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### **INVITED REVIEW**

## Coagulation and wound repair during COVID-19



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#### **KEYWORDS:**

coronavirus; SARS-CoV-2; COVID-19; coagulation; fibrinolysis While COVID-19 is best known as a respiratory infection, SARS-CoV-2 causes systemic disease manifestations including coagulopathies. Both dysregulated extracellular matrix remodeling pathways and circulating coagulation proteins are hallmarks of severe COVID-19 and often continue after the resolution of acute infection. Coagulation proteins have proven effective as biomarkers for severe disease and anticoagulants are a mainstay of COVID-19 therapeutics in hospitalized patients. While much knowledge has been gained about the role of clotting pathway activation in COVID-19, much remains to be elucidated in this complex network of signaling pathways. J Heart Lung Transplant 2021;40:1076–1081

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Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) was first detected in late 2019 in a cluster of pneumonia patients in Wuhan, China.<sup>1</sup> While some patients have had asymptomatic infections, most present with a range of symptoms from mild to lethal disease.<sup>2</sup> COVID-19, the disease caused by SARS-CoV-2 infection, most commonly causes respiratory symptoms with patients experiencing fever, shortness of breath, hypoxia, cough and in severe cases respiratory failure, coagulopathies and multiple organ failure leading to death.<sup>3-6</sup> While the overall case fatality rate of COVID-19 is relatively low ( $\sim 1\%$ ), the extraordinarily rapid spread of this new disease has resulted in overwhelmed medical facilities and exhausted medical providers.<sup>7</sup> Of increasingly recognized importance, COVID-19 patients do not all return to their previous baseline health status with 'long COVID' patients continuing to experience muscle weakness, shortness of breath, mental fogginess, and other symptoms 9+ months after their initial infection.<sup>8</sup> While novel SARS-CoV-2 vaccines and other therapeutics are expected to end the pandemic, the impact of COVID-19 will extend beyond this primary period of infection.

### Infection

SARS-CoV-2 utilizes the ACE2 receptor found on human epithelial cells in the airways as well as type II pneumocytes in the lung.<sup>9</sup> SARS-CoV-2 also has a broad affinity for ACE2 of other animal species and has been linked to progenitor CoV strains found in bats.<sup>10</sup> Although virus tropism is predominantly limited to ACE2 expressing respiratory epithelial cells, there has been evidence of virus replication outside of the respiratory tract including shedding of viral RNA in the feces, and occasional positive virus detection in the brain, heart, kidney and other organs.<sup>11-13</sup> Both in vitro and ex vivo infection of non-epithelial cells has been demonstrated, although most in vivo data has been for viral RNA and true in vivo replication data has been limited to date. While infection of other tissues has been reported, airway and lung ciliated epithelial cells represent the primary site of SARS-CoV-2 replication (Figure 1A). SARS-CoV-2, like SARS-CoV and influenza, preferentially infects type II pneumocytes versus type I C While type I pneumocytes make up most of the alveolar

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surface and are responsible for gas exchange, type II cells have a thick cuboidal shape and produce pulmonary surfactants necessary for lubricating the lung, thus reducing surface tension to allow for respiration.<sup>14</sup> Importantly, surfactant expression dramatically decreases following SARS-CoV-2 infection.<sup>15,16</sup> In addition, type II cells are the progenitor cells of the alveoli and differentiate into type I cells. Therefore, the loss of type II cells due to SARS-CoV-2 infection has a lasting impact and leaves the lung without a direct means to restore the alveoli. While type I pneumocytes can revert to type II cells, in vitro experimental systems suggest that the process can take weeks to occur.<sup>17</sup> This fact may contribute to the long recovery time required for COVID-19 patients.

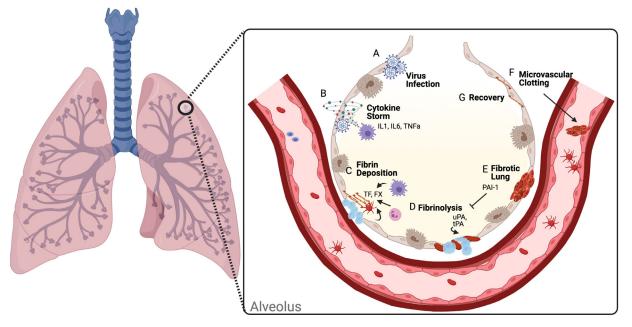
### Cytokine storm and inflammation

Following SARS-CoV-2 infection, the host immune response produces a robust and large cytokine storm characterized by inflammatory mediators (Figure 1B). Responding to damage at the infection site, local alveolar macrophages and infiltrating neutrophils produce a cascade of inflammatory cytokines including IL-1, IL-6, and TNFa.<sup>17</sup> Type I interferons, drivers of the classical antiviral response, are produced at lower levels and later timepoints than observed in influenza infection.<sup>18,19</sup> Interferon autoantibodies have also been detected in patients with severe COVID-19, but not those with milder disease.<sup>20</sup> Mutations in interferon signaling pathways appear to be enriched in severe COVID-19.<sup>21</sup> High levels of C-reactive protein,

complement activation, and lactate dehydrogenase all predict severe disease and contribute to tissue damage.<sup>22</sup> However, absent a strong and productive type I interferon response, these cytokines and inflammatory mediators have limited impact on SARS-CoV-2 replication and cause diffuse alveolar damage (DAD).<sup>23-25</sup> While many coronavirus proteins have interferon antagonist abilities,<sup>26</sup> the CoV E protein has been showed to exacerbate the inflammatory cascade via NFkB<sup>27</sup> in mouse models. Notably, other host conditions including age, obesity, and diabetes have been associated with increased inflammation and subsequent disease.<sup>28</sup> Lacking antiviral effect, the inflammatory cascade plays a detrimental role during COVID-19 infection, exacerbating lung damage and disease. Several COVID9 treatment approaches have focused on disrupting this inflammatory cascade including drugs like dexamethasone, IL-1 and IL-6 antagonists with modest effect.<sup>29</sup>

### Wound healing

The lung epithelium is part of the larger epithelial barrier that protects our tissues from the external environment, and it can be damaged chemical injury or infectious agents such as SARS-CoV-2. Wound healing or tissue repair is the complex processes by which injured cells are identified, removed, and eventually replaced with new, healthy tissue. While healthy lung cells have little cell turnover, the lung epithelium is capable of impressive regeneration after injury.<sup>30</sup> Following injury, epithelial cells can de-differentiate to replicate and replace specialized cells such as type I



**Figure 1** Activation and modulation of coagulation pathways following SARS-CoV-2 infection. Schematic of the lung and an alveolus following SARS-CoV-2 infection. A) Virus infection of type I and type II pneumocytes. B) Cytokine storm including IL1, IL6, and TNFa induced in response to viral replication. C) Inflammation induced damage activates release of FX, tissue factor (TF) and other coagulation factors to activate fibrin deposition to limit fluid accumulation in alveolar spaces. D) Release of uPA and tPA initiate breakdown of fibrinous structures. E) PAI-1 blocks activity of uPA and tPA and leaves fibrinous structures intact and the lung more rigid. F) Release of pro-clotting factors into the circulatory systems that may impact other target organs. G) Return to baseline lung function with evidence of scarring and damage still in place. Figure generated using Biorender software.

and type II pneumocytes and secretory cells. Recent COVID-19 autopsy reports have noted a high percentage of proliferating AT2 cells,<sup>31</sup> indicating that epithelial cell regeneration occurs after SARS-CoV-2 induced lung injury. Crosstalk between epithelial cells and immune cells is critical for this process with activated neutrophils and macrophages producing TGF-b and other chemokines that promote epithelial cell migration to the site of tissue injury. Immune cells also clear apoptotic debris from the site of injury and release growth factors like EGF, VEGF and IGF that are important for the tissue regeneration process. Little human data on growth factor expression in COVID-19 patients is available<sup>32,33</sup>; however, previous work using a mouse model of SARS-CoV infection demonstrated that overactive EGFR signaling leads to enhanced lung disease pulmonary fibrosis,<sup>34</sup> indicating that an appropriate wound healing response is important for resolution of coronavirusinduced lung disease.

# Coagulation and extracellular matrix remodeling pathways

In response to infection and inflammation-induced damage, the host must take actions to maintain respiratory function through activation of extracellular matrix remodeling and coagulation pathways.<sup>35</sup> Loss of pneumocytes and diffuse alveolar damage caused by inflammation raises the risk of vascular leakage, fluid accumulation (edema), and hemorrhage in the alveolar spaces, thus preventing oxygen exchange<sup>36</sup> (Figure 1C). This tissue damage leads to cytokine production, stimulating increased expression of tissue factor on endothelial cells and exposure of TF to activate the coagulation pathway, leading to the cleavage of prothrombin to thrombin.<sup>37,38</sup> Interestingly, while tissue factor protein levels are increased in COVID-19 patients, the transcripts are not elevated,<sup>39,40</sup> indicating that regulation of SARS-CoV-2 induced thrombotic events is complex. Thrombin subsequently cleaves fibringen into fibrin,<sup>41</sup> a major component of clots. Fibrin is incorporated with collagen into hyaline membranes to seal the alveoli from fluid accumulation.<sup>42</sup> However, these sealing processes thicken the alveolar walls, limit oxygen exchange, and may lead to pulmonary fibrosis, endangering respiratory function.<sup>43</sup>

Additionally, epithelial damage, production of profibrotic cytokines, and chemokines such as TGFb and MCP-1 stimulate collagen and fibronectin production leading to a pro-fibrotic state.<sup>44,45</sup> This fibrotic lung stage also stimulates the fibrinolytic pathway, a process that breaks down fibrinous deposits through release of uPa and tPA<sup>46</sup> (Figure 1D). UPa and tPA activate plasminogen into plasmin which targets fibrin for breakdown.<sup>47</sup> The activity of tPA and uPa is regulated by plasminogen activator inhibitor-1 (PAI-1),<sup>46</sup> and alpha 2-antiplasmin. Plasmin itself is regulated by several serine protease inhibitors including a2antiplasmin and there are extensive interactions between the complement and coagulation proteolytic pathways<sup>35,48</sup> (Figure 1E). Together, the fibrinolytic and coagulation pathways govern a delicate balance between hemorrhage/edema and fibrosis in order to maintain lung respiratory function. Once this axis is disrupted by coronavirus-induced acute respiratory distress (ARDS), patients are at high risk of respiratory failure from either pulmonary fibrosis or edema and DAD.

During SARS-CoV-2 infection, the balance of the coagulation signaling, including the fibrinolytic pathway can be disrupted in either direction leading to adverse outcomes. COVID-19 patients have been reported to have high levels of PAI-1 and D-dimers in their blood, 49,50 consistent with the microthrombi observed in COVID-19 patient autopsies. Confoundingly, intra-alveolar hemorrhage has also been observed in COVID-19 lungs and elevated levels of pro-fibrinolytic uPA and tPA have been associated with reduced respiratory function and more severe disease.<sup>37</sup>) Excessive levels of uPA and tPA can lead to breakdown of fibrin before damaged areas have been sufficiently repaired (Figure 1D). The results can be fluid accumulation in the alveolar spaces that disrupts oxygen exchange. However, during SARS-CoV-2 infection, coagulation has more often been identified as a persistent issue and a major factor contributing to mortality.<sup>3</sup> An increase in PAI-1 prevents the breakdown of fibrin by uPa and tPA, leaving a thickening of the alveolar walls that reduces respiratory function and makes it more difficult to breath<sup>49</sup> (Figure 1E). The pro-coagulation cascade also has an impact beyond the lung with the formation of microvascular clots in other organs and in the circulatory system<sup>43</sup> (Figure 1F). In addition, PAI-1 levels are increased in patients who are elderly or have hypertension, obesity, diabetes, and cardiovascular disease, consistent with increased susceptibility to COVID in these populations.<sup>51</sup> Even after resolution of infection, the lung can maintain fibrin and other scarring from the induced damage<sup>52</sup> (Figure 1G). Overall, both sides of the fibrinolytic/ coagulation pathway are critical to the SARS-CoV-2 response.

While experimental data is limited for SARS-CoV-2, disruption of uPa signaling had significant impact on susceptibility to the original SARS-CoV in vivo.53 Mice deficient in PAI-1 had increased weight loss and mortality following challenge with SARS-CoV.<sup>54</sup> The absence of PAI-1 resulted in an increase in ARDS related gene signatures and extensive hemorrhage in the lung. Notably, the loss of PAI-1 had no significant impact on viral load. Conversely, mice deficient in tPA (PLAT) were also more susceptible to lethal SARS-CoV challenge.<sup>54</sup> tPA<sup>-/-</sup> mice had increased mortality compared to control animals following SARS-CoV challenge. While the tPA KO mice trended to less overall hemorrhage, the presence of exudates and increased lethality indicate the delicate balance required to recover from infection. It is anticipated that the fibrinolytic signaling pathway governs similar processes following SARS-CoV-2 infection.

# Monitoring and targeting COVID induced coagulation

Given the link to severe disease and mortality,<sup>55</sup> activated pathways associated with fibrinolysis and coagulation have

been used as a biomarkers for determining COVID-19 intervention strategies. Retrospective studies have determined that lethal SARS-CoV-2 cases had higher D-dimer and fibrin degradation products in their blood.<sup>55,56</sup> These patients also had longer prothrombin time and met criteria consistent with disseminated intravascular coagulopathy.<sup>37</sup> Similarly, low platelet counts and prolonged activated partial thromboplastin time were associated with more severe disease.<sup>57</sup>

Finally, lupus anticoagulant antibodies have also been identified in a subset of patients.<sup>58</sup> Together, the results suggest that monitoring coagulation metrics can predict disease severity and dictate intervention strategies.<sup>29</sup> Similarly, improvement in these coagulation metrics may signal appropriate waning of aggressive treatment approaches.

Prophylactic targeting of the coagulation pathways is now the routine treatment approach for hospitalized COVID-19 patients.<sup>59</sup> While not employed early during the outbreak, the combination of excess thrombin production, fibrinolysis shutdown, and evidence of micro thrombotic occlusions demonstrated the need to control coagulation pathways.<sup>60</sup> The standard treatment utilizes low-molecular weight heparin (LMWH) which inhibits heparinase activity, neutralizes cytokine storm, and interferes with leukocyte trafficking.<sup>61,62</sup> An alternative approach utilized inhalation of plasminogen to improve lung lesion and hypoxemia.63 To counteract fibrin accumulation, tPA treatment and drugs that target PAI-1 have been attempted to improve outcomes.<sup>49, 63</sup> Together, these approaches to disrupt an exuberant coagulation response has produced improved outcome in hospitalized patients.

Despite being the standard of care, targeting the coagulation pathways offers a mitigation rather than preventive response. The disruption of the coagulation factors does not resolve the underlying inflammation and lung damage that initiated the response. Instead, treatments that disrupt the damage cascade may have the most significant impact on coagulation pathways activation. For example, antiviral drugs like remdesivir and EIDD-2801 and monoclonal antibodies target viral replication with best effect at early times points post infection<sup>64, 65</sup>; remdesivir has also been shown to reduce inflammatory responses and may reduce overall disease by diminishing damage.<sup>66</sup> Similarly, treatments that disrupt the inflammatory cascade may also change downstream coagulation activation. Anakinra and tocilizumab, drugs that target IL1 and IL6 respectively, have been utilized to treat COVID-19 patients.<sup>67</sup> These inflammation pathways have also been shown to activate coagulation pathways and treatments that target these inflammation cascades may reduce damage stimulating coagulation responses. Broad immune suppression drugs like dexamethosome may also produce the same results.<sup>29</sup> Further research into coagulation triggers and cascade activation in COVID-19 patients will help highlight areas for targeted therapeutics. Overall, efforts to prevent inflammation related damage

### Conclusion

As SARS-CoV-2 has spread around the world, the connections between viral infection, inflammation, and the coagulation cascade have been further illuminated. While the emergent SARS-CoV-2 causes significant disease and death, damage from viral replication appears secondary to exuberant host responses. In an effort to maintain respiratory function, the delicate balance between hemorrhage and fibrosis in the lung is at the nexus of COVID-19 disease. Focusing on the coagulation and fibrinolytic pathways provides a means to evaluate the severity of disease in patients and potentially mitigate its damage with therapeutic treatments. However, preventing the inflammatory cascade that initiates and necessitates the coagulation response may be the only means prevent severe COVID-19 disease. Importantly, these observations of the coagulation pathways may have implications for other infections like influenza or Ebola. Overall, we need a better understanding of the coagulation host response to effectively treat and overcome SARS-CoV-2 and future emergent pathogens.

### Disclosure statement and acknowledgments

The authors have no conflicts of interest to declare.

Research was supported by grants from NIA and NIAID of the NIH to (AI153602 and AG049042 to VDM; AI145372 to LEG). Research was also supported by STARs Award provided by the University of Texas System to VDM.

#### References

- 1. Gralinski LE, Menachery VD. Return of the Coronavirus: 2019-nCoV. Viruses 2020;12.
- Gandhi RT, Lynch JB, Del Rio C. Mild or moderate Covid-19. N Engl J Med 2020;383:1757-66.
- **3.** Huang C, Huang L, Wang Y, et al. 6-month consequences of COVID-19 in patients discharged from hospital: a cohort study. Lancet 2021;397:220-32.
- Chandra A, Chakraborty U, Pal J, Karmakar P. Silent hypoxia: a frequently overlooked clinical entity in patients with COVID-19. BMJ Case Rep 2020;13:e237207. https://doi.org/10.1136/bcr-2020-237207 3.
- Goh KJ, Choong MC, Cheong EH, et al. Rapid progression to acute respiratory distress syndrome: review of current understanding of critical illness from COVID-19 infection. Ann Acad Med Singap 2020;49:108-18.
- Wang F, Qu M, Zhou X, et al. The timeline and risk factors of clinical progression of COVID-19 in Shenzhen. China. J Transl Med 2020;18:270.
- 7. Petersen E, Koopmans M, Go U, et al. Comparing SARS-CoV-2 with SARS-CoV and influenza pandemics. Lancet Infect Dis 2020;20: e238-44.
- Oronsky B, Larson C, Hammond TC, et al. A review of persistent post-COVID syndrome (PPCS). Clin Rev Allergy Immunol 2021. https://doi.org/10.1007/s12016-021-08848-3:1-9.
- Zhou P, Yang XL, Wang XG, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 2020;579:270-3.

- Hu B, Guo H, Zhou P, Shi ZL. Characteristics of SARS-CoV-2 and COVID-19. Nat Rev Microbiol 2021;19:141-54.
- 11. Bradley BT, Maioli H, Johnston R, et al. Histopathology and ultrastructural findings of fatal COVID-19 infections in Washington State: a case series. Lancet 2020;396:320-32.
- Lindner D, Fitzek A, Brauninger H, et al. Association of cardiac infection with SARS-CoV-2 in confirmed COVID-19 autopsy cases. JAMA Cardiol 2020;5:1281-5.
- 13. Bhatnagar J, Gary J, Reagan-Steiner S, et al. Evidence of severe acute respiratory syndrome coronavirus 2 replication and tropism in the lungs, airways, and vascular endothelium of patients with fatal coronavirus disease 2019: an autopsy case series. J Infect Dis 2021;223:752-64.
- Olajuyin AM, Zhang X, Ji HL. Alveolar type 2 progenitor cells for lung injury repair. Cell Death Discov 2019;5:63.
- 15. Islam A, Khan MA. Lung transcriptome of a COVID-19 patient and systems biology predictions suggest impaired surfactant production which may be druggable by surfactant therapy. Sci Rep 2020;10:19395.
- 16. Katsura H, Sontake V, Tata A, et al. Human lung stem cell-based alveolospheres provide insights into SARS-CoV-2-mediated interferon responses and pneumocyte dysfunction. Cell Stem Cell 2020;27:890-904. e8.
- Yang J, Hernandez BJ, Martinez Alanis D, et al. The development and plasticity of alveolar type 1 cells. Development 2016;143:54-65.
- Galani IE, Rovina N, Lampropoulou V, et al. Untuned antiviral immunity in COVID-19 revealed by temporal type I/III interferon patterns and flu comparison. Nat Immunol 2021;22:32-40.
- Ombrello MJ, Schulert GS. COVID-19 and cytokine storm syndrome: are there lessons from macrophage activation syndrome? Transl Res 2021. https://doi.org/10.1016/j.trsl.2021.03.002.
- Bastard P, Rosen LB, Zhang Q, et al. Autoantibodies against type I IFNs in patients with life-threatening COVID-19. Science 2020;370 (6515):eabd4585.
- 21. Zhang Q, Bastard P, Liu Z, et al. Inborn errors of type I IFN immunity in patients with life-threatening COVID-19. Science 2020;370(6515): eabd4570.
- Li L, Chen C. Contribution of acute-phase reaction proteins to the diagnosis and treatment of 2019 novel coronavirus disease (COVID-19). Epidemiol Infect 2020;148:e164.
- Torres Acosta MA, Singer BD. Pathogenesis of COVID-19-induced ARDS: implications for an ageing population. Eur Respir J 2020;56:2002049.
- Batah SS, Fabro AT. Pulmonary pathology of ARDS in COVID-19: a pathological review for clinicians. Respir Med 2020;176:106239.
- **25.** Borczuk AC, Salvatore SP, Seshan SV, et al. COVID-19 pulmonary pathology: a multi-institutional autopsy cohort from Italy and New York City. Mod Pathol 2020;33:2156-68.
- 26. Lei X, Dong X, Ma R, et al. Activation and evasion of type I interferon responses by SARS-CoV-2. Nat Commun 2020;11:3810.
- DeDiego ML, Nieto-Torres JL, Regla-Nava JA, et al. Inhibition of NF-kappaB-mediated inflammation in severe acute respiratory syndrome coronavirus-infected mice increases survival. J Virol 2014;88:913-24.
- Rashedi J, Mahdavi Poor B, Asgharzadeh V, et al. Risk factors for COVID-19. Infez Med 2020;28:469-74.
- van Eijk LE, Binkhorst M, Bourgonje AR, et al. COVID-19: immunopathology, pathophysiological mechanisms, and treatment options. J Pathol 2021. https://doi.org/10.1002/path.5642.
- Ruaro B, Salton F, Braga L, et al. The history and mystery of alveolar epithelial type II cells: focus on their physiologic and pathologic role in lung. Int J Mol Sci 2021;22:2566.
- Chen J, Wu H, Yu Y, Tang N. Pulmonary alveolar regeneration in adult COVID-19 patients. Cell Res 2020;30:708-10.
- 32. Xu J, Xu X, Jiang L, Dua K, Hansbro PM, Liu G. SARS-CoV-2 induces transcriptional signatures in human lung epithelial cells that promote lung fibrosis. Respir Res 2020;21:182.
- 33. Winn BJ. Is there a role for insulin-like growth factor inhibition in the treatment of COVID-19-related adult respiratory distress syndrome? Med Hypotheses 2020;144:110167.

- 34. Venkataraman T, Coleman CM, Frieman MB. Overactive epidermal growth factor receptor signaling leads to increased fibrosis after severe acute respiratory syndrome coronavirus infection. J Virol 2017;91: e00182-17.
- Kwaan HC, Lindholm PF. The central role of fibrinolytic response in COVID-19-A hematologist's perspective. Int J Mol Sci 2021;22:1283.
- Venkataraman T, Frieman MB. The role of epidermal growth factor receptor (EGFR) signaling in SARS coronavirus-induced pulmonary fibrosis. Antiviral Res 2017;143:142-50.
- Salabei JK, Fishman TJ, Asnake ZT, Ali A, Iyer UG. COVID-19 coagulopathy: current knowledge and guidelines on anticoagulation. Heart Lung 2021;50:357-60.
- Mahmood N, Mihalcioiu C, Rabbani SA. Multifaceted role of the urokinase-type plasminogen activator (uPA) and its receptor (uPAR): diagnostic, prognostic, and therapeutic applications. Front Oncol 2018;8:24.
- Mast AE, Wolberg AS, Gailani D, et al. SARS-CoV-2 suppresses anticoagulant and fibrinolytic gene expression in the lung. Elife 2021;10: e64330.
- 40. Sandeep Subrahmanian P, Alain Borczuk M, Steven P. Salvatore M, Jeffrey Laurence M, Jasimuddin Ahamed P. 2020. Higher tissue factor (TF) expression in the lungs of COVID-19 pneumonia patients than patients with acute respiratory distress syndrome: association with thrombi formation blood 136.
- **41.** Batah SS, Fabro AT. Pulmonary pathology of ARDS in COVID-19: a pathological review for clinicians. Respir Med 2021;176:106239.
- Mason RJ. Pathogenesis of COVID-19 from a cell biology perspective. Eur Respir J 2020;55:2000607.
- Iba T, Levy JH, Levi M, Thachil J. Coagulopathy in COVID-19. J Thromb Haemost 2020;18:2103-9.
- 44. Crosby LM, Waters CM. Epithelial repair mechanisms in the lung. Am J Physiol Lung Cell Mol Physiol 2010;298:L715-31.
- Delpino MV, Quarleri J. SARS-CoV-2 pathogenesis: imbalance in the renin-angiotensin system favors lung fibrosis. Front Cell Infect Microbiol 2020;10:340.
- 46. Gharaee-Kermani M, Hu B, Phan SH, Gyetko MR. The role of urokinase in idiopathic pulmonary fibrosis and implication for therapy. Expert Opin Investig Drugs 2008;17:905-16.
- Idell S. Coagulation, fibrinolysis, and fibrin deposition in acute lung injury. Crit Care Med 2003;31:S213-20.
- **48.** Fumagalli S, De Simoni MG. Lectin complement pathway and its bloody interactions in brain ischemia. Stroke 2016;47:3067-73.
- 49. Zuo Y, Warnock M, Harbaugh A, et al. Plasma tissue plasminogen activator and plasminogen activator inhibitor-1 in hospitalized COVID-19 patients. Sci Rep 2021;11:1580.
- Mackman N, Antoniak S, Wolberg AS, Kasthuri R, Key NS. Coagulation abnormalities and thrombosis in patients infected with SARS-CoV-2 and other pandemic viruses. Arterioscler Thromb Vasc Biol 2020;40:2033-44.
- 51. Yamamoto K, Takeshita K, Kojima T, Takamatsu J, Saito H. Aging and plasminogen activator inhibitor-1 (PAI-1) regulation: implication in the pathogenesis of thrombotic disorders in the elderly. Cardiovasc Res 2005;66:276-85.
- Bharat A, Querrey M, Markov NS, et al. Lung transplantation for patients with severe COVID-19. Sci Transl Med 2020;12:eabe4282.
- Gralinski LE, Baric RS. Molecular pathology of emerging coronavirus infections. J Pathol 2015;235:185-95.
- Gralinski LE, Bankhead A, Jeng S, et al. Mechanisms of severe acute respiratory syndrome coronavirus-induced acute lung injury. Mbio 2013;4:e00271-13.
- 55. Rentsch CT, Beckman JA, Tomlinson L, et al. Early initiation of prophylactic anticoagulation for prevention of coronavirus disease 2019 mortality in patients admitted to hospital in the United States: cohort study. Bmj 2021;372:n311.
- 56. Mori H, Ohkawara H, Togawa R, Rikimaru M, Shibata Y, Ikezoe T. Diagnosis and treatment of disseminated intravascular coagulation in COVID-19 patients: a scoping review. Int J Hematol 2021;113:320-9.
- Lin J, Yan H, Chen H, et al. COVID-19 and coagulation dysfunction in adults: a systematic review and meta-analysis. J Med Virol 2021;93:934-44.

- Gazzaruso C, Mariani G, Ravetto C, et al. Lupus anticoagulant and mortality in patients hospitalized for COVID-19. J Thromb Thrombolysis 2020. https://doi.org/10.1007/s11239-020-02335-w:1-7.
- 59. Ortega-Paz L, Capodanno D, Montalescot G, Angiolillo DJ. Coronavirus disease 2019-associated thrombosis and coagulopathy: review of the pathophysiological characteristics and implications for antithrombotic management. J Am Heart Assoc 2021;10:e019650.
- Hadid T, Kafri Z, Al-Katib A. Coagulation and anticoagulation in COVID-19. Blood Rev 2020. https://doi.org/10.1016/j. blre.2020.100761:100761.
- Buijsers B, Yanginlar C, Maciej-Hulme ML, de Mast Q, van der Vlag J. Beneficial non-anticoagulant mechanisms underlying heparin treatment of COVID-19 patients. EBioMedicine 2020;59:102969.
- Arslan Y, Yilmaz G, Dogan D, et al. The effectiveness of early anticoagulant treatment in Covid-19 patients. Phlebology 2020. https://doi. org/10.1177/0268355520975595:268355520975595.

- **63.** Wu Y, Wang T, Guo C, et al. Plasminogen improves lung lesions and hypoxemia in patients with COVID-19. Qjm 2020;113:539-45.
- 64. Sheahan TP, Sims AC, Graham RL, et al. Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses. Sci Transl Med 2017;9:eaal3653.
- **65.** Sheahan TP, Sims AC, Zhou S, et al. An orally bioavailable broadspectrum antiviral inhibits SARS-CoV-2 in human airway epithelial cell cultures and multiple coronaviruses in mice. Sci Transl Med 2020;12:eabb5883.
- **66.** Fiege JK, Thiede JM, Nanda HA, et al. Single cell resolution of SARS-CoV-2 tropism, antiviral responses, and susceptibility to therapies in primary human airway epithelium. PLoS Pathog 2021;17: e1009292.
- Magro G. COVID-19: Review on latest available drugs and therapies against SARS-CoV-2. Coagulation and inflammation cross-talking. Virus Res 2020;286:198070.