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## Supporting information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

**Appendix S1.** Materials and methods.

**Table S1** Number of offspring for each of the 17 Murciano–Granadina sires.

**Table S2** Single nucleotide polymorphisms displaying transmission ratio distortion ( $\alpha$ -value above 0.15 or below  $-0.15$ ,  $q$ -value  $< 0.05$ ) in a population composed of 17 families of Murciano–Granadina goats.

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## Genetic evaluation of small ruminant lentivirus susceptibility in Valais blacknose sheep

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### Background

Maedi-Visna is a complex production-limiting ovine lentiviral disease caused by small ruminant lentivirus (SRLV) infection widespread throughout the world characterised by long immunological and clinical latencies and chronic progressive inflammatory pathology.<sup>1</sup> Previously, a SNP in the ovine *TMEM154* gene (Oar\_rambouillet\_v1.0: chr 17:g.5776842G>A) as well as a short indel variant in the promoter region of ovine *CCR5* (Oar\_rambouillet\_v1.0: chr 19:g.54980459\_54980460ins) were reported as markers associated with resistance/susceptibility to SRLV in several sheep breeds globally (OMIA 001694-9940).<sup>2,3</sup> Multiple other genomic regions associated with susceptibility to or control of infection were also reported indicating genetic complexity.<sup>4</sup> In Switzerland the Valais blacknose sheep (VBS) breed was found to have the highest prevalence of Maedi-Visna among Swiss sheep breeds.<sup>5</sup>

**Table 1** Genotypes of the 67 Valais blacknose sheep in the two previously reported markers<sup>2,3</sup>

SRLV serostatus	<i>TMEM154</i>			<i>CCR5</i>		
	GG	GA	AA	wt/wt	wt/del	del/del
Positive (n=34)	0	10	24	22	9	3
Negative (n=33)	0	1	32	24	4	5

### Own analysis

We collected pairs of SRLV-infected and non-infected individuals from this native Swiss sheep breed (Table S1). All initially positive tested sheep for SRLV were determined seropositive by two commercial ELISA tests: (1) Checkit CAEV/MVV<sup>®</sup>ELISA (IDEXX Laboratories, Liebefeld, Switzerland), a whole-virus antigen-based indirect ELISA; and (2) Small Ruminant Lentivirus Antibody Test Kit (VMRD, Pullman, WA, USA), a genotype B gp135 competitive ELISA. Positive samples were confirmed subsequently by western blotting focusing on the detection of the viral capsid (p25), matrix (p18), and nucleocapsid (p15) proteins<sup>6</sup> in the Swiss national reference laboratory for lentiviruses in small ruminants. The controls were ELISA-based negative tested only. The average age of the confirmed SRLV-positively tested sheep designated as cases was 5 years ranging from 2 to 9 years, whereas the SRLV-negative flock mates selected as controls were on average 4.6 years old (ranging from 1 to 11 years; Table S1). Subsequently all 67 animals were genotyped for the previously described *TMEM154*<sup>2</sup> and *CCR5*<sup>3</sup> variants by direct Sanger sequencing using the ABI 3730 DNA Analyzer (Thermo Fisher Scientific, Reinach, Switzerland). Therefore, genomic DNA was extracted from EDTA-stabilized blood samples using the Maxwell RSC instrument (Promega, Dübendorf, Switzerland). Interestingly, no homozygous wild type genotypes were observed for the *TMEM154* marker, while 29% of cases and 3% of controls were heterozygous. For the *CCR5* marker, 65% of cases and 73% of controls had the homozygous wild type genotype, while 26% of cases and 12% of controls were heterozygous (Table 1). We observed neither a significant association nor a trend while comparing the genotypes with the SRLV infection status.

Subsequently all 67 animals (34 cases and 33 controls) were genotyped using the Illumina ovine HD BeadChip.<sup>7</sup> After quality control (call rate  $>90\%$ , minor allele frequency  $>0.05$ ), 67 animals and 416,454 SNPs were retained. Genotyping data can be retrieved at <https://osf.io/b35ud/> (<https://doi.org/10.17605/OSF.IO/FSRPW>). Using GEMMA v0.98,<sup>8</sup> a genome-wide association study (GWAS) was conducted to look for additional genome regions associated with susceptibility to MV in VBS. To show the genetic distances among the studied animals, a relatedness matrix was generated (Fig. S1). GWAS

revealed the best-associated SNP ( $P$ -value =  $8.2 \times 10^{-5}$ ) on chromosome 9 at 69 843 937 bp (Fig. S1) although it did not reach the Bonferroni corrected genome-wide significance level ( $-\log(P\text{-value}) = 6.9$ ). Eight out of the 10 best-associated SNPs map to this genome region from 62 to 72 Mb at chromosome 9 (Table S2). Based on the NCBI *Ovis aries* annotation release 103, a total of 42 genes and loci are annotated in that 10-Mb region of the Oar\_ram-bouillet\_v1.0 assembly (Table S3).

#### Conclusions

No association between genotypes in the *TMEM154* and *CCR5* and SRLV susceptibility in the studied local Swiss breed VBS could be determined showing that these two previously reported genetic markers do not affect individual susceptibility to infection in this particular breed. Despite that our study was performed on a limited number of individuals, similar to a recent GWAS performed comparing 21 serologically positive with 27 negative tested goats of an Italian native breed,<sup>9</sup> it also suggests further genetic complexity underlying the resistance/susceptibility to SRLV in sheep.

**Acknowledgements:** We thank the Oberwalliser Schwarznasenzuchtverband and all participating farmers for providing the samples of their animals as well as the Schweizerischer Schafzuchtverband for financial support. **References** Gomez-Lucia E. *et al.* (2018) Vet Med Res

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**Figure S1.** (a) Multidimensional scaling plots of genetic relationships among the 67 VBS in the first three coordinates. (b) Manhattan plot of  $-\log(P\text{-values})$  for the genome wide association study.

**Table S1.** Features of the 67 animals used for this study.

**Table S2.** GWAS results.

**Table S3.** List of annotated genes and loci in the associated genome region.