# REVIEW Open Access



# Interleukin-22: a potential therapeutic target in atherosclerosis

Jin-Wen Luo<sup>1</sup>, Yuan Hu<sup>2</sup>, Jian Liu<sup>1</sup>, Huan Yang<sup>3\*</sup> and Peng Huang<sup>1\*</sup>

### **Abstract**

**Background:** Atherosclerosis is recognized as a chronic immuno-inflammatory disease that is characterized by the accumulation of immune cells and lipids in the vascular wall. In this review, we focus on the latest advance regarding the regulation and signaling pathways of IL-22 and highlight its impacts on atherosclerosis.

**Main body:** IL-22, an important member of the IL-10 family of cytokines, is released by cells of the adaptive and innate immune system and plays a key role in the development of inflammatory diseases. The binding of IL-22 to its receptor complex can trigger a diverse array of downstream signaling pathways, in particular the JAK/STAT, to induce the expression of chemokines and proinflammatory cytokines. Recently, numerous studies suggest that IL-22 is involved in the pathogenesis of atherosclerosis by regulation of VSMC proliferation and migration, angiogenesis, inflammatory response, hypertension, and cholesterol metabolism.

**Conclusion:** IL-22 promotes the development of atherosclerosis by multiple mechanisms, which may be a promising therapeutic target in the pathogenesis of atherosclerosis.

Keywords: Atherosclerosis, IL-22, Inflammatory response, Cytokine, Cholesterol metabolism

#### Introduction

Cardiovascular diseases (CVDs) represent the major factor of disability and premature death worldwide. Recently, the World Health Organization (WHO) revealed that CVD accounts for > 17 million deaths per year and this figure will rise to over 23 million by the year 2030 (GBD 2015 Risk Factors Collaborators 2016; Li et al. 2021a). It is well known that atherosclerosis is an inflammatory disease of the vascular wall, which is recognized as the underlying basis of many CVDs, such as hypertension, myocardial infarction, stroke, and peripheral arterial disease. The development and pathogenesis of atherogenesis are closely associated with many pathophysiological factors, such as inflammatory response,

angiogenesis, hypertension, vascular smooth muscle cell (VSMC), and cholesterol metabolism.

Interleukin-22 (IL-22), discovered in 2000, is known as a crucial member of the IL-10 cytokine family (Dumoutier et al. 2000). Since the discovery of IL-22, its biological functions have been extensively studied. IL-22 can be produced by lymphoid lineage cells (Th1, Th17, Th22, γδT, etc.) and innate lymphocyte cells. Also, IL-22 was reported to have an important function in inflammation, cell proliferation, tissue repair, and host defense (Rutz et al. 2013; Mühl et al. 2013; Eidenschenk et al. 2014; Sonnenberg et al. 2011). Although Th1 and Th17 cells play a role in the development of atherosclerosis, Th22 cellderived IL-22 is more important in atherosclerosis (Shi et al. 2020a). Several lines of evidence have demonstrated that IL-22 aggravates myocardial infarction, hypertension, cardiac hypertrophy, and myocarditis (Zhang et al. 2013; Ye et al. 2017; Kong et al. 2013; Kong et al. 2014). Recently, numerous studies suggested that IL-22 promotes atherosclerosis through multiple mechanisms

<sup>&</sup>lt;sup>3</sup> Department of Respiratory Medicine, Hunan Provincial People's Hospital, Changsha, Hunan 410001, People's Republic of China Full list of author information is available at the end of the article



<sup>\*</sup>Correspondence: 18265723@qq.com; drhuangpeng@qq.com

<sup>&</sup>lt;sup>1</sup> Department of Cardio-Thoracic Surgery, Hunan Children's Hospital, Changsha 410007, People's Republic of China

Luo et al. Mol Med (2021) 27:88 Page 2 of 13

including promotion of inflammation, regulation of cholesterol metabolism, and induction of VSMC proliferation and migration (Rattik et al. 2015; Chellan et al. 2014), indicating this cytokine as a promising and novel target to treat or prevent atherosclerotic cardiovascular disease. In the present review, we summarize the current knowledge about the regulation and signaling pathways of IL-22 and highlight its roles in atherogenesis.

#### Structural features of IL-22

The IL-22 gene is located on human chromosome 12 at q15 and encodes a protein with a length of 179 amino acids. After enzymatic modification of its signal peptide (33 amino acids), the cytokine is secreted as a 146-amino-acid protein (Nagem et al. 2002; Bleicher et al. 2008). Human IL-22 shares 79% amino acid sequence identity with the mouse type. The precursor gene of IL-22 is approximately 5257 base-pair long and consists of 5 exons encoding IL-22, and then converted into a secreted form (Xu et al. 2005; Manna et al. 2017). The crystallographic structure of recombinant human IL-22 (expressed in Drosophila melanogaster and Escherichia coli) was solved using X-ray diffraction and crystallization (Nagem et al. 2002; Xu et al. 2005). The IL-22 monomer has a compact bundle of a single domain that is composed of six interconnected  $\alpha$ -helices (A to F) and connecting loops (Manna et al. 2017; Wei et al. 2019; Nagem et al. 2006). Its basic structure contains two intramolecular disulfide bonds and four cysteines.

IL-22 is biologically active as a monomer. However, it has also been reported that there are functional tetramer and dimer forms of IL-22 under high concentrations in solution (Nagem et al. 2002; Bleicher et al. 2008; Oliveira et al. 2008). The molecular weight of IL-22 is approximately 16.7 kDa, with three potential N-linked glycosylation sites including loop AB (Asn68-Asn69-Thr70), helix A (Asn68-Arg55-Thr56), and helix C (Asn97-Phe98-Thr99). It is believed that glycosylation is only related to minor changes in the tertiary structure of IL-22. However, various genetic polymorphisms including insertion-deletion (indel), single nucleotide polymorphism, somatic single nucleotide variants, and nucleotide deletion, have been identified as the functional modulators of IL-22 gene (Shohan et al. 2020). When producing therapeutically usable neutralizing IL-22-specific monoclonal antibodies (MAbs), the glycosylation form of the gene should be considered.

# IL-22 receptors and signaling

The IL-22 receptor (IL-22R) is part of the family of class II cytokine receptors and is composed of two heterodimeric subunits, IL-10R2 (also named IL10RB) and IL-22R1 (Xie et al. 2000; Sabat et al. 2014; Li et al. 2004).

The human genes for IL-10R2 and IL-22R1 proteins are located on chromosomes 21q22.11 and 1p36.11, respectively. Both receptors have three similar domains: transmembrane, extracellular, and an intracellular signaling region (Bleicher et al. 2008; Xie et al. 2000). Extracellular motifs of the IL-22R complex consists of N-linked glycosylation sites, two fibronectin type III domains, and conserved amino acids (Xie et al. 2000; Langer et al. 2004). Moreover, it has been reported that the longer intracellular moiety of IL-10R2 (79 amino acid sequence) and IL-22R1 (325 amino acid sequence) consist of four Tyr-x-x-Gln motifs, which are putative signal transducer and activator of transcription (STAT) recruitment sites (Kotenko et al. 1997; Kotenko et al. 2001). Of note, IL-22 protein has a strong binding affinity for IL-22R1 ( $K_D$  ranging from  $1 \sim 20$  nM) but no affinity for IL-10R2 (Logsdon et al. 2002; Logsdon et al. 2004; Jones et al. 2008a). However, studies have observed that the IL-10R2 subunit has a high interaction affinity for the IL-22/IL-22R1 complex (Jones et al. 2008b). The binding of IL-22 to IL-22R1 would cause the conformational change of IL-22 protein, which promotes the secondary binding of the IL-22/IL22R1 complex to IL-10R (Bleicher et al. 2008; Li et al. 2004). Of note, a study by Weathington et al. found that the serine residue at positions 414 and 410 of the IL-22R1 gene sequence are phosphorylated under the enzymatic reaction of the kinase glycogen synthase kinase 3β (GSK3β), which promotes the stability of IL-22R and inhibits its degradation via the ubiquitin proteasome (Weathington et al. 2014).

Besides the cell surface-bound IL-22R complex, a soluble IL-22-binding receptor, known as IL-22 binding protein (IL-22BP) or L-22RA2, has been identified to bind IL-22 with high affinity. IL-22BP gene is located on human chromosome 6q23.3. This gene encodes a protein with 210 amino acid residues that share 34% sequence homology to the IL-22R1 subunit (Gruenberg et al. 2001; Dumoutier et al. 2001; Wei et al. 2003; Xu et al. 2001). It has been reported that IL-22BP interacts with IL-22 at the binding sites overlapping with IL-22R1, which directly blocks the binding of IL-22 to its receptor complex (Wu et al. 2008). In addition, the binding affinity of IL-22 to IL-22BP is up to 1000-fold higher than that between IL-22R1 and IL-22 (Wolk et al. 2007; Dudakov et al. 2015). Therefore, IL-22BP is extensively used as an effective competitive inhibitor of IL-22 signaling in vitro (Mühl and Bachmann 2019; Trevejo-Nunez et al. 2019; Fukaya et al. 2018).

After IL-22 forms a complex with IL-22R1 and IL-10R2, STAT3 phosphorylation is induced by activated Janus kinase (JAK) signaling molecules tyrosine kinase 2 (IL-10R2 activator) and JAK1 (IL-22R1 activator) (Sabat et al. 2014; Ziesché et al. 2007). In primary cells exposed

Luo et al. Mol Med (2021) 27:88 Page 3 of 13

to IL-22, STAT3 at residues Ser727 and Tyr705 is phosphorylated (Sestito et al. 2011; Xue et al. 2017). Besides STAT3, phosphorylation of STAT5 and STAT1 has also been reported (Lejeune et al. 2002; Wolk et al. 2004). Interestingly, STAT3 acetylation at lysine 686 is essential for the phosphorylation of Tyr705, and the deacetylase sirtuin 1 suppresses IL-22-induced activation of STAT3 and cellular effects (Sestito et al. 2011; Yuan et al. 2005; Nie et al. 2009). These modifications of STATs promote their molecular dimerization, allowing them to translocate to the nucleus and regulate the expression of their target genes. It has been reported that STAT molecules including STAT3, STAT1, and STAT5 play key roles in lipid metabolism and inflammatory response (Piaszyk-Borychowska et al. 2019; Chen et al. 2019a; Shi et al. 2020b). Recently, Chen et al. summarized the crucial roles of STAT3 in macrophage polarization, endothelial cell dysfunction, immunity, and inflammation during atherosclerosis (Chen et al. 2019a). Thus, targeted inhibition of STAT signaling molecules may be an important potential therapeutic strategy for atherosclerosis. In addition, IL-22 also utilizes Tyk2 and Jak1 to transmit other downstream phosphorylation signals, such as protein kinase B (PKB, also called Akt), mitogen-activated protein kinase (MAPK), P38, c-Jun N-terminal kinase (JNK), and extracellular signal-regulated kinase-1/2 (ERK1/2) (Lejeune et al. 2002; Wolk et al. 2004; Wolk et al. 2010; Andoh et al. 2005; Pickert et al. 2009). Of note, these signaling molecules are also closely associated with the progression of atherosclerosis (Linton et al. 2019; Reustle and Torzewski 2018; Chen et al. 2015).

#### Role of IL-22 in atherosclerosis

Atherosclerosis is known as a chronic and progressive pathologic process underlying cardiovascular disease. The precise role of IL-22 in atherosclerosis is still controversial, although most studies suggest a proatherogenic function of IL-22. apoE/IL-22 double knockout mice exhibit decreased plaque size both in the aortic root and the aorta compared with apoE knockout controls (Rattik et al. 2015). Moreover, in another study of atherosclerosis, IL-22R1 and IL-22 are expressed in mouse atherosclerotic plaques, and their expression levels are significantly elevated in apoE knockout mice (Shi et al. 2020a). Furthermore, the authors also showed that treatment with recombinant mouse IL-22 (rIL-22) markedly aggravates atherosclerosis development in apoE<sup>-/-</sup> mice fed a highfat diet (HFD), whereas blocking circulating IL-22 with a neutralizer of IL-22 (IL-22 mAb) substantially reduces the atherosclerotic lesion area (Shi et al. 2020a). S100/ calgranulin, a pro-inflammatory protein, promotes the development of atherosclerosis with excessive cholesterol accumulation in atherosclerotic lesions in apoE-null mice (Chellan et al. 2014). Further study revealed that myeloidderived S100/calgranulin upregulates IL-22 in a receptor for advanced glycation endproducts (RAGE)-dependent manner, resulting in impaired cholesterol homeostasis in macrophages (Chellan et al. 2014). Besides, circulating IL-22 levels are markedly higher in peripheral blood of patients with unstable angina and acute myocardial infarction (AMI) compared to stable angina and healthy control subjects (Zhang et al. 2013; Xia et al. 2012). These studies suggest that IL-22 contributes to plaque growth and promotes plaque instability during lesion formation. However, in the LDLR<sup>-/-</sup> mouse model, IL-22 plays a protective role in atherosclerosis (Fatkhullina et al. 2018). Inactivation of IL-22 exacerbates atherosclerosis in LDLR<sup>-/-</sup> mice exposed to the western diet (WD) by increasing pro-atherogenic metabolites such as trimethylamine N-oxide (TMAO) and lipopolysaccharide (LPS) (Fatkhullina et al. 2018). The contradictory results may be due to the different experimental conditions, including diets and mouse models.

# IL-22 promotes atherosclerosis through multiple mechanisms

The pathophysiology of atherosclerosis is a multifactorial and complex process involving inflammation, angiogenesis, dysregulation of cholesterol metabolism, VSMC proliferation and migration, and hypertension (Chistiakov et al. 2015; Jaipersad et al. 2014; Kostov and Halacheva 2018). As a unique and pleiotropic cytokine, IL-22 promotes the development of atherosclerosis through multiple mechanisms (Fig. 1).

#### IL-22 in inflammatory response

Atherosclerosis is considered to be a progressive chronic inflammatory disease of the vascular wall. It is known that T cells can trigger the inflammatory response in the vascular wall by secreting numerous pro-inflammatory mediators (Libby and Hansson 2018). In particular, cytokines are important contributors to complex inflammatory reactions in the vascular wall and atherosclerotic plaques.

# IL-22 regulates adhesion molecules

Adhesion molecules play a key role in atherogenesis by promoting the adhesion of monocytes and leukocytes to activated endothelial cells and inducing subsequent vascular inflammation (Galkina and Ley 2007). Intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1) are considered important risk factors involved in the development of atherosclerosis. Upon IL-22 interacts with its receptor, it can induce cells to produce a variety of adhesion molecules and chemokines to participate in the inflammatory

Luo et al. Mol Med (2021) 27:88 Page 4 of 13

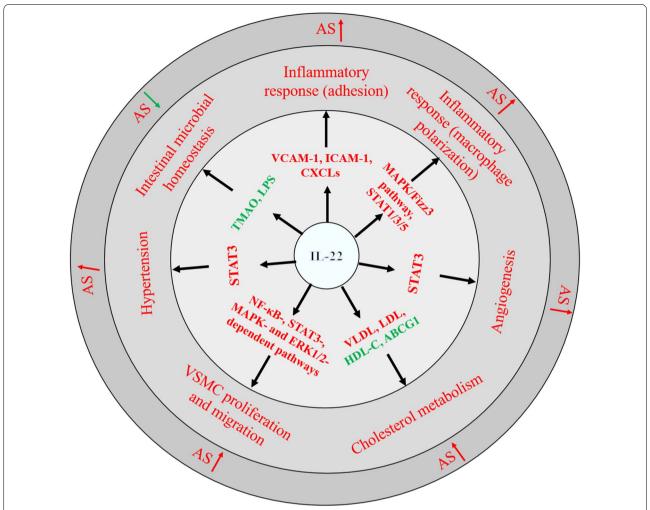


Fig. 1 IL-22-mediated regulation and the major cardiometabolic risk factors of atherosclerosis. VCAM-1 vascular cell adhesion molecule-1, ICAM-1 Intercellular adhesion molecule-1, CXCLs CXC chemokine ligands, MAPK mitogen-activated protein kinase, STAT signal transducer and activator of transcription, VLDL very low-density lipoprotein, LDL low-density lipoprotein, HDL-C high-density lipoprotein cholesterol, ABCG1 ATP-binding cassette transporter G1, NF-κB nuclear transcription factor-κB, ERK extracellular signal-regulated kinase, VSMC vascular smooth muscle cell, TMAO trimethylamine N-oxide, LPS lipopolysaccharide, AS atherosclerosis. Arrows in red: promote; Arrows in green: inhibit; text in red: proatherogenic changes; text in green: antiatherogenic changes

response. It has been reported that IL-22 induces cells to produce ICAM-1 and VCAM-1 (Hoeven et al. 2018). In addition to ICAM-1 and VCAM-1, CXC chemokine ligands (CXCL) have also been identified as key adhesion molecules in adhesion, migration, and recruitment of monocytes during the pathogenesis of atherosclerosis. Some clinical data have demonstrated that IL-22 levels are strikingly elevated in patients with atherosclerosis cardiovascular disease, which positively correlated with the proinflammatory cytokine levels (CXCL-11, CXCL-10, and CXCL-9) in carotid plaques (Xia et al. 2012). Moreover, as a pro-inflammatory cytokine, IL-22 was also reported to synergy with TNF-α in promoting the induction of CXCL1, CCL2, CCL5, CXCL5, CCL20, and

CCL26 expression (Aujla and Kolls 2009). In addition, STAT1 and STAT5 were reported to act as a proinflammatory signal in the regulation of expression of adhesion molecules and chemokines (Coccia et al. 1999; Lian et al. 2016; Soriano et al. 2003; Yang et al. 2007). Given that IL-22 is a critical regulator of STAT signaling molecules. STAT1 and STAT5 may be important mechanisms involved in the expression of adhesion molecules regulated by IL-22.

#### IL-22 regulates macrophage phenotypes

It is well known that tissue-resident and monocytederived recruited macrophages are pivotal regulators of tissue repair, regeneration, and fibrosis. Macrophages Luo et al. Mol Med (2021) 27:88 Page 5 of 13

can be functionally and phenotypically modified to adopt pro-wound-healing, pro-resolving, pro-inflammatory, anti-inflammatory, anti-fibrotic, tissue-regenerating, and pro-fibrotic properties in response to microenvironmental stimuli (Wynn and Vannella 2016). Of note, pro-inflammatory macrophages are primarily associated with inflammatory activity and have proatherogenic properties, whereas anti-inflammatory macrophages are mainly responsible for inhibiting inflammatory responses and increasing atherosclerotic plaque stability (Moore and Tabas 2011; Tabas and Bornfeldt 2016; Li et al. 2018). Several lines of evidence suggest that IL-22 is closely associated with pro-inflammatory macrophage phenotype (Hou et al. 2018; Yang et al. 2019). Deletion of IL-22 was reported to markedly induce cardiac macrophage toward an anti-inflammatory phenotype (Ye et al. 2020). The underlying mechanism involves the mitogen-activated protein kinase (MAPK)/Fizz3 pathway. Moreover, IL-22 levels increase significantly during the differentiation of bone marrow-derived macrophages into pro-inflammatory phenotype under the stimulation of lipopolysaccharide (LPS) (Yang et al. 2019). The tolllike receptor 4 (TLR4) signaling pathway plays an important role in the regulation of macrophage inflammation. It has been reported that TLR4 deficiency induces the differentiation of alveolar macrophages toward an antiinflammatory phenotype, accompanied by a decrease in IL-22 expression (Shen and Liu 2015). Besides, STAT molecules such as STAT1 and STAT5 were found to drive macrophage differentiation toward a pro-inflammatory phenotype (Gan, et al. 2017; Lawrence and Natoli 2011). STAT1 and STAT5 are also known to trigger inflammatory cascade (e.g., NF-κB, TLR4, MAPK) that promotes macrophage inflammatory response and atherogenesis (Chmielewski et al. 2016; Wu et al. 2020; Shen-Orr et al. 2016). In contrast, inhibition of STAT1 and STAT5 attenuates inflammatory response in atherosclerosis (Sikorski et al. 2011a, b; Wang et al. 2021). Of note, IL22R1 is widely expressed in macrophages. The IL-22/IL22R1 complex is a key activator of STAT signaling molecules. Thus, the IL-22/IL22R1 complex may direct macrophages towards a pro-inflammatory phenotype via regulating STAT1 and STAT5.

# IL-22 in angiogenesis

Angiogenesis, defined as the formation of new microvessels (small venules and capillaries) from pre-existing blood vasculature, plays a crucial role in plaque growth and instability (Camaré et al. 2017). IL-22 and its receptor IL-22R1 were significantly upregulated in endothelial cells following ischemic injury (Hu et al. 2018). In vivo studies further showed that IL-22 promotes the formation of new blood vessels in a mouse hindlimb ischemia

model. Also, IL-22 directly acts on human umbilical vein endothelial cells (HUVECs) and induces HUVEC proliferation, migration, and tube formation in vitro (Hu et al. 2018). Blocking IL-22 with IL-22 mAb reversed the above effects. In addition to regulating angiogenesis in peripheral arterial disease, IL-22 was also found to promote neovascularization in a mouse tumor model (Protopsaltis et al. 2019; Shang et al. 2015). Importantly, IL-22 treatment strikingly increased phosphorylation of STAT3 in HUVECs (Hu et al. 2018; Shang et al. 2015). In contrast, treatment with STAT3 inhibitor or knockdown of IL-22R1 attenuated IL-22-induced endothelial cell survival or tube formation (Hu et al. 2018; Shang et al. 2015). These studies indicate that STAT3 is an important signaling molecule required for IL-22/IL22R1 complex-induced angiogenesis. However, the key proteins involved in IL-22-induced angiogenesis still need to be further confirmed.

#### IL-22 in cholesterol metabolism

Lipid metabolic disorder, especially hypercholesterolemia, is an important risk factor for atherosclerosis. High-density lipoprotein cholesterol (HDL-C) exerts an atheroprotective effect, but very low-density lipoprotein (VLDL) and low-density lipoprotein cholesterol (LDL-C) are proatherogenic. It has been reported that circulating IL-22 is negatively associated with HDL-C levels, while it was positively associated with LDL and VLDL levels (Torquati et al. 2019; Lodge et al. 2021). Thus, IL-22 may contribute to the development of hypercholesterolemia.

The accumulation of foam cells (cholesterol-laden macrophages) in the arterial walls plays an important role in the pathogenesis of atherosclerosis. ATP-binding cassette transporter A1 (ABCA1) and ABCG1, two important transmembrane transporters, are widely expressed by macrophages and play a pivotal role in promoting the efflux of cholesterol onto apoA-I and HDL (Tarling and Edwards 2011; Ren et al. 2019). CD36, lectin-like oxidized low-density lipoprotein receptor-1 (LOX-1), and scavenger receptor class A (SRA) are primarily responsible for intracellular cholesterol uptake (Crucet et al. 2013). A recent study showed that recombinant IL-22 decreases ABCG1 expression and impairs macrophage cholesterol efflux (Chellan et al. 2014). However, IL-22 does not affect the expression of LOX-1, CD36, and ABCA1. Of note, the activity of ABCG1 in macrophages is largely modulated by the nuclear transcription factor liver X receptor (LXR) (Larrede et al. 2009). Interestingly, IL-22 does not decrease cholesterol efflux in macrophages pretreated with TO-901317 (an LXR agonist) (Chellan et al. 2014), suggesting that endogenous upregulation of ABCG1 diminishes the harmful effect of IL-22 on macrophage cholesterol efflux. In addition,

Luo et al. Mol Med (2021) 27:88 Page 6 of 13

Agrawal et al. found that STAT1 is essential for optimal foam cell formation and the development of atherosclerotic lesions (Agrawal et al. 2007). Inhibition of STAT1 in macrophages significantly suppresses oxidized low-density lipoprotein (oxLDL)-induced foam cell formation (Agrawal et al. 2007). In addition to STAT1, STAT5 has also been reported to participate in foam cell formation. STAT5 inhibitor markedly reduces cholesterol accumulation in oxLDL-stimulated macrophages (Wang et al. 2021). Therefore, it is possible that IL-22 promotes cholesterol accumulation via the regulation of STAT1 and STAT5.

#### IL-22 in VSMC proliferation and migration

It is well known that VSMC proliferation and migration from the media to the intima are pivotal cellular events in the progression of atherosclerosis and the VSMC phenotypic regulation (transformation from contractile type to synthetic type) plays a crucial role in this process (Basatemur et al. 2019). Rattik et al. found that IL-22R1 is expressed by VSMCs in the mouse vascular wall and deletion of IL-22 leads to the development of less collagen-rich and smaller plaques in apoE<sup>-/-</sup> mice (Rattik et al. 2015). Furthermore, increased expression of genes related to VSMC contraction (vinculin, calmodulin, α-actin) was observed in arteries from HFD fed apoE/IL-22 double-knockout mice (Rattik et al. 2015). However, there is no difference in the expression of markers related to VSMC migration or synthesis phenotype such as vimentin, platelet-derived growth factor (PDGF), smoothelin, or matrix metalloproteinase 9 (MMP9) (Rattik et al. 2015). Thus, IL-22 mainly affects the early stage in the VSMC phenotypic switch but not VSMC functional features such as migration. In contrast, apoE<sup>-/-</sup> mice treated with recombinant mouse IL-22 showed reduced VSMC  $\alpha$ -actin expression and increased collagen contents (Shi et al. 2020a). Several studies demonstrated that IL-22 enhances the proliferation and migration of SMCs through NF-κB-, STAT3-, MAPK- and ERK1/2-dependent pathways (Bansal et al. 2013; Chang et al. 2011). Collectively, IL-22 acts as an important cytokine in the regulation of VSMC proliferation and migration and may be a promising therapeutic target for atherosclerosis.

# IL-22 in hypertension

Hypertension is known as an important risk factor for the development of atherosclerosis. Recent studies showed that IL-22 has a close relationship with human hypertension, and IL-22 is regarded as a promising therapeutic target for hypertension (Ye et al. 2017; Zhong et al. 2020). The plasma levels of IL-22 were markedly higher in patients with hypertension than in health participants,

and IL-22 levels were strikingly elevated and positively correlated with diastolic blood pressure and systolic blood pressure (Ye et al. 2017). Moreover, after treatment with recombinant mouse IL-22 in mice injected with angiotensin II, the interaction of IL-22 with IL-10R2 and IL-22R1 can activate the STAT3 signaling pathway, aggravating angiotensin II-induced inflammatory responses, endothelial dysfunction, and hypertension (Ye et al. 2017). Conversely, treatment with the anti-IL-22 neutralizing MAbs has an opposite biological function (Ye et al. 2017).

# Positive regulators of IL-22 in atherosclerosis

IL-23 plays a pro-inflammatory role in atherosclerosis. IL-23 aggravates atherosclerosis by increasing the release of inflammatory cytokines and the production of reactive oxygen species (ROS) in macrophages (Abbas et al. 2015; Döring 2015). Also, IL-23 is a crucial marker of proinflammatory macrophages. IL-23 is one of the major inducers of IL-22 production and expression (Kastelein et al. 2007). IL-23 promotes IL-22 expression via affecting a variety of cells including macrophages. It has been reported that IL-23 increases the expression of IL-22, which results in enhanced IL-23R expression (Sano et al. 2015). This leads to enhanced interaction between IL-23 and its receptor, and therefore promotes the production of IL-22. Also, IL-23 promotes the phosphorylation of phosphatidylinositol-3-kinase (PI3K)/Akt and inhibitor  $\kappa B-\alpha$  ( $I\kappa B-\alpha$ ), which activate the transcription factors STAT3- and NF-kB-dependent signaling in target cells, respectively (Cho et al. 2006). Thus, IL-23 may be an important regulator that synergistically enhances the pro-atherosclerotic role of IL-22.

# IL-7

IL-7, a key cytokine for the proliferation and development of T cells, drives inflammation in atherogenesis and promotes plaque instability in coronary artery disease involving interactions between chemokines, platelets, and monocytes (Damås et al. 2003; Arya et al. 2012). IL-7 can positively modulate the expression and production of IL-22 (Cella et al. 2010). However, IL-7 is unlikely to directly regulate IL-22 expression. IL-7 enhances the expression of retinoid-related orphan receptor gammat (RORyt), which is an important transcription factor for IL-22 expression (Vonarbourg et al. 2010). Furthermore, blockade of IL-7R signaling or IL-7Rα deficiency reduces RORyt expression in IL-22-producing cells (Vonarbourg et al. 2010). RORyt was reported to augment IL-22 enhancer activity by directly binding to the conserved noncoding sequence 32 (CNS 32) (Sekimata et al. 2019). Also, RORyt regulates the differentiation of Luo et al. Mol Med (2021) 27:88 Page 7 of 13

IL-22-producing cells to achieve the optimal conditions for IL-22 expression (Qiu et al. 2012; Nurieva et al. 2007; Sanos et al. 2009). Taken together, the IL-7/IL-7R pathway plays an important role in the regulation of IL-22 production by enhancing RORγt expression.

# Aryl hydrocarbon receptor

The aryl hydrocarbon receptor (AhR), an important ligand-activated transcription factor, has been reported to accelerate atherosclerosis through triggering vascular inflammation and promoting foam cell formation (Dahlem et al. 2020; Wu et al. 2011). AhR can directly modulate the expression and production of IL-22 gene or regulate the development and production of Th17 and innate lymphoid cells (ILCs) (IL-22-producing cells) (Qiu et al. 2012; Veldhoen et al. 2008). AhR is typically located in the cytoplasm in a complex with Hsp90 chaperones until the binding of its ligand induces a conformational change, leading to the separation of AhR from heat shock protein 90 (HSP90) (Esser et al. 2009). Then, AhR enters the nucleus together with its ligand and directly increases the expression of IL-22 gene. Of note, this transcriptional regulation does not depend on RORyt but may require the generation of endogenous AhR ligands by the Notch signaling pathway (Alam et al. 2010). Potential endogenous AhR ligands could be derived from the gut microbiota, or from the diet (Lee et al. 2011), including 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) and indol-3-carbinol, 6-formylindolo[3,2-b] carbazole (FICZ) (Nguyen and Bradfield 2008; Denison and Nagy 2003). It has been reported that FICZ is involved in Notch-mediated IL-22 expression and production (Oberg et al. 2005). In addition, several lines of evidence indicate that AhR also acts independently of IL-23 (Veldhoen et al. 2008, 2009). However, it is unclear whether AhR can act downstream or upstream of the IL-23/IL-23R pathway.

## IL-1β

IL-1β, a classic proinflammatory cytokine, has been shown to promote atherosclerotic plaque progression through increasing activation and proliferation of VSMCs and enhancing the synthesis of adhesion molecules on endothelial cells (Khan et al. 2018; Tumurkhuu et al. 2018). Also, high IL-1β concentrations are associated with an increased risk of hypertension (Krishnan et al. 2014; Alfaidi et al. 2018). Similar to IL-22, IL-1β is expressed in a variety of cell types related to the development of atherosclerosis including endothelial cells, macrophages, and VSMCs (Sims and Smith 2010). Interestingly, IL-1β was found to promote IL-22 production independently of IL-23, although they can synergistically induce the production of IL-22 (Sutton et al. 2009). Notably, constant IL-1β signaling is essential for sustained

expression of AHR and IL-22, which is different from IL-23 (Hughes et al. 2010). Moreover, ablation of IL-1R1 signaling diminishes IL-22 expression and production (Hughes et al. 2010). Taken together, these data suggest that the IL-1 $\beta$ / IL-1R1/IL-22 axis may play a key role in the aetiology of atherosclerosis.

# Notch

The Notch signaling pathways were found to promote atherosclerosis by regulating pro-inflammatory macrophage differentiation, endothelial dysfunction, and angiogenesis (Fukuda et al. 2012; Mao and Jiang 2018; Pitulescu et al. 2017). The Notch signaling is essential for the specific differentiation and development of IL-22-producing cells (Rankin et al. 2013; Mielke et al. 2013). The Notch can stimulate CD4<sup>+</sup> T cells to increase IL-22 production (Murano et al. 2014). Moreover, the Notch enhances IL-22-induced STAT3 expression through its target gene Hes1, thereby affecting IL-22 targets and IL-22 production (Murano et al. 2014). In contrast, IL-22 signaling and production are abrogated in Notchdeficient mice (Alam et al. 2010; Murano et al. 2014). In addition, it has been found that the Notch signaling promotes the expression and production of IL-22 through upregulating transcription factors AhR (Lee et al. 2011; Weidenbusch et al. 2017) and recombination signal binding protein-Jk (RBP-J) (Alam et al. 2010). These studies suggest that multiple transcription factors are involved in Notch signaling-mediated IL-22 expression.

# Negative regulators of IL-22 in atherosclerosis $\mathsf{TGF}\text{-}\beta$

Transforming growth factor-β (TGF-β) signaling plays a crucial role in the development and progression of atherosclerosis. Many studies demonstrated that activation of TGF-β signaling suppresses the development of atherosclerosis via regulating a variety of processes, including angiogenesis, VSMC proliferation and migration, inflammation, and foam cell formation (Grainger 2007; Chen et al. 2019b). TGF-β signaling was reported to serve as a suppressor of IL-22 expression and production in a dosedependent manner (Rutz et al. 2011; Volpe et al. 2009; Penel-Sotirakis et al. 2012). However, IL-23 can override the effect of TGF- $\beta$  on IL-22, thus promoting IL-22 expression and production (Volpe et al. 2009; Zhou et al. 2008). c-musculoaponeurotic fibrosarcoma (c-Maf) is a basic region leucine zipper (bZIP)-type transcription factor, which belongs to the Maf family. It has been reported that c-Maf specifically binds to the promoter of IL-22 gene and serves as a transcriptional repressor for IL-22 (Rutz et al. 2011). Moreover, in human T and invariant natural killer T (iNKT) cells, TGF-β signaling was found to suppress IL-22 expression by activating c-Maf (Rutz Luo et al. Mol Med (2021) 27:88 Page 8 of 13

et al. 2011). Of note, TGF- $\beta$  signaling does not suppress the production of IL-22 through the inhibition of AhR, BATF, or RORyt (Rutz et al. 2011). Thus, c-Maf is an important transcription factor for TGF- $\beta$  signaling-mediated IL-22 expression in T cells. It is necessary to determine whether c-Maf functions in a similar manner in other cell types for which TGF- $\beta$  was shown to inhibit IL-22.

#### IL-27

IL-27 was reported to prevent atherosclerosis in LDLR<sup>-/-</sup> mice by inhibiting the activation of macrophages (Hirase et al. 2013). Furthermore, knockout of IL-27 or IL-27 receptor in bone marrow-derived macrophages aggravates atherosclerosis and facilitates the activation of bone marrow-derived cells in the arterial wall (Hirase et al. 2013; Koltsova et al. 2012). IL-27 has also been identified as a negative modulator of IL-22 expression. IL-27 was found to inhibit the expression and production of IL-22 by upregulating suppressor of cytokine signaling 1 (SOCS1) expression (Wang et al. 2013). SOCS proteins are known as negative regulators of the Jak/STAT signal transduction (Yoshimura et al. 2007). AG490, an inhibitor of Jak2/STAT signaling, significantly blocks the inhibition of IL-22 production mediated by IL-27 (Wang et al. 2013). Besides, IL-27 also induces c-Maf expression, which inhibits the expression of IL-22 (Pot et al. 2009). Thus, IL-27 may attenuate the pro-atherosclerotic role of IL-22 by upregulating c-Maf and SOCS1 expression.

#### IL-38

IL-38 is known as an anti-inflammatory factor that suppresses pro-inflammatory cytokine production by its receptors and protects against cardiovascular disease (Graaf et al. 2021; Esmaeilzadeh et al. 2019). Recently, IL-38 overexpression was found to inhibit hyperlipidemia and atherosclerosis in apoE<sup>-/-</sup> by alleviating inflammation (Yang et al. 2018). IL-38 can regulate IL-22 expression in a dose-dependent manner. At low concentrations, IL-38 suppresses IL-22 production, whereas, at high concentrations, IL-38 modestly increases IL-22 production (Veerdonk et al. 2012; Tortola et al. 2012). Thus, further studies are needed to explore the exact mechanism by which IL-38 regulates IL-22 expression and production in atherosclerosis.

#### IL-25

IL-25 was reported to inhibit the development of atherosclerosis by decreasing inflammation and oxidized LDL-specific antibodies (Mantani et al. 2015; Mantani et al. 2018a). Also, IL-25 appears to be expressed by cells commonly found in atherosclerotic plaques such as endothelial cells, VSMCs, and macrophages (Mantani

et al. 2018a, b). It has been reported that serum levels of IL-25 are significantly correlated with IL-22 in rheumatoid arthritis patients (Min et al. 2020). Addition of IL-25 markedly reduces IL-22 expression but not changes mRNA levels of IL-22 receptors, IL-10RB and IL-22R1 (Min et al. 2020), suggesting that the suppressive role of IL-25 on IL-22 expression is independent with IL-22 receptors. Further studies showed that IL-25 also inhibits IL-22-induced STAT3 and p38 MAPK/IkB $\alpha$  pathway (Min et al. 2020). Thus, upregulation of IL-25 may alleviate atherosclerosis by reducing IL-22 expression.

# **Conclusions and future directions**

IL-22, an important member of the IL-10 cytokine family, has attracted growing interest in recent years. IL-22 plays a critical role in the development of atherosclerosis through its involvement in the modulation of angiogenesis, inflammatory response, hypertension, VSMC proliferation and migration, and cholesterol metabolism (Fig. 1). Also, IL-22 may be regulated by other cytokines and proteins during atherosclerosis (Table 1). However, the immunoregulatory role of IL-22 appears complicated. Although numerous data have established the proatherogenic effects, a recent study revealed a paradoxical result that IL-22 protects against WD-induced atherosclerosis in LDLR<sup>-/-</sup> mice by regulating intestinal microbial homeostasis (Fatkhullina et al. 2018). The paradoxical effects of IL-22 on immunoregulation may be diet- or mouse model-specific effects, which should be carefully considered when we use drugs to modulate IL-22 expression and production in vivo. In addition, it has been reported that IL-22R1 is expressed in mouse atherosclerotic plagues, and its expression levels are increased in Apo $E^{-/-}$  mice (Shi et al. 2020a). However, the expression of other IL-22 receptors (IL-10R and IL-22BP) in atherosclerosis remains largely unknown. Overall, increasing studies support the view that targeting IL-22 may be a promising therapeutic strategy for atherosclerosis.

Numerous studies demonstrated that endothelial cells, and macrophages are the major effector cells involving atherosclerosis (Cho et al. 2011; Menon and Fisher 2015; Raman and Khanal 2021). Importantly, IL-22 and IL-22R1 are widely expressed in these cells. Also, increasing evidence suggests that IL-22 may promote atherosclerosis through multiple mechanisms by these effector cells. Thus, these cells may be the main target for IL-22 in atherosclerosis. In the future, more data are required to reveal the exact role of IL-22 in these effector cells during the development of atherosclerosis. Although many studies have demonstrated the beneficial effects of decreased expression, it is currently difficult to speculate on the most effective method to inhibit IL-22 expression in atherosclerosis. Targeting IL-22 or Luo et al. Mol Med (2021) 27:88 Page 9 of 13

**Table 1** Potential regulators of IL-22 expression and production in atherosclerosis

Molecules	Type of regulation	Transcription factor	Pathway	Potential role in AS	References
IL-23	Positive	STAT3, NF-κB	IL-23/IL-23R, PI3K/Akt, IκB-α	Promotes inflammatory responses	Sano et al. 2015; Cho et al. 2006)
IL-7	Positive	RORyt	IL-7/IL-7R	Promotes inflammatory responses	Vonarbourg et al. 2010; Seki- mata et al. 2019)
AhR	Positive	-	Notch/FICZ	Promotes inflammatory responses and cholesterol accumulation	Qiu et al. 2012; Veldhoen et al. 2008)
IL-1β	Positive	AHR	IL-1β/IL-1R1	Promotes inflammatory responses and VSMC proliferation and migration	Sutton et al. 2009; Hughes et al. 2010)
Notch	Positive	AhR, RBP-J, STAT3	Notch-Hes1	Promotes inflammatory responses, endothelial dysfunction, and angiogenesis	Rankin et al. 2013; Mielke et al. 2013; Murano et al. 2014)
TGF-β	Negative	c-Maf	TGF-β signaling	Suppresses angiogenesis, VSMC proliferation and migration, inflammation, and cholesterol accumula- tion	Rutz et al. 2011; Volpe et al. 2009; Penel-Sotirakis et al. 2012)
IL-27	Negative	SOCS1, c-Maf	Jak/STAT signaling	Suppresses inflammatory responses	Wang et al. 2013; Pot et al. 2009)
IL-38	Negative	-	IL-38- IL-36R	Suppresses inflammatory responses	Veerdonk et al. 2012; Tortola et al. 2012)
IL-25	Negative	STAT3	p38 MAPK/IĸBa	Suppresses inflammatory responses	Min et al. 2020)

<sup>-</sup> Not determined, AS atherosclerosis

its receptor complex is the most direct method to inhibit IL-22-mediated functions. Preclinical and clinical studies have demonstrated that a human anti-IL-22-mAb (fezakinumab) and IL-22R1-targeting mAbs are currently tested as a treatment for several inflammatory diseases such as rheumatoid arthritis and severe atopic dermatitis, without obvious adverse safety concerns observed (Sabat et al. 2014; Kragstrup et al. 2018; Guttman-Yassky et al. 2018). Atherosclerosis is also known as an inflammatory disease; however, it is not clear whether these therapeutic strategies can protect against the development of atherosclerosis. Therefore, both clinical and basic studies are needed to validate this possibility. Besides, there are several crucial and insightful questions that require to be answered. First, both innate and adaptive immunity play important roles in the development of atherosclerosis (Hansson et al. 2002; Miteva et al. 2018). The crosstalk of IL-22 and innate and adaptive immunity in atherosclerosis is worthy of attention. In particular, monocytes/macrophages are known as key immune cells, which have important proatherogenic effects. Th1 response has a potent proatherogenic effect, but regulatory T cells and some Th2-related cytokines play antiatherosclerotic protective roles. Thus, it is necessary to explore which immune cells-derived IL-22 affects these atherosclerosis-related immune cells. Second, how to most effectively target IL-22, either by transcriptional or posttranscriptional modulation? Third, as an important proinflammatory cytokine, does IL-22 play a role in pyroptosis (a pivotal driver of atherosclerosis)? Fourth, do a specific and efficient method to counteract the detrimental actions of IL-22 on atherogenesis exist in vivo? Finally, reverse cholesterol transport (RCT), an important protective mechanism against the development of atherogenesis, is a dynamic process in which excessive cholesterol from extrahepatic tissues is delivered to the liver for further metabolism and biliary excretion (Rader et al. 2009; Li et al. 2021). Therefore, it is of importance to detect whether IL-22 can increase the efficiency of RCT. The answers to these sorts of questions will undoubtedly provide unique insights into the roles of IL-22 in atherosclerosis and make IL-22 an attractive therapeutic target for therapy aiming at the reduction of atherogenesis.

#### Abbreviations

CVDs: Cardiovascular diseases; JAK: Janus kinase; VSMC: Vascular smooth muscle cell; IL-22: Interleukin-22; MMP9: Matrix metalloproteinase 9; VLDL: Very low-density lipoprotein; MAbs: Monoclonal antibodies; STAT: Signal transducer and activator of transcription; HFD: High-fat diet; GSK3ß: Glycogen synthase kinase 3ß; IL-22BP: IL-22 binding protein; VCAM-1: Vascular cell adhesion molecule-1; WD: Western diet; JNK: C-Jun N-terminal kinase; TLR4: The toll-like receptor 4; rIL-22: Recombinant mouse IL-22; SRA: Scavenger receptor class A; RAGE: Receptor for advanced glycation endproducts; MAPK: Mitogen-activated protein kinase; AMI: Acute myocardial infarction; TMAO: Trimethylamine N-oxide; LPS: Lipopolysaccharide; LOX-1: Lectin-like oxidized low-density lipoprotein receptor-1; ICAM-1: Intercellular adhesion molecule-1; bZIP: Basic

Luo et al. Mol Med (2021) 27:88 Page 10 of 13

region leucine zipper; HUVECs: Human umbilical vein endothelial cells; HDL-C: High-density lipoprotein cholesterol; CXCL: CXC chemokine ligands; ERK1/2: Extracellular signal-regulated kinase-1/2; PDGF: Platelet-derived growth factor; LXR: Liver X receptor; AhR: Aryl hydrocarbon receptor; ILCs: Innate lymphoid cells; TGF- $\beta$ : Transforming growth factor- $\beta$ ; LDL-C: Low-density lipoprotein cholesterol; c-Maf: C-musculoaponeurotic fibrosarcoma; ABCA1: ATP-binding cassette transporter A1; iNKT: Invariant natural killer T; SOCS1: Suppressor of cytokine signaling 1; RCT: Reverse cholesterol transport; WHO: World Health Organization.

#### Acknowledgements

Not applicable.

#### Authors' contributions

Conception and design: PH and HY. Manuscript writing: J-WL and YH. Collection and assembly of data: JL. Final approval of manuscript: All authors. All authors read and approved the final manuscript.

#### **Funding**

The authors gratefully acknowledge the financial support from the Hunan Province Technology Innovation Guidance Program-Clinical Medical Technology Innovation Guidance Project (No. 2018SK50410).

#### Availability of data and materials

Not applicable.

#### **Declarations**

# Ethical approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

# Competing interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

#### **Author details**

<sup>1</sup>Department of Cardio-Thoracic Surgery, Hunan Children's Hospital, Changsha 410007, People's Republic of China. <sup>2</sup>Department of Ultrasound Medicine, Hunan Children's Hospital, Changsha 410007, People's Republic of China. <sup>3</sup>Department of Respiratory Medicine, Hunan Provincial People's Hospital, Changsha, Hunan 410001, People's Republic of China.

Received: 14 May 2021 Accepted: 7 August 2021 Published online: 13 August 2021

#### References

- Abbas A, et al. Interleukin 23 levels are increased in carotid atherosclerosis: possible role for the interleukin 23/interleukin 17 axis. Stroke. 2015;46:793–9.
- Agrawal S, et al. Signal transducer and activator of transcription 1 is required for optimal foam cell formation and atherosclerotic lesion development. Circulation. 2007;115:2939–47.
- Alam MS, et al. Notch signaling drives IL-22 secretion in CD4+T cells by stimulating the aryl hydrocarbon receptor. Proc Natl Acad Sci USA. 2010;107:5943–8.
- Alfaidi MA, et al. Dietary docosahexaenoic acid reduces oscillatory wall shear stress, atherosclerosis, and hypertension, most likely mediated via an IL-1-mediated mechanism. J Am Heart Assoc. 2018;7:008757.
- Andoh A, et al. Interleukin-22, a member of the IL-10 subfamily, induces inflammatory responses in colonic subepithelial myofibroblasts. Gastroenterology. 2005;129:969–84.
- Arya AK, Pokharia D, Bhan S, Tripathi R, Tripathi K. Correlation between IL-7 and MCP-1 in diabetic chronic non healing ulcer patients at higher risk of coronary artery disease. Cytokine. 2012;60:767–71.

- Aujla SJ, Kolls JK. IL-22: a critical mediator in mucosal host defense. J Mol Med (berl). 2009;87:451–4.
- Bansal G, et al. IL-22 activates oxidant signaling in pulmonary vascular smooth muscle cells. Cell Signal. 2013;25:2727–33.
- Basatemur GL, Jørgensen HF, Clarke MCH, Bennett MR, Mallat Z. Vascular smooth muscle cells in atherosclerosis. Nat Rev Cardiol. 2019;16:727–44.
- Bleicher L, et al. Crystal structure of the IL-22/IL-22R1 complex and its implications for the IL-22 signaling mechanism. FEBS Lett. 2008;582:2985–92.
- Camaré C, Pucelle M, Nègre-Salvayre A, Salvayre R. Angiogenesis in the atherosclerotic plaque. Redox Biol. 2017;12:18–34.
- Cella M, Otero K, Colonna M. Expansion of human NK-22 cells with IL-7, IL-2, and IL-1beta reveals intrinsic functional plasticity. Proc Natl Acad Sci USA. 2010;107:10961–6.
- Chang Y, et al. TH17 cytokines induce human airway smooth muscle cell migration. J Allergy Clin Immunol. 2011;127:1046–53 (e1041-1042).
- Chellan B, Yan L, Sontag TJ, Reardon CA, Hofmann Bowman MA. IL-22 is induced by S100/calgranulin and impairs cholesterol efflux in macrophages by downregulating ABCG1. J Lipid Res. 2014;55:443–54.
- Chen Y, et al. Inhibition of ERK1/2 and activation of LXR synergistically reduce atherosclerotic lesions in ApoE-deficient mice. Arterioscler Thromb Vasc Biol. 2015;35:948–59.
- Chen Q, et al. Targeted inhibition of STAT3 as a potential treatment strategy for atherosclerosis. Theranostics. 2019a;9:6424–42.
- Chen PY, et al. Endothelial TGF- $\beta$  signalling drives vascular inflammation and atherosclerosis. Nat Metab. 2019b;1:912–26.
- Chistiakov DA, Orekhov AN, Bobryshev YV. Vascular smooth muscle cell in atherosclerosis. Acta Physiol (oxf). 2015;214:33–50.
- Chmielewski S, Piaszyk-Borychowska A, Wesoly J, Bluyssen HA. STAT1 and IRF8 in vascular inflammation and cardiovascular disease: diagnostic and therapeutic potential. Int Rev Immunol. 2016;35:434–54.
- Cho ML, et al. STAT3 and NF-kappaB signal pathway is required for IL-23-mediated IL-17 production in spontaneous arthritis animal model IL-1 receptor antagonist-deficient mice. J Immunol (baltimore, Md: 1950). 2006:176:5652-61.
- Cho Y, et al. Adipokine resistin is a key player to modulate monocytes, endothelial cells, and smooth muscle cells, leading to progression of atherosclerosis in rabbit carotid artery. J Am Coll Cardiol. 2011;57:99–109.
- Coccia EM, et al. STAT1 activation during monocyte to macrophage maturation: role of adhesion molecules. Int Immunol. 1999;11:1075–83.
- Crucet M, et al. Hypoxia enhances lipid uptake in macrophages: role of the scavenger receptors Lox1, SRA, and CD36. Atherosclerosis. 2013;229:110–7.
- Dahlem C, et al. AHR signaling interacting with nutritional factors regulating the expression of markers in vascular inflammation and atherogenesis. Int J Mol Sci. 2020;21:8287.
- Damås JK, et al. Interleukin-7-mediated inflammation in unstable angina: possible role of chemokines and platelets. Circulation. 2003;107:2670–6.
- de Graaf DM, et al. Reduced concentrations of the B cell cytokine interleukin 38 are associated with cardiovascular disease risk in overweight subjects. Eur J Immunol. 2021;51:662–71.
- de Oliveira NM, et al. Interleukin-22 forms dimers that are recognized by two interleukin-22R1 receptor chains. Biophys J. 2008;94:1754–65.
- Denison MS, Nagy SR. Activation of the aryl hydrocarbon receptor by structurally diverse exogenous and endogenous chemicals. Annu Rev Pharmacol Toxicol. 2003;43:309–34.
- Döring Y. Not growth but death: GM-CSF/IL-23 axis drives atherosclerotic plaque vulnerability by enhancing macrophage and DC apoptosis. Circ Res. 2015;116:222–4.
- Dudakov JA, Hanash AM, van den Brink MR. Interleukin-22: immunobiology and pathology. Annu Rev Immunol. 2015;33:747–85.
- Dumoutier L, Louahed J, Renauld JC. Cloning and characterization of IL-10-related T cell-derived inducible factor (IL-TIF), a novel cytokine structurally related to IL-10 and inducible by IL-9. J Immunol (baltimore, Md: 1950). 2000;164:1814–9.
- Dumoutier L, Lejeune D, Colau D, Renauld JC. Cloning and characterization of IL-22 binding protein, a natural antagonist of IL-10-related T cell-derived inducible factor/IL-22. J Immunol (baltimore, Md: 1950). 2001;166:7090–5.
- Eidenschenk C, Rutz S, Liesenfeld O, Ouyang W. Role of IL-22 in microbial host defense. Curr Top Microbiol Immunol. 2014;380:213–36.

Luo et al. Mol Med (2021) 27:88 Page 11 of 13

- Esmaeilzadeh A, Pouyan S, Erfanmanesh M. Is interleukin-38 a key player cytokine in atherosclerosis immune gene therapy? Med Hypotheses. 2019;125:139–43.
- Esser C, Rannug A, Stockinger B. The aryl hydrocarbon receptor in immunity. Trends Immunol. 2009;30:447–54.
- Fatkhullina AR, et al. An Interleukin-23-Interleukin-22 axis regulates intestinal microbial homeostasis to protect from diet-induced atherosclerosis. Immunity. 2018;49:943-957.e949.
- Fukaya T, et al. Pivotal role of IL-22 binding protein in the epithelial autoregulation of interleukin-22 signaling in the control of skin inflammation. Front Immunol. 2018;9:1418.
- Fukuda D, et al. Notch ligand delta-like 4 blockade attenuates atherosclerosis and metabolic disorders. Proc Natl Acad Sci USA. 2012;109:E1868-1877.
- Galkina E, Ley K. Vascular adhesion molecules in atherosclerosis. Arterioscler Thromb Vasc Biol. 2007;27:2292–301.
- Gan ZS, et al. Iron reduces M1 macrophage polarization in RAW264.7 macrophages associated with inhibition of STAT1. Mediators Inflamm 2017: 8570818.
- GBD 2015 Risk Factors Collaborators. Global, regional, and national comparative risk assessment of 79 behavioural, environmental and occupational, and metabolic risks or clusters of risks, 1990–2015: a systematic analysis for the Global Burden of Disease Study 2015. Lancet (london, England). 2016;388:1659–724.
- Grainger DJ. TGF-beta and atherosclerosis in man. Cardiovasc Res. 2007;74:213–22.
- Gruenberg BH, et al. A novel, soluble homologue of the human IL-10 receptor with preferential expression in placenta. Genes Immun. 2001;2:329–34.
- Guttman-Yassky E, et al. Efficacy and safety of fezakinumab (an IL-22 monoclonal antibody) in adults with moderate-to-severe atopic dermatitis inadequately controlled by conventional treatments: a randomized, double-blind, phase 2a trial. J Am Acad Dermatol. 2018;78:872-881.
- Hansson GK, Libby P, Schönbeck U, Yan ZQ. Innate and adaptive immunity in the pathogenesis of atherosclerosis. Circ Res. 2002;91:281–91.
- Hirase T, et al. Interleukin 27 inhibits atherosclerosis via immunoregulation of macrophages in mice. Am J Physiol Heart Circ Physiol. 2013;305:H420-429.
- Hou Y, et al. IL-23-induced macrophage polarization and its pathological roles in mice with imiquimod-induced psoriasis. Protein Cell. 2018;9:1027–38.
- Hu H, Li L, Yu T, Li Y, Tang Y. Interleukin-22 receptor 1 upregulation and activation in hypoxic endothelial cells improves perfusion recovery in experimental peripheral arterial disease. Biochem Biophys Res Commun. 2018;505:60–6.
- Hughes T, et al. Interleukin-1 beta selectively expands and sustains interleukin-22+ immature human natural killer cells in secondary lymphoid tissue. Immunity. 2010;32:803–14.
- Jaipersad AS, Lip GY, Silverman S, Shantsila E. The role of monocytes in angiogenesis and atherosclerosis. J Am Coll Cardiol. 2014;63:1–11.
- Jones BC, Logsdon NJ, Walter MR. Structure of IL-22 bound to its high-affinity IL-22R1 chain. Structure. 2008a;16:1333–44.
- Jones BC, Logsdon NJ, Walter MR. Crystallization and preliminary X-ray diffraction analysis of human IL-22 bound to the extracellular IL-22R1 chain. Acta Crystallogr Sect F Struct Biol Cryst Commun. 2008b;64:266–9.
- Kastelein RA, Hunter CA, Cua DJ. Discovery and biology of IL-23 and IL-27: related but functionally distinct regulators of inflammation. Annu Rev Immunol. 2007;25:221–42.
- Khan R, Rheaume E, Tardif JC. Examining the role of and treatment directed at IL-1ß in atherosclerosis. Curr Atheroscler Rep. 2018;20:53.
- Koltsova EK, et al. Interleukin-27 receptor limits atherosclerosis in Ldlr-/- mice. Circ Res. 2012;111:1274–85.
- Kong Q, et al. IL-22 exacerbates the severity of CVB3-induced acute viral myocarditis in IL-17A-deficient mice. Mol Med Rep. 2013;7:1329–35.
- Kong Q, et al. Increased circulating T-helper 22 cells in patients with dilated cardiomyopathy. Mol Med Rep. 2014;10:359–64.
- Kostov K, Halacheva L. Role of magnesium deficiency in promoting atherosclerosis, endothelial dysfunction, and arterial stiffening as risk factors for hypertension. Int J Mol Sci. 2018;19:1724.
- Kotenko SV, et al. Identification and functional characterization of a second chain of the interleukin-10 receptor complex. EMBO J. 1997;16:5894–903.

- Kotenko SV, et al. Identification, cloning, and characterization of a novel soluble receptor that binds IL-22 and neutralizes its activity. J Immunol (baltimore, Md: 1950). 2001;166:7096–103.
- Kragstrup TW, et al. The IL-20 cytokine family in rheumatoid arthritis and spondyloarthritis. Front Immunol. 2018;9:2226.
- Krishnan SM, Sobey CG, Latz E, Mansell A, Drummond GR. IL-1β and IL-18: inflammatory markers or mediators of hypertension? Br J Pharmacol. 2014;171:5589–602.
- La Manna S, et al. Characterization of linear mimetic peptides of Interleukin-22 from dissection of protein interfaces. Biochimie. 2017;138:106–15.
- Langer JA, Cutrone EC, Kotenko S. The class II cytokine receptor (CRF2) family: overview and patterns of receptor-ligand interactions. Cytokine Growth Factor Rev. 2004;15:33–48.
- Larrede S, et al. Stimulation of cholesterol efflux by LXR agonists in cholesterol-loaded human macrophages is ABCA1-dependent but ABCG1-independent. Arterioscler Thromb Vasc Biol. 2009;29:1930–6.
- Lawrence T, Natoli G. Transcriptional regulation of macrophage polarization: enabling diversity with identity. Nat Rev Immunol. 2011;11:750–61.
- Lee JS, et al. AHR drives the development of gut ILC22 cells and postnatal lymphoid tissues via pathways dependent on and independent of Notch. Nat Immunol. 2011:13:144–51.
- Lejeune D, et al. Interleukin-22 (IL-22) activates the JAK/STAT, ERK, JNK, and p38 MAP kinase pathways in a rat hepatoma cell line. Pathways that are shared with and distinct from IL-10. J Biol Chem. 2002;277:33676–82.
- Li J, et al. Temporal associations between interleukin 22 and the extracellular domains of IL-22R and IL-10R2. Int Immunopharmacol. 2004;4:693–708.
- Li H, Jiang T, Li MQ, Zheng XL, Zhao GJ. Transcriptional regulation of macrophages polarization by MicroRNAs. Front Immunol. 2018;9:1175.
- Li H, Zou J, Yu XH, Ou X, Tang CK. Zinc finger E-box binding homeobox 1 and atherosclerosis: new insights and therapeutic potential. J Cell Physiol. 2021a:236:4216–30.
- Li H, Yu XH, Ou X, Ouyang XP, Tang CK. Hepatic cholesterol transport and its role in non-alcoholic fatty liver disease and atherosclerosis. Prog Lipid Res. 2021;83:101109.
- Lian S, et al. S-nitrosocaptopril interrupts adhesion of cancer cells to vascular endothelium by suppressing cell adhesion molecules via inhibition of the NF-κB and JAK/STAT signal pathways in endothelial cells. Eur J Pharmacol. 2016;791:62–71.
- Libby P, Hansson GK. Taming immune and inflammatory responses to treat atherosclerosis. J Am Coll Cardiol. 2018;71:173–6.
- Linton MF, Moslehi JJ, Babaev VR. Akt signaling in macrophage polarization, survival, and atherosclerosis. Int J Mol Sci. 2019;20:2703.
- Lodge S, et al. NMR spectroscopic windows on the systemic effects of SARS-CoV-2 infection on plasma lipoproteins and metabolites in relation to circulating cytokines. J Proteome Res. 2021;20:1382–96.
- Logsdon NJ, Jones BC, Josephson K, Cook J, Walter MR. Comparison of inter-leukin-22 and interleukin-10 soluble receptor complexes. J Interferon Cytokine Res. 2002;22:1099–112.
- $\label{logs} Logs don NJ, et al. The IL-10R2 binding hot spot on IL-22 is located on the N-terminal helix and is dependent on N-linked glycosylation. J Mol Biol. 2004;342:503–14.$
- Mantani PT, et al. IL-25 inhibits atherosclerosis development in apolipoprotein E deficient mice. PLoS ONE. 2015;10:0117255.
- Mantani PT, et al. Interleukin-25 (IL-25) has a protective role in atherosclerosis development in the aortic arch in mice. J Biol Chem. 2018a;293:6791–801.
- Mantani PT, et al. Interleukin-25 reduces Th17 cells and inflammatory responses in human peripheral blood mononuclear cells. Hum Immunol. 2018b;79:685–92.
- Mao YZ, Jiang L. Effects of Notch signalling pathway on the relationship between vascular endothelial dysfunction and endothelial stromal transformation in atherosclerosis. Artif Cells Nanomed Biotechnol. 2018;46:764–72.
- Menon P, Fisher EA. Immunostaining of macrophages, endothelial cells, and smooth muscle cells in the atherosclerotic mouse aorta. Methods Mol Biol. 2015;1339:131–48.
- Mielke LA, et al. TCF-1 controls ILC2 and NKp46+RORγt+ innate lymphocyte differentiation and protection in intestinal inflammation. J Immunol (baltimore, Md: 1950). 2013;191:4383–91.

Luo et al. Mol Med (2021) 27:88 Page 12 of 13

- Min HK, et al. Interleukin (IL)-25 suppresses IL-22-induced osteoclastogenesis in rheumatoid arthritis via STAT3 and p38 MAPK/IκBα pathway. Arthritis Res Ther. 2020;22:222.
- Miteva K, Madonna R, De Caterina R, Van Linthout S. Innate and adaptive immunity in atherosclerosis. Vascul Pharmacol. 2018;107:67–77.
- Moore KJ, Tabas I. Macrophages in the pathogenesis of atherosclerosis. Cell. 2011;145:341–55.
- Mühl H, Bachmann M. IL-18/IL-18BP and IL-22/IL-22BP: two interrelated couples with therapeutic potential. Cell Signal. 2019;63:109388.
- Mühl H, et al. IL-22 in tissue-protective therapy. Br J Pharmacol. 2013;169:761–71.
- Murano T, et al. Hes1 promotes the IL-22-mediated antimicrobial response by enhancing STAT3-dependent transcription in human intestinal epithelial cells. Biochem Biophys Res Commun. 2014;443:840–6.
- Nagem RA, et al. Crystal structure of recombinant human interleukin-22. Structure. 2002;10:1051–62.
- Nagem RA, Ferreira Júnior JR, Dumoutier L, Renauld JC, Polikarpov I. Interleukin-22 and its crystal structure. Vitam Horm. 2006;74:77–103.
- Nguyen LP, Bradfield CA. The search for endogenous activators of the aryl hydrocarbon receptor. Chem Res Toxicol. 2008;21:102–16.
- Nie Y, et al. STAT3 inhibition of gluconeogenesis is downregulated by SirT1. Nat Cell Biol. 2009;11:492–500.
- Nurieva R, et al. Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. Nature. 2007;448:480–3.
- Oberg M, Bergander L, Håkansson H, Rannug U, Rannug A. Identification of the tryptophan photoproduct 6-formylindolo[3,2-b]carbazole, in cell culture medium, as a factor that controls the background aryl hydrocarbon receptor activity. Toxicol Sci. 2005;85:935–43.
- Penel-Sotirakis K, Simonazzi E, Péguet-Navarro J, Rozières A. Differential capacity of human skin dendritic cells to polarize CD4+ T cells into IL-17, IL-21 and IL-22 producing cells. PLoS ONE. 2012;7:45680.
- Piaszyk-Borychowska A, et al. Signal integration of IFN-I and IFN-II with TLR4 involves sequential recruitment of STAT1-complexes and NFkB to enhance pro-inflammatory transcription. Front Immunol. 2019;10:1253.
- Pickert G, et al. STAT3 links IL-22 signaling in intestinal epithelial cells to mucosal wound healing. J Exp Med. 2009;206:1465–72.
- Pitulescu ME, et al. Dll4 and Notch signalling couples sprouting angiogenesis and artery formation. Nat Cell Biol. 2017;19:915–27.
- Pot C, et al. Cutting edge: IL-27 induces the transcription factor c-Maf, cytokine IL-21, and the costimulatory receptor ICOS that coordinately act together to promote differentiation of IL-10-producing Tr1 cells. J Immunol Baltimore, Md: 1950. 2009;183:797–801.
- Protopsaltis NJ, Liang W, Nudleman E, Ferrara N. Interleukin-22 promotes tumor angiogenesis. Angiogenesis. 2019;22:311–23.
- Qiu J, et al. The aryl hydrocarbon receptor regulates gut immunity through modulation of innate lymphoid cells. Immunity. 2012;36:92–104.
- Rader DJ, Alexander ET, Weibel GL, Billheimer J, Rothblat GH. The role of reverse cholesterol transport in animals and humans and relationship to atherosclerosis. J Lipid Res. 2009;50(Suppl):S189-194.
- Raman P, Khanal S. Leptin in atherosclerosis: focus on macrophages, endothelial and smooth muscle cells. Int J Mol Sci. 2021;22:5446.
- Rankin LC, et al. The transcription factor T-bet is essential for the development of NKp46+ innate lymphocytes via the Notch pathway. Nat Immunol. 2013;14:389–95.
- Rattik S, et al. IL-22 affects smooth muscle cell phenotype and plaque formation in apolipoprotein E knockout mice. Atherosclerosis. 2015;242:506–14.
- Ren K, et al. Mangiferin promotes macrophage cholesterol efflux and protects against atherosclerosis by augmenting the expression of ABCA1 and ABCG1. Aging. 2019;11:10992–1009.
- Reustle A, Torzewski M. Role of p38 MAPK in atherosclerosis and aortic valve sclerosis. Int J Mol Sci. 2018;19:3761.
- Rutz S, et al. Transcription factor c-Maf mediates the TGF- $\beta$ -dependent suppression of IL-22 production in T(H)17 cells. Nat Immunol. 2011;12:1238–45.
- Rutz S, Eidenschenk C, Ouyang W. IL-22, not simply a Th17 cytokine. Immunol Rev. 2013;252:116–32.
- Sabat R, Ouyang W, Wolk K. Therapeutic opportunities of the IL-22-IL-22R1 system. Nat Rev Drug Discovery. 2014;13:21–38.
- Sano T, et al. An IL-23R/IL-22 circuit regulates epithelial serum amyloid A to promote local effector Th17 responses. Cell. 2015;163:381–93.

- Sanos SL, et al. RORgammat and commensal microflora are required for the differentiation of mucosal interleukin 22-producing NKp46+ cells. Nat Immunol. 2009;10:83–91.
- Sekimata M, et al. Runx1 and ROR $\gamma$ t cooperate to upregulate IL-22 expression in Th cells through its distal enhancer. J Immunol (baltimore, Md: 1950). 2019;202:3198–210.
- Sestito R, et al. STAT3-dependent effects of IL-22 in human keratinocytes are counterregulated by sirtuin 1 through a direct inhibition of STAT3 acetylation. FASEB j: off Publ Federation Am Soc Exp Biol. 2011;25:916–27.
- Shang WQ, et al. Blocking IL-22, a potential treatment strategy for adenomyosis by inhibiting crosstalk between vascular endothelial and endometrial stromal cells. Am J Transl Res. 2015;7:1782–97.
- Shen L, Liu Y. Prostaglandin E2 blockade enhances the pulmonary anti-Cryptococcus neoformans immune reaction via the induction of TLR-4. Int Immunopharmacol. 2015;28:376–81.
- Shen-Orr SS, et al. Defective signaling in the JAK-STAT pathway tracks with chronic inflammation and cardiovascular risk in aging humans. Cell Syst. 2016;3:374-384.e374.
- Shi L, et al. IL-22 produced by Th22 cells aggravates atherosclerosis development in ApoE(-/-) mice by enhancing DC-induced Th17 cell proliferation. J Cell Mol Med. 2020a;24:3064–78.
- Shi X, et al. Xanthoplanine attenuates macrophage polarization towards M1 and inflammation response via disruption of CrkL-STAT5 complex. Arch Biochem Biophys. 2020b;683:108325.
- Shohan M, et al. Interleukin-22 and intestinal homeostasis: protective or destructive? IUBMB Life. 2020;72:1585–602.
- Sikorski K, et al. STAT1-mediated signal integration between IFNγ and LPS leads to increased EC and SMC activation and monocyte adhesion. Am J Physiol Cell Physiol. 2011a;300:C1337-1344.
- Sikorski K, Czerwoniec A, Bujnicki JM, Wesoly J, Bluyssen HA. STAT1 as a novel therapeutical target in pro-atherogenic signal integration of IFNY, TLR4 and IL-6 in vascular disease. Cytokine Growth Factor Rev. 2011 b;22:211–9.
- Sims JE, Smith DE. The IL-1 family: regulators of immunity. Nat Rev Immunol. 2010;10:89–102.
- Sonnenberg GF, Fouser LA, Artis D. Border patrol: regulation of immunity, inflammation and tissue homeostasis at barrier surfaces by IL-22. Nat Immunol. 2011;12:383–90.
- Soriano SF, et al. Chemokines integrate JAK/STAT and G-protein pathways during chemotaxis and calcium flux responses. Eur J Immunol. 2003;33:1328–33.
- Sutton CE, et al. Interleukin-1 and IL-23 induce innate IL-17 production from gammadelta T cells, amplifying Th17 responses and autoimmunity. Immunity. 2009;31:331–41.
- Tabas I, Bornfeldt KE. Macrophage phenotype and function in different stages of atherosclerosis. Circ Res. 2016;118:653–67.
- Tarling EJ, Edwards PA. ATP binding cassette transporter G1 (ABCG1) is an intracellular sterol transporter. Proc Natl Acad Sci USA. 2011;108:19719–24.
- Torquati L, et al. Fibre intake is independently associated with increased circulating interleukin-22 in individuals with metabolic syndrome. Nutrients. 2019:11:815.
- Tortola L, et al. Psoriasiform dermatitis is driven by IL-36-mediated DC-keratinocyte crosstalk. J Clin Investig. 2012;122:3965–76.
- Trevejo-Nunez G, et al. Interleukin-22 (IL-22) binding protein constrains IL-22 activity, host defense, and oxidative phosphorylation genes during pneumococcal pneumonia. Infect Immun. 2019;87:e00550-e619.
- Tumurkhuu G, et al. Chlamydia pneumoniae Hijacks a host autoregulatory IL-1β loop to drive foam cell formation and accelerate atherosclerosis. Cell Metab. 2018;28:432-448.e434.
- van de Veerdonk FL, et al. IL-38 binds to the IL-36 receptor and has biological effects on immune cells similar to IL-36 receptor antagonist. Proc Natl Acad Sci USA. 2012;109:3001–5.
- van Hoeven V, et al. Mesenchymal stromal cells stimulate the proliferation and IL-22 production of group 3 innate lymphoid cells. J Immunol (baltimore, Md: 1950). 2018;201:1165–73.
- Veldhoen M, et al. The aryl hydrocarbon receptor links TH17-cell-mediated autoimmunity to environmental toxins. Nature. 2008;453:106–9.
- Veldhoen M, Hirota K, Christensen J, O'Garra A, Stockinger B. Natural agonists for aryl hydrocarbon receptor in culture medium are essential for optimal differentiation of Th17 T cells. J Exp Med. 2009;206:43–9.

Luo et al. Mol Med (2021) 27:88 Page 13 of 13

- Volpe E, et al. Multiparametric analysis of cytokine-driven human Th17 differentiation reveals a differential regulation of IL-17 and IL-22 production. Blood. 2009;114:3610–4.
- Vonarbourg C, et al. Regulated expression of nuclear receptor RORyt confers distinct functional fates to NK cell receptor-expressing RORyt(+) innate lymphocytes. Immunity. 2010;33:736–51.
- Wang H, Li Z, Yang B, Yu S, Wu C. IL-27 suppresses the production of IL-22 in human CD4(+) T cells by inducing the expression of SOCS1. Immunol Lett. 2013:152:96–103.
- Wang X, et al. STAT5 inhibitor attenuates atherosclerosis via inhibition of inflammation: the role of STAT5 in atherosclerosis. Am J Transl Res. 2021:13:1422–31.
- Weathington NM, et al. Glycogen synthase kinase- $3\beta$  stabilizes the interleukin (IL)-22 receptor from proteasomal degradation in murine lung epithelia. J Biol Chem. 2014;289:17610–9.
- Wei CC, Ho TW, Liang WG, Chen GY, Chang MS. Cloning and characterization of mouse IL-22 binding protein. Genes Immun. 2003;4:204–11.
- Wei H, Li B, Sun A, Guo F. Interleukin-10 family cytokines immunobiology and structure. Adv Exp Med Biol. 2019;1172:79–96.
- Weidenbusch M, et al. Gene expression profiling of the Notch-AhR-IL22 axis at homeostasis and in response to tissue injury. Biosci Rep. 2017;37:BSR20170099.
- Wolk K, et al. IL-22 increases the innate immunity of tissues. Immunity. 2004;21:241–54.
- Wolk K, et al. IL-22 induces lipopolysaccharide-binding protein in hepatocytes: a potential systemic role of IL-22 in Crohn's disease. J Immunol (baltimore, Md: 1950). 2007;178:5973–81.
- Wolk K, Witte E, Witte K, Warszawska K, Sabat R. Biology of interleukin-22. Semin Immunopathol. 2010;32:17–31.
- Wu PW, et al. IL-22R, IL-10R2, and IL-22BP binding sites are topologically juxtaposed on adjacent and overlapping surfaces of IL-22. J Mol Biol. 2008;382:1168–83.
- Wu D, et al. Activation of aryl hydrocarbon receptor induces vascular inflammation and promotes atherosclerosis in apolipoprotein E-/- mice. Arterioscler Thromb Vasc Biol. 2011;31:1260–7.
- Wu QY, et al. A novel STAT3 inhibitor attenuates angiotensin II-induced abdominal aortic aneurysm progression in mice through modulating vascular inflammation and autophagy. Cell Death Dis. 2020;11:131.
- Wynn TA, Vannella KM. Macrophages in tissue repair, regeneration, and fibrosis. Immunity. 2016;44:450–62.
- Xia Q, et al. Characterisation of IL-22 and interferon-gamma-inducible chemokines in human carotid plaque. Int J Cardiol. 2012;154:187–9.
- Xie MH, et al. Interleukin (IL)-22, a novel human cytokine that signals through the interferon receptor-related proteins CRF2-4 and IL-22R. J Biol Chem. 2000;275:31335–9.

- Xu W, et al. A soluble class II cytokine receptor, IL-22RA2, is a naturally occurring IL-22 antagonist. Proc Natl Acad Sci USA. 2001;98:9511–6.
- Xu T, Logsdon NJ, Walter MR. Structure of insect-cell-derived IL-22. Acta Crystallogr D Biol Crystallogr. 2005;61:942–50.
- Xue M, et al. IL-22 suppresses the infection of porcine enteric coronaviruses and rotavirus by activating STAT3 signal pathway. Antiviral Res. 2017;142:68–75.
- Yang XP, et al. Fractalkine upregulates intercellular adhesion molecule-1 in endothelial cells through CX3CR1 and the Jak Stat5 pathway. Circ Res. 2007:101:1001–8
- Yang N, et al. Elevated interleukin-38 level associates with clinical response to atorvastatin in patients with hyperlipidemia. Cell Physiol Biochem: Int J Exp Cell Physiol, Biochem, Pharmacol. 2018;49:653–61.
- Yang FC, Chiu PY, Chen Y, Mak TW, Chen NJ. TREM-1-dependent M1 macrophage polarization restores intestinal epithelium damaged by DSSinduced colitis by activating IL-22-producing innate lymphoid cells. J Biomed Sci. 2019:26:46.
- Ye J, et al. Interleukin 22 promotes blood pressure elevation and endothelial dysfunction in angiotensin II-treated mice. J Am Heart Assoc. 2017:6:005875.
- Ye J, et al. Interleukin-22 deficiency alleviates doxorubicin-induced oxidative stress and cardiac injury via the p38 MAPK/macrophage/Fizz3 axis in mice. Redox Biol. 2020;36:101636.
- Yoshimura A, Naka T, Kubo M. SOCS proteins, cytokine signalling and immune regulation. Nat Rev Immunol. 2007;7:454–65.
- Yuan ZL, Guan YJ, Chatterjee D, Chin YE. Stat3 dimerization regulated by reversible acetylation of a single lysine residue. Science (new York, N.y). 2005;307:269–73.
- Zhang L, et al. Elevated frequencies of circulating Th22 cell in addition to Th17 cell and Th17/Th1 cell in patients with acute coronary syndrome. PLoS ONE. 2013;8:71466.
- Zhong Y, et al. Irbesartan may relieve renal injury by suppressing Th22 cells chemotaxis and infiltration in Ang Il-induced hypertension. Int Immunopharmacol. 2020:87:106789.
- Zhou L, et al. TGF-beta-induced Foxp3 inhibits T(H)17 cell differentiation by antagonizing RORgammat function. Nature. 2008;453:236–40.
- Ziesché E, Bachmann M, Kleinert H, Pfeilschifter J, Mühl H. The interleukin-22/ STAT3 pathway potentiates expression of inducible nitric-oxide synthase in human colon carcinoma cells. J Biol Chem. 2007;282:16006–15.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

# Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$  thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

#### At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

