

SCIENTIFIC REPORTS



OPEN

Highly-accurate metabolomic detection of early-stage ovarian cancer

David A. Gaul^{1,2}, Roman Mezencev², Tran O. Long³, Christina M. Jones¹, Benedict B. Benigno⁴, Alexander Gray³, Facundo M. Fernández^{1,5,*} & John F. McDonald^{2,4,5,*}

Received: 08 July 2015

Accepted: 13 October 2015

Published: 17 November 2015

High performance mass spectrometry was employed to interrogate the serum metabolome of early-stage ovarian cancer (OC) patients and age-matched control women. The resulting spectral features were used to establish a linear support vector machine (SVM) model of sixteen diagnostic metabolites that are able to identify early-stage OC with 100% accuracy in our patient cohort. The results provide evidence for the importance of lipid and fatty acid metabolism in OC and serve as the foundation of a clinically significant diagnostic test.

Ovarian cancer (OC) is the most lethal of all gynecological malignancies and the fifth leading cause of death among women living in the United States¹. The disease is essentially asymptomatic until late stages when the 5-year relative survival rate is <44%². If detected and treated early in its progression, the 5-year survival rate is ~90%. For this reason, considerable effort has been focused on the development of a screening test to diagnose OC early in its progression³. This challenge is confounded by the fact that because the disease is in low prevalence in the general population (~0.1% in USA), a screening test must attain a positive predictive value (PPV) of >10%, with a specificity \geq 99.6% and a sensitivity \geq 75% to be of clinical relevance in the general population⁴.

The current standard screening method for OC involves trans-vaginal ultrasound and measurement of serum CA-125 levels³. Combined, these tests result in a positive predictive value of only 24%⁵. A recent study reports that monitoring changes in serum levels of CA-125 over time rather than reliance on a single predefined threshold level of significance can increase accuracy of detection up to 86%⁶. In addition, a variety of proteomic⁷ and microarray⁸ based tests are currently under development but, thus far, no assay has attained the stringent level of accuracy required to be of clinical relevance in the general population.

We report here on the combined use of ultra-performance liquid chromatography, high-resolution mass spectrometry (UPLC-MS) and tandem MS (MS/MS), combined with a customized support vector machine (SVM)-based learning algorithm for identification of 16 diagnostic metabolites that collectively are able to distinguish early-stage OC with 100% accuracy in our cohort. The results provide the foundation for clinically-significant diagnostic tests and evidence for the importance of alterations in lipid and fatty acid metabolism in the onset and progression of the disease.

Results

Negative ion mode UPLC-MS interrogation of the serum metabolome from 46 early stage (I/II) serous epithelial ovarian cancer (EOC) patients and 49 age-matched normal healthy controls (Supplementary Table S1) resulted in the detection of >4000 spectral features (R_x , m/z pairs). After filtering and curation to remove inconsistent and/or ambiguous features, a remaining pool of 255 (Supplementary Table S2)

¹School of Chemistry and Biochemistry, Georgia Institute of Technology, Atlanta GA 30332 (USA). ²School of Biology, Integrated Cancer Research Center, Georgia Institute of Technology, Atlanta GA 30332 (USA). ³College of Computing, Georgia Institute of Technology, Atlanta GA 30332 (USA). ⁴Ovarian Cancer Institute, Atlanta GA 30342 (USA). ⁵Parker H. Petit Institute of Bioengineering and Biosciences, Georgia Institute of Technology, Atlanta GA 30332 (USA). *These authors contributed equally to this work. Correspondence and requests for materials should be addressed to J.F.M. (email: john.mcdonald@biology.gatech.edu)

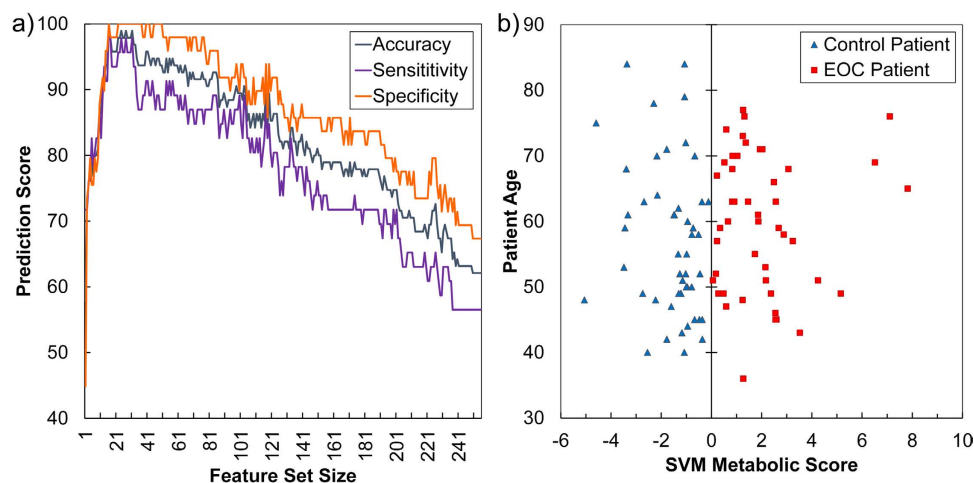


Figure 1. Recursive feature elimination (RFE) selects 16 metabolic features that distinguish early stage serous epithelial ovarian cancer (EOC) serum samples with high accuracy. (a) Evolution of accuracy using support vector machine (SVM)-RFE feature selection for metabolic classifiers. The initial 255 metabolic features identified by ultra-performance liquid chromatography-mass spectrometry (UPLC-MS) provided only moderate predictive accuracy (accuracy 62%; specificity 57%; sensitivity 67%) in distinguishing EOC from control samples. SVM-RFE selected a minimum of 16 metabolic features that provided 100% accuracy (100% sensitivity; 100% specificity) in distinguishing between EOC and control samples. (b) Visualization of the optimal separation between EOC and control samples by the SVM model. The X-axis is the optimal weight vector of the SVM model; the Y-axis is the age of donors (EOC patients or normal control women at the time of sample collection). The vertical line is the projection of the separating hyperplane generated by the SVM model. The discriminant linear SVM model was evaluated by leave-one-out cross-validation (LOOCV).

was used to build a discriminant linear support vector machine (SVM) model that was evaluated by leave-one-out cross-validation (LOOCV)⁹. Binary classifications (cancer/normal) were established through a previously described “metabolic score” decision function that numerically separates the predicted cancer (positive score) from control (negative score) samples¹⁰.

Using all 255 metabolic features, a first SVM model was generated displaying moderate predictive accuracy (accuracy 62%; specificity 57%; sensitivity 67%). Since SVM models built upon large datasets typically contain uninformative features, a number of feature selection methods have been developed to identify subsets with optimal predictive accuracy¹¹. We employed a previously described recursive feature elimination (RFE) method⁹ to select features that distinguished the early-staged EOC samples from controls with optimal accuracy. As shown in the Fig. 1 (see also Supplementary Figs S1–2), 100% accuracy (100% sensitivity, and 100% specificity) was obtained with a minimum of 16 features. The high predictive accuracy of these 16 metabolites was independently validated by orthogonal partial least squares-discriminant analysis (oPLS-DA) using a variety of cross-validation approaches (Supplementary Table S3, Supplementary Figs S3–4).

The high resolution MS technology employed allowed generation of accurate masses and isotopic patterns for each discriminant feature and therefore establishment of candidate elemental formulas. These proposed metabolite identities were confirmed by UPLC-MS/MS, and the resultant tandem MS spectra were compared to those in databases or literature, resulting in chemical identification of 11 of the 16 discriminating features (Table 1, Supplementary Fig. S5, Supplementary Table S4). Two feature identities were further supported by comparison to a standard (R_f and ion fragmentation pattern). Relative concentration levels of about half of the 16 features were elevated and half reduced in cancer samples relative to controls (Fig. 2).

Many of the identified features were lipids or fatty acids. An emerging body of evidence has implicated changes in lipid and fatty acid metabolism with the onset and progression of ovarian¹² and other types of cancer¹³. In many cases, these changes have been linked to the aberrant expression of genes involved in lipid/fatty acid synthesis. For example, the well-known tumor suppressor gene p53 is mutated in >95% of high-grade serous ovarian cancers¹⁴. It has recently been reported that the protein encoded by p53 (TP53) interacts with sterol regulatory element-binding proteins (SREBPs) and guanidinoacetate N-methyltransferase (GAMT) resulting in the elevated expression of enzymes involved in fatty acid and cholesterol biosynthesis and the inhibition of fatty acid oxidation leading to lipid anabolism and accelerated tumor growth and progression¹⁵.

Two of the identified metabolites are lysophospholipids (LPLs) [lysophosphatidylethanolamine (LPE) and lysophosphatidylinositol (LPI)]. Serum levels of LPLs have been previously reported to be elevated in

| Feature | Average m/z | Average Retention Time (min) | Ion Type | Ion Theoretical m/z | Mass Error (ppm) | Neutral Elemental Formula | Tentative Metabolite Identification [Database: #] | SVM Model Weight |
|---------|-------------|------------------------------|------------------------|---------------------|------------------|---|---|------------------|
| 279 | 552.2327 | 0.70 | [M-H] ²⁻ | 552.2335 | -1.4 | C ₄₂ H ₈₀ N ₃ O ₃₀ | [NA] | 1.1521 |
| | | | | 552.2342 | -2.7 | C ₄₃ H ₇₆ N ₇ O ₂₆ | | |
| | | | | 552.2398 | -12.9 | C ₄₂ H ₇₆ N ₉ O ₂₅ | | |
| 571 | 329.1733 | 4.81 | [M-CHO-H] ⁻ | 329.1753 | -6.1 | C ₂₁ H ₂₈ O ₅ | cortisone [CAS: 53-06-5] | 1.0810 |
| 286 | 597.3029 | 10.89 | [M-H] ⁻ | 597.3040 | -1.8 | C ₂₇ H ₅₁ O ₁₂ P | lysophatidylinositol(18:1) [CAS: 1246298-13-4] | 0.4762 |
| 683 | 261.0727 | 1.01 | [M-H] ⁻ | 261.0723 | 1.5 | C ₉ H ₁₃ N ₂ O ₇ | aspartyl-glutamic acid [CAS: 6157-06-8] | 0.3967 |
| 226 | 539.4301 | 14.86 | [M-H] ⁻ | 539.4312 | -2.0 | C ₃₂ H ₆₀ O ₆ | 16-(6-butoxy-3-hydroxy-4,5-dimethylcyclohex-1-en-1-yl)-6,10-dihydroxy-2,6,10,14-tetramethyl hexadecanoic acid [USPTO: document #20100086960] ^a | 0.1748 |
| 45 | 536.5042 | 18.56 | [M-H] ⁻ | 536.5043 | -0.2 | C ₃₄ H ₆₇ NO ₃ | ceramide(d18:1/16:0) [CAS: 24696-26-2] | 0.1076 |
| 64 | 365.3413 | 16.97 | [M-H] ⁻ | 365.3425 | 1.2 | C ₂₄ H ₄₆ O ₂ | [NA] | 0.0847 |
| 28 | 524.2778 | 12.79 | [M-H] ⁻ | 524.2777 | 0.2 | C ₂₇ H ₄₄ NO ₇ P | lysophosphatidylethanolamine(22:6) [PUBCHEM: 52925132] | 0.0445 |
| 105 | 195.1016 | 1.01 | [M+Na-2H] ⁻ | 195.0997 | 9.7 | C ₉ H ₁₈ O ₃ | 2-hydroxyl nonanoic acid [CAS: 617-31-2] ^a | 0.0195 |
| 14 | 307.2633 | 14.80 | [M+Na-2H] ⁻ | 307.2613 | 6.5 | C ₁₈ H ₃₆ O ₂ | iso-1,2-octadecanediol [PUBCHEM: 42607317] ^a | -0.0200 |
| 79 | 245.1378 | 0.99 | [M-H] ⁻ | 245.1389 | -4.5 | C ₁₂ H ₂₂ O ₅ | 3-hydroxyl dodecanedioic acid [CAS: 34574-69-1] ^a | -0.0205 |
| 80 | 883.5358 | 15.40 | [M-H] ⁻ | 883.5337 | 2.4 | C ₄₇ H ₈₁ O ₁₃ P | phosphatidylinositol(20:4/18:1) [HMDB: 09901] | -0.0308 |
| 123 | 467.3727 | 14.61 | [M-H] ⁻ | 467.3737 | -2.1 | C ₂₈ H ₃₂ O ₅ | 7,9,13-trihydroxyoctacos-16,22-dienoic acid [USPTO: document #20120136057] ^a | -0.0803 |
| 231 | 429.2997 | 11.49 | [M-H] ⁻ | 429.3010 | 1.3 | C ₂₇ H ₄₂ O ₄ | [NA] | -0.3420 |
| 261 | 451.2275 | 1.58 | [M-H] ²⁻ | 451.2242 | 7.3 | C ₄₀ H ₆₈ N ₆ O ₁₇ | [NA] | -0.4834 |
| | | | | 451.2249 | 5.8 | C ₄₁ H ₆₄ N ₁₀ O ₁₃ | | |
| | | | | 451.2305 | -6.6 | C ₄₀ H ₆₄ N ₁₂ O ₁₂ | | |
| 620 | 129.0909 | 1.14 | [M-H] ⁻ | 129.0916 | -2.1 | C ₇ H ₁₄ O ₂ | [NA] | -0.9938 |

Table 1. Chemical identification of 16 features that distinguish early-staged ovarian cancer sera from the sera of normal healthy controls with high accuracy (m/z = mass-to-charge ratio, min = minutes, ppm = part per million, CAS = chemical abstract service, USPTO = United States Patent and Trademark Office, HMDB = the human metabolome database, SVM = support vector machine, NA = not available).
^aAll possible metabolite isomers are not listed. Where indicated, the species in the Table are those for which MS/MS data was available in the literature.

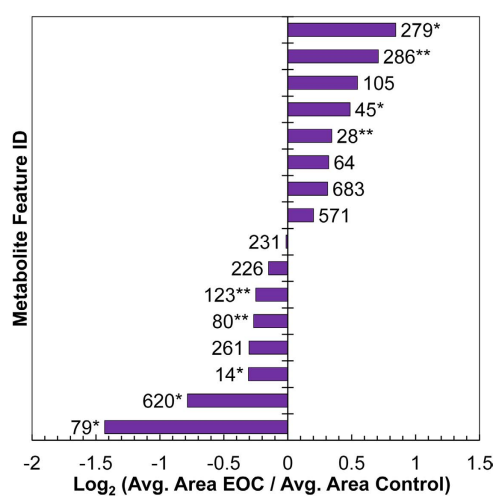


Figure 2. Fold-change of average peak areas of each discriminant feature. Positive values indicate higher levels of metabolite observed on average for EOC patients compared to control patients, while negative values indicate inverse relationship (*p < 0.05; **p < 0.10, Mann Whitney U test).

OC patients and in matched sets of samples isolated from preoperative vs. postoperative patients¹⁶. LPIs are also known to bind and activate the orphan G-protein coupled receptor GPR55, which triggers proliferation and anchorage-independent growth of OC cells, as well as activation of Akt and ERK1/2 kinase¹⁷.

Phosphatidylinositol is one of several inositol membrane phospholipids known to be responsible for recruitment of the serine/threonine kinase Akt to the plasma membrane and its subsequent phosphorylation and activation¹⁸. Phosphorylation of the inositol ring 3'-OH group in inositol phospholipids is carried out by the enzyme phosphatidylinositol 3-kinase (PI3K). A broad range of functions related to cancer onset and progression have been associated with PI3K activity, including proliferation, cell adhesion, apoptosis, and transformation¹⁹. Our identification of the sphingolipid ceramide as a differentiating metabolite is consistent with its previously proposed roles in ovarian and other cancers²⁰.

Discussion

As demonstrated by the presented work, SVM machine learning is a powerful computational tool for the identification of correlated patterns in large datasets. We have previously shown that combining this computational approach with high-resolution mass spectrometry of patient sera is a minimally invasive and highly accurate method for the detection of prostate¹⁰ and late-stage ovarian cancers²¹.

Because of the extensive genetic diversity known to exist among individual patient tumors of even the same type of OC²², it is not surprising that it has proven extremely difficult to identify a single set of biomarkers capable of diagnosing the disease with high accuracy²³. Although there may be multiple genetic lesions and alternative molecular pathways leading to the development of even the same type of OC, all of these mutations and pathways converge on a similar cancer phenotype. Thus, molecular features closely associated with the cancer phenotype, like metabolites, may be expected to be less variable across patients than the broader spectrum of individual mutations and disrupted pathways underlying the disease²⁴.

The predictive accuracy of SVM-derived biomarkers is heavily dependent upon the representative nature of the biological samples used in building the model. For this reason, we designed our study to include samples collected from a broad spectrum of geographic locations in the United States and Canada. However, more extensive sampling of patients across a wider diversity of racial and ethnic groups will be needed to determine the general robustness of the diagnostic biomarkers presented here. Regardless, our results demonstrate that this evidence-based approach to metabolic biomarker discovery is conceptually unbiased for the establishment of highly accurate biomarker panels of early-staged OC across a broad geographic area. If deemed appropriate by future studies, the method can be equally well applied to racial or ethnic sub-populations to obtain optimally accurate panels of metabolic features in these cohorts. When combined with experimental chemical identification of these diagnostic features, our approach provides valuable insight into the metabolic alterations accompanying the disease and can serve as the foundation for clinically significant diagnostic tests.

Methods

Chemicals. Ultrapure water with 18.2 M Ω cm resistivity (Barnstead Nanopure UV ultrapure water system, USA) was used to prepare all mobile phase components. Chromasolv[®] (Fluka) LC-MS grade methanol was purchased from Sigma-Aldrich Corp. (St. Louis, MO, USA). Lysophatidylinositol (18:1) and ceramide (d18:1/16:0) were purchased from Avanti Polar Lipids, Inc. (Alabaster, AL, USA).

Sample Preparation. All samples were collected after informed consent under approved IRB protocols. Serum samples were thawed on ice, and protein precipitation was performed by the addition of methanol in a 3:1 volume ratio to 50 μ L of serum. Aliquots of 10 μ L from each sample were combined to create a pooled sample, which was split into 50 μ L portions before protein precipitation. Samples were vortex-mixed for 10 s and centrifuged at 13,000 g for 7 min. After centrifugation, 150 μ L of supernatant was mixed with 400 μ L of ultrapure water prior to solvent removal using a VirTis benchtop freeze dryer (Warminster, PA). Samples were stored at -80°C until analysis. Samples were separated into 8 batches with equal representation of epithelial ovarian cancer (EOC) and control samples from each collection site in each group. All samples were thawed, reconstituted with 80:20 (v:v) H₂O: MeOH, and analyzed in duplicate. Samples were run in alternating fashion so that duplicate runs for a specific sample were not consecutive. Pooled quality control serum samples were analyzed every eight sample runs. The mass spectrometer was mass calibrated before analysis; and solvent, sample preparation blanks, and pooled samples were analyzed jointly with the EOC and control samples.

UPLC-MS. UPLC-MS was performed using a Waters ACQUITY Ultra-Performance LC system (Waters Corporation, Manchester, UK), fitted with a Waters ACQUITY UPLC BEH C18 column (2.1 \times 50 mm, 1.7 μ m particle size), coupled to a high-resolution accurate mass Synapt G2 high-definition mass spectrometry system (Waters Corporation, Manchester, UK). The Synapt G2 HDMS is a hybrid quadrupole-ion mobility-orthogonal acceleration time-of-flight instrument with a typical resolving power of 20,000 FWHM and mass accuracy of 9 ppm at m/z 544.2615. The instrument was operated in negative ion mode with a probe capillary of 2.0 kV and a sampling cone voltage of 35 V. The source and desolvation temperatures were 150 and 500 $^{\circ}\text{C}$, respectively, and the nitrogen desolvation flow rate was 1000 L h⁻¹. The mass spectrometer was calibrated across the range of m/z 50–1200 using a 0.5 mM

sodium formate solution prepared in 90:10 (v/v) 2-propanol:water. Data were mass corrected during acquisition using a leucine enkephalin reference spray (LockSpray) infused at $3\mu\text{L min}^{-1}$. Data were acquired in the 50–1200 m/z range, and the scan time was set to 1 s. Data acquisition and processing were carried out using MassLynx V4.1 and MZmine V2.0, respectively. The chromatographic method for sample analysis involved elution with ultrapure water (mobile phase A) and methanol (mobile phase B) using the following gradient program: 0–15 min 80–10% A; 15–23 min 10% A. The flow rate was constant at 0.40 mL min^{-1} for 23 min. The gradient was returned to its initial conditions with a solvent blank run of 11 min. The column temperature was set to $60\text{ }^\circ\text{C}$, the autosampler tray was set to $5\text{ }^\circ\text{C}$, and the injection volume was $8\mu\text{L}$. UPLC-MS/MS experiments were performed by acquiring mass spectra with applied voltages between 5 and 50 V in the trap cell, using ultra high purity grade argon ($\geq 99.999\%$) as the collision gas.

Data processing/analysis. Metabolic features (retention time (R_t), m/z pairs) were extracted from chromatograms using MZmine V2.0 software and Excel. A five point Savitzky Golay smoothing function was applied to each scan of the raw data prior to peak detection. After chromatogram alignment, the subsequent peak list was conservatively filtered by elimination of peaks that were not present in at least 40 of the 237 collected runs prior to gap filling. The exported peak areas for each sample were normalized by division of the total peak area sum for that sample in Excel. Potential features in which the slope of the peak area of the pooled samples vs. time changed more than one standard deviation away from zero were removed from the peak list. The potential features list was further constrained by purging features that were not present in 50% of sample groups (all samples, EOC samples, or control samples) at ten times the baseline, defined as the maximum peak area observed in the sample blank and ten mobile phase runs (one from each day of analysis). The duplicate sample peak areas were averaged to create a matrix containing sample peak areas for each feature (average R_t , average m/z).

The data set was scrutinized for the presence of experimental and instrument bias with principal component analysis (PCA) using MATLAB R2012b (Version 8.0.0.783 The MathWorks, Inc., Natick, MA, USA) and the PLS Toolbox (v.6.71, Eigenvector Research, Inc., Wenatchee, WA, USA). Peak area data were labeled with the corresponding collection day, analysis batch, or sample origin. Data were preprocessed by autoscaling, and PCA run with leave-one-out cross-validation. Sample clustering was assessed with the plot of the first versus second principal component.

Linear support vector machine (SVM) analysis of the feature matrix was performed with in-house-developed code utilizing liblinearSVM²⁵. Recursive feature elimination (RFE) was used to find the minimum set of discriminant features that maximized accuracy in the classification⁹. For a binary classification problem, linearly-separable samples represented as a row vector \mathbf{x} , had membership of two classes g ($=N$ or C), where N stands for normal or control patient samples and C represents EOC disease patient samples with class value c ($=-1$ for class N , and $+1$ for class C). The decision function that separated the two classes, defined here as the “EOC metabolic score”, was as follows:

$$\text{EOC metabolic score} = b + \sum_{j=1}^J w_j x_{ij} \quad [1]$$

where w and b are the weight and bias parameters that were determined from the training set and J is the total number of features. The sign of the EOC metabolic score determined which class a sample was assigned to: class N if negative and class C if positive. In this classification function, the two classes were divided in the dataspace by a hyperplane $\mathbf{w}\mathbf{x}' + b = 0$ that maximized the margins between samples of different classes. The margin between the two classes was defined such that:

$$\mathbf{w}\mathbf{x}' + b \geq 1, \quad c = +1 \quad [2]$$

$$\mathbf{w}\mathbf{x}' + b \leq -1, \quad c = -1 \quad [3]$$

The RFE method involved an “outer” matrix with 95 columns (equal to the number of serum samples) and 255 rows (equal to the number of features). Each row in the outer matrix represents a subset of features to be tested for discriminatory power. Each subsequent row examines a feature subset that contains one less feature than the previous row. For each row, a set of 95 “inner” matrices is constructed, each one containing a subset of samples to build an SVM model and one sample left out for testing, following standard LOOCV practice. After model building, the SVM feature weights were calculated for each inner matrix, and these weights were summed across the inner matrix. The average feature weight was then calculated across the outer matrix row. The least important feature was then discarded. This process was subsequently repeated for every outer matrix row. A panel of optimal features was determined by examining which feature set had maximized accuracy, sensitivity, and specificity. Data were preprocessed by autoscaling the features across the samples prior to SVM-RFE.

Orthogonal partial least squares discriminant analysis (oPLS-DA⁹) was performed to inspect data after discriminant feature selection via SVM-RFE. oPLS-DA models were internally cross validated using leave-one-out, venetian blinds (10 data splits and 10 samples per blind), contiguous block (10 splits),

or random subsets (10 data splits and 10 iterations) approaches. Permutation testing was performed by randomizing the class labels for all samples. Data were preprocessed by autoscaling the features' peak areas across the samples.

Metabolite Identification. Compound identification was carried out for the 16 discriminant features obtained after the feature selection processes. Elemental formulas were generated based on the mass accuracy of the peak of interest and isotopic patterns with a mass error of 10 mDa using MassLynx 4.1. The chemical formulas were searched against the following publically-available databases: Metlin, the human metabolome database (HMDB), Metabolomics Workshop, LIPID Metabolites and Pathways Strategy (LIPID MAPS), and MassBank to determine possible endogenous metabolite candidates. Entries in the MS/MS Metlin database, MassBank, and Lipid Maps, together with literature searches subsequently confirmed the identity of putative candidates. When available, metabolite standards were analyzed to support identification. Identification of metabolites was pursued according to established criteria²⁶.

References

- Jelovac, D. & Armstrong, D. K. Recent progress in the diagnosis and treatment of ovarian cancers. *CA. Cancer J. Clin.* **61**, 183–203 (2011).
- Cancer Fact & Figures 2013. American Cancer Society, Atlanta, GA, USA (2013).
- Muñoz, K. A. Harlan, L. C. & Trimble, E. I. Patterns of care for women with ovarian cancer in the United States. *J. Clin. Oncol.* **15**, 3408–3415 (1997).
- Clark-Pearson, D. L. Screening for ovarian cancer. *New Engl. J. Med.* **361**, 170–177 (2009).
- Bosse, K. *et al.* Screening for ovarian cancer by transvaginal ultrasound and serum CA125 measurement in women with a familial predisposition: a prospective cohort study. *Gynecol. Oncol.* **103**, 1077–1082 (2006).
- Menon, U. *et al.* Risk algorithm using serial biomarker measurements doubles the number of screen-detected cancers compared with a single-threshold rule in the United Kingdom collaborative trial of ovarian cancer screening. *J. Clin. Oncol.* **33**, 2062–71 (2015).
- Elzek, M. A. & Rodland, K. D. Proteomics of ovarian cancer: functional insights and clinical applications. *Cancer Metastasis Rev.* **34**, 83–96 (2015).
- Zhen, H., Liu, J. Y., Song, F. J. & Chen K. X. Advances in circulating microRNAs as diagnostic and prognostic markers for ovarian cancer. *Cancer Biol. Med.* **10**, 123–130 (2013).
- Guan, W. *et al.* Ovarian cancer detection from metabolomic liquid chromatography/mass spectrometry data by support vector machines. *BMC Bioinformatics.* **10**, 259–274 (2009).
- Zang, X. *et al.* Feasibility of detecting prostate cancer by ultraperformance liquid chromatography–mass spectrometry serum metabolomics. *J. Proteome Res.* **13**, 3444–3454 (2014).
- Rakotomamonjy, A. Variable selection using SVM-based criteria. *J. Machine Learn. Res.* **3**, 1357–1370 (2003).
- Pyragius, C. E., Fuller, M., Ricciardelli, C. & Oehler, M. K. Aberrant lipid metabolism: an emerging diagnostic and therapeutic target in ovarian cancer. *Int. J. Mol. Sci.* **14**, 7742–7756 (2013).
- Omabe, M. Ezeani, M. & Omabe, K. N. Lipid metabolism and cancer progression: the missing target in metastatic cancer treatment. *J. App. Biomed.* **13**, 47–59 (2015).
- Havrilesky, L. *et al.* Prognostic significance of p53 mutations and p53 overexpression in advanced epithelial ovarian cancer: a gynecological oncology group study. *J. Clin. Oncol.* **21**, 3814–3825 (2003).
- Hu, J., Liu, Z. & Wang, X. Does TP53 mutation promote ovarian cancer metastasis to omentum by regulating lipid metabolism? *Med. Hyp.* **81**, 515–520 (2013).
- Sutphen, R. *et al.* Lysophospholipids are potential biomarkers of ovarian cancer. *Cancer Epidemiol. Biomarkers Prev.* **13**, 1185–1191 (2004).
- Pineiro, R., Maffucci, T. & Falasca, M. The putative cannabinoid receptor GPR55 defines a novel autocrine loop in cancer cell proliferation. *Oncogene* **30**, 142–152 (2011).
- Vivanco, I. & Sawyers, C. L. The phosphatidylinositol 3-Kinase–AKT pathway in human cancer. *Nat. Rev. Can.* **2**, 489–501 (2002).
- Fruman, D. A. & Rommel, C. PI3K and cancer: lessons, challenges and opportunities. *Nat. Rev. Drug Disc.* **13**, 140–156 (2014).
- Morad, S. A. F. & Cabot, M. C. Ceramide-orchestrated signaling in cancer cells. *Nat. Rev. Can.* **13**, 51–65 (2013).
- Zhou, M., *et al.* Rapid mass spectrometric metabolic profiling of blood sera detects ovarian cancer with high accuracy. *Can. Epidemiol. Biomark. Prev.* **19**, 2262–2271 (2010).
- The Cancer Genome Atlas Research Network. Integrated genomic analyses of ovarian carcinoma. *Nature* **474**, 609–615 (2011).
- Nguyen, L. *et al.* Biomarkers for early detection of ovarian cancer. *Women's Health* **9**, 171–187 (2013).
- Arakaki, A. K., Skolnick, J. & McDonald, J. F. Marker metabolites can be therapeutic targets as well. *Nature* **456**, 443 (2008).
- Fan, R. E., Chang, K. W., Hsieh, C. J., Wang, X. R. & Lin, C. J. LIBLINEAR: A library for large linear classification. *J. Machine Learn. Res.* **9**, 1871–1874 (2008).
- Sumner, L. W. *et al.* Proposed minimum reporting standards for chemical analysis chemical analysis working group (CAWG) metabolomics standards initiative (MSI). *Metabolomic* **3**, 211–221 (2007).

Acknowledgements

We are grateful to Fox Chase Cancer Center (Philadelphia, PA, USA), Northside Hospital (Atlanta, GA, USA) and Alberta Health Services (Edmonton, Alberta, Canada) for providing serum samples for this study. The research was supported by grants from The Laura Crandall Brown Ovarian Cancer Foundation, The Ovarian Cancer Research Fund, The Ovarian Cancer Institute, Northside Hospital (Atlanta), The Robinson Family Fund, and the Deborah Nash Endowment Fund. We thank Martin R.L. Paine for discussions regarding metabolite identification and Maria Eugenia Monge for assistance in performing measurements.

Author Contributions

D.A.G. led all experimental and analytical aspects of the study. D.A.G., F.M.F. & J.F.M. wrote the paper, R.M. carried out biological analysis of the results, T.Q.L. & A.G. performed SVM analysis, C.M.J.

participated in MS analysis, B.B.B. collected the Northside samples and consulted on medical aspects of the study. J.E.M. & F.M.F. designed and supervised all aspects of the study.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Gaul, D. A. *et al.* Highly-accurate metabolomic detection of early-stage ovarian cancer. *Sci. Rep.* **5**, 16351; doi: 10.1038/srep16351 (2015).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>