REVIEW ARTICLE



Cholinergic System and Post-translational Modifications: An Insight on the Role in Alzheimer's Disease



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Abstract: *Background*: Alzheimer's disease (AD) is the most common form of old age dementia. The formation of amyloid plaques (A β), neurofibrillary tangles and loss of basal forebrain cholinergic neurons are the hallmark events in the pathology of AD.

ARTICLE HISTORY

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DOI: 10.2174/1570159X14666160325121145 *Literature Review*: Cholinergic system is one of the most important neurotransmitter system involved in learning and memory which preferentially degenerates in the initial stages of AD. Activation of cholinergic receptors (muscarinic and nicotinic) activates multiple pathways which result in post translational modifications (PTMs) in multiple proteins which bring changes in nervous system. Cholinergic receptors-mediated PTMs "in-part" substantially affect the biosynthesis, proteolysis, degradation and expression of many proteins and in particular, amyloid precursor protein (APP). APP is subjected to several PTMs (proteolytic processing, glycosylation, sulfation, and phosphorylation) during its course of processing, resulting in $A\beta$ deposition, leading to AD. $A\beta$ also alters the PTMs of tau which is a microtubule associated protein. Therefore, post-translationally modified tau and $A\beta$ collectively aggravate the neuronal loss that leads to cholinergic hypofunction.

Conclusion: Despite the accumulating evidences, the interaction between cholinergic neurotransmission and the physiological significance of PTM events remain speculative and still needs further exploration. This review focuses on the role of cholinergic system and discusses the significance of PTMs in pathological progression of AD and highlights some important future directions.

Keywords: Acetylcholine, Alzheimer's disease, muscarinic receptors, nicotinic receptors, post translational modifications.

ALZHEIMER'S DISEASE

Neurodegenerative diseases are devastating conditions with progressive degeneration of nerve cells resulting in abnormal mental functioning specially dementia [1]. Degenerative diseases of the brain were long considered among the most ambiguous and troublesome of all diseases [2]. In 1906, Alois Alzheimer for the first time described the neuropathological features in the brain of a patient Auguste D., suffering from dementia. Later on, Emil Kraepelin in 1910 renamed the same pathology as "Alzheimer's Disease" to differentiate the general memory impairment from the common senile dementia [3].

PATHOLOGICAL FEATURES OF ALZHEIMER'S DISEASE

Major hallmarks of AD include basal forebrain cholinergic hypofunction [4], extracellular accumulation of beta-amyloid known as amyloid or "senile" plaques [5] and intracellular neurofibrillary tangles (NFTs) accumulation [6, 7]. Senile plaques are the extracellular aggregates of betaamyloid protein (AB) derived from cleavage of amyloid precursor protein (APP) via the action of β - and γ -secretase [8], while neurofibrillary tangles (NFTs) consist of hyperphosphorylated tau protein, present inside the neurons [9]. Beside plaques and NFTs, synaptic dysfunction is one of the most critical aspects of dementia [10, 11]. It has been found that synapses involving acetylcholine (ACh), glutamate and serotonin are primarily impaired in AD [12, 13]. The loss of basal cholinergic neurons is associated with severe neurodegeneration and cell loss in the nucleus basalis complex [14]. Cortex and hippocampus receive their major cholinergic input from nucleus basalis of Meynert and diagonal band of broncha, respectively [15]. The degeneration

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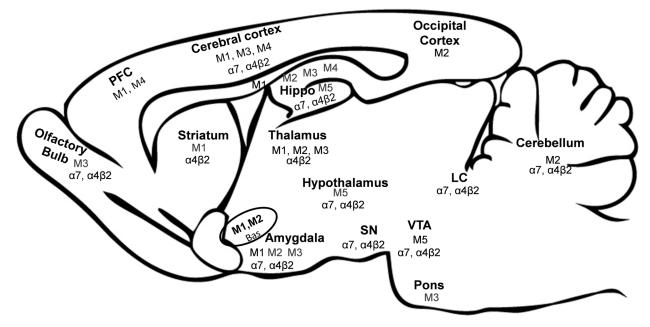


Fig. (1). Expression of Muscarinic and Nicotinic Receptors in Brain. Abbreviations: bas: Nucleus basalis, Hippo: hippocampus, LC: locus coeruleus, PFC: Prefrontal cortex, SN: substantia nigra, VTA: ventral tegmental area [39, 40, 44, 51, 52, 76].

of basal forebrain cholinergic neurons is considered to be the earliest pathological event along with plaque and tangle formation [16, 17]. Lesions of the cholinergic basal nuclei in rats result in a number of memory deficits [18] and affect memory and cognition in primates [19]. The basal forebrain cholinergic deficits positively correlate with cognitive [20] and non cognitive behavioral deficits [21] observed in AD patients. But the dilemma, that why the basal forebrain cholinergic neurons are among the first targets in AD pathology, still needs to be solved [22].

CHOLINERGIC RECEPTORS

The ACh receptor (AChR) is a vital membrane protein on which ACh acts as a neurotransmitter. The cholinergic receptors are broadly categorized as muscarinic ACh receptors (mAChR) and nicotinic ACh receptors (nAChR) on the basis of their exogenous agonists. The expression of these receptors varies in different brain areas (Fig. 1). Nicotinic receptors are found in the neuromuscular junction, autonomic ganglia and various places in the CNS, though, with different composition. While muscarinic receptors are found to be expressed in the brain both at the pre-synaptic and post-synaptic nerve terminals and parasympathetic effector organs [23].

MUSCARINIC RECEPTORS: CLASSIFICATION, LOCATION AND BRAIN FUNCTIONS

The mAChR belong to the family of seven transmembrane receptors coupled to G-proteins (GPCRs), regulating a variety of physiological processes [24]. These receptors are comprised of single polypeptides which form seven transmembrane domains forming a central pore. ACh binds at a site inside this pore to activate the signaling cascade *via* G-proteins [25]. In the central nervous system (CNS), the muscarinic system plays important role in the

regulation of many sensory, motor and autonomic processes [26]. Moreover, mAChRs have established roles in cholinergic transmission as well as learning and memory [12].

Muscarinic receptors are further sub-divided into five types M1-M5 encoded by five genes, m1-m5 [27]. The five mAChR subtypes are similar with the exception of the third intracellular loop while their signaling features are different, so these subtypes are further categorized into two groups [28-30] which determine the specific coupling preferences of these receptors [31]. The M1-like subfamily (M1, M3 and M5) is coupled to Gag/11 protein which causes activation of phospholipase C. Stimulation of M1-like subfamily receptors leads to regulation of different proteins and their functions by the process of phosphorylation. Whereas the M2-like subfamily (M2 and M4) is coupled to Gi/o, which inhibits adenylate cyclase [32]. The stimulation of M2 and M4 receptors causes a reduced cytosolic cAMP level [33]. The intracellular muscarinic signaling responses include activation of protein kinases, phospholipases A2 and D (releasing arachidonic acid and choline, respectively) and regulation of calcium and potassium channels [34].

The mAChRs are widely distributed throughout the body peripherally as well as centrally. In the hippocampus and cerebral cortex, mAChRs are involved in cognitive processes such as memory [35-37]. While in the striatum and motor cortex, these receptors are involved in motor function [38].

M1 receptors are abundantly present in the hippocampus, neocortex, striatum, amygdala, thalamus [39] and prefrontal cortex [40]. M1 receptor knock-out mouse showed elevated levels of A β peptides in brain [41] and increased aggregation of amyloid plaques which leads to impaired memory consolidation in this model [42].

M2 receptors are auto-receptors for ACh release [43], present on presynaptic cholinergic neurons, and abundantly

found in the cerebellum, thalamus [44] and nucleus basalis while lower levels are found in the hippocampus, amygdala and caudate putamen [39, 45].

The expression of M3 is relatively low i-e; 5-10% of total mAChRs in brain [46]. It is found in several brain regions, including cortex, amygdala, hippocampus, olfactory bulb, striatum, thalamus and pons [39]. M3 is involved in the regulation of neurotransmitter release, including dopamine, GABA and glycine as well as endocannabinoids [47, 48], suggesting its critical role in regulating other types of neurotransmission and learning and memory.

Relatively low levels of M4 receptors are expressed in brain as compared to other mAChR subtypes and are localized to hippocampus, including CA1 region and dentate gyrus [39], while the highest levels are in the caudate putamen [45] and prefrontal cortex [40] where they play role in the feedback control of neurotransmitter release [49] and cognitive processes [50].

M5 mAChRs has been found at low levels in the brain, particularly in the ventral tegmental area [51, 52], hippocampus and substantia nigra [13]. These receptors play an important role in facilitating muscarinic agonist-induced dopamine release from the nerve terminal [53]. So as a consequence, presence of M5 receptor is important for dopamine release and might be involved in facilitating dopamine-mediated and reward related physiological functions.

INVOLVEMENT IN ALZHEIMER'S DISEASE

Normal processing of the APP is under the control of cholinergic inputs [54, 55]. So altered cholinergic innervations could lead to abnormal processing of β-amyloid and possible formation of potentially neurotoxic fragments leading to neuritic plaque formation [56-58]. It has been observed that ACh esterase (AChE) accelerates the aggregation of AB into insoluble amyloid fibrils via unknown mechanism [59]. Basal forebrain cholinergic neuronal fibres are lost in AD at a later stage [60, 61]. This neuronal loss may be a result of $A\beta$ neurotoxicity to the cholinergic terminals followed by retrograde degeneration [62]. Long-term exposure to micromolar concentration of $A\beta$ is toxic to cholinergic neurons [63]. There are several reports on loss of cholinergic fibers and nerve terminals in AD and reduced cholinergic receptors [12, 64] but the relationship between AB and cholinergic deficit is poorly understood [62]. Due to the deterioration of cholinergic neurons in the brain of AD patients, there is a considerable loss of nicotinic receptors and certain muscarinic receptors particularly in the cortex and hippocampus [65-67], leading to impaired neurotransmitter release. In the cortical pyramidal neurons the activation of muscarinic receptors is known to enhance GABAergic transmission. The GABAergic inhibition is cruical for execution of certain memory forms by controlling the information flow in cortical circuits. Therefore, cholinergic hypofunction leads to cognitive impairment in AD patients [68].

Cholinergic hypofunction is a hallmark of AD [69, 70]. Specially, M1 and, M2 are down-regulated in hippocampus and cortex [45] and M4 appears to be down-regulated in cortex [71]. Down-regulation of M1, M2 and M4 is responsible for cognition deficits as well as impaired ACh release which exacerbates the AD symptoms. In another study, M1 receptors remained unchanged in AD patients, but M1/G-protein coupling was considerably decreased in the frontal cortex which was linked with the progression of cognitive impairment [72]. M1 receptors being involved in modulation of cognition, and are found to be the therapeutic targets for AD treatment. M1 agonists may alter the proteolysis of APP resulting in significantly reduced AB levels in cortex and hippocampus [73]. The M1/M3 activation increases non amyloidogenic pathway of APP processing. Therefore, the hypofunction of these receptors might increase $A\beta$ generation leading to severe AD pathology [74]. It is reported that M1 receptor signal transduction-related functions are compromised in AD [72, 75, 76] but in another study cortical M1 receptor was increased in AD [77]. M2 receptor is increased in AD suggesting, that the presynaptic M2 receptors are preserved or up-regulated resulting in reduction in neurotransmitter release [71], but in another study M2 receptors were significantly reduced in hippocampus of AD brains [78]. Learning and memory associated with fear conditioning was declined in M3 knock-out mice [79]. M3 receptor levels were found to be decreased in the entorrhinal cortex and hippocampus [78]. Alteration in mAChR subtypes has important implications in cognitive control as well as ACh regulation. Impaired M1 receptor exacerbates AD-related cognitive decline, while disruption of M2/M4 receptors negatively regulates the ACh release as well the cognitive learning.

Among the cholinergic markers, the activity of choline acetyltransferase (ChAT) is greatly reduced in AD and is related to severity of disease [62, 80, 81]. Reduction in ChAT activity has been correlated with the numbers of neurofibrillary tangles in AD [82], suggesting a strong relationship between cholinergic transmission and AD. Loss of cholinergic neurons causes a significant reduction of ChAT activity (up to 95%) in the neocortex [83, 84] and hippocampus [85], that has been related to a marked decrease of ACh levels in these regions [86].

Agents that block mAChRs disturb cognitive functions and cause temporary loss of short term memory [12, 87-89]. Recently, muscarinic receptor family has shown clinical effectiveness in recovering cognitive impairment associated with AD [90, 91]. The role of Gq-coupled M1 and M3 receptors has been confirmed in modifying non-amyloidogenic pathway of APP processing while Gi-coupled M2 and M4 receptors promote amyloidogenic pathway [54, 92]. It reflects that the muscarinic receptors are among the excellent targets for the treatment of AD.

MUSCARINIC AGONISTS FOR ALZHEIMER'S DISEASE TREATMENT

mAChR are considered to be of prime importance among key drug discovery targets for the cure of AD (Table 1) [93-95]. Muscarinic agonists delay the progression of AD by decreasing β -amyloid aggregation, reducing tau phosphorylation and improving cognitive behavior [73, 94, 96, 97]. Cholinergic system modulation improves synaptic function by increasing synaptic protein expression at various stages of disease [98], improves synaptic plasticity [99] and also suppresses inflammatory response [100].

M1 receptor is considered to be an important therapeutic target as it is abundant in the hippocampus and cerebral cortex, where the cholinergic hypofunction is well-defined in AD. This receptor subtype is concerned with short-term memory [13]. Moreover, stimulation of M1 muscarinic receptors reduces the production of β -amyloid by activating α -secretase as this leads to non-amyloidogenic pathway [101].

NICOTINIC ACETYLCHOLINE RECEPTORS

The nicotinic ACh receptors (nAChRs) are ligand gated ion channels. These receptors are formed by assembly of five subunits, around a central pore, in homomeric or heteromeric conformation [106]. The standard subunits include $\alpha 2 - \alpha 9$ and $\beta_{2-\beta_{4}}$ [107]. The neural subunits capable of forming heteromeric nAChRs with $\alpha\beta$ subunit combinations are α^2 - $\alpha 6$ and $\beta 2$ - $\beta 4$. Whereas subunits $\alpha 7$ - $\alpha 9$ make functional homomeric nAChRs [106]. The β subunits alone are incapable of forming functional receptor while $\alpha 2-\alpha 6$ alone can only make receptors with very weak response to ligand. This indicates that only the combination of α and β receptors make a fully functional receptor [108]. It is also reported that α subunits contain agonist recognition and binding site. The β subunits are helpful to increase affinity towards agonist and to stabilize the whole receptor [108]. Individual nAChR subunits can combine in different stichiometries but, $(\alpha 7)_5$, $(\alpha 4)_2(\beta 2)_3$ and $(\alpha 4)_3(\beta 2)_2$ nAChR are the most common receptor types in central nervous system [109]. Each subunit of nAChRs contains four transmembrane domains (M1-M4), two hydrophilic extracellular segments (N- and C- terminals) and an intracellular loop between M3 and M4 transmembrane domains [110]. This intracellular loop has putative phosphorylation sites [110]. The second transmembrane domain, M2, aligns along the centre to make the central pore [106]. Two molecules of ligand must bind to the receptor to allow opening of the central pore and permeation of cations (Ca^{+2}, Na^{+}, K^{+}) [110].

LOCATION AND FUNCTION OF NICOTINIC ACETYLCHOLINE RECEPTORS

Among many different possible combinations of the nicotinic receptor subunits the $\alpha 4\beta 2$ and $\alpha 7$ type receptors are most abundant in the mammalian brain [107]. In the rodent brain $\alpha 4\beta 2$ receptor type is reported to be the most abundant of all the nicotinic receptor subtypes and is found to be expressed in all layers of cerebral cortex and hippocampus in rat [111]. The $\alpha 7$ receptor subtype is highly expressed by basal forebrain neurons, and the innervations projecting towards hippocampus from basal forebrain [112, 113].

The function of the nAChRs is regulated by the binding of ligand, nicotine or ACh, to the receptor. ACh binds to extracellular N-terminal domain of receptor at boundary between α and non- α subunits [114]. The binding causes an influx of different cations, especially Ca⁺² ion, inside the cell [115]. This nAChR mediated Ca⁺² entry causes a marked increase in intracellular Ca⁺² concentration, which is adequate to initiate Ca⁺² sensitive processes [110].

INVOLVEMENT IN ALZHEIMER'S DISEASE

There are contradictory reports about the expression of most abundant nAChR subtypes, $\alpha 4\beta 2$ and $\alpha 7$, during AD in different brain areas. Some studies report an increase in the expression of nAChRs in AD [116-118] while others report nAChRs decrease during progression of AD [108, 119-121]. It was reported in a study that at mRNA level, the expression of both receptor types remains the same in control and AD patient brain cortices while at protein level there is a 30% decrease in the expression of $\alpha 4\beta 2$ and $\alpha 7$ receptor subtypes. The difference observed in $\alpha 4\beta 2$ and $\alpha 7$ receptor expression at protein level and mRNA level might be due to a change at translational or post translational level during nAChR biosynthesis [122], but the exact mechanism is not known and needs to be investigated. Similar observation was made when autopsy samples of cerebral cortex from AD patients were studied [123]. But they reported a 40% decrease in $\alpha 4$ receptor expression and 17% decrease in a7 receptor expression. Decreased expression of nAChRs causes a

M1 Agonist	Therapeutic Effects	Refs.
AF150(S)	Decreases β-amyloid levels in CSF	[102]
AF267B	Restores cognitive and behavioural impairments, decreases A β aggregation and tau hyperphosphorylation	[61]
AF102B	Decreases CSF β-amyloid level in AD	[84]
TBPB	Activation of non-amyloidogenic pathway for APP processing and reduced Aß synthesis in vitro.	[81]
BQCA	Restores discrimination-based learning in a transgenic mouse model of AD, control non-amyloidogenic pathway of APP in vitro	[103]
AC-260584	Improves cognitive performance in animal model	[104]
77-LH-28-1	An agonist at rat hippocampal M1 receptors, increases cell firing	[105]

 Table 1.
 M1 Allosteric modulators/agonists under clinical trials.

AD: Alzheimer's disease, APP: Amyloid Precursor Protein, CSF: Cerebrospinal fluid, M1: Muscarinic receptor 1, BQCA: Benzylquinolone carboxylic acid.

deficiency in the binding sites for nicotine and ACh which leads to cognitive deficit in AD [123]. Contrary to these observations that report a decrease in nicotinic receptor expression, another study [124] reported an increase in α 7 mRNA expression in patients of AD, while no difference in the expression of α 4 was observed by them. An increased expression of nicotinic receptor protein was also reported in animal models [125, 126]. This difference in expression studies might be due to age dependent biphasic effect on nicotinic receptor expression, during AD, in animal model. As it is reported that nAChR expression increases 3-4 fold at 9 months of age and then a decrease in expression is observed at 12 months of age [127].

It is reported that in early AD the initial A β aggregation overlaps with the α 7 receptor expression in basal forebrain cholinergic system [128]. This early co-localization of the α 7 receptors and A β may be due to high affinity binding between these two components [117, 129]. Receptor binding experiments have also shown co-precipitation of A β 1-42 and α 7 nAChRs [113]. Because these co-precipitates are resistant to detergent treatment which shows that a high affinity binding takes place between A β 1-42 and α 7 nAChRs [117]. A β 1-42 also binds to heteromeric nAChRs but with 5000 times lesser affinity as compared to α 7- and is known to block whole cell and single channel currents in CA1 stratum radiatum interneurons in rat hippocampal slices [130].

It is now known that $A\beta 1-42$ at high (nM) concentrations leads to nAChR inactivation and thus can disrupt synaptic plasticity and cognitive functions [131]. However, at low (pM) concentration the A β 1-42 plays neuromodulatory role and activate nAChRs, thus modulating synaptic plasticity and enhancing cognitive functions [132]. Low levels of A β and short exposure time help to activate different neuromodulatory pathways in nAChR dependant manner but extended exposure at higher doses causes a dysregulation of these signal transduction pathways, possibly through desensitization of receptor, leading to cell death which in turn impairs learning and memory [133].

α7 nAChRs are reported to be involved in induction of long term potentiation (LTP) and long term depression (LTD), two forms of synaptic plasticity. As reported by Gu and Yakel schaffer collateral (SC) CA1 plasticity is dependent on α7 receptor. In this experiment when septal cholinergic input was activated 100ms or 10ms prior to SC stimulation caused induction of LTP or LTD which was blocked by α7 antagonist MLA but not by non-α7 antagonist DHβE. Moreover, this synaptic plasticity was disrupted by 10nM or 100nM Aβ. These results suggest that inactivation of α7 receptors by Aβ has negative effects on synaptic plasticity [134] which results in impaired learning and memory.

The fact that α 7 nAChRs are present in glial cells, particularly astrocytes, suggests that these receptors also have an important role in inflammation process. Work by Nagele *et al.* show that α 7 nAChRs and A β were found to be intensely co-localized with green fluorescence activated protein (GFAP) positive (activated) astrocytes in AD brains. Since the authors also found ChAT, they proposed a model that α 7 and A β are phagocytized by activated astrocytes in

the vicinity of neural remnants. As a result astrocyte viability is compromised with increased accumulation of neuronal debris in astrocytes. This results in selective lysis of the astrocytes lead to astrocyte derived amyloid plaque formation [135]. The A β peptide is also reported to activate caspase 3 and induce astrocyte apoptosis [136] leading to higher rate of astrocyte apoptosis as compared to neuronal cells [137]. Thus, apoptosis of astrocytes may positively contribute to pathogenesis of AD [138].

α7 NICOTINIC ACETYLCHOLINE RECEPTOR TARGETING DRUGS FOR TREATMENT OF ALZHEIMER'S DISEASE

Although there is no preventive treatment available for AD but there is a continuous urge in the scientific community for the search of novel therapeutic strategies that can alleviate pathological symptoms of AD. The α 7 nAChRs are pentameric ligand gated ion channels with selective permeability to Na⁺ and Ca⁺² ions [139]. These receptors have exceptionally high Ca⁺² permebility [140] as compared to other ligand gated ion channels. The Ca⁺² acts as a second messenger and activates many signaling pathways in the cell and also mediates neurotransmitter release [141]. Due to high vulnerability of cholinergic neurons (specifically those having high amount of α 7 nAChRs), high affinity binding between α 7 nAChRs these receptors have become an attractive target for the treatment of AD [140].

For drug target both α 7 nAChR agonists and antagonists are under investigation. The α 7 nAChR agonists are of more therapeutic interest for the pharmacologist. Antagonists of α 7 nAChRs have lower practical impact as compared to its agonists [109]. Table **2** lists some of the agonists, targeting α 7 nAChR, under clinical trials for treatment of AD.

AMYLOID PRECURSOR PROTEIN AND ITS POST TRANSLATIONAL MODIFICATIONS AND PROCESSING IN ALZHEIMER'S DISEASE

Although the relationship between protein dysfunction and neurodegeneration remains elusive [146-148], protein aggregation has evolved as an emerging theme in diseases such as AD. Several heavily debated hypotheses exist to sequentially interlink all these phenomena under one event; aggregation of toxic A β is considered to be the driving force of AD pathology. A β , peptides of 40 or 42 amino acids, are derived from the sequential proteolytic cleavage of β amyloid precursor protein (APP). Two mutually exclusive pathways exist for proteolytic processing of APP; while cleavage at residue Lys 16 by α -secretase results in the generation of soluble APP (sAPP) peptides, altered cleavage by β - and γ -secretases results in the formation of the 40-42 amino acid which coalesces to form insoluble, extracellular A β [149].

With recent paradigm shift, post-translational modifications (PTMs) of pathology associated proteins have become a valuable tool in the evaluation of the structural and functional alterations governing the neurodegenerative diseases [150]. PTMs significantly contribute to proteome expansion with each variant displaying a starkly different

a7 Nicotinic Acetylcholine Receptor Agonists			
Name	Therapeutic effects	Refs.	
EVP-6124	Activates a7 nAChRs and is used for the treatment of mild to moderate AD, under phase 3 clinical trials	[142]	
AZD-0328	Activates α7 nAChRs and enhances dopamine release. Used for the treatment of AD and is under clinical testing.	[143]	
ABT-107	Treatment of AD and cognitive deficits associated with schizophrenia, under testing, not comercially available	[144]	
GTS-21	Treatment of AD and cognitive deficits associated with schizophrenia, experimental testing for anti-inflammatory potency, under clinical testing	[145]	

Table 2. a7 Nicotinic acetylcholine receptor agonists and antagonists under clinical testing for the treatment of AD.

property such as phosphorylation of Tau protein in neurofibrillary tangles (NFT) or alternative cleavage of posttranslationally modified APP into different forms of A β [150]. It comes in good agreement with studies implicating aberrant PTMs in AD pathogenesis [151].

APP is post-translationally modified by sulfation, phosphorylation, glycosylation, including both N- and Olinked glycosylation and proteolytic processing. In fact it is the O-glycosylated version of APP that is preferentially secreted. The most interesting correlation of PTMs with AD pathophysiology is "glycosylation" whereby oligosaccharide side chains attach themselves at N&O- linked sites on the nascent APP, in the endoplasmic reticulum. Two putative Nlinked oligosaccharide attachment sites (Asn 467 and Asn 496) have been identified. Accumulating evidence proposes that only the former is occupied under normal conditions [152]. It has been suggested that the oligosaccharide side chains have a pivotal role in the protein processing. In AD, the major lesion associated proteins, APP and Tau, and their respective metabolites undergo altered N- and Oglycosylation [153].

Since $A\beta$ can be produced by cultured cells, this has left us with a powerful model system for analyzing the aberrant PTMs, leading to $A\beta$ formation in cells. Mutant Lec 8 strain, CHO cell lines are reported to have a defect in the CMP-NeuNAc transport system, which has shown an increase in asialo-oligosaccharide expression [154]. Tunicamycin and brefeldin A, soluble inhibitors of glycosylation have demonstrated to reduce APP secretion when N-glycosylation and sialylation were inhibited [152]. Altered protein glycosylation, via tunicamycin or mannosidase inhibition, has shown the disruption of axonal sorting in both in vitro and in vivo [155]. A similar result was observed with another model, when the asparagine residues, i.e. the sites of N Glycosylation, were removed [156]. McFarlane et al., [157] used mannosidase I and II inhibitors, 2-deosxymannojirimycin (dMan) and swainsonine respectively, for the investigation of the different effects of mannose and other complex sugars on APP processing. They observed that the treatment of AtT-20 mouse pituitary cells with dMan or swainsonine in vitro, prevented the N-linked sugars to mature which resulted in a significant decrease in APP secretion from the cell to the cell membrane.

Taken together these results confirm the previous lectin studies that suggested the preferential transfer of mature APP

from the perinuclear region to cell membrane, where the high-mannose containing forms were retained in the ER/Golgi complex. If the APP is retained in the perinuclear region, it may have implications in its processing and the generation of A β [158]. Hence we can say that the impairment of APP maturation of the oligosaccharide chains, causing the retention of APP in the perinuclear region leads to an uprise in A β concentration in the cell due to holoprotein buildup in cellular compartments [159].

Decreased secretion of sAPP has been associated with generation of oligomannosyl oligosaccharides mediated altered APP glycosylation state. This was observed to be coupled with a parallel increase in the deposition of the cellular protein within the cell [154, 160]. Conversely, conjugation of terminal sialic acid residues to the glycan was shown to increase sAPP levels [161, 162]. Activation of Protein kinase C (PKC) has been widely reported to alter APP processing [163, 164]. Sialyltransferase enzyme transfected cells have demonstrated a direct relationship between the sialylation potential of APP and the fold stimulation of sAPP, following PKC activation [154]. Mutations altering the APP glycosylation state have been linked to an increased A β 42/A β 40 ratio, such as Swedish and London mutations. Both of these mutations account for altered N-glycosylation of APP, with an increased content of bisecting GlcNAc [165]. In accordance with this, GlcNActransferase III mRNA expression has reportedly been increased in AD brains [161].

Several studies have reported the presence of Oglycosylation sites and their functional role in APP [166-169]. Though elusive, the role of O-Glycosylastion has been proposed in proteolytic processing of APP by α -secretase, β secretase and γ -secretase. In addition, studies have shown that it is the O-glycosylated APP that is preferentially secreted [168, 170]. Tyrosine O-glycosylation has been reported in A β 1–15 to Ab1–20 but not in full-length (A β 1– 38 to A β 1–42) A β fragments [167]. An increase in the shorter AB fragments cerebrospinal fluid (CSF) from AD patients and non-demented controls showed to carry the tyrosine-linked glycan in AD patients. APP is also O-GlcNAcylated [171], which affects proteolytic processing of APP, thereby increasing sAPP and decreasing $A\beta$ secretion [172]. These results suggest that the post-translational modification of APP by glycosylation is a key event in determining the processing of the protein and may have significant implications in understanding the initial

deposition and kinetics of amyloid aggregation in a pathological situation like AD.

ROLE OF CHOLINERGIC SYSTEM IN APP POST TRANSLATIONAL MODIFICATIONS

It has been established that APP, γ -secretase and altered glycosylation can lead towards misfolding and AD pathology. Moreover, cholinergic system has a pivotal role in learning and memory, and its deficits are also part of AD pathology. But the relationship between APP and cholinergic neurons has not been elucidated. However, recent evidence obtained from mice and cell lines implies that the cognitive decline occurs due to loss of cholinergic neurons and APP processing [173]. Experimental evidence obtained from the studies on these model organisms suggests that activity of cholinergic neurotransmission might have an impact on APP processing. Moreover, the APP phosphorylation on threonine 668 (P-APP) may also influence the APP metabolism. The Aß production significantly reduced due to mutation or inhibition of T668 kinase inhibitors. It is suggested that the T668 phosphorylation may facilitate the β -secretase (BACE) 1 cleavage of APP to increase $A\beta$ generation [174]. In addition, p35- and p25-mediated Cdk5 activities lead to discrete APP (Thr668) phosphorylation, where the overexpression of both p35 and p25, increases the secretion of A_β, as well sAPP (beta), and sAPP (alpha) [175]. The APP T688 phosphorylation also regulates the nuclear translocation of APP intracellular domain that also contributes towards neurodegeneration [176].

Alterations in APP metabolism significantly aids in the long-lasting effects of AChE inhibitors. However, complete inhibition is lethal as the natural physiology of the neuron will also be inhibited [177]. The potentiation of the central cholinergic system can be a potential tool and a promising strategy for therapeutics by modulating AChE that ultimately increases the ACh concentration in the brain [178]. However, despite the accumulating evidences, the interaction between cholinergic neurotransmission and APP processing and the physiological significance of PTM events remain speculative and still needs further exploration.

POST TRANSLATIONAL MODIFICATIONS AND TAU

Several studies have suggested and proposed two major hypotheses about the relationship of tau and A β through which tau may facilitate A β -induced impairments [179]. According to the first hypothesis, physiological forms of tau may cause abnormal neural network activity *via* diverse pathogenic triggers [180, 181], while the second hypothesis proposed that the A β might change the PTM or distribution of tau, making it an active mediator of A β -induced neuronal dysfunction [182]. A detailed characterization of tau PTMs may assist to strategize the plausible mechanisms to combat the consequences of the pathological processes associated with tau.

Although the phosphorylation of tau is well understood, with identified phosphorylated sites *i.e.* Ser-68, Thr-69, and Thr-71 [183], it also involves cross talk among diverse and sometimes competing PTMs which have still not been well studied including O-glycosylation, ubiquitination, acetylation and methylation [179].

Methylation of tau on lysine and arginine residues has been recently reported [184, 185]; however, the functional effects are still unknown. Methylation inhibits tau aggregation by increasing the amount of proteins required to form aggregates by increasing tau's dissociation rate from fibrils and decreasing the flexible extension rate. Moreover, methylation also delays the aggregation rate by diminishing the filament nucleation, which is the rate limiting step in the formation of neurofibrillary tangles [184]. Interestingly, the increased demethylation of protein phosphatase 2A (PP2A) (L309) results in reduced PP2A activity in AD brain, mediated by A β overproduction (or estrogen deficiency in mice), leading to compromised dephosphorylation of abnormally hyperphosphorylated tau [186].

In addition, tau phosphorylation which is directly mediated by phosphotransferases, there is a complex regulatory control on tau aggregation by competing modifications like O-linked β -N-acetylglucosaminylation (OGlcNAcylation). The OGlcNAcylation significantly regulates tau phosphorylation by decreasing the phosphorylation levels, thus depressing the neurofibrillary lesion formation and aggregation [187].

Furthermore, the ubiquitination of tau at Lys-6, Lys-11 and Lys-48 also modulates the intracellular tau levels in AD brains [188]. Studies have also shown significant regulating effects of increased acetylation on tau in AD which also counteracts the ubiquitination and degradation of phosphorylated tau [189].

MUSCARINIC ACETYLCHOLINE RECEPTOR AND TAU HYPERPHOSPHORYLATION

Post-translational modifications play an important role in the structure and function of GPCRs. Although N-linked glycosylation is the most common posttranslational modification of GPCRs but limited data is available regarding their role in mAChRs [190]. Among all mAChRs, M3 undergoes few important modifications, such as Nglycosylation and disulfide bond formation [191]. The absence of M3 N-glycosylation promotes receptor trafficking impairment, generates ER stress and thus leads to an increased susceptibility for cell disruption [191]. Impaired neurotransmission which is highly evident in many neurodegenerative disorders is also perturbed due to lack of N-glycosylation of M3 [192]. On the contrary, Nglycosylation of the M2 is not required for cell surface localization or ligand binding [193].

It has been observed that activation of M1 receptor decreases tau phosphorylation. Fisher *et al.*, demonstrated the plausible mechanism of M1-mediated decrease in tau phosphorylation [194]. M1- agonists improved cognition and behavior, decreased the hyperphosphorylated tau and the number of neurons containing aggregated tau and paired helical filaments (PHFs), and decreased the inflammation [194].

NICOTINIC ACETYLCHOLINE RECEPTOR AND TAU HYPERPHOSPHORYLATION

The α 7-A β binding activates α 7 nAChR and increases tau hyperphosphorylation *via* ERK-MAPK and JNK-1-MAPK activation [195]. The ERK-MAPK activation leads to

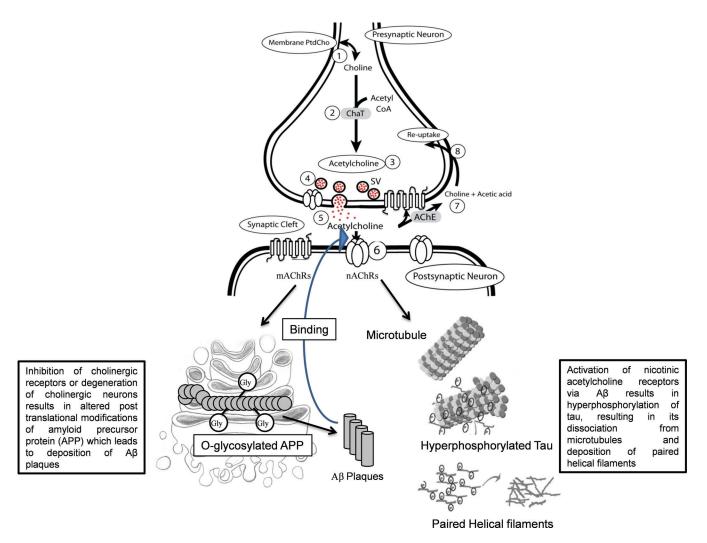


Fig. (2). Cholinergic Synapse and PTMs of APP and Tau. Abbreviations: ChaT: Choline acetylTransferase, PtdCho: Phosphotidylcholine, SV: Synaptic vesicles, AChE: Acetylcholine esterase, ACh: acetylcholine, mAChR: muscarinic ACh receptor, nAChR: nicotinic ACh receptor, 1) PtdCho provides the choline precursor and acetyl moiety from acetyl CoA, 2) in the presence of ChaT 3) the ACh is synthesized, 4) Storage of ACh in synaptic vesicles, 5) Release of ACh in the synaptic cleft, 6) Action of ACh on postsynaptic cholinergic receptor, 7) Degradation of ACh *via* AChE into choline and acetate ion, 8) Reuptake of choline into the presynaptic nerve terminal by choline transporter.

phosphorylation of two proline directed MAPK-targeted serine and threonine residues (S202, T181) while the activation of JNK-1-MAPK pathway phosphorylates the T231 along with S202 and T181 residues on tau [195]. Activation of α 7 nAChR *via* A β mediates the phosphorylation of GSK 3 β at tyrosine 216 which also results in phosphorylation of S202 on tau [196]. The phosphorylated S202 and T181 residues can lead to microtubule instability as they are involved in binding kinetics of tau-microtubule, resulting in the formation of NFTs [197]. The A β induced tau hyperphosphorylation can be blocked by α 7 nAChR selective antagonist, methyllycaconitine (MLA) [133].

Moreover, increase in tau phosphorylation was observed after nAChR activation *via* application of nAChR agonists. It is evident that chronic nicotine treatment in transgenic model of AD causes an upregulation of nAChRs which results in activation of p38 MAP kinase which in turn phosphorylates tau and exacerbates tau pathology [198]. Similarly, increase in tau phosphorylation was also observed after activation of nAChRs *via* AChE inhibitors and nAChR agonists, however, this requires nAChR mediated Ca⁺² entry inside the cell as Ca⁺² removal by ethylene glycol tetra acetic acid (EGTA) prevents increased tau hyperphosphorylation [199]. Taken together these studies highlight the nAChR-A β interaction, mediating tau hyperphosphorylation and neurofibrillary tangle formation (Fig. **2**).

CONCLUSION

AD, despite been extensively studied for the last many years, is a constellation of consequences which is still to be explored, to devise effective therapeutic strategies. The dysfunctional cholinergic system in AD has been linked to ACh deficits, which emphasizes therapeutic research to focus on effective approaches for maintaining its level in brain. Muscarinic cholinergic receptors play a significant role in regulating CNS circuits, particularly involving learning and memory. Muscarinic receptor subtypes with different distributions in CNS, provides the opportunity to take each receptor as a drug target. The available cholinergic compounds lack the subtype-specificity and effectiveness that favors the side effects and may influence cognitive effects because of weak or differing actions. However, some selective allosteric modulators of ACh have demonstrated therapeutic potential that can be used as a better therapeutic approach. Additionally, a rather unique aspect of involvement of PTMs and their potential effect on muscarinic and nicotinic cholinergic receptors, gives a new dimension to study the pathological consequences where these cholinergic receptors are involved. The distinct and influential association of several competing PTMs with cholinergic receptors provide a detailed understanding of complex regulation of cholinergic system under several PTMs that may govern the diverse pathological mechanisms and associated consequences in AD. Future studies are required to focus on the specific role of cholinergic neurotransmission and PTMs in AD, to further validate drug targets and effective therapeutics.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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REFERENCES

- Jucker, M.; Walker, L.C. Neurodegeneration: Amyloid-β pathology induced in humans. *Nature*, 2015, 525(7568), 193-194. [http://dx. doi.org/10.1038/525193a] [PMID: 26354478]
- [2] Selkoe, D.J.; Schenk, D. Alzheimers disease: molecular understanding predicts amyloid-based therapeutics. Annu. Rev. Pharmacol. Toxicol., 2003, 43, 545-584. [http://dx.doi.org/ 10.1146/annurev.pharmtox.43.100901.140248] [PMID: 12415125]
- Zilka, N.; Novak, M. The tangled story of Alois Alzheimer. Bratisl. Lek Listy (Tlacene Vyd), 2006, 107(9-10), 343-345. [PMID: 17262985]
- [4] Fu, W.; Jhamandas, J.H. β-amyloid peptide activates non-α7 nicotinic acetylcholine receptors in rat basal forebrain neurons. J. Neurophysiol., 2003, 90(5), 3130-3136. [http://dx.doi.org/10.1152/ jn.00616.2003] [PMID: 12890800]
- [5] Glenner, G.; Wong, C. Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Alzheimer Dis. Assoc. Disord.*, **1988**, 2(2), 134.
- [6] Zheng, W-H.; Bastianetto, S.; Mennicken, F.; Ma, W.; Kar, S. Amyloid β peptide induces tau phosphorylation and loss of cholinergic neurons in rat primary septal cultures. *Neuroscience*, **2002**, *115*(1), 201-211. [http://dx.doi.org/10.1016/S0306-4522(02) 00404-9] [PMID: 12401334]
- [7] Grundke-Iqbal, I.; Iqbal, K.; Tung, Y-C.; Quinlan, M.; Wisniewski, H.M.; Binder, L.I. Abnormal phosphorylation of the microtubuleassociated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc. Natl. Acad. Sci. USA*, **1986**, *83*(13), 4913-4917. [http://dx. doi.org/10.1073/pnas.83.13.4913] [PMID: 3088567]
- [8] Walsh, D.M.; Selkoe, D.J. A beta oligomers a decade of discovery. J. Neurochem., 2007, 101(5), 1172-1184. [http://dx.doi. org/10.1111/j.1471-4159.2006.04426.x] [PMID: 17286590]
- [9] Akiyama, H.; Barger, S.; Barnum, S.; Bradt, B.; Bauer, J.; Cole, G.M.; Cooper, N.R.; Eikelenboom, P.; Emmerling, M.; Fiebich, B.L.; Finch, C.E.; Frautschy, S.; Griffin, W.S.; Hampel, H.; Hull, M.; Landreth, G.; Lue, L.; Mrak, R.; Mackenzie, I.R.; McGeer,

P.L.; OBanion, M.K.; Pachter, J.; Pasinetti, G.; Plata-Salaman, C.; Rogers, J.; Rydel, R.; Shen, Y.; Streit, W.; Strohmeyer, R.; Tooyoma, I.; Van Muiswinkel, F.L.; Veerhuis, R.; Walker, D.; Webster, S.; Wegrzyniak, B.; Wenk, G.; Wyss-Coray, T. Inflammation and Alzheimers disease. *Neurobiol. Aging*, **2000**, *21*(3), 383-421. [http://dx.doi.org/10.1016/S0197-4580(00)00124-X] [PMID: 10858586]

- [10] Terry, R.D.; Masliah, E.; Salmon, D.P.; Butters, N.; DeTeresa, R.; Hill, R.; Hansen, L.A.; Katzman, R. Physical basis of cognitive alterations in Alzheimers disease: synapse loss is the major correlate of cognitive impairment. *Ann. Neurol.*, **1991**, *30*(4), 572-580. [http://dx.doi.org/10.1002/ana.410300410] [PMID: 1789684]
- [11] DeKosky, S.T.; Scheff, S.W. Synapse loss in frontal cortex biopsies in Alzheimers disease: correlation with cognitive severity. *Ann. Neurol.*, **1990**, *27*(5), 457-464. [http://dx.doi.org/10.1002/ana. 410270502] [PMID: 2360787]
- [12] Coyle, J.T.; Price, D.L.; DeLong, M.R. Alzheimers disease: a disorder of cortical cholinergic innervation. *Science*, **1983**, 219(4589), 1184-1190. [http://dx.doi.org/10.1126/science.6338589]
 [PMID: 6338589]
- [13] Levey, A.I. Muscarinic acetylcholine receptor expression in memory circuits: implications for treatment of Alzheimer disease. *Proc. Natl. Acad. Sci. USA*, **1996**, *93*(24), 13541-13546. [http://dx. doi.org/10.1073/pnas.93.24.13541] [PMID: 8942969]
- Geula, C.; Mesulam, M.M. Cortical cholinergic fibers in aging and Alzheimers disease: a morphometric study. *Neuroscience*, **1989**, 33(3), 469-481. [http://dx.doi.org/10.1016/0306-4522(89)90399-0]
 [PMID: 2636703]
- [15] Mufson, E.J.; Ginsberg, S.D.; Ikonomovic, M.D.; DeKosky, S.T. Human cholinergic basal forebrain: chemoanatomy and neurologic dysfunction. J. Chem. Neuroanat., 2003, 26(4), 233-242. [http://dx. doi.org/10.1016/S0891-0618(03)00068-1] [PMID: 14729126]
- Pearson, R.C.; Sofroniew, M.V.; Cuello, A.C.; Powell, T.P.; Eckenstein, F.; Esiri, M.M.; Wilcock, G.K. Persistence of cholinergic neurons in the basal nucleus in a brain with senile dementia of the Alzheimers type demonstrated by immunohistochemical staining for choline acetyltransferase. *Brain Res.*, 1983, 289(1-2), 375-379. [http://dx.doi.org/10.1016/0006-8993(83) 90046-X] [PMID: 6362777]
- [17] Bowen, D.M.; White, P.; Spillane, J.A.; Goodhardt, M.J.; Curzon, G.; Iwangoff, P.; Meier-Ruge, W.; Davison, A.N. Accelerated ageing or selective neuronal loss as an important cause of dementia? *Lancet*, **1979**, *1*(8106), 11-14. [PMID: 83462]
- [18] Sarter, M.; Bruno, J.P. Developmental origins of the age-related decline in cortical cholinergic function and associated cognitive abilities. *Neurobiol. Aging*, 2004, 25(9), 1127-1139. [http://dx.doi. org/10.1016/j.neurobiolaging.2003.11.011] [PMID: 15312959]
- [19] Ridley, R.M.; Murray, T.K.; Johnson, J.A.; Baker, H.F. Learning impairment following lesion of the basal nucleus of Meynert in the marmoset: modification by cholinergic drugs. *Brain Res.*, **1986**, *376*(1), 108-116. [http://dx.doi.org/10.1016/0006-8993(86)90904-2] [PMID: 3087582]
- [20] DeKosky, S.T.; Harbaugh, R.E.; Schmitt, F.A.; Bakay, R.A.; Chui, H.C.; Knopman, D.S.; Reeder, T.M.; Shetter, A.G.; Senter, H.J.; Markesbery, W.R. Cortical biopsy in Alzheimers disease: diagnostic accuracy and neurochemical, neuropathological, and cognitive correlations. *Ann. Neurol.*, **1992**, *32*(5), 625-632. [http://dx.doi.org/ 10.1002/ana.410320505] [PMID: 1360195]
- [21] Minger, S.L.; Esiri, M.M.; McDonald, B.; Keene, J.; Carter, J.; Hope, T.; Francis, P.T. Cholinergic deficits contribute to behavioral disturbance in patients with dementia. *Neurology*, **2000**, 55(10), 1460-1467. [http://dx.doi.org/10.1212/WNL.55.10.1460] [PMID: 11094098]
- [22] Davies, P.; Maloney, A.J. Selective loss of central cholinergic neurons in Alzheimers disease. *Lancet*, **1976**, 2(8000), 1403. [http:// dx.doi.org/10.1016/S0140-6736(76)91936-X] [PMID: 63862]
- [23] Kato, G.; Agid, Y. Acetylcholine receptors. Nouv. Presse Med., 1979, 8(29), 2407-2411. [PMID: 226931]
- [24] Lefkowitz, R.J. Seven transmembrane receptors: a brief personal retrospective. *Biochim. Biophys. Acta*, 2007, 1768(4), 748-755. [http:// dx.doi.org/10.1016/j.bbamem.2006.11.001] [PMID: 17173855]
- [25] Hulme, E.C.; Curtis, C.A.; Wheatley, M.; Aitken, A.; Harris, A.C. Localization and structure of the muscarinic receptor ligand binding site. *Trends Pharmacol. Sci.*, **1989**(Suppl.), 22-25. [PMID: 2694518]

- [26] Koshimizu, H.; Leiter, L.M.; Miyakawa, T. M4 muscarinic receptor knockout mice display abnormal social behavior and decreased prepulse inhibition. *Mol. Brain*, 2012, 5, 10. [http://dx. doi.org/10.1186/1756-6606-5-10] [PMID: 22463818]
- [27] Bonner, T.I.; Buckley, N.J.; Young, A.C.; Brann, M.R. Identification of a family of muscarinic acetylcholine receptor genes. *Science*, **1987**, *237*(4814), 527-532. [http://dx.doi.org/10. 1126/science.3037705] [PMID: 3037705]
- Hulme, E.C.; Birdsall, N.J.; Buckley, N.J. Muscarinic receptor subtypes. Annu. Rev. Pharmacol. Toxicol., 1990, 30, 633-673. [http://dx.doi.org/10.1146/annurev.pa.30.040190.003221] [PMID: 2188581]
- [29] Wess, J. Molecular biology of muscarinic acetylcholine receptors. *Crit. Rev. Neurobiol.*, **1996**, 10(1), 69-99. [http://dx.doi.org/10. 1615/CritRevNeurobiol.v10.i1.40] [PMID: 8853955]
- [30] Wess, J.; Liu, J.; Blin, N.; Yun, J.; Lerche, C.; Kostenis, E. Structural basis of receptor/G protein coupling selectivity studied with muscarinic receptors as model systems. *Life Sci.*, **1997**, *60*(13-14), 1007-1014. [http://dx.doi.org/10.1016/S0024-3205(97)00041-6] [PMID: 9121341]
- [31] Wess, J. Molecular basis of muscarinic acetylcholine receptor function. *Trends Pharmacol. Sci.*, **1993**, *14*(8), 308-313. [http://dx. doi.org/10.1016/0165-6147(93)90049-P] [PMID: 8249149]
- [32] Matsui, M.; Yamada, S.; Oki, T.; Manabe, T.; Taketo, M.M.; Ehlert, F.J. Functional analysis of muscarinic acetylcholine receptors using knockout mice. *Life Sci.*, 2004, 75(25), 2971-2981. [http://dx.doi.org/10.1016/j.lfs.2004.05.034] [PMID: 15474550]
- [33] Caulfield, M.P.; Birdsall, N.J. International Union of Pharmacology. XVII. Classification of muscarinic acetylcholine receptors. *Pharmacol. Rev.*, **1998**, 50(2), 279-290. [PMID: 9647869]
- [34] Lanzafame, A.A.; Christopoulos, A.; Mitchelson, F. Cellular signaling mechanisms for muscarinic acetylcholine receptors. *Recept. Channels*, 2003, 9(4), 241-260. [http://dx.doi.org/10.1080/ 10606820308263] [PMID: 12893537]
- [35] Bartus, R.T.; Dean, R.L., III; Beer, B.; Lippa, A.S. The cholinergic hypothesis of geriatric memory dysfunction. *Science*, **1982**, 217 (4558), 408-414. [http://dx.doi.org/10.1126/science.7046051] [PMID: 7046051]
- [36] Bartus, R.T.; Johnson, H.R. Short-term memory in the rhesus monkey: disruption from the anti-cholinergic scopolamine. *Pharmacol. Biochem. Behav.*, **1976**, 5(1), 39-46. [http://dx.doi.org/ 10.1016/0091-3057(76)90286-0] [PMID: 825880]
- [37] Everitt, B.J.; Robbins, T.W. Central cholinergic systems and cognition. Annu. Rev. Psychol., 1997, 48, 649-684. [http://dx.doi.org/10.1146/ annurev.psych.48.1.649] [PMID: 9046571]
- [38] Howe, A.R.; Surmeier, D.J. Muscarinic receptors modulate N-, P-, and L-type Ca²⁺ currents in rat striatal neurons through parallel pathways. J. Neurosci., **1995**, 15(1 Pt 1), 458-469. [PMID: 7823150]
- [39] Levey, A.I.; Kitt, C.A.; Simonds, W.F.; Price, D.L.; Brann, M.R. Identification and localization of muscarinic acetylcholine receptor proteins in brain with subtype-specific antibodies. *J. Neurosci.*, **1991**, *11*(10), 3218-3226. [PMID: 1941081]
- [40] Crook, J.M.; Tomaskovic-Crook, E.; Copolov, D.L.; Dean, B. Low muscarinic receptor binding in prefrontal cortex from subjects with schizophrenia: a study of Brodmanns areas 8, 9, 10, and 46 and the effects of neuroleptic drug treatment. Am. J. Psychiatry, 2001, 158(6), 918-925. [http://dx.doi.org/10.1176/appi.ajp.158.6.918] [PMID: 11384900]
- [41] Anagnostaras, S.G.; Murphy, G.G.; Hamilton, S.E.; Mitchell, S.L.; Rahnama, N.P.; Nathanson, N.M.; Silva, A.J. Selective cognitive dysfunction in acetylcholine M1 muscarinic receptor mutant mice. *Nat. Neurosci.*, 2003, 6(1), 51-58. [http://dx.doi.org/10.1038/nn992] [PMID: 12483218]
- [42] Davis, A.A.; Fritz, J.J.; Wess, J.; Lah, J.J.; Levey, A.I. Deletion of M1 muscarinic acetylcholine receptors increases amyloid pathology *in vitro* and *in vivo*. J. Neurosci., 2010, 30(12), 4190-4196. [http:// dx.doi.org/10.1523/JNEUROSCI.6393-09.2010] [PMID: 20335454]
- [43] Gamberini, M.T.; Bolognesi, M.L.; Nasello, A.G. The modulatory role of M2 muscarinic receptor on apomorphine-induced yawning and genital grooming. *Neurosci. Lett.*, **2012**, *531*(2), 91-95. [http://dx.doi.org/10.1016/j.neulet.2012.09.052] [PMID: 23041487]
- [44] Piggott, M.; Owens, J.; OBrien, J.; Paling, S.; Wyper, D.; Fenwick, J.; Johnson, M.; Perry, R.; Perry, E. Comparative distribution of binding of the muscarinic receptor ligands pirenzepine, AF-DX 384, (R,R)-I-QNB and (R,S)-I-QNB to human brain. J. Chem.

Neuroanat., **2002**, *24*(3), 211-223. [http://dx.doi.org/10.1016/ S0891-0618(02)00066-2] [PMID: 12297267]

- [45] Flynn, D.D.; Ferrari-DiLeo, G.; Mash, D.C.; Levey, A.I. Differential regulation of molecular subtypes of muscarinic receptors in Alzheimers disease. J. Neurochem., 1995, 64(4), 1888-1891. [http://dx.doi.org/10.1046/j.1471-4159.1995.64041888.x] [PMID: 7891119]
- [46] Levey, A.I.; Edmunds, S.M.; Heilman, C.J.; Desmond, T.J.; Frey, K.A. Localization of muscarinic m3 receptor protein and M3 receptor binding in rat brain. *Neuroscience*, **1994**, *63*(1), 207-221. [http://dx.doi.org/10.1016/0306-4522(94)90017-5] [PMID: 7898649]
- [47] Zhang, W.; Yamada, M.; Gomeza, J.; Basile, A.S.; Wess, J. Multiple muscarinic acetylcholine receptor subtypes modulate striatal dopamine release, as studied with M1-M5 muscarinic receptor knock-out mice. J. Neurosci., 2002, 22(15), 6347-6352. [PMID: 12151512]
- [48] Ohno-Shosaku, T.; Matsui, M.; Fukudome, Y.; Shosaku, J.; Tsubokawa, H.; Taketo, M.M.; Manabe, T.; Kano, M. Postsynaptic M1 and M3 receptors are responsible for the muscarinic enhancement of retrograde endocannabinoid signalling in the hippocampus. *Eur. J. Neurosci.*, **2003**, *18*(1), 109-116. [http://dx. doi.org/10.1046/j.1460-9568.2003.02732.x] [PMID: 12859343]
- [49] Zhang, W.; Basile, A.S.; Gomeza, J.; Volpicelli, L.A.; Levey, A.I.; Wess, J. Characterization of central inhibitory muscarinic autoreceptors by the use of muscarinic acetylcholine receptor knock-out mice. J. Neurosci., 2002, 22(5), 1709-1717. [PMID: 11880500]
- [50] Tzavara, E.T.; Bymaster, F.P.; Felder, C.C.; Wade, M.; Gomeza, J.; Wess, J.; McKinzie, D.L.; Nomikos, G.G. Dysregulated hippocampal acetylcholine neurotransmission and impaired cognition in M2, M4 and M2/M4 muscarinic receptor knockout mice. *Mol. Psychiatry*, 2003, 8(7), 673-679. [http://dx.doi.org/10.1038/sj.mp.4001270] [PMID: 12874603]
- [51] Vilaró, M.T.; Palacios, J.M.; Mengod, G. Localization of m5 muscarinic receptor mRNA in rat brain examined by in situ hybridization histochemistry. *Neurosci. Lett.*, **1990**, *114*(2), 154-159. [http://dx.doi.org/10.1016/0304-3940(90)90064-G] [PMID: 2395528]
- [52] Yasuda, R.P.; Ciesla, W.; Flores, L.R.; Wall, S.J.; Li, M.; Satkus, S.A.; Weisstein, J.S.; Spagnola, B.V.; Wolfe, B.B. Development of antisera selective for m4 and m5 muscarinic cholinergic receptors: distribution of m4 and m5 receptors in rat brain. *Mol. Pharmacol.*, **1993**, 43(2), 149-157. [PMID: 8429821]
- [53] Yamada, M.; Lamping, K.G.; Duttaroy, A.; Zhang, W.; Cui, Y.; Bymaster, F.P.; McKinzie, D.L.; Felder, C.C.; Deng, C.X.; Faraci, F.M.; Wess, J. Cholinergic dilation of cerebral blood vessels is abolished in M(5) muscarinic acetylcholine receptor knockout mice. *Proc. Natl. Acad. Sci. USA*, **2001**, *98*(24), 14096-14101. [http://dx.doi.org/10.1073/pnas.251542998] [PMID: 11707605]
- [54] Nitsch, R.M.; Slack, B.E.; Wurtman, R.J.; Growdon, J.H. Release of Alzheimer amyloid precursor derivatives stimulated by activation of muscarinic acetylcholine receptors. *Science*, **1992**, 258(5080), 304-307. [http://dx.doi.org/10.1126/science.1411529] [PMID: 1411529]
- [55] Buxbaum, J.D.; Oishi, M.; Chen, H.I.; Pinkas-Kramarski, R.; Jaffe, E.A.; Gandy, S.E.; Greengard, P. Cholinergic agonists and interleukin 1 regulate processing and secretion of the Alzheimer beta/A4 amyloid protein precursor. *Proc. Natl. Acad. Sci. USA*, **1992**, 89(21), 10075-10078. [http://dx.doi.org/10.1073/pnas.89.21. 10075] [PMID: 1359534]
- [56] Kosik, K.S. Alzheimers disease: a cell biological perspective. *Science*, **1992**, 256(5058), 780-783. [http://dx.doi.org/10.1126/ science.1589757] [PMID: 1589757]
- [57] Mullan, M.; Crawford, F. Genetic and molecular advances in Alzheimers disease. *Trends Neurosci.*, **1993**, *16*(10), 398-403. [http://dx.doi.org/10.1016/0166-2236(93)90007-9] [PMID: 7504354]
- [58] Selkoe, D.J. Physiological production of the beta-amyloid protein and the mechanism of Alzheimers disease. *Trends Neurosci.*, 1993, *16*(10), 403-409. [http://dx.doi.org/10.1016/0166-2236(93)90008-A] [PMID: 7504355]
- [59] Alvarez, A.; Alarcón, R.; Opazo, C.; Campos, E.O.; Muñoz, F.J.; Calderón, F.H.; Dajas, F.; Gentry, M.K.; Doctor, B.P.; De Mello, F.G.; Inestrosa, N.C. Stable complexes involving acetylcholinesterase and amyloid-beta peptide change the biochemical properties of the enzyme and increase the neurotoxicity of Alzheimers fibrils. J. Neurosci., 1998, 18(9), 3213-3223. [PMID: 9547230]

- [60] Whitehouse, P.J.; Price, D.L.; Struble, R.G.; Clark, A.W.; Coyle, J.T.; Delon, M.R. Alzheimers disease and senile dementia: loss of neurons in the basal forebrain. *Science*, **1982**, *215*(4537), 1237-1239. [http://dx.doi.org/10.1126/science.7058341] [PMID: 7058341]
- [61] Mufson, E.J.; Bothwell, M.; Kordower, J.H. Loss of nerve growth factor receptor-containing neurons in Alzheimers disease: a quantitative analysis across subregions of the basal forebrain. *Exp. Neurol.*, **1989**, *105*(3), 221-232. [http://dx.doi.org/10.1016/0014-4886(89)90124-6] [PMID: 2548888]
- [62] Boncristiano, S.; Calhoun, M.E.; Kelly, P.H.; Pfeifer, M.; Bondolfi, L.; Stalder, M.; Phinney, A.L.; Abramowski, D.; Sturchler-Pierrat, C.; Enz, A.; Sommer, B.; Staufenbiel, M.; Jucker, M. Cholinergic changes in the APP23 transgenic mouse model of cerebral amyloidosis. *J. Neurosci.*, **2002**, *22*(8), 3234-3243. [PMID: 11943824]
- [63] Kar, S.; Slowikowski, S.P.; Westaway, D.; Mount, H.T. Interactions between β-amyloid and central cholinergic neurons: implications for Alzheimers disease. J. Psychiatry Neurosci., 2004, 29(6), 427-441. [PMID: 15644984]
- [64] Perry, E.K.; Johnson, M.; Kerwin, J.M.; Piggott, M.A.; Court, J.A.; Shaw, P.J.; Ince, P.G.; Brown, A.; Perry, R.H. Convergent cholinergic activities in aging and Alzheimers disease. *Neurobiol. Aging*, **1992**, *13*(3), 393-400. [http://dx.doi.org/10.1016/0197-4580 (92)90113-C] [PMID: 1625768]
- [65] Giacobini, E. Cholinergic receptors in human brain: effects of aging and Alzheimer disease. J. Neurosci. Res., 1990, 27(4), 548-560. [http://dx.doi.org/10.1002/jnr.490270416] [PMID: 2079716]
- [66] Greenamyre, J.T.; Maragos, W.F. Neurotransmitter receptors in Alzheimer disease. *Cerebrovasc. Brain Metab. Rev.*, **1993**, 5(2), 61-94. [PMID: 8392361]
- [67] Perry, E.K.; Morris, C.M.; Court, J.A.; Cheng, A.; Fairbairn, A.F.; McKeith, I.G.; Irving, D.; Brown, A.; Perry, R.H. Alteration in nicotine binding sites in Parkinsons disease, Lewy body dementia and Alzheimers disease: possible index of early neuropathology. *Neuroscience*, **1995**, *64*(2), 385-395. [http://dx.doi.org/10.1016/ 0306-4522(94)00410-7] [PMID: 7700528]
- [68] Ahmed, T.; Gilani, A-H. Inhibitory effect of curcuminoids on acetylcholinesterase activity and attenuation of scopolamineinduced amnesia may explain medicinal use of turmeric in Alzheimers disease. *Pharmacol. Biochem. Behav.*, **2009**, *91*(4), 554-559. [http://dx.doi.org/10.1016/j.pbb.2008.09.010] [PMID: 18930076]
- [69] Barrantes, F.J.; Borroni, V.; Vallés, S. Neuronal nicotinic acetylcholine receptor-cholesterol crosstalk in Alzheimers disease. *FEBS Lett.*, **2010**, *584*(9), 1856-1863. [http://dx.doi.org/10.1016/j. febslet.2009.11.036] [PMID: 19914249]
- [70] Medeiros, R.; Kitazawa, M.; Caccamo, A.; Baglietto-Vargas, D.; Estrada-Hernandez, T.; Cribbs, D.H.; Fisher, A.; LaFerla, F.M. Loss of muscarinic M1 receptor exacerbates Alzheimers diseaselike pathology and cognitive decline. *Am. J. Pathol.*, 2011, *179*(2), 980-991. [http://dx.doi.org/10.1016/j.ajpath.2011.04.041] [PMID: 21704011]
- [71] Shiozaki, K.; Iseki, E.; Uchiyama, H.; Watanabe, Y.; Haga, T.; Kameyama, K.; Ikeda, T.; Yamamoto, T.; Kosaka, K. Alterations of muscarinic acetylcholine receptor subtypes in diffuse lewy body disease: relation to Alzheimers disease. J. Neurol. Neurosurg. Psychiatry, 1999, 67(2), 209-213. [http://dx.doi.org/10.1136/ jnnp.67.2.209] [PMID: 10406992]
- [72] Tsang, S.W.; Lai, M.K.; Kirvell, S.; Francis, P.T.; Esiri, M.M.; Hope, T.; Chen, C.P.; Wong, P.T. Impaired coupling of muscarinic M1 receptors to G-proteins in the neocortex is associated with severity of dementia in Alzheimers disease. *Neurobiol. Aging*, 2006, 27(9), 1216-1223. [http://dx.doi.org/10.1016/j.neurobiolaging.2005. 07.010] [PMID: 16129514]
- [73] Caccamo, A.; Oddo, S.; Billings, L.M.; Green, K.N.; Martinez-Coria, H.; Fisher, A.; LaFerla, F.M. M1 receptors play a central role in modulating AD-like pathology in transgenic mice. *Neuron*, 2006, 49(5), 671-682. [http://dx.doi.org/10.1016/j.neuron.2006.01. 020] [PMID: 16504943]
- [74] Hashmi, A.N.; Yaqinuddin, A.; Ahmed, T. Pharmacological effects of Ibuprofen on learning and memory, Muscarinic receptors genes expression and APP isoforms levels in Pre-frontal cortex of AlCl3induced toxicity mouse model. *Int. J. Neurosci.*, **2014**, (0), 1-37. [PMID: 24825584]
- [75] Tsang, S.W.; Pomakian, J.; Marshall, G.A.; Vinters, H.V.; Cummings, J.L.; Chen, C.P.; Wong, P.T.; Lai, M.K. Disrupted

muscarinic M1 receptor signaling correlates with loss of protein kinase C activity and glutamatergic deficit in Alzheimers disease. *Neurobiol. Aging*, **2007**, *28*(9), 1381-1387. [http://dx.doi.org/ 10.1016/j.neurobiolaging.2006.06.001] [PMID: 16828202]

- [76] Ferrari-DiLeo, G.; Mash, D.C.; Flynn, D.D. Attenuation of muscarinic receptor-G-protein interaction in Alzheimer disease. *Mol. Chem. Neuropathol.*, **1995**, *24*(1), 69-91. [http://dx.doi.org/10. 1007/BF03160113] [PMID: 7755848]
- [77] Harrison, P.J.; Barton, A.J.; Najlerahim, A.; McDonald, B.; Pearson, R.C. Increased muscarinic receptor messenger RNA in Alzheimers disease temporal cortex demonstrated by in situ hybridization histochemistry. *Brain Res. Mol. Brain Res.*, **1991**, 9(1-2), 15-21. [http://dx.doi.org/10.1016/0169-328X(91)90125-H] [PMID: 1673214]
- [78] Rodríguez-Puertas, R.; Pascual, J.; Vilaró, T.; Pazos, A. Autoradiographic distribution of M1, M2, M3, and M4 muscarinic receptor subtypes in Alzheimers disease. *Synapse*, **1997**, *26*(4), 341-350. [http://dx.doi.org/10.1002/(SICI)1098-2396(199708)26:4 <341::AID-SYN2>3.0.CO;2-6] [PMID: 9215593]
- [79] Poulin, B.; Butcher, A.; McWilliams, P.; Bourgognon, J.M.; Pawlak, R.; Kong, K.C.; Bottrill, A.; Mistry, S.; Wess, J.; Rosethorne, E.M.; Charlton, S.J.; Tobin, A.B. The M3-muscarinic receptor regulates learning and memory in a receptor phosphorylation/arrestindependent manner. *Proc. Natl. Acad. Sci. USA*, **2010**, *107*(20), 9440-9445. [http://dx.doi.org/10.1073/pnas.0914801107] [PMID: 20439723]
- [80] Pedersen, W.A.; Kloczewiak, M.A.; Blusztajn, J.K. Amyloid betaprotein reduces acetylcholine synthesis in a cell line derived from cholinergic neurons of the basal forebrain. *Proc. Natl. Acad. Sci.* USA, **1996**, 93(15), 8068-8071. [http://dx.doi.org/10.1073/pnas.93. 15.8068] [PMID: 8755604]
- [81] Bierer, L.M.; Haroutunian, V.; Gabriel, S.; Knott, P.J.; Carlin, L.S.; Purohit, D.P.; Perl, D.P.; Schmeidler, J.; Kanof, P.; Davis, K.L. Neurochemical correlates of dementia severity in Alzheimers disease: relative importance of the cholinergic deficits. J. Neurochem., 1995, 64(2), 749-760. [http://dx.doi.org/10.1046/j. 1471-4159.1995.64020749.x] [PMID: 7830069]
- [82] Wilcock, G.K.; Esiri, M.M.; Bowen, D.M.; Smith, C.C. Alzheimers disease. Correlation of cortical choline acetyltransferase activity with the severity of dementia and histological abnormalities. J. Neurol. Sci., 1982, 57(2-3), 407-417. [http://dx.doi.org/10.1016/ 0022-510X(82)90045-4] [PMID: 7161627]
- [83] Davies, P.; Maloney, A.J. Selective loss of central cholinergic neurons in Alzheimers disease. *Lancet*, **1976**, 2(8000), 1403. [http:// dx.doi.org/10.1016/S0140-6736(76)91936-X] [PMID: 63862]
- [84] Bowen, D.M.; Smith, C.B.; White, P.; Davison, A.N. Neurotransmitter-related enzymes and indices of hypoxia in senile dementia and other abiotrophies. *Brain*, **1976**, *99*(3), 459-496. [http://dx.doi.org/10.1093/brain/99.3.459] [PMID: 11871]
- [85] Araujo, D.M.; Lapchak, P.A.; Robitaille, Y.; Gauthier, S.; Quirion, R. Differential alteration of various cholinergic markers in cortical and subcortical regions of human brain in Alzheimers disease. J. Neurochem., 1988, 50(6), 1914-1923. [http://dx.doi.org/10.1111/ j.1471-4159.1988.tb02497.x] [PMID: 3373218]
- [86] Bartus, R.T.; Dean, R.L., III; Beer, B.; Lippa, A.S. The cholinergic hypothesis of geriatric memory dysfunction. *Science*, **1982**, 217(4558), 408-414. [http://dx.doi.org/10.1126/science.7046051] [PMID: 7046051]
- [87] Drachman, D.A. Memory and cognitive function in man: does the cholinergic system have a specific role? *Neurology*, **1977**, *27*(8), 783-790. [http://dx.doi.org/10.1212/WNL.27.8.783] [PMID: 560649]
- [88] Sutherland, R.J.; Whishaw, I.Q.; Regehr, J.C. Cholinergic receptor blockade impairs spatial localization by use of distal cues in the rat. *J. Comp. Physiol. Psychol.*, **1982**, *96*(4), 563-573. [http://dx.doi. org/10.1037/h0077914] [PMID: 7119176]
- [89] Roldán, G.; Bolaños-Badillo, E.; González-Sánchez, H.; Quirarte, G.L.; Prado-Alcalá, R.A. Selective M1 muscarinic receptor antagonists disrupt memory consolidation of inhibitory avoidance in rats. *Neurosci. Lett.*, **1997**, *230*(2), 93-96. [http://dx.doi.org/ 10.1016/S0304-3940(97)00489-8] [PMID: 9259472]
- [90] Bodick, N.C.; Offen, W.W.; Levey, A.I.; Cutler, N.R.; Gauthier, S.G.; Satlin, A.; Shannon, H.E.; Tollefson, G.D.; Rasmussen, K.; Bymaster, F.P.; Hurley, D.J.; Potter, W.Z.; Paul, S.M. Effects of xanomeline, a selective muscarinic receptor agonist, on cognitive

function and behavioral symptoms in Alzheimer disease. *Arch. Neurol.*, **1997**, *54*(4), 465-473. [http://dx.doi.org/10.1001/ archneur.1997.00550160091022] [PMID: 9109749]

- [91] Shekhar, A.; Potter, W.Z.; Lightfoot, J.; Lienemann, J.; Dubé, S.; Mallinckrodt, C.; Bymaster, F.P.; McKinzie, D.L.; Felder, C.C. Selective muscarinic receptor agonist xanomeline as a novel treatment approach for schizophrenia. *Am. J. Psychiatry*, **2008**, *165*(8), 1033-1039. [http://dx.doi.org/10.1176/appi.ajp.2008. 06091591] [PMID: 18593778]
- [92] Farber, S.A.; Nitsch, R.M.; Schulz, J.G.; Wurtman, R.J. Regulated secretion of beta-amyloid precursor protein in rat brain. J. Neurosci., 1995, 15(11), 7442-7451. [PMID: 7472496]
- [93] Fisher, A. Cholinergic treatments with emphasis on m1 muscarinic agonists as potential disease-modifying agents for Alzheimers disease. *Neurotherapeutics*, 2008, 5(3), 433-442. [http://dx. doi.org/10.1016/j.nurt.2008.05.002] [PMID: 18625455]
- [94] Jones, C.K.; Brady, A.E.; Davis, A.A.; Xiang, Z.; Bubser, M.; Tantawy, M.N.; Kane, A.S.; Bridges, T.M.; Kennedy, J.P.; Bradley, S.R.; Peterson, T.E.; Ansari, M.S.; Baldwin, R.M.; Kessler, R.M.; Deutch, A.Y.; Lah, J.J.; Levey, A.I.; Lindsley, C.W.; Conn, P.J. Novel selective allosteric activator of the M1 muscarinic acetylcholine receptor regulates amyloid processing and produces antipsychotic-like activity in rats. *J. Neurosci.*, 2008, 28(41), 10422-10433. [http://dx.doi.org/10.1523/JNEUROSCI. 1850-08.2008] [PMID: 18842902]
- [95] Bridges, T.M.; Reid, P.R.; Lewis, L.M.; Dawson, E.S.; Weaver, C.D.; Wood, M.R.; Lindsley, C.W. Discovery and development of a second highly selective M1 Positive Allosteric Modulator; PAM, 2010. PMID: 21433387.
- [96] Beach, T.G.; Walker, D.G.; Potter, P.E.; Sue, L.I.; Fisher, A. Reduction of cerebrospinal fluid amyloid beta after systemic administration of M1 muscarinic agonists. *Brain Res.*, 2001, 905(1-2), 220-223. [http://dx.doi.org/10.1016/S0006-8993(01)02484-2] [PMID: 11423097]
- [97] Nitsch, R.M.; Deng, M.; Tennis, M.; Schoenfeld, D.; Growdon, J.H. The selective muscarinic M1 agonist AF102B decreases levels of total Abeta in cerebrospinal fluid of patients with Alzheimers disease. Ann. Neurol., 2000, 48(6), 913-918. [http://dx.doi.org/ 10.1002/1531-8249(200012)48:6<913::AID-ANA12>3.0.CO;2-8] [PMID: 11117548]
- [98] Ahmed, T.; Enam, S.A.; Gilani, A.H. Curcuminoids enhance memory in an amyloid-infused rat model of Alzheimers disease. *Neuroscience*, **2010**, *169*(3), 1296-1306. [http://dx.doi.org/10. 1016/j.neuroscience.2010.05.078] [PMID: 20538041]
- [99] Ahmed, T.; Gilani, A.H.; Hosseinmardi, N.; Semnanian, S.; Enam, S.A.; Fathollahi, Y. Curcuminoids rescue long-term potentiation impaired by amyloid peptide in rat hippocampal slices. *Synapse*, **2011**, 65(7), 572-582. [http://dx.doi.org/10.1002/syn.20876] [PMID: 20963814]
- [100] Ahmed, T.; Gilani, A-H. A comparative study of curcuminoids to measure their effect on inflammatory and apoptotic gene expression in an Aβ plus ibotenic acid-infused rat model of Alzheimers disease. *Brain Res.*, **2011**, *1400*, 1-18. [http://dx.doi.org/10. 1016/j.brainres.2011.05.022] [PMID: 21640982]
- [101] Wolf, B.A.; Wertkin, A.M.; Jolly, Y.C.; Yasuda, R.P.; Wolfe, B.B.; Konrad, R.J.; Manning, D.; Ravi, S.; Williamson, J.R.; Lee, V.M. Muscarinic regulation of Alzheimers disease amyloid precursor protein secretion and amyloid beta-protein production in human neuronal NT2N cells. *J. Biol. Chem.*, **1995**, *270*(9), 4916-4922. [http://dx.doi.org/10.1074/jbc.270.9.4916] [PMID: 7876266]
- [102] Buiter, H.J.; Windhorst, A.D.; Huisman, M.C.; Yaqub, M.; Knol, D.L.; Fisher, A.; Lammertsma, A.A.; Leysen, J.E. [11C]AF150(S), an agonist PET ligand for M1 muscarinic acetylcholine receptors. *EJNMMI Res.*, **2013**, 3(1), 19. [http://dx.doi.org/10.1186/2191-219X-3-19] [PMID: 23514539]
- [103] Shirey, J.K.; Brady, A.E.; Jones, P.J.; Davis, A.A.; Bridges, T.M.; Kennedy, J.P.; Jadhav, S.B.; Menon, U.N.; Xiang, Z.; Watson, M.L.; Christian, E.P.; Doherty, J.J.; Quirk, M.C.; Snyder, D.H.; Lah, J.J.; Levey, A.I.; Nicolle, M.M.; Lindsley, C.W.; Conn, P.J. A selective allosteric potentiator of the M1 muscarinic acetylcholine receptor increases activity of medial prefrontal cortical neurons and restores impairments in reversal learning. *J. Neurosci.*, 2009, 29(45), 14271-14286. [http://dx.doi.org/10.1523/JNEUROSCI. 3930-09.2009] [PMID: 19906975]

- [104] Bradley, S.R.; Lameh, J.; Ohrmund, L.; Son, T.; Bajpai, A.; Nguyen, D.; Friberg, M.; Burstein, E.S.; Spalding, T.A.; Ott, T.R.; Schiffer, H.H.; Tabatabaei, A.; McFarland, K.; Davis, R.E.; Bonhaus, D.W. AC-260584, an orally bioavailable M(1) muscarinic receptor allosteric agonist, improves cognitive performance in an animal model. *Neuropharmacology*, **2010**, *58*(2), 365-373. [http:// dx.doi.org/10.1016/j.neuropharm.2009.10.003] [PMID: 19835892]
- [105] Langmead, C.J.; Austin, N.E.; Branch, C.L.; Brown, J.T.; Buchanan, K.A.; Davies, C.H.; Forbes, I.T.; Fry, V.A.; Hagan, J.J.; Herdon, H.J.; Jones, G.A.; Jeggo, R.; Kew, J.N.; Mazzali, A.; Melarange, R.; Patel, N.; Pardoe, J.; Randall, A.D.; Roberts, C.; Roopun, A.; Starr, K.R.; Teriakidis, A.; Wood, M.D.; Whittington, M.; Wu, Z.; Watson, J. Characterization of a CNS penetrant, selective M1 muscarinic receptor agonist, 77-LH-281. Br. J. Pharmacol., 2008, 154(5), 1104-1115. [http://dx.doi.org/10.1038/ bjp.2008.152] [PMID: 18454168]
- [106] Dani, J.A.; Bertrand, D. Nicotinic acetylcholine receptors and nicotinic cholinergic mechanisms of the central nervous system. *Annu. Rev. Pharmacol. Toxicol.*, **2007**, *47*, 699-729. [http://dx.doi. org/10.1146/annurev.pharmtox.47.120505.105214] [PMID: 17009926]
- [107] Dani, J.A. Overview of nicotinic receptors and their roles in the central nervous system. *Biol. Psychiatry*, 2001, 49(3), 166-174. [http://dx.doi.org/10.1016/S0006-3223(00)01011-8] [PMID: 11230867]
- [108] Tohgi, H.; Utsugisawa, K.; Yoshimura, M.; Nagane, Y.; Mihara, M. Age-related changes in nicotinic acetylcholine receptor subunits α4 and β2 messenger RNA expression in postmortem human frontal cortex and hippocampus. *Neurosci. Lett.*, **1998**, *245*(3), 139-142. [http://dx.doi.org/10.1016/S0304-3940(98)00205-5] [PMID: 9605475]
- [109] Pohanka, M. Alpha7 nicotinic acetylcholine receptor is a target in pharmacology and toxicology. *Int. J. Mol. Sci.*, **2012**, *13*(2), 2219-2238. [http://dx.doi.org/10.3390/ijms13022219] [PMID: 22408449]
- [110] Lucas-Meunier, E.; Fossier, P.; Baux, G.; Amar, M. Cholinergic modulation of the cortical neuronal network. *Pflugers Arch.*, 2003, 446(1), 17-29. [http://dx.doi.org/10.1007/s00424-002-0999-2] [PMID: 12690458]
- [111] Tohgi, H.; Utsugisawa, K.; Yoshimura, M.; Nagane, Y.; Mihara, M. Age-related changes in nicotinic acetylcholine receptor subunits alpha4 and beta2 messenger RNA expression in postmortem human frontal cortex and hippocampus. *Neurosci. Lett.*, **1998**, *245*(3), 139-142. [http://dx.doi.org/10.1016/S0304-3940(98)00205-5] [PMID: 9605475]
- [112] Hernandez, C.M.; Dineley, K.T. α7 nicotinic acetylcholine receptors in Alzheimers disease: neuroprotective, neurotrophic or both? *Curr. Drug Targets*, **2012**, *13*(5), 613-622. [http://dx. doi.org/10.2174/138945012800398973] [PMID: 22300028]
- [113] Wang, H.; Lee, D.H.; DAndrea, M.R.; Peterson, P.A.; Shank, R.P.; Reitz, A.B. β-Amyloid1–42 binds to α7 Nicotinic Acetylcholine Receptor with High Affinity. J. Biol. Chem., 2000, 275(8), 5626-5632. [http://dx.doi.org/10.1074/jbc.275.8.5626] [PMID: 10681545]
- [114] Resende, R.R.; Adhikari, A. Cholinergic receptor pathways involved in apoptosis, cell proliferation and neuronal differentiation. *Cell Commun. Signal.*, **2009**, *7*, 20. [http://dx.doi.org/10.1186/1478-811X-7-20] [PMID: 19712465]
- [115] Itier, V.; Bertrand, D. Neuronal nicotinic receptors: from protein structure to function. *FEBS Lett.*, **2001**, *504*(3), 118-125. [http:// dx.doi.org/10.1016/S0014-5793(01)02702-8] [PMID: 11532443]
- [116] Liu, Q.; Xie, X.; Lukas, R.J.; St John, P.A.; Wu, J. A novel nicotinic mechanism underlies β-amyloid-induced neuronal hyperexcitation. J. Neurosci., 2013, 33(17), 7253-7263. [http://dx.doi. org/10.1523/JNEUROSCI.3235-12.2013] [PMID: 23616534]
- [117] Jones, I.W.; Westmacott, A.; Chan, E.; Jones, R.W.; Dineley, K.; ONeill, M.J.; Wonnacott, S. α7 nicotinic acetylcholine receptor expression in Alzheimers disease. J. Mol. Neurosci., 2006, 30(1), 83-84. [http://dx.doi.org/10.1385/JMN:30:1:83] [PMID: 17192639]
- [118] Counts, S.E.; He, B.; Čhe, S.; Ikonomovic, M.D.; DeKosky, S.T.; Ginsberg, S.D.; Mufson, E.J. α7 nicotinic receptor up-regulation in cholinergic basal forebrain neurons in Alzheimer disease. *Arch. Neurol.*, **2007**, *64*(12), 1771-1776. [http://dx.doi.org/10.1001/ archneur.64.12.1771] [PMID: 18071042]
- [119] Pandya, A.; Yakel, J.L. Allosteric modulator Desformylflustrabromine relieves the inhibition of α2β2 and α4β2 nicotinic acetylcholine receptors by β-amyloid(142) peptide. J. Mol. Neurosci., 2011,

- [120] Lee, J.; Durst, R.; Wrolstad, R. 4 but not 3 and 7 nicotinic acetylcholine receptor subunits are lost from the temporal cortex in alzheimer's disease. J. Neurochem., 2002. [http://dx.doi.org/ 10.1023/A:1014832520809]
- [121] Banerjee, C.; Nyengaard, J.R.; Wevers, A.; de Vos, R.A.; Jansen Steur, E.N.; Lindstrom, J.; Pilz, K.; Nowacki, S.; Bloch, W.; Schröder, H. Cellular expression of α7 nicotinic acetylcholine receptor protein in the temporal cortex in Alzheimers and Parkinsons diseasea stereological approach. *Neurobiol. Dis.*, **2000**, 7(6 Pt B), 666-672. [http://dx.doi.org/10.1006/nbdi.2000.0317] [PMID: 11114264]
- [122] Wevers, A.; Monteggia, L.; Nowacki, S.; Bloch, W.; Schütz, U.; Lindstrom, J.; Pereira, E.F.; Eisenberg, H.; Giacobini, E.; de Vos, R.A.; Steur, E.N.; Maelicke, A.; Albuquerque, E.X.; Schröder, H. Expression of nicotinic acetylcholine receptor subunits in the cerebral cortex in Alzheimers disease: histotopographical correlation with amyloid plaques and hyperphosphorylated-tau protein. *Eur. J. Neurosci.*, **1999**, *11*(7), 2551-2565. [http://dx.doi.org/10.1046/ j.1460-9568.1999.00676.x] [PMID: 10383644]
- Burghaus, L.; Schütz, U.; Krempel, U.; de Vos, R.A.; Jansen, S.E.N.; Wevers, A.; Lindstrom, J.; Schröder, H. Quantitative assessment of nicotinic acetylcholine receptor proteins in the cerebral cortex of Alzheimer patients. *Brain Res. Mol. Brain Res.*, 2000, 76(2), 385-388. [http://dx.doi.org/10.1016/S0169-328X(00) 00031-0] [PMID: 10762715]
- [124] Hellström-Lindahl, E.; Mousavi, M.; Zhang, X.; Ravid, R.; Nordberg, A. Regional distribution of nicotinic receptor subunit mRNAs in human brain: comparison between Alzheimer and normal brain. *Brain Res. Mol. Brain Res.*, **1999**, *66*(1-2), 94-103. [http://dx. doi.org/10.1016/S0169-328X(99)00030-3] [PMID: 10095081]
- [125] Bednar, I.; Paterson, D.; Marutle, A.; Pham, T.M.; Svedberg, M.; Hellström-Lindahl, E.; Mousavi, M.; Court, J.; Morris, C.; Perry, E.; Mohammed, A.; Zhang, X.; Nordberg, A. Selective nicotinic receptor consequences in APP(SWE) transgenic mice. *Mol. Cell. Neurosci.*, **2002**, 20(2), 354-365. [http://dx.doi.org/10.1006/mcne. 2002.1112] [PMID: 12093166]
- [126] Dineley, K.T.; Westerman, M.; Bui, D.; Bell, K.; Ashe, K.H.; Sweatt, J.D. Beta-amyloid activates the mitogen-activated protein kinase cascade via hippocampal alpha7 nicotinic acetylcholine receptors: *In vitro* and *in vivo* mechanisms related to Alzheimers disease. *J. Neurosci.*, **2001**, 21(12), 4125-4133. [PMID: 11404397]
- [127] Jones, I.W.; Westmacott, A.; Chan, E.; Jones, R.W.; Dineley, K.; ONeill, M.J.; Wonnacott, S. α7 nicotinic acetylcholine receptor expression in Alzheimers disease: receptor densities in brain regions of the APP(SWE) mouse model and in human peripheral blood lymphocytes. J. Mol. Neurosci., 2006, 30(1-2), 83-84. [http://dx.doi.org/10.1385/JMN:30:1:83] [PMID: 17192639]
- [128] Parri, H.R.; Hernandez, C.M.; Dineley, K.T. Research update: Alpha7 nicotinic acetylcholine receptor mechanisms in Alzheimers disease. *Biochem. Pharmacol.*, 2011, 82(8), 931-942. [http://dx. doi.org/10.1016/j.bcp.2011.06.039] [PMID: 21763291]
- [129] Wang, H-Y.; Lee, D.H.; DAndrea, M.R.; Peterson, P.A.; Shank, R.P.; Reitz, A.B. β-Amyloid(142) binds to α7 nicotinic acetylcholine receptor with high affinity. Implications for Alzheimers disease pathology. J. Biol. Chem., 2000, 275(8), 5626-5632. [http://dx. doi.org/10.1074/jbc.275.8.5626] [PMID: 10681545]
- [130] Pettit, D.L.; Shao, Z.; Yakel, J.L. Beta-Amyloid(142) peptide directly modulates nicotinic receptors in the rat hippocampal slice. *J. Neurosci.*, 2001, 21(1), RC120-RC120. [PMID: 11150356]
- [131] Cleary, J.P.; Walsh, D.M.; Hofmeister, J.J.; Shankar, G.M.; Kuskowski, M.A.; Selkoe, D.J.; Ashe, K.H. Natural oligomers of the amyloid-β protein specifically disrupt cognitive function. *Nat. Neurosci.*, 2005, 8(1), 79-84. [http://dx.doi.org/10.1038/nn1372] [PMID: 15608634]
- [132] Puzzo, D.; Privitera, L.; Leznik, E.; Fà, M.; Staniszewski, A.; Palmeri, A.; Arancio, O. Picomolar amyloid-β positively modulates synaptic plasticity and memory in hippocampus. *J. Neurosci.*, 2008, 28(53), 14537-14545. [http://dx.doi.org/10.1523/JNEUROSCI.2692-08.2008] [PMID: 19118188]
- [133] Dineley, K. T. Beta-amyloid peptide--nicotinic acetylcholine receptor interaction: the two faces of health and disease. *Front. Biosci.*, 2006, 12, 5030-5038.

- [134] Gu, Z.; Yakel, J.L. Timing-dependent septal cholinergic induction of dynamic hippocampal synaptic plasticity. *Neuron*, **2011**, *71*(1), 155-165. [http://dx.doi.org/10.1016/j.neuron.2011.04.026] [PMID: 21745645]
- [135] Nagele, R.G.; DAndrea, M.R.; Lee, H.; Venkataraman, V.; Wang, H-Y. Astrocytes accumulate A β 42 and give rise to astrocytic amyloid plaques in Alzheimer disease brains. *Brain Res.*, 2003, 971(2), 197-209. [http://dx.doi.org/10.1016/S0006-8993(03)02361-8] [PMID: 12706236]
- [136] Wang, G.; Dinkins, M.; He, Q.; Zhu, G.; Poirier, C.; Campbell, A.; Mayer-Proschel, M.; Bieberich, E. Astrocytes secrete exosomes enriched with proapoptotic ceramide and prostate apoptosis response 4 (PAR-4): potential mechanism of apoptosis induction in Alzheimer disease (AD). J. Biol. Chem., 2012, 287(25), 21384-21395. [http://dx.doi.org/10.1074/jbc.M112.340513] [PMID: 22532571]
- [137] Smale, G.; Nichols, N.R.; Brady, D.R.; Finch, C.E.; Horton, W.E., Jr. Evidence for apoptotic cell death in Alzheimers disease. *Exp. Neurol.*, **1995**, *133*(2), 225-230. [http://dx.doi.org/10.1006/exnr. 1995.1025] [PMID: 7544290]
- [138] Liu, Y.; Hu, J.; Wu, J.; Zhu, C.; Hui, Y.; Han, Y.; Huang, Z.; Ellsworth, K.; Fan, W. α7 nicotinic acetylcholine receptormediated neuroprotection against dopaminergic neuron loss in an MPTP mouse model *via* inhibition of astrocyte activation. J. Neuroinflammation, **2012**, 9(98), 2094-2099.
- [139] Wang, H.Y.; Lee, D.H.; Davis, C.B.; Shank, R.P. Amyloid peptide Abeta(142) binds selectively and with picomolar affinity to α7 nicotinic acetylcholine receptors. J. Neurochem., 2000, 75(3), 1155-1161. [http://dx.doi.org/10.1046/j.1471-4159.2000.0751155.
 x] [PMID: 10936198]
- [140] Kem, W.R. The brain α7 nicotinic receptor may be an important therapeutic target for the treatment of Alzheimers disease: studies with DMXBA (GTS-21). *Behav. Brain Res.*, **2000**, *113*(1-2), 169-181. [http://dx.doi.org/10.1016/S0166-4328(00)00211-4] [PMID: 10942043]
- [141] Shen, J.X.; Yakel, J.L. Nicotinic acetylcholine receptor-mediated calcium signaling in the nervous system. *Acta Pharmacol. Sin.*, 2009, 30(6), 673-680. [http://dx.doi.org/10.1038/aps.2009.64] [PMID: 19448647]
- [142] Preskorn, S.H.; Gawryl, M.; Dgetluck, N.; Palfreyman, M.; Bauer, L.O.; Hilt, D.C. Normalizing effects of EVP-6124, an α-7 nicotinic partial agonist, on event-related potentials and cognition: a proof of concept, randomized trial in patients with schizophrenia. J. Psychiatr. Pract., 2014, 20(1), 12-24. [http://dx.doi.org/10.1097/ 01.pra.0000442935.15833.c5] [PMID: 24419307]
- [143] Sydserff, S.; Sutton, E.J.; Song, D.; Quirk, M.C.; Maciag, C.; Li, C.; Jonak, G.; Gurley, D.; Gordon, J.C.; Christian, E.P.; Doherty, J.J.; Hudzik, T.; Johnson, E.; Mrzljak, L.; Piser, T.; Smagin, G.N.; Wang, Y.; Widzowski, D.; Smith, J.S. Selective a7 nicotinic receptor activation by AZD0328 enhances cortical dopamine release and improves learning and attentional processes. *Biochem. Pharmacol.*, **2009**, *78*(7), 880-888. [http://dx.doi.org/10.1016/ j.bcp.2009.07.005] [PMID: 19615981]
- [144] Bitner, R.S.; Bunnelle, W.H.; Decker, M.W.; Drescher, K.U.; Kohlhaas, K.L.; Markosyan, S.; Marsh, K.C.; Nikkel, A.L.; Browman, K.; Radek, R.; Anderson, D.J.; Buccafusco, J.; Gopalakrishnan, M. *In vivo* pharmacological characterization of a novel selective a7 neuronal nicotinic acetylcholine receptor agonist ABT-107: preclinical considerations in Alzheimers disease. *J. Pharmacol. Exp. Ther.*, **2010**, *334*(3), 875-886. [http://dx.doi.org/ 10.1124/jpet.110.167213] [PMID: 20504913]
- [145] Stevens, K.E.; Cornejo, B.; Adams, C.E.; Zheng, L.; Yonchek, J.; Hoffman, K.L.; Christians, U.; Kem, W.R. Continuous administration of a selective α7 nicotinic partial agonist, DMXBA, improves sensory inhibition without causing tachyphylaxis or receptor upregulation in DBA/2 mice. *Brain Res.*, **2010**, *1352*, 140-146. [http:// dx.doi.org/10.1016/j.brainres.2010.06.063] [PMID: 20599427]
- [146] Hardy, J. Expression of normal sequence pathogenic proteins for neurodegenerative disease contributes to disease risk: permissive templating as a general mechanism underlying neurodegeneration. *Biochem. Soc. Trans.*, 2005, 33(Pt 4), 578-581. [http://dx.doi.org/ 10.1042/BST0330578] [PMID: 16042548]
- [147] Schedin-Weiss, S.; Winblad, B.; Tjernberg, L.O. The role of protein glycosylation in Alzheimer disease. *FEBS J.*, 2014, 281(1), 46-62. [http://dx.doi.org/10.1111/febs.12590] [PMID: 24279329]

- [148] Taylor, J.P.; Hardy, J.; Fischbeck, K.H. Toxic proteins in neurodegenerative disease. *Science*, 2002, 296(5575), 1991-1995. [http://dx.doi.org/10.1126/science.1067122] [PMID: 12065827]
- [149] Poulsen, S-A.; Watson, A.A.; Fairlie, D.P.; Craik, D.J. Solution structures in aqueous SDS micelles of two amyloid β peptides of A β(128) mutated at the α-secretase cleavage site (K16E, K16F). J. Struct. Biol., 2000, 130(2-3), 142-152. [http://dx.doi.org/10.1006/ jsbi.2000.4267] [PMID: 10940222]
- [150] Ren, R.J.; Dammer, E.B.; Wang, G.; Seyfried, N.T.; Levey, A.I. Proteomics of protein post-translational modifications implicated in neurodegeneration. *Transl. Neurodegener.*, **2014**, 3(1), 23. [http:// dx.doi.org/10.1186/2047-9158-3-23] [PMID: 25671099]
- [151] Zahid, S.; Khan, R.; Oellerich, M.; Ahmed, N.; Asif, A.R. Differential S-nitrosylation of proteins in Alzheimers disease. *Neuroscience*, 2014, 256, 126-136. [http://dx.doi.org/10.1016/ j.neuroscience.2013.10.026] [PMID: 24157928]
- [152] McFarlane, I.; Georgopoulou, N.; Coughlan, C.M.; Gillian, A.M.; Breen, K.C. The role of the protein glycosylation state in the control of cellular transport of the amyloid β precursor protein. *Neuroscience*, **1999**, *90*(1), 15-25. [http://dx.doi.org/10.1016/ S0306-4522(98)00361-3] [PMID: 10188930]
- [153] Mohorko, E.; Glockshuber, R.; Aebi, M. Oligosaccharyltransferase: the central enzyme of N-linked protein glycosylation. J. Inherit. Metab. Dis., 2011, 34(4), 869-878. [http://dx.doi.org/10.1007/ s10545-011-9337-1] [PMID: 21614585]
- [154] Georgopoulou, N.; McLaughlin, M.; McFarlane, I.; Breen, K.C. The role of post-translational modification in beta-amyloid precursor protein processing. *Biochem. Soc. Symp.*, 2001, (67), 23-36. [http://dx.doi.org/10.1042/bss0670023] [PMID: 11447837]
- [155] Haass, C.; Kaether, C.; Thinakaran, G.; Sisodia, S. Trafficking and proteolytic processing of APP. *Cold Spring Harb. Perspect. Med.*, 2012, 2(5), a006270. [http://dx.doi.org/10.1101/cshperspect.a006270]
 [PMID: 22553493]
- [156] Yazaki, M.; Tagawa, K.; Maruyama, K.; Sorimachi, H.; Tsuchiya, T.; Ishiura, S.; Suzuki, K. Mutation of potential N-linked glycosylation sites in the Alzheimers disease amyloid precursor protein (APP). *Neurosci. Lett.*, **1996**, 221(1), 57-60. [http://dx. doi.org/10.1016/S0304-3940(96)13285-7] [PMID: 9014180]
- [157] McFarlane, I.; Georgopoulou, N.; Coughlan, C.M.; Gillian, A.M.; Breen, K.C. The role of the protein glycosylation state in the control of cellular transport of the amyloid beta precursor protein. *Neuroscience*, **1999**, *90*(1), 15-25. [http://dx.doi.org/10.1016/ S0306-4522(98)00361-3] [PMID: 10188930]
- [158] Yang, D.S.; Tandon, A.; Chen, F.; Yu, G.; Yu, H.; Arawaka, S.; Hasegawa, H.; Duthie, M.; Schmidt, S.D.; Ramabhadran, T.V.; Nixon, R.A.; Mathews, P.M.; Gandy, S.E.; Mount, H.T.; St George-Hyslop, P.; Fraser, P.E. Mature glycosylation and trafficking of nicastrin modulate its binding to presenilins. *J. Biol. Chem.*, **2002**, *277*(31), 28135-28142. [http://dx.doi.org/10.1074/ jbc.M110871200] [PMID: 12032140]
- [159] LaFerla, F.M.; Green, K.N.; Oddo, S. Intracellular amyloid-beta in Alzheimers disease. *Nat. Rev. Neurosci.*, 2007, 8(7), 499-509. [http://dx.doi.org/10.1038/nrn2168] [PMID: 17551515]
- [160] Menéndez-González, M.; Pérez-Pinera, P.; Martínez-Rivera, M.; Calatayud, M.T.; Blázquez, M.B. APP processing and the APP-KPI domain involvement in the amyloid cascade. *Neurodegener. Dis.*, 2005, 2(6), 277-283. [http://dx.doi.org/10.1159/000092315] [PMID: 16909010]
- [161] Akasaka-Manya, K.; Manya, H.; Sakurai, Y.; Wojczyk, B.S.; Kozutsumi, Y.; Saito, Y.; Taniguchi, N.; Murayama, S.; Spitalnik, S.L.; Endo, T. Protective effect of N-glycan bisecting GlcNAc residues on beta-amyloid production in Alzheimers disease. *Glycobiology*, **2010**, 20(1), 99-106. [http://dx.doi.org/10.1093/ glycob/cwp152] [PMID: 19776078]
- [162] Nakagawa, K.; Kitazume, S.; Oka, R.; Maruyama, K.; Saido, T.C.; Sato, Y.; Endo, T.; Hashimoto, Y. Sialylation enhances the secretion of neurotoxic amyloid-beta peptides. *J. Neurochem.*, **2006**, *96*(4), 924-933. [http://dx.doi.org/10.1111/j.1471-4159.2005. 03595.x] [PMID: 16412100]
- [163] Mills, J.; Reiner, P.B. Regulation of amyloid precursor protein cleavage. J. Neurochem., 1999, 72(2), 443-460. [http://dx.doi.org/ 10.1046/j.1471-4159.1999.0720443.x] [PMID: 9930716]
- [164] Zhang, H.Y.; Yan, H.; Tang, X.C. Huperzine A enhances the level of secretory amyloid precursor protein and protein kinase C-alpha in intracerebroventricular beta-amyloid-(140) infused rats and

human embryonic kidney 293 Swedish mutant cells. *Neurosci. Lett.*, **2004**, *360*(1-2), 21-24. [http://dx.doi.org/10.1016/j.neulet. 2004.01.055] [PMID: 15082169]

- [165] Akasaka-Manya, K.; Manya, H.; Sakurai, Y.; Wojczyk, B.S.; Spitalnik, S.L.; Endo, T. Increased bisecting and core-fucosylated N-glycans on mutant human amyloid precursor proteins. *Glycoconj. J.*, **2008**, *25*(8), 775-786. [http://dx.doi.org/10.1007/ s10719-008-9140-x] [PMID: 18521746]
- [166] Brinkmalm, G.; Portelius, E.; Öhrfelt, A.; Mattsson, N.; Persson, R.; Gustavsson, M.K.; Vite, C.H.; Gobom, J.; Månsson, J.E.; Nilsson, J.; Halim, A.; Larson, G.; Rüetschi, U.; Zetterberg, H.; Blennow, K.; Brinkmalm, A. An online nano-LC-ESI-FTICR-MS method for comprehensive characterization of endogenous fragments from amyloid β and amyloid precursor protein in human and cat cerebrospinal fluid. J. Mass Spectrom. 2012, 47(5), 591-603. [http://dx.doi.org/10.1002/jms.2987] [PMID: 22576872]
- [167] Halim, A.; Brinkmalm, G.; Rüetschi, U.; Westman-Brinkmalm, A.; Portelius, E.; Zetterberg, H.; Blennow, K.; Larson, G.; Nilsson, J. Site-specific characterization of threonine, serine, and tyrosine glycosylations of amyloid precursor protein/amyloid beta-peptides in human cerebrospinal fluid. *Proc. Natl. Acad. Sci. USA*, 2011, *108*(29), 11848-11853. [http://dx.doi.org/10.1073/pnas.1102664108] [PMID: 21712440]
- [168] Kitazume, S.; Tachida, Y.; Kato, M.; Yamaguchi, Y.; Honda, T.; Hashimoto, Y.; Wada, Y.; Saito, T.; Iwata, N.; Saido, T.; Taniguchi, N. Brain endothelial cells produce amyloid beta from amyloid precursor protein 770 and preferentially secrete the Oglycosylated form. J. Biol. Chem., 2010, 285(51), 40097-40103. [http://dx.doi.org/10.1074/jbc.M110.144626] [PMID: 20952385]
- [169] Perdivara, I.; Petrovich, R.; Allinquant, B.; Deterding, L.J.; Tomer, K.B.; Przybylski, M. Elucidation of O-glycosylation structures of the beta-amyloid precursor protein by liquid chromatography-mass spectrometry using electron transfer dissociation and collision induced dissociation. J. Proteome Res., 2009, 8(2), 631-642. [http://dx.doi.org/10.1021/pr800758g] [PMID: 19093876]
- [170] Tomita, S.; Kirino, Y.; Suzuki, T. Cleavage of Alzheimers amyloid precursor protein (APP) by secretases occurs after O-glycosylation of APP in the protein secretory pathway. Identification of intracellular compartments in which APP cleavage occurs without using toxic agents that interfere with protein metabolism. J. Biol. Chem., 1998, 273(11), 6277-6284. [http://dx.doi.org/10.1074/ jbc.273.11.6277] [PMID: 9497354]
- [171] Griffith, L.S.; Mathes, M.; Schmitz, B. Beta-amyloid precursor protein is modified with O-linked N-acetylglucosamine. J. Neurosci. Res., 1995, 41(2), 270-278. [http://dx.doi.org/10.1002/jnr.490410214]
 [PMID: 7650762]
- [172] Jacobsen, K.T.; Iverfeldt, K. O-GlcNAcylation increases nonamyloidogenic processing of the amyloid-β precursor protein (APP). *Biochem. Biophys. Res. Commun.*, **2011**, 404(3), 882-886. [http://dx.doi.org/10.1016/j.bbrc.2010.12.080] [PMID: 21182826]
- [173] Zhang, X. Cholinergic activity and amyloid precursor protein processing in aging and Alzheimers disease. *Curr. Drug Targets CNS Neurol. Disord.*, **2004**, *3*(2), 137-152. [http://dx.doi.org/ 10.2174/1568007043482499] [PMID: 15078189]
- [174] Lee, M-S.; Kao, S-C.; Lemere, C.A.; Xia, W.; Tseng, H-C.; Zhou, Y.; Neve, R.; Ahlijanian, M.K.; Tsai, L-H. APP processing is regulated by cytoplasmic phosphorylation. J. Cell Biol., 2003, 163(1), 83-95. [http://dx.doi.org/10.1083/jcb.200301115] [PMID: 14557249]
- [175] Liu, F.; Su, Y.; Li, B.; Zhou, Y.; Ryder, J.; Gonzalez-DeWhitt, P.; May, P.C.; Ni, B. Regulation of amyloid precursor protein (APP) phosphorylation and processing by p35/Cdk5 and p25/Cdk5. *FEBS Lett.*, **2003**, *547*(1-3), 193-196. [http://dx.doi.org/10.1016/S0014-5793(03)00714-2] [PMID: 12860412]
- [176] Chang, K-A.; Kim, H-S.; Ha, T-Y.; Ha, J-W.; Shin, K.Y.; Jeong, Y.H.; Lee, J-P.; Park, C-H.; Kim, S.; Baik, T-K.; Suh, Y.H. Phosphorylation of amyloid precursor protein (APP) at Thr668 regulates the nuclear translocation of the APP intracellular domain and induces neurodegeneration. *Mol. Cell. Biol.*, 2006, 26(11), 4327-4338. [http://dx.doi.org/10.1128/MCB.02393-05] [PMID: 16705182]
- [177] Castro, A.; Martinez, A. Targeting beta-amyloid pathogenesis through acetylcholinesterase inhibitors. *Curr. Pharm. Des.*, 2006, *12*(33), 4377-4387. [http://dx.doi.org/10.2174/138161206778792985]
 [PMID: 17105433]
- [178] Pakaski, M.; Kasa, P. Role of acetylcholinesterase inhibitors in the metabolism of amyloid precursor protein. *Curr. Drug Targets CNS*

Neurol. Disord., **2003**, *2*(3), 163-171. [http://dx.doi.org/10.2174/1568007033482869] [PMID: 12769797]

- [179] Morris, M.; Knudsen, G.M.; Maeda, S.; Trinidad, J.C.; Ioanoviciu, A.; Burlingame, A.L.; Mucke, L. Tau post-translational modifications in wild-type and human amyloid precursor protein transgenic mice. *Nat. Neurosci.*, **2015**, *18*(8), 1183-1189. [http://dx.doi.org/10.1038/nn.4067] [PMID: 26192747]
- [180] Roberson, E.D.; Scearce-Levie, K.; Palop, J.J.; Yan, F.; Cheng, I.H.; Wu, T.; Gerstein, H.; Yu, G-Q.; Mucke, L. Reducing endogenous tau ameliorates amyloid beta-induced deficits in an Alzheimers disease mouse model. *Science*, 2007, *316*(5825), 750-754. [http://dx.doi.org/10.1126/science.1141736] [PMID: 17478722]
- [181] Ittner, L.M.; Ke, Y.D.; Delerue, F.; Bi, M.; Gladbach, A.; van Eersel, J.; Wölfing, H.; Chieng, B.C.; Christie, M.J.; Napier, I.A.; Eckert, A.; Staufenbiel, M.; Hardeman, E.; Götz, J. Dendritic function of tau mediates amyloid-β toxicity in Alzheimers disease mouse models. *Cell*, **2010**, *142*(3), 387-397. [http://dx.doi.org/ 10.1016/j.cell.2010.06.036] [PMID: 20655099]
- [182] Zempel, H.; Thies, E.; Mandelkow, E.; Mandelkow, E-M. Abeta oligomers cause localized Ca(2+) elevation, missorting of endogenous Tau into dendrites, Tau phosphorylation, and destruction of microtubules and spines. J. Neurosci., 2010, 30(36), 11938-11950. [http:// dx.doi.org/10.1523/JNEUROSCI.2357-10.2010] [PMID: 20826658]
- [183] Hanger, D.P.; Byers, H.L.; Wray, S.; Leung, K-Y.; Saxton, M.J.; Seereeram, A.; Reynolds, C.H.; Ward, M.A.; Anderton, B.H. Novel phosphorylation sites in tau from Alzheimer brain support a role for casein kinase 1 in disease pathogenesis. *J. Biol. Chem.*, 2007, 282(32), 23645-23654. [http://dx.doi.org/10.1074/jbc.M703269200] [PMID: 17562708]
- [184] Funk, K.E.; Thomas, S.N.; Schafer, K.N.; Cooper, G.L.; Liao, Z.; Clark, D.J.; Yang, A.J.; Kuret, J. Lysine methylation is an endogenous post-translational modification of tau protein in human brain and a modulator of aggregation propensity. *Biochem. J.*, 2014, 462(1), 77-88.
- [185] Guo, A.; Gu, H.; Zhou, J.; Mulhern, D.; Wang, Y.; Lee, K.A.; Yang, V.; Aguiar, M.; Kornhauser, J.; Jia, X.; Ren, J.; Beausoleil, S.A.; Silva, J.C.; Vemulapalli, V.; Bedford, M.T.; Comb, M.J. Immunoaffinity enrichment and mass spectrometry analysis of protein methylation. *Mol. Cell. Proteom.*, **2014**, *13*(1), 372-387. [http://dx.doi.org/10.1074/mcp.O113.027870] [PMID: 24129315]
- [186] Zhou, X-W.; Gustafsson, J-Å.; Tanila, H.; Bjorkdahl, C.; Liu, R.; Winblad, B.; Pei, J-J. Tau hyperphosphorylation correlates with reduced methylation of protein phosphatase 2A. *Neurobiol. Dis.*, **2008**, *31*(3), 386-394. [http://dx.doi.org/10.1016/j.nbd.2008.05.013]
 [PMID: 18586097]
- [187] Liu, F.; Iqbal, K.; Grundke-Iqbal, I.; Hart, G.W.; Gong, C-X. O-GlcNAcylation regulates phosphorylation of tau: a mechanism involved in Alzheimers disease. *Proc. Natl. Acad. Sci. USA*, 2004, 101(29), 10804-10809. [http://dx.doi.org/10.1073/pnas.0400348101] [PMID: 15249677]
- [188] Cripps, D.; Thomas, S.N.; Jeng, Y.; Yang, F.; Davies, P.; Yang, A.J. Alzheimer disease-specific conformation of hyperphosphorylated

paired helical filament-Tau is polyubiquitinated through Lys-48, Lys-11, and Lys-6 ubiquitin conjugation. *J. Biol. Chem.*, **2006**, *281*(16), 10825-10838. [http://dx.doi.org/10.1074/jbc.M512786200] [PMID: 16443603]

- [189] Min, S-W.; Cho, S-H.; Zhou, Y.; Schroeder, S.; Haroutunian, V.; Seeley, W.W.; Huang, E.J.; Shen, Y.; Masliah, E.; Mukherjee, C.; Meyers, D.; Cole, P.A.; Ott, M.; Gan, L. Acetylation of tau inhibits its degradation and contributes to tauopathy. *Neuron*, **2010**, *67*(6), 953-966. [http://dx.doi.org/10.1016/j.neuron.2010.08.044] [PMID: 20869593]
- [190] Nathanson, N.M. Synthesis, trafficking, and localization of muscarinic acetylcholine receptors. *Pharmacol. Ther.*, 2008, *119*(1), 33-43. [http://dx.doi.org/10.1016/j.pharmthera.2008.04.006]
 [PMID: 18558434]
- [191] Romero-Fernandez, W.; Borroto-Escuela, D.O.; Alea, M.P.; Garcia-Mesa, Y.; Garriga, P. Altered trafficking and unfolded protein response induction as a result of M3 muscarinic receptor impaired N-glycosylation. *Glycobiology*, **2011**, *21*(12), 1663-1672. [http://dx.doi.org/10.1093/glycob/cwr105] [PMID: 21798865]
- [192] Peretto, I.; Petrillo, P.; Imbimbo, B. P. Medicinal chemistry and therapeutic potential of muscarinic M3 antagonists. *Med. Res. Rev.*, 2009, 29(6), 867-902.
- [193] van Koppen, C.J.; Nathanson, N.M. Site-directed mutagenesis of the m2 muscarinic acetylcholine receptor. Analysis of the role of N-glycosylation in receptor expression and function. J. Biol. Chem., 1990, 265(34), 20887-20892. [PMID: 2249995]
- [194] Fisher, A.; Pittel, Z.; Haring, R.; Bar-Ner, N.; Kliger-Spatz, M.; Natan, N.; Egozi, I.; Sonego, H.; Marcovitch, I.; Brandeis, R. M1 muscarinic agonists can modulate some of the hallmarks in Alzheimers disease. J. Mol. Neurosci., 2003, 20(3), 349-356. [http://dx.doi.org/10.1385/JMN:20:3:349] [PMID: 14501019]
- [195] Wang, H-Y.; Li, W.; Benedetti, N.J.; Lee, D.H. α 7 nicotinic acetylcholine receptors mediate β-amyloid peptide-induced tau protein phosphorylation. J. Biol. Chem., 2003, 278(34), 31547-31553. [http://dx.doi.org/10.1074/jbc.M212532200] [PMID: 12801934]
- [196] Hu, M.; Waring, J.F.; Gopalakrishnan, M.; Li, J. Role of GSK-3β activation and α7 nAChRs in Abeta(142)-induced tau phosphorylation in PC12 cells. J. Neurochem., 2008, 106(3), 1371-1377. [http:// dx.doi.org/10.1111/j.1471-4159.2008.05483.x] [PMID: 18485099]
- [197] Xie, H.; Litersky, J.M.; Hartigan, J.A.; Jope, R.S.; Johnson, G.V. The interrelationship between selective tau phosphorylation and microtubule association. *Brain Res.*, **1998**, *798*(1-2), 173-183. [http:// dx.doi.org/10.1016/S0006-8993(98)00407-7] [PMID: 9666118]
- [198] Oddo, S.; Caccamo, A.; Green, K.N.; Liang, K.; Tran, L.; Chen, Y.; Leslie, F.M.; LaFerla, F.M. Chronic nicotine administration exacerbates tau pathology in a transgenic model of Alzheimers disease. *Proc. Natl. Acad. Sci. USA*, **2005**, *102*(8), 3046-3051. [http://dx.doi.org/10.1073/pnas.0408500102] [PMID: 15705720]
- [199] Hellström-Lindahl, E. Modulation of β-amyloid precursor protein processing and tau phosphorylation by acetylcholine receptors. *Eur. J. Pharmacol.*, **2000**, *393*(1-3), 255-263. [http://dx.doi.org/10. 1016/S0014-2999(00)00028-5] [PMID: 10771022]