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## Serotonin 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> receptors in the prefrontal cortex of Alzheimer and normal aging patients

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### Abstract

**Background:** It has been hypothesized that alterations of the serotonergic system contribute to neuropsychiatric symptoms in Alzheimer disease (AD). Cellular expressions of the two serotonergic receptors 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> have therefore been determined by immunohistochemistry in the prefrontal cortex of patients with AD (n=6) and normal age-matched controls (n = 7).

**Results:** In normal aging patients, 5-HT<sub>2A</sub> label was mainly observed in large pyramidal cells, but to a lesser extent also in small pyramidal cells and in stellate cells of cortical layers II-VI. In AD, a similar distribution was observed, but density of positive cells was significantly reduced by 33%. In aging control patients, the 5-HT<sub>6</sub> receptor was expressed by pyramidal cells and occasional stellate cells, not only of layers II-V, but also of layer I, where a distinct label was observed in neurons and surrounding fibers. 5-HT<sub>6</sub> receptor expression in AD patients had the same pattern, but was significantly decreased by 40%.

**Conclusion:** Our results indicate that a decline in neurons expressing 5-HT<sub>2A</sub>, but also 5-HT<sub>6</sub> receptors may play a role in the etiopathology of neuropsychiatric symptoms in AD.

### Background

Alzheimer disease (AD), the most common cause of dementia in the elderly, is clinically characterized by progressive cognitive impairment associated with severe neuropsychiatric disturbances. These behavioral and psychological symptoms of dementia (BPSD) include hallucinations, delusions, aggressive behavior, overactivity, anxieties and affective disturbances [1,2]. Whereas the decline in cognitive functions can be largely related to cholinergic dysfunction arising from disruption of basal forebrain cholinergic pathways (cholinergic hypothesis) [1,3,4], impaired balance between several neurotransmit-

ters has been implicated in the pathogenesis of BPSD [2,5-7], with serotonin (5-HT) playing a pivotal role [2,8-10]. The actions of 5-HT are mediated through seven major receptors classes, 5-HT<sub>1-7</sub>, comprising a total of 14 distinct mammalian 5-HT receptor subtypes (for review, see [11]). The 5-HT<sub>2A</sub> receptor has attracted most interest because of its possible participation in behavioral alterations in AD. 5-HT<sub>2A</sub> is localized in the cortex and caudate and is involved in anxiety [10]. 5-HT<sub>2A</sub> receptors mediate the psychotomimetic effects of hallucinogens [11-13], and alterations in binding characteristics to this receptor have been observed in the prefrontal cortex of patients suffer-

ing from psychiatric diseases, e.g. schizophrenia [14,15], depression [16] and suicide [17]. Electrophysiological evidence suggests that 5-HT<sub>2A</sub> receptors are involved in the 5-HT-induced increase in excitatory postsynaptic potentials [12,18] and play an important role in the working memory process [13]. There is indication that 5-HT<sub>2A</sub> also participates in the etiopathology of AD. Serotonin increases the secretion of amyloid precursor protein (APP) through activation of 5-HT<sub>2A</sub> receptors [19]. 5-HT<sub>2A</sub> receptor binding is decreased in AD (for review, see [9,10]), and polymorphic variations have been described for the 5-HT<sub>2A</sub> gene that may be risk factors for hallucinations [20], aggression [21] and major depression [22] in AD. Much less is known about the 5-HT<sub>6</sub> receptor, the most recent 5-HT receptor to be identified. Many antidepressants and antipsychotics are antagonists of the 5-HT<sub>6</sub> receptor [15,23]. It has been shown to influence acetylcholine release in the frontal cortex [24] and may play a role in cognition deficits and in some form of anxiety [23]. A recent autoradiographic study examining [<sup>125</sup>I]-SB-258585 binding in autopsy specimens of AD patients indicates lowered 5-HT<sub>6</sub> receptor density in the frontal and temporal cortices [5]. Association of a 5-HT<sub>6</sub> receptor gene polymorphic variant and late-onset AD has been observed in the Chinese population [25], but not in Germans [26].

Surprisingly, although 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> receptor binding in AD has been studied extensively [2,5,9,10], there is no information on the expression of these receptors at the cellular level. We have therefore examined the expression of the 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> receptors in the brains of AD and of normal aging control patients by immunohistochemistry. Because the prefrontal cortex is of particular importance for the etiopathology of BPSD [1,8,12,14,16,27-31] and because neuropsychiatric symptoms in AD are associated with reduced metabolism [32,33] and perfusion [34]

in Brodmann area 10, we have chosen this brain region for examination.

**Methods**

**Tissue samples**

The brains were obtained from a total of 13 autopsy cases with the consent of a close relative and with full approval by the ethical committees of Guangzhou hospital authorities and of the Chinese University of Hong Kong. Six of the specimens were from individuals with clinically and pathologically diagnosed senile dementia of the Alzheimer's type (mean age: 88 years, see table 1), seven specimens were from individuals, matched for age, gender and *postmortem* delay, who had no history of neurological diseases (normal control). Drug history was recorded for all patients, who had only received antibiotic treatment, but no psychopharmacological medication. Alzheimer Disease was clinically diagnosed according to the "National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association" (NINCDS-ADRA) criteria and the "Diagnostic and Statistical Manual of Mental Disorders", Fourth Edition, (DSM-IV-R) criteria.

Whole brains were removed with an average *postmortem* delay of six hours, and tissue samples of the anterior prefrontal cortex (Brodmann's area 10) were dissected. Specimens were fixed overnight in 4% paraformaldehyde in phosphate-buffered saline (PBS; pH 7.4), dehydrated in graded ethanol, cleared with xylene and embedded in paraffin. 6-µm-thick serial sections were cut in the coronal plane, 90° perpendicular to the surface to avoid sectioning artifacts. Two sets of slides were obtained for each individual.

**Immunohistochemistry**

All steps were carried out at room temperature unless stated otherwise. Mounted sections were dewaxed, rehy-

**Table 1: Profiles of Alzheimer (AD) and matched normal aging patients**

| Subject | Diagnosis | Sex | Age | Cause of death            |
|---------|-----------|-----|-----|---------------------------|
| 1       | Normal    | F   | 83  | Pneumonia                 |
| 2       | Normal    | F   | 86  | Gastrointestinal bleeding |
| 3       | Normal    | M   | 82  | Pneumonia                 |
| 4       | Normal    | M   | 93  | Sudden death              |
| 5       | Normal    | F   | 72  | Sudden death              |
| 6       | Normal    | F   | 85  | Pneumonia                 |
| 7       | Normal    | F   | 81  | Pneumonia                 |
| 8       | Alzheimer | M   | 82  | Subarachnoid hemorrhage   |
| 9       | Alzheimer | F   | 91  | Septicemia                |
| 10      | Alzheimer | F   | 96  | Pneumonia                 |
| 11      | Alzheimer | F   | 82  | Pneumonia                 |
| 12      | Alzheimer | F   | 86  | Pneumonia                 |
| 13      | Alzheimer | F   | 89  | Sudden death              |

drated and predigested with 0.1% trypsin (BDH Laboratory Supplies, Poole, U.K.) in 0.05M tris buffered saline containing 0.1% CaCl<sub>2</sub> (pH 7.6; 37°C; 20 min) followed by two rinses (5 min) in 0.01M PBS for antigen retrieval. The endogenous peroxidase activity was blocked with 0.3% hydrogen peroxide in absolute methanol for 30 min, followed by another three rinses in PBS (5 min each). To suppress non-specific binding, sections were then incubated for 1h in 2 % normal blocking serum (Vectastain<sup>R</sup> ABC Kit, Vector Laboratories, Burlingame, CA) in 0.3 % triton/PBS. Thereafter, sections were incubated overnight with the primary polyclonal antibodies: either goat anti-human serotonin 2A receptor (Santa Cruz sc-15073, Santa Cruz Biotechnology Inc., Sta. Cruz, CA) or goat anti-human serotonin 6 receptor (Santa Cruz sc-26668). The sections were then washed with three rinses of PBS containing 0.05 % Tween 20 (5 min each) and incubated with biotinylated secondary antibody in blocking solution (1:200) for 30 min (PK6105, anti-goat IgG, Vectastain<sup>R</sup> ABC Kit, Vector Laboratories, Burlingame, CA). Sections were washed three times in PBS again (5 min) and incubated with ABC Reagent (Vectastain<sup>R</sup> ABC Kit, Vector Laboratories, Burlingame, CA) for 30 min. The immunocytochemical staining signals were visualized by incubating the sections in 0.05% of the substrate 3'3'-diaminobenzidine tetrahydrochloride (DAB) in PBS containing 0.01% H<sub>2</sub>O<sub>2</sub>. All stainings included negative controls with omission of the primary antibody, which did not show any immunoreaction. The sections were washed with distilled water and then counterstained with 0.5% cresyl fast violet in 0.1M sodium acetate (pH 3.5, 5 min). They were differentiated with 95% ethanol, dehydrated, cleared and covered in Permount<sup>®</sup> (Fisher Scientific, Hampton, VA). Additional sections were stained with routine methods, including H&E, Nissl and Bielschowsky silver impregnation for tangle staging [35,36].

All cases underwent a standardized post-mortem neuropathological assessment of AD using established criteria including Braak staging [35]. Four brain regions were examined: prefrontal (area 10), occipital (area 17), entorhinal (area 28) and hippocampal cortices. Diagnosis was confirmed by the characteristic presence of amyloid plaques, neurofibrillary tangles (NFT) and neuronal degeneration. In addition, the presence of  $\beta$ -amyloid in the plaques was ascertained by immunohistochemistry using the rabbit polyclonal Anti-Amyloid Peptide  $\beta$ , Cleavage site 42 (A $\beta$ 42), antibody (A1976, Sigma, St. Louis, MU) [37]. All AD cases had neocortical NFT with involvement of both isocortical association areas and additional involvement of primary cortical (area 17) areas (Braak stage V-VI). In control patients, deposition of NFT was negligible in the entorhinal cortex and absent in the hippocampus as well as the neocortex (Braak stage = I). To exclude Dementia with Lewy bodies, all brain specimens

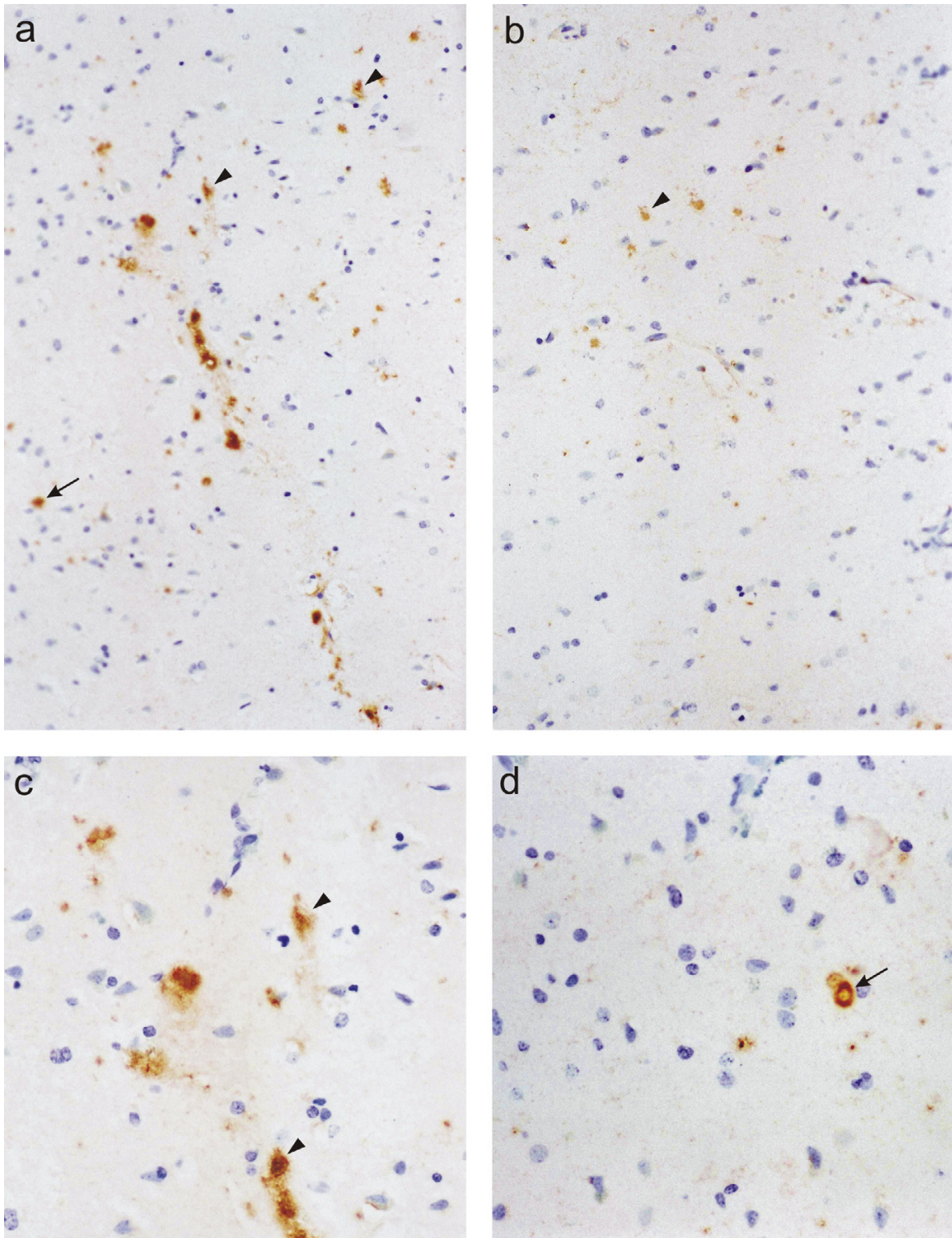
were scrutinized for Lewy bodies, and only cases devoid of Lewy bodies were included in the study.

### Statistics

For qualitative and quantitative evaluation, sections were observed under a photomicroscope (Axioplan 2 Photomicroscope, Zeiss, Germany). The number of 5-HT<sub>2A</sub>- and 5-HT<sub>6</sub>-immunoreactive cells was counted in 30 random selected 700  $\mu$ m<sup>2</sup> fields of normal aged and AD patients. 4–5 sections per individual were evaluated, sparing areas containing amyloid plaques. It has been shown in AD patients [38] that astrocytes in and around A $\beta$  plaques are strongly positive for 5-HT<sub>2A</sub> receptor protein, whereas astrocytes in control patients do not display any 5-HT<sub>2A</sub> immunoreaction. Amyloid plaques were therefore excluded from the quantitative assessment, because the aim of our study was to count neurons expressing 5-HT receptors. Measurements were expressed as means  $\pm$  SEM. For statistical comparison, the values were subjected to a one-way analysis of variance (ANOVA), demonstrating that the four data groups exhibited equal variance, but were not distributed normally. We have then applied the One Way non-parametric ANOVA (Kruskal-Wallis test) and the Mann Whitney Rank sum test, yielding the same results. A p value below 0.05 was considered significant.

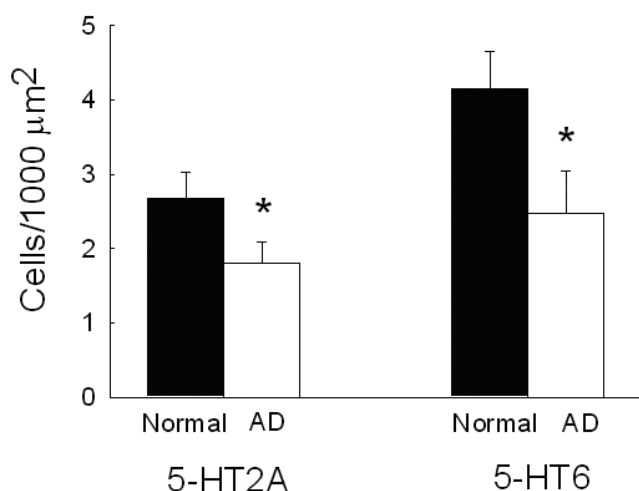
### Results

In the prefrontal cortex of normal aging patients, 5-HT<sub>2A</sub> receptor immunoreaction was observed in cortical layers II-VI. Large pyramidal neurons of layer V constituted the majority of immunoreactive cells, characterized by a strongly labeled cytoplasm and a moderately stained proximal part of the apical dendrite (Figs. 1a, c). In addition, smaller pyramidal cells and scattered stellate-shaped cells in the other layers were immunoreactive. Stellate cells had a strongly stained cytoplasm and labeled thin processes radiating to the side of the ovoid cell bodies and most likely represented interneurons. A few multipolar cells in layer VI were also 5-HT<sub>2A</sub> receptor positive. In the underlying white matter, occasional 5-HT<sub>2A</sub> receptor immunoreactive fiber bundles were observed. AD patients showed a similar distribution of 5-HT<sub>2A</sub> receptor label in the prefrontal cortex, i.e. staining in the cell bodies and apical dendrites of large pyramidal cells and in the soma and tender processes of a few scattered interneurons (Figs. 1b, d). However, numerical density of 5-HT<sub>2A</sub> receptor immunoreactive cells was significantly ( $p=0.001$ ) decreased by 33% in the frontal cortex of AD patients (Fig. 2), as compared to normal aging patients. Reduction affected both pyramidal cells and cells of stellate morphology. In addition, labeled cells were also seen within and close to amyloid plaques. An association between 5-HT<sub>2A</sub> receptors and NFT was not observed.



**Figure 1**  
**5-HT<sub>2A</sub> receptor.** Immunoreaction for the 5-HT<sub>2A</sub> receptor in the prefrontal cortex of a normal aging (a, c) and an Alzheimer patient (b, d), cresyl violet counterstain. Both large pyramidal cells (arrowheads) and small interneurons (arrows) are stained. Note the reduction in labeled cells in the cortex of the Alzheimer patient (b, d). a, b: x200; c, d: x400.





**Figure 2**  
**Cell densities.** Numerical density of 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> receptor immunoreactive neurons in the prefrontal cortex of normal aging (n=7) and Alzheimer (AD) patients (n=6). Depicted are means ± SEM. Significant differences ( $p \leq 0.05$ ) are marked by an asterisk (\*).

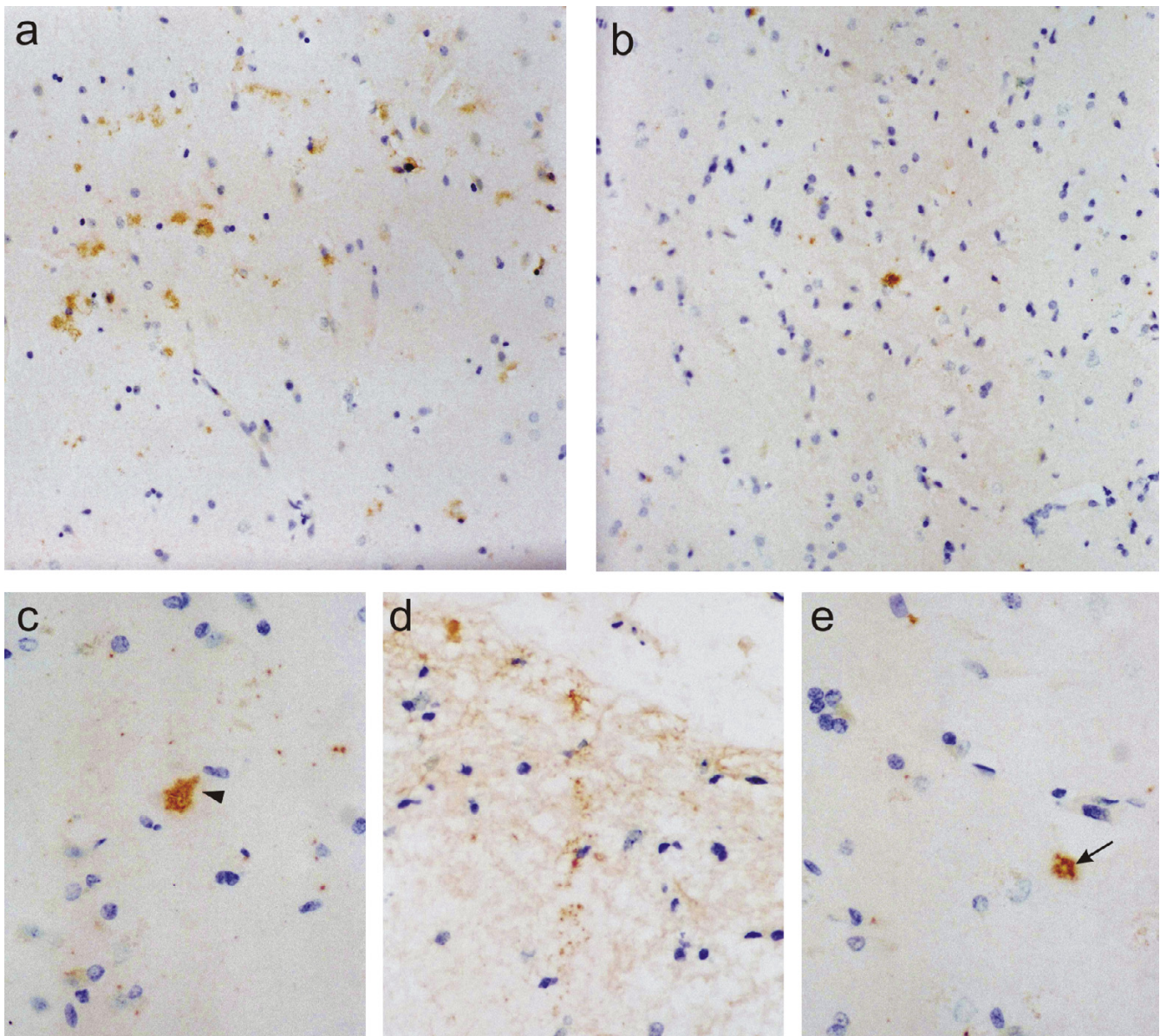
Distribution of 5-HT<sub>6</sub> receptor immunoreaction in the prefrontal cortex of normal aging patients was slightly different from that of the 5-HT<sub>2A</sub> receptor. As for the 5-HT<sub>2A</sub> receptor, the most prominent label was observed in the cell bodies and the proximal apical dendrites of large pyramidal neurons, and occasionally, the soma and fine processes of scattered stellate cells were stained as well (Figs. 3a, c). In addition, 5-HT<sub>6</sub> receptor immunoreaction was also detected in neurons of the molecular layer (layer I). These cells had a bipolar strongly labeled cell body and numerous immunoreactive processes radiating into the molecular layer (Fig. 3d). In contrast, hardly any 5-HT<sub>6</sub> receptor immunoreactive cells were observed in layer VI. Occasional immunoreactive fiber bundles were seen in the underlying white matter. In summary, density of 5-HT<sub>6</sub> receptor positive neurons was significantly higher than that of 5-HT<sub>2A</sub> receptor immunoreactive cells ( $p=0.05$ ). In the prefrontal cortex of AD patients, 5-HT<sub>6</sub> receptor label showed the same overall distribution (Figs. 3b, e), but numerical density of 5-HT<sub>6</sub> receptor immunoreactive cells was again significantly ( $p=0.001$ ) reduced, as compared to normal aging patients (Fig. 2). Both pyramidal and stellate cells were decreased by about 40%. There was no association between 5-HT<sub>6</sub> receptors and NFT.

## Discussion

The present study has been undertaken in order to determine changes in cellular distribution of two serotonin receptors, 5-HT<sub>2A</sub> and 5-HT<sub>6</sub>, in the prefrontal cortex of AD patients as compared to normal age-matched individuals. Our observation in aging (control) patients, showing

5-HT<sub>2A</sub> immunoreactivity in the cell bodies and the apical dendrites of pyramidal cells as well as in stellate-shaped cells, is consistent with previous studies in young humans [17], primates [18,39] and rodents [40,41]. However, we have detected relatively few labeled cells, and we are currently testing if the paucity in 5-HT<sub>2A</sub> receptors is attributable to the very advanced age of our patients or to the technique employed. A considerable age-related reduction in the number of cortical 5-HT<sub>2A</sub> binding sites in the frontal lobe has been described by numerous authors (see [9] for review). In contrast to 5-HT<sub>2A</sub>, relatively little is known on the distribution of the 5-HT<sub>6</sub> receptor in the mammalian neocortex. 5-HT<sub>6</sub> receptor binding sites have been detected by receptor autoradiography in the gray matter of the prefrontal cortex of healthy human adults [5,42], but there are no studies on the cellular level. We have been able to show that the 5-HT<sub>6</sub> receptor is expressed both by pyramidal cells and by stellate-shaped cells located in cortical layers I-V. We have observed very little 5-HT<sub>6</sub> immunoreaction in layer VI, but a distinct 5-HT<sub>6</sub> label in layer I, with staining detected not only in a dense network of fibers, but also in neurons. This is somewhat at variance with a study on the rat neocortex, where 5-HT<sub>6</sub> receptor mRNA has been detected by in-situ hybridization in layers II-VI, but not in layer I [43]. This discrepancy is, however, not surprising, given the marked interspecies differences described [44].

To our knowledge, this is the first immunohistochemical study of 5-HT<sub>2A</sub> receptor expression in AD. The reduction observed corroborates the results of previous 5-HT<sub>2A</sub> receptor binding studies showing decreased binding of <sup>3</sup>[H] ketanserin [45-47] and <sup>3</sup>[H] spiperone [48] in *post-mortem* specimens of the frontal cortex of AD patients and in PET imaging studies [49]. Contrasting reports describing unaltered <sup>3</sup>[H] ketanserin binding [50], which are at variance with our results, may be attributable to recent psychotropic medication, which had not been administered to our patients. Reduced 5-HT<sub>2A</sub> receptor binding in AD has been explained by a loss of interneurons [51]. In contrast, our immunohistochemical data indicate that both 5-HT<sub>2A</sub> immunoreactive pyramidal and stellate-shaped cells, i.e. interneurons, are affected in AD. Moreover, we have demonstrated that the density of neurons expressing the 5-HT<sub>6</sub> receptor is reduced to a similar extent, corroborating autoradiographic studies revealing decreased binding to 5-HT<sub>6</sub> receptors in the prefrontal cortex of AD patients [5]. Our observation of reduced density of neurons expressing 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> receptors, which is not accompanied by similar alterations in GABAergic markers [6], points at a selective vulnerability of neurons receiving serotonergic input. This decrease is associated with several other abnormalities of the serotonergic system in AD. A marked depletion in 5-HT and its metabolite 5-Hydroxyindole Acetic Acid (5-HIAA) has been



**Figure 3**

**5-HT<sub>6</sub> receptor.** 5-HT<sub>6</sub> receptor immunoreactive cells in the prefrontal cortex of normal aging (a, c, d) and Alzheimer patients (b, e), cresyl violet counterstain. Label is observed in large pyramidal cells (arrowhead) and small interneurons (arrow). Note positive cells and dense immunoreactive fibers in the molecular layer (d). In the cortex of the Alzheimer patient (b, e), significantly fewer cells are labeled than in that of the normal aging patient (a, c, d). a, b: x200; c, d, e: x400.

described in the frontal and temporal cortices of AD patients [2,28,52], which is most likely attributable to a reduction in serotonergic projection fibers. The underlying cause appears to be a significant loss of neurons in the dorsal and median raphe nuclei [53,54], which are the source of serotonergic nerve terminals and which, in addition, are a preferential site for neurofibrillary tangle formation [55]. It is very likely that the decrease in neurons expressing 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> receptors is related to this

loss of serotonergic fibers. However, it may either reflect a process of primary cortical degeneration [4,56] leading to presynaptic disturbance of the serotonergic system, or be secondary to the decrease in serotonergic afferents.

Several lines of evidence indicate that serotonergic dysfunction in AD has important functional consequences. In a study using retrospective data, loss in 5-HT<sub>2A</sub> binding has been confined to AD patients with aggressive symp-

toms [57]. When cognitive function is assessed *antemortem*, 5-HT<sub>2A</sub> receptors are lost in the frontal and temporal cortex only in patients with severe, but not with mild to moderate dementia [47]. The decrease in the expression of 5-HT<sub>6</sub> receptors in the prefrontal cortex of AD patients can be correlated to the extent of aggressive behavior [5]. The number of 5-HT uptake sites is significantly reduced in the frontal and temporal cortex of AD patients with persistent depression, anxiety and overactivity, compared with AD patients without these symptoms [52], and 5-HT levels in the prefrontal cortex (Brodmann 10) of AD correlate with overactivity [2].

### Conclusion

Our results of normal aging patients, showing that 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> receptors are expressed by both pyramidal cells and stellate-shaped neurons, indicate that serotonergic fibers exert their influence upon these two neocortical cell types not only through the 5-HT<sub>2A</sub> but also the 5-HT<sub>6</sub> receptor. The significant 33–40% reduction in cells immunoreactive for 5-HT<sub>2A</sub> and 5-HT<sub>6</sub> receptors observed in AD patients, affecting both large pyramidal cells and interneurons, points at a severely compromised serotonergic system in AD, involving not only serotonergic projection fibers, but also their corresponding receptors. This decline in receptors most likely contributes to the development of neuropsychiatric symptoms in AD.

### Authors' contributions

Dietrich E. Lorke performed the evaluation of the slides, interpreted the results and wrote the manuscript. Gang Lu performed immunohistochemistry and documented the results. Eric Cho performed the quantitative and statistical analyses. David T. Yew conceived and planned the study, performed the neuropathological examination and evaluated the slides.

### Abbreviations

AD: Alzheimer disease

BPSD: behavioral and psychological symptoms of dementia

5-HT: serotonin

5-HT<sub>2A</sub>: serotonergic receptor 2A

5-HT<sub>6</sub>: serotonergic receptor 6

NFT: neurofibrillary tangles

PBS: phosphate-buffered saline

SEM: standard error of the mean

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