


Amica C. Müller-Nedebock, BSc (Hons),^{1,2} 
 Morenikeji A. Komolafe, MBBS, FWACP,³
 Michael B. Fawale, MBBS, MSc, FMCP,³
 Jonathan A. Carr, MBChB, PhD,^{2,4}
 Francois H. van der Westhuizen, PhD,⁵

Owen A. Ross, PhD,^{6,7}  and Soraya Bardien, PhD^{1,2*} 
¹Division of Molecular Biology and Human Genetics, Department of Biomedical Sciences, Faculty of Medicine and Health Sciences, Stellenbosch University, Cape Town, South Africa, ²South African Medical Research Council, Stellenbosch University Genomics of Brain Disorders Research Unit, Cape Town, South Africa, ³Neurology Unit, Department of Medicine, College of Health Sciences, Obafemi Awolowo University, Ile-Ife, Nigeria, ⁴Division of Neurology, Department of Medicine, Faculty of Medicine and Health Sciences, Stellenbosch University, Cape Town, South Africa, ⁵Human Metabolomics, North-West University, Potchefstroom, South Africa, ⁶Department of Neuroscience, Mayo Clinic, Jacksonville, Florida, USA, and ⁷Department of Clinical Genomics, Mayo Clinic, Jacksonville, Florida, USA

References

1. La Cognata V, Morello G, D'Agata V, Cavallaro S. Copy number variability in Parkinson's disease: assembling the puzzle through a systems biology approach. *Hum Genet* 2017;136:13–37. <https://doi.org/10.1007/s00439-016-1749-4>.
2. Gentile G, La Cognata V, Cavallaro S. The contribution of CNVs to the most common aging-related neurodegenerative diseases. *Aging Clin Exp Res* 2020. <https://doi.org/10.1007/s40520-020-01485-4>.
3. Okubadejo N, Britton A, Crews C, et al. Analysis of Nigerians with apparently sporadic Parkinson disease for mutations in LRRK2, PRKN and ATXN3. *PLoS One* 2008;3. <https://doi.org/10.1371/journal.pone.0003421>.
4. Milanowski LM, Oshinaike O, Broadway BJ, et al. Early-onset Parkinson disease screening in patients from Nigeria. *Front Neurol* 2021;11. <https://doi.org/10.3389/fneur.2020.594927>.
5. Keyser RJ, Lombard D, Veikondis R, Carr J, Bardien S. Analysis of exon dosage using MLPA in South African Parkinson's disease patients. *Neurogenetics* 2010;11:305–312. <https://doi.org/10.1007/s10048-009-0229-6>.
6. Haylett WL, Keyser RJ, du Plessis MC, et al. Mutations in the parkin gene are a minor cause of Parkinson's disease in the South African population. *Parkinsonism Relat Disord* 2012;18: 89–92. <https://doi.org/10.1016/j.parkreldis.2011.09.022>.
7. van der Merwe C, Carr J, Glanzmann B, Bardien S. Exonic rearrangements in the known Parkinson's disease-causing genes are a rare cause of the disease in South African patients. *Neurosci Lett* 2016;619:168–171. <https://doi.org/10.1016/j.neulet.2016.03.028>.
8. Mahne AC, Carr JA, Bardien S, Schutte CM. Clinical findings and genetic screening for copy number variation mutations in a cohort of South African patients with Parkinson's disease. *S Afr Med J* 2016;106:623–625. <https://doi.org/10.7196/SAMJ.2016.v106i6.10340>.
9. Dekker MCJ, Suleiman JM, Bhwana D, et al. PRKN-related familial Parkinson's disease: first molecular confirmation from East Africa. *Parkinsonism Relat Disord* 2020;73:14–15. <https://doi.org/10.1016/j.parkreldis.2020.02.014>.
10. Yonova-Doing E, Atadzhanov M, Quadri M, et al. Analysis of LRRK2, SNCA, Parkin, PINK1, and DJ-1 in Zambian patients with Parkinson's disease. *Parkinsonism Relat Disord* 2012;18:567–571. <https://doi.org/10.1016/j.parkreldis.2012.02.018>.
11. Karczewski KJ, Francioli LC, Tiao G, et al. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature* 2020; 581:434–443. <https://doi.org/10.1038/s41586-020-03174-8>.
12. Shu L, Zhang Y, Sun Q, Pan H, Tang B. A comprehensive analysis of population differences in LRRK2 variant distribution in Parkinson's disease. *Front Aging Neurosci* 2019;11. <https://doi.org/10.3389/fnagi.2019.00013>.

13. du Toit N, van Coller R, Anderson DG, Carr J, Bardien S. Frequency of the LRRK2 G2019S mutation in South African patients with Parkinson's disease. *Neurogenetics* 2019;20:215–218. <https://doi.org/10.1007/s10048-019-00588-z>.
14. Rizig M, Ojo OO, Athanasiou-Fragkouli A, et al. Negative screening for 12 rare LRRK2 pathogenic variants in a cohort of Nigerians with Parkinson's disease. *Neurobiol Aging* 2021;99:101.e15–101.e19.
15. Dekker MCJ, Coulibaly T, Bardien S, Ross OA, Carr J, Komolafe M. Parkinson's disease research on the African continent: obstacles and opportunities. *Front Neurol* 2020;11. <https://doi.org/10.3389/fneur.2020.00512>.

Supporting Data

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.

Seed Amplification Assay to Diagnose Early Parkinson's and Predict Dopaminergic Deficit Progression

Parkinson's disease (PD) diagnosis relies primarily on clinical evaluation due to lack of validated tests and biomarkers. DaTscan imaging has been used to distinguish psychogenic and drug-induced parkinsonism from idiopathic PD. Some clinically diagnosed PD patients show scans without evidence of dopaminergic deficit (SWEDD), including some that respond to dopaminergic treatment. Some SWEDD patients present abnormal scans consistent with PD many years later

© 2021 Amprion Inc. *Movement Disorders* published by Wiley Periodicals LLC on behalf of International Parkinson Movement Disorder Society.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

***Correspondence to:** Dr. L. Concha-Marambio, Amprion, 11095 Flintkote Avenue Suite D, San Diego, CA 92121, USA; E-mail: luis@ampriondx.com

Relevant conflicts of interest/financial disclosures: Dr. Soto, Dr. Concha, Ms. Farris, Mr. Ma, and Mr. Holguin are inventors on several patents related to the SAA (PMCA) technology and are affiliated to Amprion Inc., a biotech company focusing on the commercial utilization of SAA (PMCA) for diagnosis. Dr. Shahnawaz is also an inventor on several patents related to SAA (PMCA) technology but he is not associated with Amprion. Dr. Kang is on the advisory board of Amprion.

Funding agency: This work was funded by grant #16712 from The Michael J. Fox Foundation for Parkinson's Research to Amprion. The results are available online in the PPMI website under project #155.

Received: 12 April 2021; **Revised:** 13 May 2021; **Accepted:** 14 June 2021

Published online 8 July 2021 in Wiley Online Library (wileyonlinelibrary.com). DOI: 10.1002/mds.28715

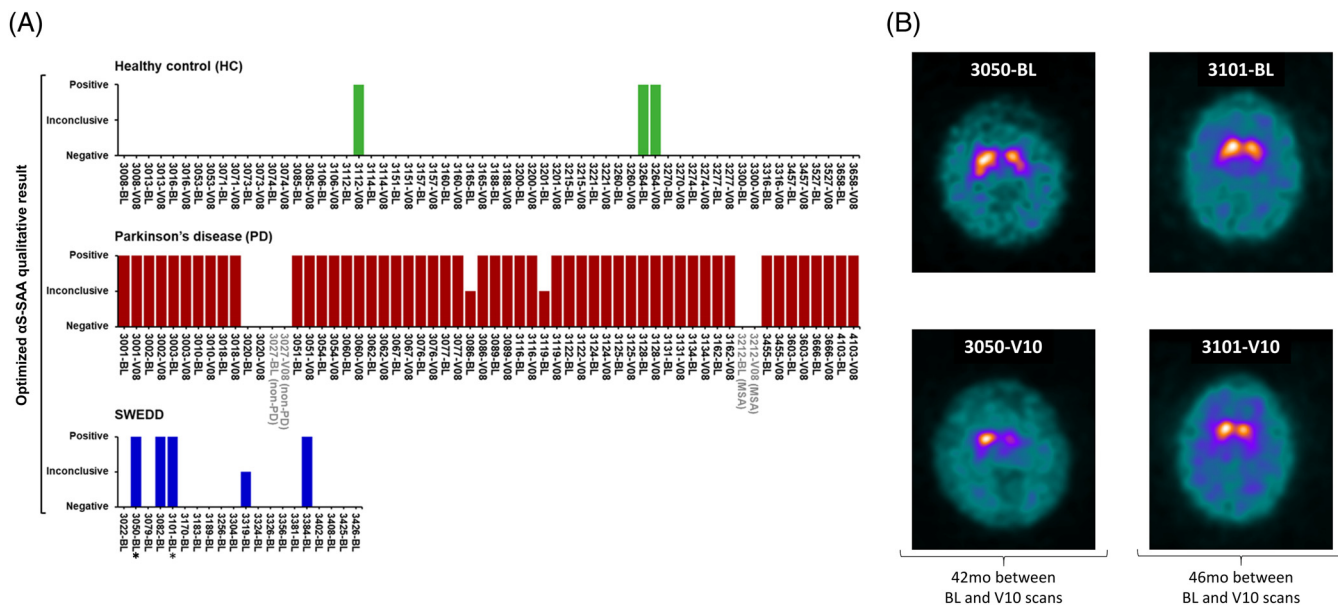


FIG. 1. α -Synuclein seed amplification assay (α S-SAA) results of the 140 cerebrospinal fluid (CSF) samples and DaTscan images of α S-SAA-positive scans without evidence of dopaminergic deficit (SWEDD) patients. **(A)** The data analysis algorithm produced a trinary qualitative outcome (positive, inconclusive, and negative), which was graphed for all 60 healthy control (HC), 60 Parkinson's disease (PD) and 20 SWEDD CSF samples. Each bar represents the α S-SAA result for a sample from a patient collected at a given time point (BL for baseline and V08 for visit 08). Reclassified PD patients are shown in grey; one was reclassified as multiple system atrophy (MSA) confirmed by postmortem pathological examination and the other one as unknown non-PD clinically. SWEDD patients whose abnormal brain scans were predicted by the optimized α S-SAA 42 and 46 months (mo) before the abnormal DaTscans are indicated with an asterisk (*). **(B)** Transverse DaTscan from patient #3050 at BL was considered borderline normal by visual inspection and abnormal 42 months later at visit 10 (V10). BL scan (2010/09/23) showed slightly asymmetric uptake with heterogenous appearance left striatum with specific binding ratio (SBR) lowest putamen equal to 1.04. V10 scan (2014/03/05) shows significant bilateral reduction, with the greatest changes in the left striatum. Within 4 years, there was a significant 25% SBR reduction (SBR-V10 = 0.78). Transverse DaTscan from patient #3101 at BL was considered borderline normal with rotation effect creating a faux reduction in the left putamen (BL-SBR lowest putamen = 1.02). V10 scan (2014/08/22) shows decided asymmetric signal loss, bilateral, with greatest involvement of signal loss on the left, with relatively preserved right caudate. Within 4 years, there was a significant 47% SBR reduction (SBR-V10 = 0.54).

and it is unknown if these patients developed PD in between scans or presented PD with low dopaminergic degeneration.

α -Synuclein seed amplification assays (α S-SAAs) detect α -synuclein (α Syn) aggregates in the cerebrospinal fluid (CSF) of PD, dementia with Lewy bodies (DLB), and isolated rapid eye movement (REM) sleep behavior disorder (iRBD) patients with high sensitivity and specificity.¹⁻⁴ We used an optimized high-throughput α S-SAA (based on a previously described α -Syn protein misfolding cyclic amplification (PMCA) assay)^{2,5,6} that detects α Syn aggregates in CSF, to evaluate 140 blinded samples from the Parkinson's Progression Markers Initiative (PPMI). Samples included baseline (BL) and 3-year follow-up (V08) from 30 PD and 30 healthy controls (HC), and BL samples from 20 SWEDD patients. PD-BL samples were collected within 2 years from diagnosis and presented abnormal DaTscans, while SWEDD patients presented normal DaTscans. PPMI classified enrollees as PD or SWEDD based on visual inspection of their baseline DaTscans.

Figure 1A shows the assay results. The assay performed with 96.2% sensitivity (95% CI: 80.4%–99.9%) and 96.7% specificity (95% CI: 82.8%–99.9%) for PD versus HC at BL, and 96.4% sensitivity (95% CI: 81.7%–99.9%) and 93.8% specificity (95% CI: 79.2%–99.2%) at V08. After α S-SAA analysis, PPMI reclassified two of the three α S-SAA-negative subjects in the PD cohort as non-PD, therefore they were excluded from the above calculation. There were three false-positive samples: #3112-V08 and both samples from patient

#3264. The latter was found to be a probable RBD case based on their RBD questionnaire score. Unfortunately, confirmatory polysomnography is not available and both samples from this patient were considered to be false-positives. Two BL-PD samples were inconclusive. Retest was not possible due to lack of sample and they were excluded from analysis.

Of the 20 BL-SWEDD samples, we found 4 positive, 15 negative, and 1 inconclusive. Second DaTscans available at V10 (4 years from BL) of the negatives were normal. Interestingly, two of the four α S-SAA-positive SWEDD subjects (#3050 and #3101) showed a substantial increase in dopaminergic degeneration by DaTscans at 42 and 46 months after enrollment, with substantial putamen deficits consistent with a PD diagnosis (Fig. 1B). After α S-SAA analysis and after reviewing all longitudinal, clinical, and imaging data, the PPMI analytic cohort consensus committee decided to change the enrollment diagnosis of #3050 and #3101 from SWEDD to PD.



Our results indicate that the optimized α S-SAA is highly accurate compared to the gold standard (longitudinal, clinical, and imaging data). The potential value of unbiased α S-SAA results in a clinical setting can be appreciated in cases with disputable diagnosis, such as the two α S-SAA-negative clinical PD patients (reclassified as non-PD) and the two α S-SAA-positive SWEDD cases (reclassified as PD). The α S-SAA-positive HC with probable RBD is in agreement with recent reports showing prodromal PD diagnosis.^{4,7} Detailed

introduction, results, methods, comparison to the original assay,^{2,5} and discussion are included as Appendix S1. ■

Acknowledgments: We thank the sample donors and their families for supporting this research by participating in the PPMI study. We also want to thank The Michael J. Fox Foundation for their assistance in accessing CSF samples and online information of the cohort.

Data Availability Statement

All data presented in this letter is available at the PPMI database (<https://ida.loni.usc.edu/login.jsp>).

Luis Concha-Marambio, PhD,^{1*}  Carly M. Farris, MS,¹ Bret Holguin, MS,¹ Yihua Ma, MS,¹ John Seibyl, MD,² Marco J. Russo, MD, PhD,³ Un J. Kang, MD,³  Samantha J. Hutten, PhD,⁴ Kalpana Merchant, PhD,⁵ Mohammad Shahnawaz, PhD,⁶ and Claudio Soto, PhD^{1,6}
¹R&D Unit, Amprion Inc., San Diego, California, 92121, USA, ²Institute for Neurodegenerative Disorders, New Haven, Connecticut, USA, ³Department of Neurology, Neuroscience Institute, NYU Grossman School of Medicine, New York, New York, USA, ⁴The Michael J. Fox Foundation for Parkinson's Research, New York, New York, USA, ⁵Northwestern University Feinberg School of Medicine, Chicago, Illinois, USA, and ⁶Mitchell Center for Alzheimer's Disease and Related Brain Disorders, University of Texas McGovern Medical School, Houston, Texas, USA

References

1. Fairfoul G, McGuire LI, Pal S, et al. Alpha-synuclein RT-QuIC in the CSF of patients with alpha-synucleinopathies. *Ann Clin Transl Neurol* 2016;3(10):812–818. <https://doi.org/10.1002/acn3.338>.
2. Shahnawaz M, Tokuda T, Waragai M, et al. Development of a biochemical diagnosis of Parkinson disease by detection of α -synuclein misfolded aggregates in cerebrospinal fluid. *JAMA Neurol* 2017;74(2):163. <https://doi.org/10.1001/jamaneurol.2016.4547>.
3. Groveman BR, Orrù CD, Hughson AG, et al. Rapid and ultra-sensitive quantitation of disease-associated α -synuclein seeds in brain and cerebrospinal fluid by α Syn RT-QuIC. *Acta Neuropathol Commun* 2018;6(1):7. <https://doi.org/10.1186/s40478-018-0508-2>.
4. Iranzo A, Fairfoul G, Ayudhaya ACN, et al. Detection of α -synuclein in CSF by RT-QuIC in patients with isolated rapid-eye-movement sleep behaviour disorder: a longitudinal observational study. *Lancet Neurol* 2021;20(3):203–212. [https://doi.org/10.1016/S1474-4422\(20\)30449-X](https://doi.org/10.1016/S1474-4422(20)30449-X).
5. Concha-Marambio L, Shahnawaz M, Soto C. Detection of misfolded α -synuclein aggregates in cerebrospinal fluid by the protein misfolding cyclic amplification platform. *Methods Mol Biol* 1948;2019:35–44. https://doi.org/10.1007/978-1-4939-9124-4_4.
6. Shahnawaz M, Mukherjee A, Pritzkow S, et al. Discriminating α -synuclein strains in Parkinson's disease and multiple system atrophy. *Nature* 2020;578(7794):273–277. <https://doi.org/10.1038/s41586-020-1984-7>.
7. Stefani A, Iranzo A, Holzkecht E, et al. Alpha-synuclein seeds in olfactory mucosa of patients with isolated REM sleep behaviour disorder. *Brain* 2021;144(4):1–9. <https://doi.org/10.1093/brain/awab005>.

Supporting Data

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.

A Role of Aging in the Progression of Cortical Excitability in Benign Adult Familial Myoclonus Epilepsy type 1 Patients

Recent molecular genetic studies have revealed that abnormal intronic TTTTA and TTTC A repeat expansion in sterile alpha motif domain containing 12 (*SAMD12*) causes benign adult familial myoclonus epilepsy (BAFME) type 1.¹ This provides evidence of an inverse correlation between the expanded repeat sizes and seizure onset age. However, the effect of non-coding expanded repeat length on symptom progression during the late lifetime stage remains obscure. This knowledge gap is of interest given that cortical excitability gradually worsens during late-stage BAFME.²

We evaluated 18 patients with genetically diagnosed BAFME type 1 and previously identified abnormal TTTTA and TTTC A repeat lengths¹ (Table S1). Partial correlation analyses were conducted for internal correlations among cortical excitability (amplitudes of early components of cortical somatosensory-evoked potential [SEP], ie, N20, P25, and N33),³ aging (current age at SEP examination), and genotype (sum of the expanded TTTTA and TTTC A repeat sizes).¹ We also investigated the associations among patients' age, genotype, and an enhanced long-loop reflex (C-reflex).

© 2021 International Parkinson and Movement Disorder Society

***Correspondence to:** Dr. A. Ikeda, Department of Epilepsy, Movement Disorders and Physiology, Kyoto University Graduate School of Medicine, Shogoin, Sakyo-ku, Kyoto 606-8507, Japan; E-mail: akio@kuhp.kyoto-u.ac.jp

Relevant conflicts of interest/financial disclosures: Shuichiro Neshige, Takefumi Hitomi, Maya Tojima, Kazuki Oi, Katsuya Kobayashi, Akihiro Shimotake, Riki Matsumoto, Masutaro Kanda, Hirofumi Maruyama, Hiroyuki Ishiura, Shoji Tsuji, and Ryosuke Takahashi report no disclosures relevant to the manuscript. Masao Matsushashi, and Akio Ikeda are the current members of the Department of Epilepsy, Movement Disorders and Physiology. This department is an endowment department supported by a grant from GlaxoSmithKline, Inc., Nihon Kohden Corporation, Otsuka Pharmaceutical Co., and UCB Japan Co. Ltd. until May 2018. Since June 2018, this department has changed to the Industry-Academia Collaboration Courses supported by Eisai Co. Ltd., Nihon Kohden Corporation, Otsuka Pharmaceutical Co., and UCB Japan Co. Ltd.

Funding agencies: This study was supported by the Kyoto University Research Fund for Senior Scientists and by a Grant-in-Aid for Scientific Research on Innovative Areas (Non-linear Neuro-oscillology: Towards Integrative Understanding of Human Nature) KAKENHI grant number 15H05874, a Grant-in-Aid for Scientific Research (C) KAKENHI grant number 17K09798, a Grant-in-Aid for Scientific Research (B) KAKENHI grant number 19H03574, and a Grant-in-Aid for Scientific Research (C) KAKENHI grant number 20K07902 from the Ministry of Education, Culture, Sports, Science and Technology (MEXT); Japan Society for the Promotion of Science (JSPS) KAKENHI grant number 26293209; Japan Society for the Promotion of Science (JSPS) KAKENHI grant number 20H03588; the Japan Agency for Medical Research and Development grant numbers 18kk0205001, 19ek0109279, and 20ek0109491; the Japan Epilepsy Research Foundation.

Received: 21 January 2021; **Revised:** 27 June 2021; **Accepted:** 29 June 2021

Published online 24 July 2021 in Wiley Online Library (wileyonlinelibrary.com). DOI: 10.1002/mds.28718