

Draft Genome Sequence of *Gordonia alkanivorans* Strain CGMCC6845, a Halotolerant Hydrocarbon-Degrading Bacterium

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***Gordonia alkanivorans* strain CGMCC6845 is a halotolerant hydrocarbon-degrading bacterium isolated from petroleum-contaminated saline soil. Here we present the 5.0-Mb draft genome sequence of this strain, which will improve our understanding of the diversity of *G. alkanivorans* and the mechanisms of microbial hydrocarbon degradation in saline environment.**

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Gordonia alkanivorans can degrade diesel (1), dibenzothiophene, benzothiophene, and other thiophene analogs (2), which may have potential applications in the bioremediation of petroleum-contaminated soils and the biodesulfurization of oil. Therefore, hydrocarbon biodegradation processes (1), biodesulfurization-related genes (3–5), and optimal conditions for the biodesulfurization (6) of *G. alkanivorans* have been well studied. Despite the increasing interest in *G. alkanivorans*, genomic information for this species is still limited. To date, only one strain, *G. alkanivorans* NBRC16433, has been sequenced. Here, the draft genome sequence of the halotolerant hydrocarbon-degrading strain *G. alkanivorans* CGMCC6845 is presented, which was isolated from petroleum-contaminated saline soil in China (7).

Genomic DNA was extracted from a 3-day culture growing in tryptic soy broth and was sequenced using the Illumina HiSeq 2000 platform with a whole-genome shotgun (WGS) strategy. The sequencing produced 8,467,694 paired-end reads with an insert size of 300 bp, yielding about 170-fold coverage. Filtered reads were assembled, scaffolded, gap filled, and validated using SOAPdenovo v2.04 (8), SSPACE v2.0 (9), GapFiller v1.10 (10), and BWA v0.7.4 (11). Final assembly consisted of 74 contigs with an N_{50} length of 217,608 bp, which were assembled into 61 scaffolds with an N_{50} length of 269,646 bp. Genome annotation was performed using the NCBI Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) (http://www.ncbi.nlm.nih.gov/genome/annotation_prok/).

The genome consists of 5.0 Mb, with a G+C content of 67.5%. A total of 4,476 coding sequences (CDS), 42 pseudogenes, 1 noncoding RNA (ncRNA), 49 tRNA genes, and 3 rRNA operons were identified. A plasmid partitioning gene, *parA*, was detected on contig 20, which suggests the occurrence of a plasmid. We detected 349 tandem repeats using Tandem Repeats Finder v4.07 (14). Neither prophage sequences nor clustered regularly interspaced short palindromic repeat (CRISPR) elements were present,

as revealed by PHAST (12) and CRISPRFinder (13). IS3 and IS481 families dominate the insertion sequence (IS) elements, as revealed by ISFinder (14). Average nucleotide identity (ANI) analysis (15) revealed that *G. alkanivorans* CGMCC6845 is phylogenetically related to *G. alkanivorans* NBRC16433 (98.0%).

Nine genes were identified as involved in hydrocarbon degradation, including 3 alkanal monooxygenase genes, 2 catechol 1,2-dioxygenase genes, 2 ring-cleavage dioxygenase genes, 2 benzoate 1,2-dioxygenase genes, and 1 dibenzothiophene desulfurization gene. Moreover, 8 genes were identified as involved in compatible solute synthesis and uptake, including 1 ectoine synthase gene, 2 betaine synthase genes, 3 trehalose synthase genes, and 2 glycine/betaine ABC transport genes, which may enhance the tolerance to osmotic stress. Copper-, arsenic-, and tellurium-resistant genes were detected, which may enhance the resistance to heavy metal. Information about the genome sequence of *G. alkanivorans* CGMCC6845 will give a better understanding of the diversity of *G. alkanivorans* and the mechanisms of microbial hydrocarbon degradation in saline environment.

Nucleotide sequence accession number. The draft genome sequence of *G. alkanivorans* CGMCC6845 has been deposited in GenBank under the accession number [AYXO000000000](https://www.ncbi.nlm.nih.gov/nuccore/AYXO000000000). The version described in this paper is the first version.

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REFERENCES

1. Young CC, Lin TC, Yeh MS, Shen FT, Chang JS. 2005. Identification and kinetic characteristics of an indigenous diesel-degrading *Gordonia*

- alkanivorans* strain. World J. Microbiol. Biotechnol. 21:1409–1414. <http://dx.doi.org/10.1007/s11274-005-5742-7>.
2. Alves L, Salgueiro R, Rodrigues C, Mesquita E, Matos J, Gírio FM. 2005. Desulfurization of dibenzothiophene, benzothiophene, and other thiophene analogs by a newly isolated bacterium, *Gordonia alkanivorans* strain 1B. Appl. Biochem. Biotechnol. 120:199–208. <http://dx.doi.org/10.1385/ABAB:120:3:199>.
 3. Alves L, Melo M, Mendonca D, Simoes F, Matos J, Tenreiro R, Gírio FM. 2007. Sequencing, cloning and expression of the *dsz* genes required for dibenzothiophene sulfone desulfurization from *Gordonia alkanivorans* strain 1B. Enzyme Microb. Technol. 40:1598–1603. <http://dx.doi.org/10.1016/j.enzmictec.2006.11.008>.
 4. Shavandi M, Sadeghizadeh M, Khajeh K, Mohebbali G, Zomorodipour A. 2010. Genomic structure and promoter analysis of the *dsz* operon for dibenzothiophene biodesulfurization from *Gordonia alkanivorans* RPI90A. Appl. Microbiol. Biotechnol. 87(4):1455–1461. <http://dx.doi.org/10.1007/s00253-010-2605-4>.
 5. Shavandi M, Sadeghizadeh M, Zomorodipour A, Khajeh K. 2009. Biodesulfurization of dibenzothiophene by recombinant *Gordonia alkanivorans* RPI90A. Bioresour. Technol. 100(1):475–479. <http://dx.doi.org/10.1016/j.biortech.2008.06.011>.
 6. Mohebbali G, Ball AS, Rasekh B, Kaytash A. 2007. Biodesulfurization potential of a newly isolated bacterium, *Gordonia alkanivorans* RPI90A. Enzyme Microb. Technol. 40:578–584. <http://dx.doi.org/10.1016/j.enzmictec.2006.05.012>.
 7. Wang X, Han Z, Bai Z, Tang J, Ma A, He J, Zhuang G. 2011. Archaeal community structure along a gradient of petroleum contamination in saline-alkali soil. J. Environ. Sci. (China) 23:1858–1864. [http://dx.doi.org/10.1016/S1001-0742\(10\)60640-7](http://dx.doi.org/10.1016/S1001-0742(10)60640-7).
 8. Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Xiaoqian Z, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. Gigascience 1:18. <http://dx.doi.org/10.1186/2047-217X-1-18>.
 9. Boetzer M, Henkel CV, Jansen HJ, Butler D, Pirovano W. 2011. Scaffolding pre-assembled contigs using SSPACE. Bioinformatics 27: 578–579. <http://dx.doi.org/10.1093/bioinformatics/btq683>.
 10. Boetzer M, Pirovano W. 2012. Toward almost closed genomes with GapFiller. Genome Biol. 13(6):R56. <http://dx.doi.org/10.1186/gb-2012-13-6-r56>.
 11. Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. Bioinformatics 25(14):1754–1760. <http://dx.doi.org/10.1093/bioinformatics/btp324>.
 12. Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. Nucleic Acids Res. 39(Web Server issue):W347–W352. <http://dx.doi.org/10.1093/nar/gkr485>.
 13. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a web tool to identify clustered regularly interspaced short palindromic repeats. Nucleic Acids Res. 35(Web Server issue):W52–W57. <http://dx.doi.org/10.1093/nar/gkm360>.
 14. Siguier P, Perochon J, Lestrade L, Mahillon J, Chandler M. 2006. ISfinder: the reference centre for bacterial insertion sequences. Nucleic Acids Res. 34(Database issue):D32–D36. <http://dx.doi.org/10.1093/nar/gkj014>.
 15. Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc. Natl. Acad. Sci. U. S. A. 106: 19126–19131. <http://dx.doi.org/10.1073/pnas.0906412106>.