uen	ces	fron	n B	razi	l, w	hic	h a	re o	omp	ared

with other sequences of subgenotypes HBV/A2 to HBV/A6 previously reported. The five specific substitutions for subgenotype A1 are shown in the figure (blue squares)

in this state, followed by subgenotype A2 and genotype D [28].

HBsAg frequency in Afro-Venezuelan communities (3.6%) and in rural populations from Venezuela were higher than those found in Brazilian studies involving Quilombo inhabitants (2.2%) [29,35].

In Frechal, seventy-two samples were collected and this sample size probably did not represent the population but our data suggest a high frequency of HBV in this community. However, further studies are needed to better evaluate the epidemiology of hepatitis B in this region.

In this study, we reported a high frequency of HBV infection and the exclusive presence of subgenotype A1 in an Afro-descendent community in the Maranhão State, Brazil. In conclusion, HBV subgenotype A1 was found in all African descendant population from South

America studied so far except in Venezuela, where subgenotype A2 was found. Also, this study shows the need to report more HBV sequences from Afro-descendant communities to complement the actual data and the establishment of an accurate substitution rate for this virus to understand the evolutionary origins. The study of HBV sequences from other Afro-descendant communities from American countries, as well as from other different African countries, will allow a better understanding on HBV spreading between these two continents.

Acknowledgements

We are deeply indebted to Maria Claudia Nascimento and Laura Sumita for provide the samples for this study. This work has been supported by Fundação de Amparo à Pesquisa do Estado de São Paulo - FAPESP 2007/ 53457-7 and 2008/50461-6 and CNPq.

Author details

¹Laboratory of Tropical Gastroenterology and Hepatology, São Paulo Institute of Tropical Medicine and Department of Gastroenterology, School of Medicine, University of São Paulo, São Paulo, Brazil. ²Laboratory of Virology, São Paulo Institute of Tropical Medicine, Department of Infectious and Parasitic Diseases, School of Medicine, University of São Paulo, Brazil.

Authors' contributions

MVAM participated in the design of the study, conducted the phylogenetic and evolutionary analysis, drafted the manuscript and in its design and coordination. LB participated in the PCR amplification and sequencing process. MSGG participated in the PCR amplification. VFS, MCN, CSP and FJC participated in the design of the study. JRRP participated in the design of the study and drafted the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 15 May 2011 Accepted: 25 August 2011 Published: 25 August 2011

References

- 1. World Health Organization: Hepatitis B. Fact Sheet 204 (Revised August 2011) World Health Organization;[http://www.who.int/mediacentre/factsheets/ fs204/en/index.html].
- Yu H, Yuan Q, Ge SX, Wang HY, Zhang YL, Chen QR, Zhang J, Chen PJ, Xia NS: Molecular and phylogenetic analyses suggest an additional hepatitis B virus genotype "I". PLoS One 2010, 5:e9297.
- Pourkarim MR, Amini-Bavil-Olyaee S, Lemey P, Maes P, Van Ranst M: HBV subgenotype misclassification expands quasi-subgenotype A3. Clin Microbiol Infect 2011, 17:947-949.
- Magnius LO, Norder H: Subtypes, genotypes and molecular epidemiology of the hepatitis B virus as reflected by sequence variability of the Sgene. Intervirology 1995, 38:24-34.
- Stuyver L, De Gendt S, Van Geyt C, Zoulim F, Fried M, Schinazi RF, Rossau R: A new genotype of hepatitis B virus: complete genome and phylogenetic relatedness. *The Journal of general virology* 2000, 81:67-74.
- Alvarado Mora MV, Romano CM, Gomes-Gouvea MS, Gutierrez MF, Botelho L, Carrilho FJ, Pinho JR: Molecular characterization of the Hepatitis B virus genotypes in Colombia: a Bayesian inference on the genotype F. Infection, genetics and evolution: journal of molecular epidemiology and evolutionary genetics in infectious diseases 2011, 11:103-108.
- Bottecchia M, Souto FJ, O KM, Amendola M, Brandao CE, Niel C, Gomes SA: Hepatitis B virus genotypes and resistance mutations in patients under long term lamivudine therapy: characterization of genotype G in Brazil. BMC microbiology 2008, 8:11.
- Arauz-Ruiz P, Norder H, Robertson BH, Magnius LO: Genotype H: a new Amerindian genotype of hepatitis B virus revealed in Central America. *The Journal of general virology* 2002, 83:2059-2073.
- Hannoun C, Norder H, Lindh M: An aberrant genotype revealed in recombinant hepatitis B virus strains from Vietnam. J Gen Virol 2000, 81:2267-2272.
- Stuyver L, De Gendt S, Van Geyt C, Zoulim F, Fried M, Schinazi RF, Rossau R: A new genotype of hepatitis B virus: complete genome and phylogenetic relatedness. J Gen Virol 2000, 81:67-74.
- Pereira LM, Martelli CM, Merchan-Hamann E, Montarroyos UR, Braga MC, de Lima ML, Cardoso MR, Turchi MD, Costa MA, de Alencar LC, et al: Population-based multicentric survey of hepatitis B infection and risk factor differences among three regions in Brazil. Am J Trop Med Hyg 2009, 81:240-247.
- Araujo NM, Mello FC, Yoshida CF, Niel C, Gomes SA: High proportion of subgroup A' (genotype A) among Brazilian isolates of Hepatitis B virus. Archives of virology 2004, 149:1383-1395.
- Sitnik R, Pinho JR, Bertolini DA, Bernardini AP, Da Silva LC, Carrilho FJ: Hepatitis B virus genotypes and precore and core mutants in Brazilian patients. *Journal of clinical microbiology* 2004, 42:2455-2460.
- Viana S, Parana R, Moreira RC, Compri AP, Macedo V: High prevalence of hepatitis B virus and hepatitis D virus in the western Brazilian Amazon. The American journal of tropical medicine and hygiene 2005, 73:808-814.

- Santos AO, Alvarado-Mora MV, Botelho L, Vieira DS, Pinho JR, Carrilho FJ, Honda ER, Salcedo JM: Characterization of hepatitis B virus (HBV) genotypes in patients from Rondonia, Brazil. Virology journal 2010, 7:315.
- Pena SD, Di Pietro G, Fuchshuber-Moraes M, Genro JP, Hutz MH, Kehdy Fde S, Kohlrausch F, Magno LA, Montenegro RC, Moraes MO, et al: The genomic ancestry of individuals from different geographical regions of Brazil is more uniform than expected. PLoS One 2011, 6:e17063.
- Rapoport Delegation on Afro-Brazilian Land Rights: Between the Law and Their Land. Afro-Brazilian Quilombo Communities' Struggle for land Rights. 2008, 2-57.
- Chomczynski P, Sacchi N: Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Analytical* biochemistry 1987, 162:156-159.
- Kwok S, Higuchi R: Avoiding false positives with PCR. Nature 1989, 339:237-238.
- Sanger F, Nicklen S, Coulson AR: DNA sequencing with chain-terminating inhibitors. Proceedings of the National Academy of Sciences of the United States of America 1977, 74:5463-5467.
- 21. Ewing B, Hillier L, Wendl MC, Green P: Base-calling of automated sequencer traces using phred. I. Accuracy assessment. *Genome research* 1998, 8:175-185.
- 22. Edgar RC: MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC bioinformatics* 2004, **5**:113.
- 23. Drummond AJ, Rambaut A: BEAST: Bayesian evolutionary analysis by sampling trees. *BMC evolutionary biology* 2007, 7:214.
- 24. Bowyer SM, van Staden L, Kew MC, Sim JG: A unique segment of the hepatitis B virus group A genotype identified in isolates from South Africa. *The Journal of general virology* 1997, **78(Pt 7)**:1719-1729.
- 25. Kramvis A, Kew MC: Molecular characterization of subgenotype A1 (subgroup Aa) of hepatitis B virus. *Hepatology research: the official journal* of the Japan Society of Hepatology 2007, **37**:S27-32.
- Kimbi GC, Kramvis A, Kew MC: Distinctive sequence characteristics of subgenotype A1 isolates of hepatitis B virus from South Africa. The Journal of general virology 2004, 85:1211-1220.
- Mello FC, Souto FJ, Nabuco LC, Villela-Nogueira CA, Coelho HS, Franz HC, Saraiva JC, Virgolino HA, Motta-Castro AR, Melo MM, *et al*: Hepatitis B virus genotypes circulating in Brazil: molecular characterization of genotype F isolates. *BMC microbiology* 2007, 7:103.
- Motta-Castro AR, Martins RM, Araujo NM, Niel C, Facholi GB, Lago BV, Mello FC, Gomes SA: Molecular epidemiology of hepatitis B virus in an isolated Afro-Brazilian community. *Archives of virology* 2008, 153:2197-2205.
- Quintero A, Martinez D, Alarcon De Noya B, Costagliola A, Urbina L, Gonzalez N, Liprandi F, Castro De Guerra D, Pujol FH: Molecular epidemiology of hepatitis B virus in Afro-Venezuelan populations. *Archives of virology* 2002, 147:1829-1836.
- 30. Alencastro LF: O trato dos viventes: formação do Brasil no Atlântico Sul. São Paulo: Companhia das Letras 2000, 9-523.
- Ribeiro NR, Campos GS, Angelo AL, Braga EL, Santana N, Gomes MM, Pinho JR, De Carvalho WA, Lyra LG, Lyra AC: Distribution of hepatitis B virus genotypes among patients with chronic infection. *Liver* international: official journal of the International Association for the Study of the Liver 2006, 26:636-642.
- 32. Ribeiro D: O Povo Brasileiro: a formação e o sentido do Brasil. *Companhia das Letras* São Paulo; 1995.
- Kimbi GC, Kramvis A, Kew MC: Distinctive sequence characteristics of subgenotype A1 isolates of hepatitis B virus from South Africa. J Gen Virol 2004, 85:1211-1220.
- Matos MA, Reis NR, Kozlowski AG, Teles SA, Motta-Castro AR, Mello FC, Gomes SA, Martins RM: Epidemiological study of hepatitis A, B and C in the largest Afro-Brazilian isolated community. *Transactions of the Royal* Society of Tropical Medicine and Hygiene 2009, 103:899-905.
- Motta-Castro AR, Martins RM, Yoshida CF, Teles SA, Paniago AM, Lima KM, Gomes SA: Hepatitis B virus infection in isolated Afro-Brazilian communities. *Journal of medical virology* 2005, 77:188-193.

doi:10.1186/1743-422X-8-415

Cite this article as: Alvarado-Mora *et al.*: **Detection of Hepatitis B virus subgenotype A1 in a Quilombo community from Maranhão, Brazil.** *Virology Journal* 2011 **8**:415.