

Survivorship and feeding preferences among size classes of outplanted sea urchins, *Tripneustes gratilla*, and possible use as biocontrol for invasive alien algae

Charley E. Westbrook¹, Rory R. Ringang¹, Sean Michael A. Cantero¹, HDAR & TNC Urchin Team and Robert J. Toonen¹

ABSTRACT

We investigate the survivorship, growth and diet preferences of hatchery-raised juvenile urchins, *Tripneustes gratilla*, to evaluate the efficacy of their use as biocontrol agents in the efforts to reduce alien invasive algae. In flow-through tanks, we measured urchin growth rates, feeding rates and feeding preferences among diets of the most common invasive algae found in Kāne'ohe Bay, Hawai'i: Acanthophora spicifera, Gracilaria salicornia, Eucheuma denticulatum and Kappaphycus clade B. Post-transport survivorship of outplanted urchins was measured in paired open and closed cages in three different reef environments (lagoon, reef flat and reef slope) for a month. Survivorship in closed cages was highest on the reef flat (\sim 75%), and intermediate in the lagoon and reef slope (\sim 50%). In contrast, open cages showed similar survivorship on the reef flat and in the lagoon, but only 20% of juvenile urchins survived in open cages placed on the reef slope. Urchins grew significantly faster on diets of G. salicornia (1.58 mm/week \pm 0.14 SE) and Kappaphycus clade B (1.69 \pm 0.14 mm/wk) than on E. denticulatum (0.97 \pm 0.14 mm/wk), with intermediate growth when fed on A. spicifera (1.23 \pm 0.11 mm/wk). Interestingly, urchins display size-specific feeding preferences. In non-choice feeding trials, small urchins (17.5-22.5 mm test diameter) consumed G. salicornia fastest $(6.08 \text{ g/day} \pm 0.19 \text{ SE})$, with A. spicifera $(4.25 \pm 0.02 \text{ g/day})$ and Kappaphycus clade B $(3.83 \pm 0.02 \text{ g/day})$ intermediate, and *E. denticulatum* was clearly the least consumed $(2.32 \pm 0.37 \text{ g/day})$. Medium-sized (29.8-43.8 mm) urchins likewise preferentially consumed G. salicornia (12.60 \pm 0.08 g/day), with less clear differences among the other species in which *E. denticulatum* was still consumed least (9.35 \pm 0.90 g/day). In contrast, large urchins (45.0–65.0 mm) showed no significant preferences among the different algae species at all (12.43–15.24 g/day). Overall consumption rates in non-choice trials were roughly equal to those in the choice trials, but differences among feeding rates on each species were not predictive of feeding preferences when urchins were presented all four species simultaneously. In the choice feeding trials, both small and medium urchins clearly preferred A. spicifera over all other algae (roughly twice as much consumed as any other species). Again, however, differences were less pronounced among adult urchins, with adults showing a significant preference for A. spicifera and Kappaphycus clade B compared to the other two algal

Submitted 7 May 2015 Accepted 18 August 2015 Published 15 September 2015

Corresponding author Charley E. Westbrook, charleye@hawaii.edu

Academic editor María Ángeles Esteban

Additional Information and Declarations can be found on page 16

DOI 10.7717/peerj.1235

© Copyright 2015 Westbrook et al.

Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

¹ Hawai'i Institute of Marine Biology, University of Hawai'i at Mānoa, USA

species. These findings indicate that outplanted urchins are surviving on the reef flats and eating a variety of alien invasive algae as intended. Although juvenile urchins show stronger feeding preferences, these animals grow quickly, and adult urchins are more generalist herbivores that consume all four alien invasive algae.

Subjects Animal Behavior, Aquaculture, Fisheries and Fish Science, Ecology, Ecosystem Science, Marine Biology

Keywords Hawai'i, Kāne'ohe Bay, Acanthophora spicifera, Gracilaria salicornia, Eucheuma denticulatum, Kappaphycus clade B, Super sucker, Kappaphycus striatum, Eucheuma striatum

INTRODUCTION

Within the last 70 years, Kāne'ohe Bay has become home to many introduced and invasive algal species, whose aggressive growth has smothered corals and overgrown many patch reefs across the bay (Coles, DeFelice & Eldredge, 2002; Conklin & Smith, 2005; Smith et al., 2004; Stimson, Larned & Conklin, 2001; Russell & Balazs, 2009; Bahr, Jokiel & Toonen, 2015). Some of these alien algae species were introduced intentionally, whereas others have unknown origins (are cryptogenic), but appear to have gained a foothold in Kāne'ohe Bay thanks to a combination of reduced grazing intensity and high nutrient influx as a result of sewage discharge into the bay (Stimson, Larned & Conklin, 2001). Among the most obvious and ecologically dominant of these invasive algal species are Kappaphycus clade B, Eucheuma denticulatum, Gracilaria salicornia, and Acanthophora spicifera. The species level taxonomy of Kappaphycus remains a subject of contention (Conklin, Kurihara & Sherwood, 2009; Sherwood et al., 2010). Due to the uncertain nomenclature of Kappaphycus in the literature, it was suggested we use the most contemporary denomination of the alga (despite the prospect of its name changing again, this is the best we could do at the time), henceforth it should and will be referred to as Kappaphycus clade B in this study (K Conklin & A Sherwood, pers. comm., 2015).

Native species of algae which once dominated the Bay (Stimson, Larned & Conklin, 2001; Smith et al., 2004; Conklin & Smith, 2005; Stimson, Cunha & Philippoff, 2007) have become comparatively rare as the rhodophytes K. clade B and G. salicornia both spread rapidly across Kāne'ohe Bay after their introduction, and are now found at high abundance throughout the Bay (Hunter & Evans, 1995; Smith, Hunter & Smith, 2002; Bahr, Jokiel & Toonen, 2015). Originally these alien species were estimated to spread at a minimum rate of 250 m yr⁻¹ (Rodgers & Cox, 1999), although this is now considered to have been a gross underestimate (Coles, DeFelice & Eldredge, 2002; Smith, Hunter & Smith, 2002). Since its introduction and release in 1974, K. clade B has resulted in phase-shifts across the bay by replacing native algae and corals with newly formed monocultures of this alien alga over large areas of reef flat and slope (Coles, DeFelice & Eldredge, 2002; Smith, Hunter & Smith, 2002). Likewise, alien algal overgrowth is smothering live corals on patch reefs, resulting in a loss of biodiversity, changes in community structure of the reef fishes, and erosion of the physical structure of the reef (Smith, Hunter & Smith, 2002). In response to the spread of

ecological impacts associated with these alien invasive species, the State of Hawaii Division of Aquatic Resources (DAR) and The Nature Conservancy (TNC) have undertaken manual removal efforts using suction-assisted divers supported by the "supersucker" barges. The "supersucker" teams selectively remove invasive algae species from reefs with sorters on the surface looking for any native species accidentally removed from the reef. Materials that are sucked up through the pumps are sorted on the deck of the barge and any non-alien algae species are returned to the reef immediately. The alien algae is bagged and given to organic farmers who use it as natural fertilizer. However, removal of alien algae by these teams is labor-intensive and only effective if the algae do not regrow, which was happening within roughly a year in the initial supersucker trials (DAR & TNC, pers. comm., 2014). Thus, the long-term solution proposed for areas where invasive algae have begun to overgrow and smother native corals has been to increase the population of native herbivores such as grazing urchins (Conklin & Smith, 2005; Stimson, Cunha & Philippoff, 2007; Weis & Butler, 2009).

Biocontrol agents have been the topic of much debate due to infamous cases during which their introduction lead to their subsequent invasion (*Howarth*, 1983; *Simberloff & Stiling*, 1996). Introduced biolcontrol agents, which turned invasive, have wreaked irreversible damage to many host ecosystems (*Howarth*, 1991). Notable instances of failed biocontrol efforts in Hawaii include the introduction of the Rosy Wolf Snail (*Howarth*, 1985; *Holland*, *Taylor & Sugiura*, 2012), as well as the Indian Mongoose (*Simberloff et al.*, 2000; *Godwin*, *Rodgers & Jokiel*, 2006). Indeed, there are precious few examples of successful biocontrol efforts with alien species (*Howarth*, 1983; *Godwin*, *Rodgers & Jokiel*, 2006). Many now argue that if biocontrol agents are to be used at all, they should be native to the ecosystem being targeted (*Howarth*, 1985). In addition to eliminating the likelihood that alien biocontrol agents become pests in a novel environment, it has been documented that native grazers suppress the establishment of exotic plants better than the introduction of exotic grazers (*Parker*, *Burkepile & Hay*, 2006; *Kimbro*, *Cheng & Grosholz*, 2013).

Following on that logic, experiments with native sea urchins have demonstrated that *T. gratilla* have reduced the biomass of the invasive *Kappaphycus* spp. within enclosure areas on the reef where alien algae were abundant (*Conklin & Smith*, 2005). Urchins are an important part of the macro-grazing fauna on many tropical reefs, including those in Hawaiii (*Chiappone et al.*, 2002; *Alves et al.*, 2003; *Mumby et al.*, 2006; *Stimson, Cunha & Philippoff*, 2007; *Sandin, Walsh & Jackson*, 2010; *Valentine & Edgar*, 2010). Although some urchins show dietary preferences in laboratory studies (e.g., *Stimson, Cunha & Philippoff*, 2007; *Seymour et al.*, 2013), others appear to be generalist herbivores that will graze on just about any algae or sea grass made available (e.g., *Vaïtilington, Rasolofonirina & Jangoux*, 2003; *Dworjanyn, Pirozzi & Wenshan*, 2007). Other potential biocontrol agents, such as fish (acanthurids and scarids) exhibit a relatively low degree of preference for the invasive algae, are far more motile, and are highly prized by local fishermen, making it difficult to rely on herbivorous fishes as a potential mechanism of biocontrol (*Conklin & Smith*, 2005). Urchins therefore make an obvious choice for a variety of algal biocontrol efforts because of their generalist feeding behavior and limited vagility as adults, coupled with

the high dispersal potential and the ubiquity of habitat that has allowed a number of tropical urchins to successfully colonize reefs across the globe (*Lessios, Kane & Robertson*, 2003; *Seymour et al.*, 2013). Likewise, *T. gratilla* was historically abundant in Kāne'ohe Bay (*Ogden, Ogden & Abbott, 1989; Thomas, 1994*), but for unknown reasons has become rare since the 1990s (*Stimson, Cunha & Philippoff, 2007; Bahr, Jokiel & Toonen, 2015*).

Studies on the ecological impacts of natural outbreaks of *T. gratilla* corroborated the efficiency with which these urchins can significantly reduce the abundance of algae (*Valentine & Edgar*, 2010). Due to their limited movement as adults and their voracious appetite for a wide variety of algae and seagrasses, *T. gratilla* has been recommended as the best species for use as a biological control agent in Kāne'ohe Bay (*Conklin & Smith*, 2005; *Stimson*, *Cunha & Philippoff*, 2007). Since 2010, DAR has been culturing juvenile *T. gratilla* for outplanting as herbivorous biocontrol agents to prevent regrowth of algal biomass once the alien algae have been manually removed from patch reefs in Kāne'ohe Bay (*Gibo*, *Letsom & Westbrook*, 2012). This study set out to investigate if tank bred urchins would eat the targeted alien algae species, and if so, to determine their potential grazing rates. The project also examined if the urchins' growth could be sustained on diets of non-native algae, and to what extent each alga facilitated growth of *T. gratilla*. Potential feeding preferences between the four alien algae were also evaluated. Lastly, urchins were caged in various habitats in order to elucidate post-transplant survival of tank bred juvenile urchins in the bay.

Currently, thousands of cultured urchins are outplanted at 20-25 mm test diameter, but comparatively few were observed in subsequent surveys of urchin density on the reef (J Blodgett, DAR, pers. comm., 2014). A major motivation of this research was that it was unknown at that time whether the missing urchins were dying from transplant stress, starvation, being eaten by predators after outplanting, or simply moving into cryptic habitats at small sizes such that they were missing in subsequent surveys. Stimson, Cunha & Philippoff (2007) conducted feeding preference trials with large T. gratilla (8–9 cm) and showed that feeding preferences were generally unchanged after 5 months on monospecific diets, except urchins that were maintained on Padina sanctae-crucis and showed enhanced preference in subsequent choice trials, whereas those maintained on G. salicornia tended to avoid it when offered five species from which to choose at the end of the trial. Further, Stimson, Cunha & Philippoff (2007) found that urchins offered a variety of algal species consume more per day than when limited to a single-species diet. This study expands on the previous work to elucidate patterns of the post-transport juvenile urchin survival, growth and diet preferences of lab cultured T. gratilla being outplanted in Kāne'ohe Bay. Together, these studies will aid both State and conservation group efforts to control alien algal overgrowth of corals on reefs in Kāne'ohe Bay and across Hawai'i.

MATERIALS AND METHODS

Study animal

The short-spined collector urchin *Tripneustes gratilla* received its common name from the habit of gathering fragments of coral rubble, rocks, or algae from the benthic environment

as camouflage while it forages. Its body is predominantly black but often possesses a pentaradial bluish or reddish hue when its tube feet are retracted close to its body. Its spines are typically black, white or cream. This echinoid is relatively common in shallow waters (0–15 m) across the Hawaiian archipelago (*Kay*, 1994; *Hoover*, 2002). Natural densities of *T. gratilla* range from 2.9–4.4 m⁻², placing it in the top three most abundant urchins in Hawaii (*Ogden*, *Ogden & Abbott*, 1989).

Although *T. gratilla* have shown significant dietary preferences for *Kappaphycus* spp. in controlled laboratory studies (Stimson, Cunha & Philippoff, 2007), in the wild these urchins are a generalist herbivore that will graze on virtually any algae or sea grass available (Vaïtilington, Rasolofonirina & Jangoux, 2003; Dworjanyn, Pirozzi & Wenshan, 2007; Stimson, Cunha & Philippoff, 2007). The generalist diet and habitat requirements of T. gratilla coupled with high dispersive potential have resulted in an extremely wide, pantropical distribution (Lessios, Kane & Robertson, 2003). It should be noted that T. gratilla was formerly a native resident of Kāne'ohe Bay. Tripneustes gratilla was once thought to be one of the most abundant urchin species within Kāne'ohe Bay (Edmonson, 1946; Alender, 1964; Banner & Bailey, 1970; Kay, 1994). Conversely, T. gratilla is now relatively rare and does not contribute significantly to herbivory on reefs within the bay (Conklin & Smith, 2005; Stimson & Conklin, 2008). However, historical outbreaks of the native alga Dictyospaeria were thought to be controlled by T. gratilla because growth accumulated mostly in calm waters of the bay where the urchin was rare (Banner & Bailey, 1970). Hence, there is considerable interest from both State and local conservation groups to replenish the natural population of *T. gratilla* in the bay and enhance natural herbivory to control these invasive alien algal species.

Tripneustes gratilla were provided by the DAR urchin hatchery as juveniles. We arbitrarily placed urchins in three non-overlapping size classes: small (17.5–22.5 mm maximum test diameter), medium (29.8–43.8 mm), and large (45.1–65.1 mm) that were then used for each of the experimental trials outlined below. Prior to the experiments, medium and large urchin size classes were raised on diets of all four alien algae. Small urchins were not fed before trials, but instead were placed directly into experiments within a few days of arriving from the hatchery. Therefore, urchins were starved 3–5 days prior to each experiment.

Algae

We chose the four most common species of alien invasive algae found on the patch reefs of Kāne'ohe Bay: Acanthophora spicifera, Gracilaria salicornia, Eucheuma denticulatum, and Kappaphycus clade B. Kappaphycus clade B (formerly identified as Kappaphycus alvarezii, K. striatum, or Eucheuma striatum) and Eucheuma denticulatum were intentionally introduced from the Philippines to Kāne'ohe Bay September 1974 by researchers from the University of Hawai'i for scientific research (Doty, 1971; Doty, 1977); fragments apparently drifted away from test sites on the north reef of Moku O Lo'e (Coconut Island) and were also collected and transplanted around the bay by local residents for personal cultivation (Russell, 1983; Batibasaga, Zertuche-González & de San, 2003; Weis & Butler, 2009; Ask et al., 2003). Despite having observed vegetative propagules being released, researchers

reported that such propagules were incapable of dispersing over deep water or finding suitable hollows on which to settle (*Doty, 1977*). This oversight led to the documented proliferation of new eucheumatoid colonies upon the introduction of the algae to test sites around Coconut Island (*Doty, 1977*). Likewise, the intentional introduction of *Gracilaria salicornia* by the same researchers to Kāne'ohe Bay occurred September of 1978, specifically for experimental aquaculture aimed at the development of commercial agar production (reviewed by *Rodgers & Cox, 1999*; *Smith et al., 2004*). The idea of a commercial agar industry in Hawai'i has long since been abandoned, but the introduced *G. salicornia* has established and spread along the shores of Waikīkī and reefs in Kāne'ohe Bay.

In contrast to these intentional introductions, a fragment of *Acanthophora spicifera* was first documented in Pearl Harbor in the fall of 1952, and was believed to have been transported on the heavily fouled hull of the barge "Yon 146" which was towed to O'ahu from Guam in 1950 (*Doty, 1961*). By February 1956, *A. spicifera* had been documented in Kāne'ohe Bay, making it the first documented accidental introduction to the Bay (*Kohn, 1961*; *Coles, DeFelice & Eldredge, 2002*). These invasive macrophytes became not only some of the most dominant benthic organisms, but they have also resulted in the most detrimental impacts to marine communities in the bay (*Coles, DeFelice & Eldredge, 2002*).

The four algae species had widespread distributions in the bay and were readily available for collection. *Acantohophora spicifera* and *G. salicornia* were easily collected nearly anywhere around Coconut Island and around the southern portion of Kāne'ohe Bay. The eucheumoids were consistently collected from patch reefs in the central portion of the Bay. We did not included native algae in this study because their abundance is so reduced in the Bay (*Stimson, Larned & Conklin, 2001; Conklin & Smith, 2005; Stimson, Cunha & Philippoff, 2007*) that we could not collect enough for this experiment without impacting the remaining population.

Growth on single-species diet

Growth rates of *T. gratilla* were measured while on single-species diets of each *A. spicifera*, *G. salicornia*, *E. denticulatum* and *K. alverezii*. For each of the four algal species, three *T. gratilla* were housed in each of three 15 L replicate aerated flow-through tanks (~1–2 L/min). In order to monitor individual growth rates of urchins without marking the animals, each tank held a single urchin of each size class: small, medium, and large. For each treatment, algae were provided *ad libitum* to reduce any resource competition, and all aquariums were cleaned twice a week during which freshly collected algae were provided to each tank. Urchin test diameter was measured using Vernier calipers (VWR) to the nearest tenth of a millimeter each week for a month. An analysis of covariance (ANCOVA) was employed to analyze growth, with initial urchin test size used as the covariate. Tukey's HSD post-hoc comparison was then performed to determine significance of pairwise differences in average growth rates of urchins on each algae diet (Fig. 1).

No-choice feeding trials

No-choice feeding trials provided juvenile *T. gratilla* in each treatment with only a single species of alga and measured differences in mean consumption among the treatments with

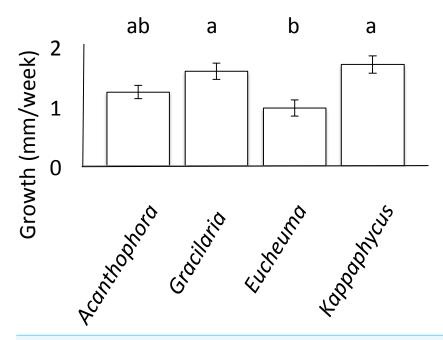


Figure 1 Urchin Growth. Mean weekly growth rate (mm/week \pm SE) of *Tripneustes gratilla* on non-choice diets of algae (*Acanthophora spicifera n* = 26, *Gracilaria salicornia n* = 28, *Eucheuma denticula-tum n* = 27, or *Kappaphycus clade B n* = 9) reared in aquaria over a 4-week period. Note letters identify significant subsets (p < 0.05, Tukey HSD).

different algal species. For each algal species, two urchins of the same size class were added to each of six replicate tanks (15 L tanks with 1-2 L/min flow rate, as above). Algae were blotted on paper towels to remove excess water and weighed before being placed in each tank. Urchins were allowed to graze for \sim 5 days and the amount of remaining algae was weighed as before to calculate the amount of each species consumed per urchin per day. In a few cases, we stopped the experiment after the 4th day because we did not want the urchins to consume more than half of the algae offered in any trial. Consumption rates (grams of algae per day) were then compared by analysis of variance (ANOVA). Tukey's HSD post-hoc comparison was used to identify significant differences between each of the algae (Fig. 2). The assumptions of Normality and homogeneity of variance of the data were tested using the Shapiro-Wilks test and the Levene's test, respectively. The null hypothesis for the Shapiro-Wilks test was that the data were Normally distributed; therefore, p-values less than 0.05 suggested that the data were not Normally distributed. For the small size class in the no-choice feeding trials the data were Normally distributed (Shapiro-Wilk, W = 0.96, p = 0.82). The data from the medium cohort from the no-choice trial was also normally distributed (Shapiro-Wilk, W = 0.90, p = 0.28). However, the data from the large urchins of the no-choice feeding trial were only marginally non-normal (Shapiro–Wilk, W = 0.80, p = 0.042). To test the homogeneity of variance among our feeding trial data, the Levene's test was employed. The null hypothesis for the Levene's test was that the variances are homogenous. For the urchins in the no-choice feeding trials, the data passed the homogeneity test (Levene's test, F(3,44) = 0.56, p = 0.64).

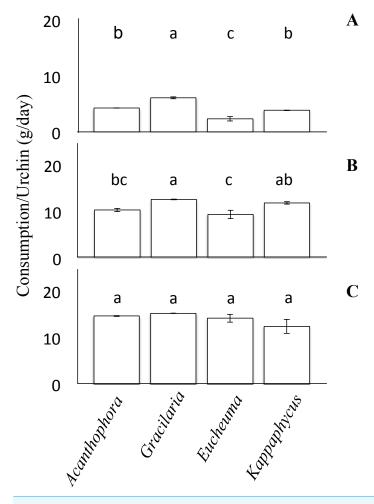


Figure 2 No-choice feeding trials. Consumption rates (g/day \pm SE) by *Tripneustes gratilla* during non-choice feeding trials of algae (*Acanthophora spicifera*, *Gracilaria salicornia*, *Eucheuma denticulatum*, or *Kappaphycus clade B*). (A) Small. (B) Medium. (C) Large. Note letters identify significant subsets (p < 0.05 Tukey HSD post-hoc pairwise comparison). For each diet of each size cohort n = 4.

Choice feeding trials

The choice feeding experiment provided all four species of algae, in equivalent amounts, simultaneously to urchins. As with no-choice experiments above, each algal species was blotted and weighed before being introduced to the experimental tanks. For this assay, larger tanks (80 L, \sim 4 L/min flow-through) were used to allow room to separate algae into the four quadrants of the experimental tank. Four urchins were then introduced to the middle of the tank and allowed to graze at will for \sim 7 days. Again, if any species of alga became low relative to the others (less than half the initial amount), we stopped the experiment a day early to avoid biasing results. At the end of each experiment, algae were removed, blotted and weighed as previously to calculate the amount of algae consumed per urchin per day for each species.

For each choice and no-choice feeding trials, small urchins were provided with ~ 100 g of algae initially, whereas medium and large urchins were offered a starting biomass

of \sim 150 g of algae. To account for any growth or decline of the algae not attributed to urchin grazing during the experiment, both choice and no-choice experiment tanks had a divider such that one half of the tank housed experimental algae and urchins whereas the other side housed only equivalent amounts of algae to serve as a no urchin control. The consumption rates of algae were then calculated as:

Consumption = $(A_i (AC_f/AC_i) - A_f)$

where A_i and A_f were the initial and final blotted masses of algae subject to grazing by urchins; while AC_i and AC_f were the initial and final masses of the algae in the no-urchin control tanks. This equation was used to account for growth of algae over the course of the experiment, but can also account for any unexpected decline in algal biomass unrelated to the grazing trial (Dworjanyn, Pirozzi & Wenshan, 2007; Seymour et al., 2013). Because all species of algae were provided simultaneously during choice feeding trials, the consumption of one species was affected by the consumption of the others, therefore the assumption of independence required to perform an ANOVA was violated. Consequently, choice feeding preference assays were analyzed using a non-parametric Friedman's rank test, and both parametric and non-parametric analyses are congruent. Relative consumption rates of each algal species were reported (Fig. 3) and ranked. Nevertheless, due to the lack of post-hoc pairwise comparison for Friedman's rank test, a Tukey's HSD post-hoc comparison was used to identify significant differences between each of the algae. Again, to test the data's distribution for Normality and homogeneity of variance the Shapiro-Wilk and Levene's test were used. The data from the small urchins of the choice feeding trials failed the normality test (Shapiro-Wilk, W = 0.86, p = 0.0036). However, the data for the medium and large sized urchins of the choice feeding trials both passed the normality test (Shapiro–Wilk, W = 0.97, p = 0.27, and W = 0.96, p = 0.17, respectively). For the small urchins of the choice feeding trial, the data passed the homogeneity test (Levene's test, F(3,20) = 2.13, p = 0.13). The data from the medium urchins in the choice feeding trial fail to reject the null hypothesis (Levene's test, F(3,36) = 0.32, p = 0.81). The data from the cohort of large urchins in the choice feeding trials also passed the test for equality of variances (Levene's test, F(3,40) = 1.98, p = 0.13).

Field caging experiment

Cages measuring roughly $50 \times 50 \times 75$ cm were constructed from 1 cm² galvanized chicken wire mesh. We constructed both open and closed cages. For closed cages, the mesh extended across all sides including the tops to prevent urchins from being able to crawl out and prevent access by fishes on the reef. In contrast, the sides of the open cages end with back-folded edges and no top to minimize escape of the juvenile urchins from the cage, but still allow open access of predatory fishes. Initial trials with urchins caged in seawater tables indicated that this back-folded edge design (approximating an upside down U) was the most effective for open cages, but urchins still escaped the cages at the rate of 1–2 animals per week. Urchins ranged from 18–22 mm at the start of the experiment. Cages were filled with *G. salicornia* to provide the juvenile urchins with food and a place to

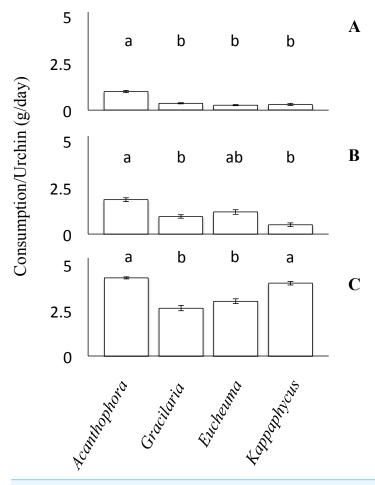


Figure 3 Choice feeding trials. Consumption rates (g/day \pm SE) of *Tripneustes gratilla* during three choice feeding trials of algae (*Acanthophora spicifera, Gracilaria Salicornia, Eucheuma denticulatum, and Kappaphycus clade B*). (A) Small, n = 6. (B) Medium, n = 10. (C) Large, n = 11.

hide, because our initial aquarium trials revealed that urchins were far more likely to escape the open cages in the absence of hiding spots and food in the cage. In the absence of any cover or food, open cages were frequently empty within 24 h in our water table trials (data not shown).

Cages were placed at 6 sites across three habitats, with four cages, three open and one closed control, at each site. 251 urchins were used during these caging experiments. Three habitats surrounding Coconut Island (map in Supplemental Information) were selected to mimic the conditions on the reef to which urchins are being currently outplanted: a protected lagoon, a shallow back-reef and a fore-reef slope each at 1–3 m depth. The protected lagoon had low coral cover, high alien algal cover and minimal water flow, whereas both the back-reef and fore-reef sites had high coral cover, relatively low alien algal cover and relatively high water flow. Each cage was checked three times a week for 30 days to count surviving urchins as well as replenish consumed algae. All studies reported here were conducted under the State of Hawaii, Department of Land and Natural Resources,

Division of Aquatic Resources Special Activity Permits sap#2012–63 and SAP#2013–47. Survivorship between treatments was compared using the Kaplan–Meier product-limit method for fitting survivorship curves and comparison by Log-rank (*Forsman, Rinkevich & Hunter, 2006*), and Wilcoxon non-parametric tests. These statistical tests were done using JMP Pro 11.

RESULTS

Growth on single-species diet

Growth rates of *T. gratilla*, measured as maximum test diameter (mm), were significantly affected by fixed algal diets (ANCOVA, initial urchin size as covariate, $F_{(5,84)}=10.80$, p<0.001, Fig. 1). Urchins that fed exclusively on diets of either *G. salicornia* or *K. clade B* grew at significantly (p=0.001 and p=0.009, respectively, Tukey HSD) higher rates (1.58 \pm 0.14 and 1.69 \pm 0.14 mm/week TD (Test Diameter), respectively) than those urchins that fed on a diet of *E. denticulatum*. Urchins that fed on a diet of *E. denticulatum* had the lowest growth rates (0.97 \pm 0.14 mm/week TD) out of the four assays, though not significantly lower to urchins on a diet of *A. spicifera* (1.23 \pm 0.11 mm/week TD).

No-choice feeding trials

When presented with no choice, urchins consumed different species of algae at different rates, but the effect varied by urchin size (Fig. 2). On average, large urchins ate G. salicornia at the highest rate of all algal species offered (15.24 \pm 0.001 g day⁻¹), and K. clade B at the lowest rate $(12.43 \pm 1.51 \text{ g day}^{-1})$ although these trends were not significant (ANOVA, $F_{(3,12)} = 1.94$, p = 1.78, Fig. 2C). Medium sized urchins showed similar trends, but with significant differences in the amounts of algae they consumed on a daily basis (ANOVA, $F_{(3,12)} = 8.49$, p < 0.05, Fig. 2B). For the medium size class, *Gracilaria salicornia* was eaten at a significantly higher rate $(12.60 \pm 0.08 \text{ g day}^{-1})$ than either A. spicifera $(10.33 \pm 0.36 \,\mathrm{g \, day^{-1}})$ or E. denticulatum $(9.35 \pm 0.90 \,\mathrm{g \, day^{-1}})$ $(p = 0.034 \,\mathrm{and}\, p = 0.003,$ respectively, Tukey HSD); K. clade B was also eaten at a higher rate $(11.87 \pm 0.27 \text{ g day}^{-1})$ than E. denticulatum (p = 0.018, Tukey HSD), but not A. spicifera (p > 0.05, Tukey HSD). Among the small collector urchins, feeding rate patterns were comperable to those of the medium urchins, but with more significant disparities among algal species, (ANOVA, $F_{(3,12)} = 51.30$, p < 0.001, Fig. 2A). Small urchins offered only G. salicornia had a significantly higher mean consumption rate $(6.08 \pm 0.19 \text{ g day}^{-1})$ than any other algal assay in the non-choice feeding trial (p < 0.05 for each pairwise comparison, Tukey HSD). Small urchins likewise consumed Eucheuma denticulatum at the lowest rate $(2.32 \pm 0.39 \text{ g day}^{-1})$, which was significantly lower than both A. spicifera and K. clade B (p < 0.05, Tukey HSD).

Choice feeding trials

Feeding trials in which urchins were offered multiple species of algae simultaneously revealed different patterns than those observed in the non-choice feeding assays. Small urchins significantly preferred to feed on *A. spicifera* than any of the other three available algae species (Friedman's rank test, p < 0.05; $p \le 0.01$, Tukey HSD, Fig. 3A). Small urchins

Table 1 Analysis of the Kaplan-Meier Survivorship curves for the Urchin Caging Experiment. A comparison of survivorship distributions between open and closed cages placed in the lagoon, the reef flat and the reef slope.

		df	Log rank		Wilcoxin		Verdict
			Statistic	P	Statistic	P	
I. Between all treatments		5	47.15	< 0.0001	41.36	< 0.0001	
II. Between open cages		2	38.62	< 0.0001	33.59	< 0.0001	
	R vs L	1	4.43	0.0352	3.17	0.0750	$R \ge L$
	R vs S	1	37.79	< 0.0001	32.66	< 0.0001	R > S
	S vs L	1	13.01	0.0003	12.06	0.0005	L > S
III. Between closed cages		2	6.73	0.0346	6.16	0.0460	
	R vs L	1	4.08	0.0434	3.62	0.0571	$R \ge L$
	R vs S	1	6.13	0.0133	5.73	0.0166	R > S
	S vs L	1	0.05	0.8315	0.15	0.6959	ns

Notes.

R, reef flat; S, reef slope; L, lagoon; ns, not significant.

did not display any patterns of preference among G. salicornia, E. denticulatum or K. clade B(p > 0.05, Tukey HSD). Likewise medium urchins showed a significant preference (Friedman's rank tests, p < 0.001, Fig. 3B) for A. spicifera over G. salicornia and K. clade B (p < 0.05 and p < 0.001, respectively, Tukey HSD), as it was consumed at the highest rate (3.69 \pm 0.21 g day⁻¹), whereas G. salicornia and E. denticulatum were consumed at intermediate rates (0.95 \pm 0.1 and 1.19 \pm 0.13 g day⁻¹, respectively). In contrast, medium urchins consumed K. clade B at the lowest rate $(0.51 \pm 0.10 \text{ g day}^{-1})$. Unlike the small and medium urchins, which avoided K. clade B in the choice trials, large urchins exhibited significant preferences for both A. spicifera and K. clade B (4.30 \pm 0.09 g day⁻¹ and 4.31 ± 0.14 g day⁻¹, respectively) in the choice feeding trials (Freidman's rank test, P < 0.001, Fig. 3C). Large urchins significantly preferred A. spicifera to both G. salicornia and E. denticulatum (p < 0.0001 and p < 0.001, respectively, Tukey HSD). Kappaphycus clade B was also significantly preferred to G. salicornia and E. denticulatum (p < 0.01and p < 0.05, respectively, Tukey HSD). Whereas G. salicornia and E. denticulatum were consumed at intermediate rates by small and medium urchins, these species tend to be avoided by the large urchins $(3.11 \pm 0.20 \text{ g day}^{-1} \text{ and } 3.20 \pm 0.20 \text{ g day}^{-1}, \text{ respectively})$ when given a choice of algae on which to feed (Fig. 3).

Caging experiment

Considerable differences in urchin survivorship among the three habitats that they were caged in were found to be significant (Table 1). On the reef flat, 84.4% of urchins deployed in closed cages remained and 75% of urchins remained in the open cages after 29 days (Fig. 4A). Urchins caged in the lagoon had a substantially higher rate of loss than those on the reef flat, with only 56.3% of urchins remaining after 29 days, regardless of cage type (Fig. 4B). After 29 days, 55.2% of urchins remained in closed cages, but only 20% survived in the open cages, giving urchins deployed on the reef slope the highest rate of loss (Fig. 4C).

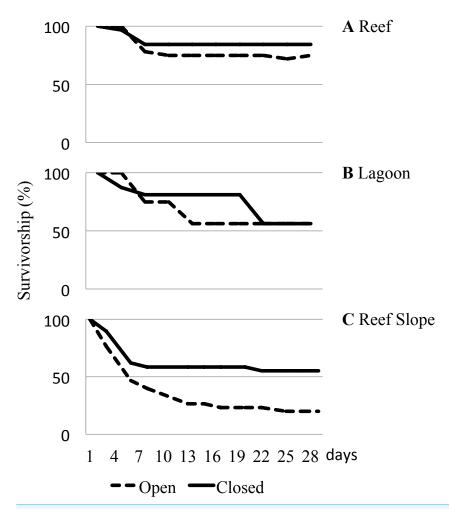


Figure 4 Caged urchin survivorship. Survivorship (%) curves reported for urchins deployed in open and closed cages in various underwater habitats for a month. (A) Reef. (B) Lagoon. (C) Reef Slope. Two hundred and fifty-one urchins were used across 6 sites.

DISCUSSION

Increasing the abundance of native grazers would not only control and remove current alien invasive algae species, but it would also serve to increase the degree of biotic resistance to novel invasive species (*Kimbro, Cheng & Grosholz, 2013*). Consistent with previous studies (e.g., *Lawrence & Agatsuma, 2001*; *Seymour et al., 2013*), *Tripneustes gratilla* is a generalist herbivore that managed to grow on every species of algae tested, and fed on all algal species offered to them without exception. However, *T. gratilla* did not interact with every species of algae indiscriminately. The urchins experienced variable growth depending on the diet they fed on. They did not feed on all rhodophytes at the same rate, and even exhibited preferences between the available species when presented with a choice of all four alien algae species. All of the algal diets supported growth, but the algae species that supported the highest growth rates were *G. salicornia* and *K. clade B*, whilst urchins that fed solely on *E. denticulatum* had the lowest growth rates, and *A. spicifera* sustained

intermediate growth (Fig. 1). The reason for differential growth was not investigated, but could possibly be attributed to variation in nutritional content, consumption rate, digestibility or assimilation efficiency (*Sterner & Hessen, 1994*). For example, growth rates may be higher on *G. salicornia* than *A. spicifera* because *Gracilaria* contains more protein than does *Acanthophora* (*McDermid, Stuercke & Balazs, 2007*). Likewise, the low growth rate of urchins on a diet of *E. denticulatum* could result from the comparatively high dietary fiber of this alga, a compound that resists digestion and lowers assimilation efficiency (*McDermid, Stuercke & Haleakala, 2005*).

As with consumption rates, feeding preferences were also observed to vary among size classes (Fig. 3). The differences in feeding preference and consumption rates could translate into urchins of different size classes having differential impacts on alien algae, and argue that biocontrol efficiency could be increased by outplanting urchins of the correct size class for the dominant algal species to be controlled. For example, urchins of all size classes consumed A. spicifera preferentially in choice experiments (Fig. 3). Acanthophora spicifera is known to be consumed by other native grazers (Wylie & Paul, 1988; Russell & Balazs, 1994), but the effects of alien algal ingestion on the diets of native herbivores remains unknown (Smith et al., 2004). Our results show a greater disparity of preference among smaller than larger urchins (Fig. 3), suggesting that the role of alien algae in the diets of native grazers may vary ontogenetically. Nevertheless, small urchins showed a significant preference for A. spicifera relative to all other species, whereas larger urchins showed a higher affinity for both A. spicifera and K. clade B. Despite being a preferred species in all choice feeding trials, urchins did not seem to grow at a significantly different rate $(1.23 \pm 0.11 \text{ mm/week test diameter})$ when fed only A. spicifera relative to any other algal diet (Fig. 1). In fact, growth rates appeared maximal on a diet of G. salicornia which was less preferred in all choice feeding trials (Fig. 3).

Our results contrast those presented by Stimson, Cunha & Philippoff (2007), which focused solely on large adult urchins (7–8 cm test diameter), and showed a significant preference of urchins for Kappaphycus spp. but without including Eucheuma denticulatum in the feeding trials. Here we find that consumption rates of each algal species vary by size class, and we see no significant differences among consumption rates of algal species for urchins in our largest size class (Fig. 2). Given the strong differences seen in diet preference among urchins of different size classes, the discrepancy between our results and those of Stimson, Cunha & Philippoff (2007), could result from a continued change in diet preference and their use of significantly larger urchins than used in our feeding experiments. Among medium urchins, however, significantly more G. salicornia and K. clade B were consumed, and the smallest urchins consumed significantly more G. salicornia than the other three species (Fig. 2). Although the general trend is similar among urchin size classes, the biggest differences among the consumption rates were seen in the smallest urchins, which consume slightly more than twice as much G. salicornia as E. denticulatum (Fig. 2A). However, as urchin test diameter increased, there was a reduction in the difference between feeding assays, until all four species were consumed at statistically indistinguishable rates among adult urchins in the no-choice trials; larger

urchins eat more algae, and grazing rates become more homogeneous among the four species of red algae (Fig. 2). Algal palatability can be reduced by increased algal toughness in a range of herbivorous species (e.g., *Peters et al., 2002*). For some species of echinoderms, it has been documented that larger sizes (body diameter) are associated with greater jaw strength (*Ellers & Telford, 1991*). Thus, it may be that larger urchins are better able to masticate a wider variety of algae, including species with larger and tougher thalli, such as *E. denticulatum* and *K. clade B*.

It is noteworthy that despite the widespread distribution of *T. gratilla*, caged juvenile collector urchins did not fare equally well in every environment. The caging experiment highlighted significant differences in survivorship among potential outplanting locations (Table 1). Urchins placed in high algal cover, comparatively low-flow lagoonal habitats fared poorly, with only \sim 50% survivorship (Fig. 4B). Survivorship was equal for animals in both open and closed cages in the lagoon, and the removal of empty tests (clear evidence of mortality) accounted for all but a couple animals by the end of the month deployment. Thus, we are confident that the decline in urchin number in the lagoon was a result of low survival as opposed to predation or escape. Urchins caged on the reef flat consistently had the highest overall survivorship rates, with nearly 80% of urchins remaining at the end of the experiment. In these sites, we found only a single urchin test in any cage, and we had a few urchins somehow escape the closed cages in our initial water table trials, so we cannot be sure of whether the missing urchins died or escaped, but the high overall survival in both open and closed cages provides evidence that placement of urchins in these habitats is likely to increase the population of herbivorous urchins over time. In contrast, juvenile urchins placed on the reef slope suffered mortality rates as high as 80% in open cages (Fig. 4C). Even in closed cages, survival over the course of the experiment was only slightly above 50%, and again we recovered only 25 total tests in the cages over the course of the experiment. This seems likely to be a result of predation by reef fishes because we see a much more dramatic decline on the deeper water reef slope than in the shallows of the reef flats (Fig. 4). The decline of urchins in the closed cages on the reef slope below what is seen in open cages on the reef flat is somewhat puzzling. However, we noted during the experiment that the saddleback wrasse, *Thalassoma duperrey*, was particularly abundant around the urchin cages along the reef slope. Wrasses have been documented harassing juvenile T. gratilla urchins (Dafni & Tobo, 1987), and in a couple of cases, fish were even found to have somehow squeezed themselves inside one of the closed cages along the reef slope and were subsequently trapped in the cage on the next day. Although the mesh size was intended to exclude predatory fish, it is possible that wrasses small enough to fit through 1 cm² mesh may have been responsible for the decline of urchins in closed cages along the reef slope, and the loss of 80% of animals from open cages within a month suggests that outplanting urchins into deeper waters of the bay is simply generating a feeding station.

CONCLUSION

The estimated overall productivity of these alien invasive eucheumatoids in Kāne'ohe Bay is 20.8 tonnes dry wt/ha/yr, which translates to 5.7 g dry wt/m²/day (*Glenn & Doty*,

1990). Given this approximation of productivity and the data compiled on consumption rates, a rough estimate of the ideal urchin density can be derived. Here we find that a large urchin (45–65 mm TD) could graze \sim 7.5 g of alien algae per day when presented with a mixed diet. This back-of-the-envelope calculation suggests that the grazing rate of one adult urchin/m² may be just about equal to the predicted growth rates of these algae. Additionally, our caging experiments indicate that mortality rates for juvenile urchins in open cages on the reef flat are on the order of 25%. Thus, a target density of two urchins/m² is recommended to overcome growth and reduce the biomass of alien algae if urchin grazing is to be effective for biocontrol. Given the high mortality rates for juvenile urchins on the deeper reef slope and the protected lagoon habitats, it is not advisable to invest the effort to culture and outplant juvenile urchins in either environment. Although there is variability in growth rates, juvenile urchins tend to grow quickly and the largest size class of urchins we tested showed no significant preferences among any of the target alien invasive algal species. Given that current efforts by conservation groups aim to manually remove alien algae and then outplant native urchins to the reef flats, where survivorship was high, this study increases confidence that intentional outplanting of juvenile urchins is likely to be an effective means of biocontrol for these invasive alien algae.

ACKNOWLEDGEMENTS

Thank you to Brian Bowen, Mary Hagedorn, Ginnie Carter and Richard Coleman. The members of the HDAR & TNC Urchin Team are David Cohen, Jono Blodgett, Brian Neilson, Andrew Purvus, Frank Mancini, Tristan Walker, Brad Stubbs, Derek LeVault, Cathy Gewecke, Vince Calabrese, Karen Brittain, Neil Rodriguez, Shealin Johnson, Ryan Carr, Hank Lynch, Justin Dennis, and Kirsten Fujitani.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This project was funded in part by the National Science Foundation (NSF) Grant #OCE-0623678 and NOAA National Marine Sanctuaries Partnership MOA#2005-008/66882 to RJT, RR & SC were supported as research interns under the Undergraduate Research Mentoring in Biological Sciences (URM) Grant (NSF #0829272). This is contribution X from the Hawaii Institute of Marine Biology, Y from the School of Ocean and Earth Science and Technology. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

National Science Foundation (NSF): #OCE-0623678.

NOAA National Marine Sanctuaries Partnership: MOA#2005-008/66882.

Undergraduate Research Mentoring in Biological Sciences (URM): NSF #0829272.

Hawai'i Institute of Marine Biology.

School of Ocean and Earth Science and Technology.

Competing Interests

Robert J. Toonen is an Academic Editor for PeerJ.

Author Contributions

- Charley E. Westbrook and Robert J. Toonen conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Rory R. Ringang and Sean Michael A. Cantero conceived and designed the experiments, performed the experiments, contributed reagents/materials/analysis tools, reviewed drafts of the paper.
- HDAR & TNC Urchin Team conceived and designed the experiments, contributed reagents/materials/analysis tools, reviewed drafts of the paper.

Field Study Permissions

The following information was supplied relating to field study approvals (i.e., approving body and any reference numbers):

Division of Aquatic Resources Special Activity Permits SAP#2012-63 and SAP#2013-47.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.1235#supplemental-information.

REFERENCES

Alender CB. 1964. The venom from the heads of the globiferous pedicellariae of the sea urchin *Tripneustes gratilla* (Linnaeus). PhD dissertation, University of Hawaii at Mānoa, Honolulu, USA. 126 pp.

Alves FMA, Chícharo LM, Serrão E, Abreu AD. 2003. Grazing by *Diadema antillarum* (Philippi) upon algal communities of rocky substrates. *Scientia Marina* **67**:307–311 DOI 10.3989/scimar.2003.67n3307.

Ask EI, Batibasaga A, Zertuche-Gonzalez JA, De San M. 2003. Three decades of *Kappaphycus alvarezii* (Rhodophyta) introduction to non-endemic locations. In: Chapman ARO, Anderson RJ, Vreeland VJ, Davison IR, eds. *Proceedings of the 17th international seaweed symposium*. Oxford: Oxford University Press, 49–57.

Bahr KD, Jokiel PL, Toonen RJ. 2015. The unnatural history of Kāne'ohe Bay: coral reef resilience in the face of centuries of anthropogenic impacts. *PeerJ* 3:e950. DOI 10.7717/peerj.950.

Banner AH, Bailey JH. 1970. The effects of urban pollution upon a coral reef system: a preliminary report. *HIMB Technical Bulletin*, #25. University of Hawai'I at Mānoa. 66 pp.

Batibasaga EI, Zertuche-González JA, De San M. 2003. Three decades of *Kappaphycus alvarezii* (Rhodophyta) introduction to non-endemic locations. *Proceedings of the International Seaweed Symposium* 17:49–58.

Chiappone M, Swanson DW, Miller SL, Smith SG. 2002. Large-scale surveys on the Florida reef tract indicate poor recovery of the long-spined sea urchin. *Diadema antillarum. Coral Reefs* **21**:155–159.

- Coles SL, DeFelice RC, Eldredge LG. 2002. Nonindigenous Marine Species in Kāne'ohe Bay, Oʻahu, Hawaiʻi. In: *Technical Report 24*. Honolulu: Bernice Pauahi Bishop Museum. *Available at http://hbs.bishopmuseum.org/pdf/kbay-report.pdf*.
- Conklin KY, Kurihara A, Sherwood AR. 2009. A molecular method for identification of the morphologically plastic invasive algal genera Eucheuma and Kappaphycus (Rhodophyta, Gigartinales) in Hawaii. *Journal of Applied Phycology* 21(6):691–699 DOI 10.1007/s10811-009-9404-2.
- Conklin EJ, Smith JE. 2005. Abundance and spread of the invasive red algae, *Kappaphycus* spp., in Kāne'ohe Bay, Hawai'i and an experimental assessment of management options. *Biological Invasions* 7:1029–1039 DOI 10.1007/s10530-004-3125-x.
- **Dafni J, Tobo R. 1987.** Population structure patterns of a common red sea echinoid *Tripneustes-gratilla-elatensis. Israel Journal of Zoology* **34(3–4)**:191–204.
- **Doty MS. 1961.** *Acanthophora*, a Possible Invader of the Marine Flora of Hawaïi. *Pacific Science* **15**:547–552.
- **Doty MS. 1971.** Antecedent event influence on benthic marine algal standing crops in Hawaii. *Journal of Experimental Marine Biology and Ecology* **6(3)**:161–166 DOI 10.1016/0022-0981(71)90015-3.
- **Doty MS. 1977.** *Eucheuma*-current marine agronomy. In: Krauss RW, ed. *The marine plant biomass of the Pacific Northwest Coast.* Corvallis: Oregon State University Press, 203–214.
- **Dworjanyn SA, Pirozzi I, Wenshan L. 2007.** The effect of the addition of algae feeding stimulants to artificial diets for the sea urchin *Tripneustes gratilla*. *Aquaculture* **273**:624–633 DOI 10.1016/j.aquaculture.2007.08.023.
- **Edmonson CH. 1946.** *Reef and shore fauna of Hawai'i*. Vol. 22. Honolulu: Bernice Pauahi Bishop Museum, 381.
- **Ellers O, Telford M. 1991.** Forces generated by the jaws of clypeasteroids (Echinodermata, Echinoidea). *Journal of Experimental Marine Biology* **155**:585–603.
- **Forsman ZH, Rinkevich B, Hunter CL. 2006.** Investigating fragment size for culturing reef-building corals (*Porites lobata* and *P. compressa*) in *ex situ* nurseries. *Aquaculture* **261**:89–97 DOI 10.1016/j.aquaculture.2006.06.040.
- **Gibo C, Letsom T, Westbrook C. 2012.** Effects of temperature, salinity, pH, reef size, and *Tripneustes gratilla* on the distribution of *Montipora dilatata* in Kāne'ohe Bay. Final class report to NOAA species of Concern, Honolulu. *Available at http://www.fpir.noaa.gov/Library/PRD/SOC/M.*%20dilatata%20SOC%20Final%20Report%207-2012.pdf.
- **Glenn EP, Doty MS. 1990.** Growth of the seaweeds *Kappaphycus alvarezii, K. striatum* and *Eucheuma denticulatum* as affected by the environment in Hawaii. *Aquaculture* **84**:245–255 DOI 10.1016/0044-8486(90)90090-A.
- **Godwin S, Rodgers KS, Jokiel PL. 2006.** Reducing Potential Impacts of Invasive Marine Species in the Northwestern Hawaiian Islands Marine National Monument. Report to: Northwestern Hawaiian Islands Marine National Monument Administration. 1–66.
- **Holland BS, Taylor C, Sugiura S. 2012.** Tracking behavior in the snail *Euglandina rosea*: First evidence of preference for endemic vs. biocontrol target pest species in Hawaiï. *American Malacological Bulletin* **30(1)**:153–157 DOI 10.4003/006.030.0113.
- **Hoover JP. 2002.** *Hawaii's Sea Creatures: a guide to Hawaii's marine invertebrates.* Honolulu: Mutual Publishing, 319.
- **Howarth FG. 1983.** Classical biocontrol: panacea or pandora's box. *Proceedings, Hawaiian Entomological Society* **24**:239–244.

- **Howarth FG. 1985.** Impact of alien land arthropods and mollusks on native plants and animals in Hawaii. In: Stone CP, Scott JM, eds. *Hawaii's terrestrial ecosystems: preservation and management.* Honolulu: University of Hawaii's Cooperative National Park Resources Study Unit, 149–178.
- **Howarth FG. 1991.** Environmental impacts of classical biological control. *Annual Review of Entomology* **36**:485–509 DOI 10.1146/annurev.en.36.010191.002413.
- **Hunter CL, Evans CW. 1995.** Coral reefs in Kaneohe Bay, Hawaii: two centuries of western influence and two decades of data. *Bulletin of Marine Science* **57**(2):501–515.
- **Kay EA. 1994.** Marine ecosystems in the Hawaiian Islands. In: Kay EA, ed. *A natural history of the Hawaiian Islands: selected reading II.* Honolulu: University of Hawaii Press, 187–195.
- **Kimbro DL, Cheng BS, Grosholz ED. 2013.** Biotic resistance in marine environments. *Ecology Letters* **16**:821–833 DOI 10.1111/ele.12106.
- **Kohn AJ. 1961.** Studies on spawning behavior, egg masses, and larval development in the gastropod genus *conus*, Part I observations on nine species in Hawaiï. *Pacific Science* **15**:163–179.
- **Lawrence JM, Agatsuma Y. 2001.** The Ecology of *Tripneustes gratilla*. In: Lawrence JM, ed. *Edible Sea Urchins: biology and ecology*. Amsterdam: Elsevier, 514.
- **Lessios HA, Kane J, Robertson DR. 2003.** Phylogeography of the pantropical sea urchin *Tripneustes*: contrasting patterns of population structure between oceans. *Evolution* 57:2026–2036 DOI 10.1111/j.0014-3820.2003.tb00382.x.
- McDermid KJ, Stuercke B, Balazs GH. 2007. Nutritional composition of marine plants in the diet of the green sea turtle (*Chelonian mydas*) in the Hawaiian Islands. *Bulletin of Marine Science* 81:55–71.
- McDermid KJ, Stuercke B, Haleakala OJ. 2005. Total dietary fiber content in Hawaiian marine algae. *Botanica Marina* 48:437–440 DOI 10.1515/bot.2005.057.
- Mumby PJ, Hedley JD, Zychaluk K, Harborne AR, Blackwell PG. 2006. Revisiting the catastrophic die-off of the urchin *Diadema antillarum* on Caribbean coral Reefs: fresh insights on resilience from a simulation model. *Ecological Modeling* 196:131–148 DOI 10.1016/j.ecolmodel.2005.11.035.
- **Ogden NB, Ogden JC, Abbott IA. 1989.** Distribution, abundance and food of sea urchins on a leeward Hawaiian reef. *Bulletin of Marine Science* **45**(2):539–549.
- **Parker JD, Burkepile DE, Hay ME. 2006.** Opposing effects of native and exotic herbivores on plant invasions. *Science* **301**:1459–1561 DOI 10.1126/science.1121407.
- Peters KJ, Amsler CD, Amsler MO, McClintock JB, Iken KB, Baker WJ. 2002. The effects of thallus toughness and nutritive quality on palatability of antarctic macroalgae. *Journal of Phycology* **38**:29–30 DOI 10.1046/j.1529-8817.38.s1.84.x.
- **Rodgers SK, Cox EF. 1999.** Rate of Spread of Introduced Rhodophytes *Kappaphycus alvarezii*, *kappaphycus striatum*, and *Gracilaria salicornia* and Their Current Distribution in Kāne'ohe Bay, Oʻahu, Hawai'i. *Pacific Science* **53**:232–241.
- **Russell DJ. 1983.** Ecology of the Imported Red Seaweed *Eucheuma striatum* Schmitz on Coconut Island, Oʻahu, Hawaiʻi. *Pacific Science* **37**:87–107.
- Russell DJ, Balazs GH. 1994. Colonization by the alien marine alga *Hypnea* musciformis (Wulfen) J.Ag.(Rhodophyta: Gigartinales) in the Hawaiian islands and its utilization by the green turtle, *Chelonia mydas* L. *Aquatic Botany* 47(1):53–60 DOI 10.1016/0304-3770(94)90048-5.
- Russell DJ, Balazs GH. 2009. Dietary Shifts by Green Turtles (*Chelonia mydas*) in Kāne'ohe Bay Region of the Hawaiian Islands: a 28-year study. *Pacific Science* 63:181–192 DOI 10.2984/049.063.0202.

- Sandin SA, Walsh SM, Jackson JBC. 2010. Prey release, trophic cascades, and phase shifts in tropical nearshore marine ecosystems. In: Terborgh J, Estes JA, eds. *Trophic cascades: predators, prey, and the changing dynamics of nature.* Honolulu: Island Press, 71–90.
- Seymour S, Paul NA, Dworjanyn SA, De Nys R. 2013. Feeding preference and performance in the tropical sea urchin. *Tripneustes gratilla*. *Aquaculture* 400–401:6–13 DOI 10.1016/j.aquaculture.2013.02.030.
- Sherwood AR, Kurihara A, Conklin KY, Sauvage T, Presting GG. 2010. The Hawaiian Rhodophyta Biodiversity Survey (2006–2010): a summary of principal findings. *BMC Plant Biology* 10(1):258 DOI 10.1186/1471-2229-10-258.
- Simberloff D, Dayan T, Jones C, Ogura G. 2000. Character Displacement and Release in the Small Indian Mongoose *Herpestes javanicus*. *Ecology* 81(8):2086–2099 DOI 10.1890/0012-9658(2000)081[2086:CDARIT]2.0.CO;2.
- Simberloff D, Stiling P. 1996. How risky is biological control? *Ecology* 77(7):1965–1974 DOI 10.2307/2265693.
- Smith JE, Hunter CL, Conklin EJ, Most R, Sauvage T, Squair C, Smith CM. 2004. Ecology of the Invasive Red Alga *Gracilaria salicornia* (Rhodophyta) on Oʻahu, Hawaiʻi. *Pacific Science* 58:325–343 DOI 10.1353/psc.2004.0023.
- Smith JE, Hunter CL, Smith CM. 2002. Distribution and reproductive characteristics of nonindigenous and invasive marine algae in the Hawaiian Islands. *Pacific Science* **56(3)**:299–315 DOI 10.1353/psc.2002.0030.
- **Sterner RW, Hessen DO. 1994.** Algal nutrient limitation and the nutrition of aquatic herbivores. *Annual Review of Ecology and Systematics* **25**:1–29 DOI 10.1146/annurev.es.25.110194.000245.
- Stimson J, Conklin E. 2008. Potential reversal of a phase shift: the rapid decrease in the cover of the invasive green macroalgae Dictyosphaeria cavernosa Forsskål on coral reefs in Kāne'ohe Bay, Oahu, Hawai'i. *Coral Reefs* 27:717–726 DOI 10.1007/s00338-008-0409-0.
- **Stimson J, Cunha T, Philippoff J. 2007.** Food preference and related behavior of the browsing sea urchin *Tripneustes gratilla* (Linnaeus) and its potential for use as a biological control agent. *Marine Biology* **151**:1761–1772 DOI 10.1007/s00227-007-0628-x.
- **Stimson J, Larned ST, Conklin E. 2001.** Effects of herbivory, nutrient levels, and introduced algae on the distribution and abundance of the invasive macroalga *Dictyophaeria caevernosa* in Kāne'ohe Bay, Hawai'i. *Coral Reefs* **19**:343–357 DOI 10.1007/s003380000123.
- **Thomas F. 1994.** Physical properties of gametes in three sea urchin species. *The Journal of Experimental Biology* **194(1)**:263–284.
- Valentine JP, Edgar GJ. 2010. Impacts of a population outbreak of the urchin *Tripneustes* gratilla amongst Lord Howe Island coral communities. *Coral Reefs* 29:399–410 DOI 10.1007/s00338-010-0610-9.
- Vaïtilington D, Rasolofonirina R, Jangoux M. 2003. Feeding preferences, seasonal gut repletion indices, and diel feeding patterns of the sea urchin *Tripneustes gratilla* (Echinodermata: Echinoidea) on a coastal habitat off Toliara (Madagascar). *Marine Biology* 143:451–458 DOI 10.1007/s00227-003-1111-y.
- **Weis JS, Butler CA. 2009.** *Salt Marshes: a natural and unnatural history.* New Brunswick: Rutgers University Press, 159–160.
- Wylie CR, Paul VJ. 1988. Feeding preferences of the surgeonfish *Zebrasoma flavescens* in relation to chemical defenses of tropical algae. *Marine Ecology Progress Series* 45:23–32 DOI 10.3354/meps045023.