

Research Article

Increased Circulatory Interleukin-17A Levels in Patients with Progressive and Leukotrichial Vitiligo

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Background. Vitiligo is a chronic condition characterized by skin depigmentation. Although not life-threatening, it significantly impacts quality of life. The pathophysiology of vitiligo remains poorly understood, and treatment options are limited. Mounting evidence supports the importance of autoreactive T cells and, particularly interleukin-17A- (IL-17A-) secreting Th17 cells, in vitiligo. IL-17A targeting has been proven successful in various inflammatory dermatological conditions, including psoriasis and lupus erythematosus. **Objective.** We evaluated the relationship between serum levels of IL-17A and the clinicopathological characteristics of Vietnamese vitiligo patients. **Methods.** In this cross-sectional study, we analyzed data from 52 nonsegmental vitiligo patients and 50 age- and sex-matched healthy individuals. Serum levels of IL-17A were measured using an enzyme-linked immunosorbent assay. We evaluated the correlation between IL-17A levels and clinical characteristics including leukotrichia, disease duration, vitiligo activity, and body surface area involvement. **Results.** Patients with progressive vitiligo had significantly higher IL-17A levels than patients with stable vitiligo ($P = 0.014$) or healthy individuals ($P = 0.002$). In addition, serum IL-17A levels were higher in vitiligo patients with leukotrichia than in patients without it ($P = 0.04$). Furthermore, serum IL-17A levels were negatively correlated with age ($r = -0.39$, $P = 0.004$) and age of onset ($r = -0.33$, $P = 0.016$) in vitiligo patients. **Conclusions.** Higher serum levels of IL-17A in patients with progressive vitiligo and leukotrichia suggest a potential role of IL-17A in melanocyte destruction in the epidermis and the follicular matrix.

1. Introduction

Vitiligo is characterized by the progressive destruction of pigmentary cells in the epidermis and the hair follicles, which leads to white macules that are challenging to cure. The disease has an estimated prevalence of 0.1–2% of the world's population [1–3]. Although not life-threatening, vitiligo significantly impacts patients' quality of life, particularly those with facial lesions. The precise etiology of vitiligo remains unclear although (a) biochemical [4], (b) neural [5], and (c) autoimmune [6, 7] mechanisms seem to play a role in its onset. Notably, the immune-related mechanisms underlying nonsegmental vitiligo have been comprehensively investigated.

Changes in CD4+ T cell functions and the presence of autoreactive melanocyte-specific cytotoxic T cells play important roles in vitiligo pathogenesis [8, 9]. Importantly, the role of the CD8-CXCR3-CXCL9/10-IFN γ axis in the pathogenesis of nonsegmental vitiligo is becoming increasingly evident [10, 11]; however, targeting this pathway may lead to long-term adverse effects, such as skin cancer, as it plays a vital role in maintaining the cutaneous immunity [12]. On the contrary, T helper 17 (Th17) cells exert their biological functions by producing interleukin (IL)-17, which may exacerbate autoimmune inflammation in vitiligo. IL-17 also stimulates keratinocytes to produce several chemokines, resulting in

recruitment of T cell, neutrophil, macrophage [13], and potential loss of melanocytes [14]. IL-17 is a potent stimulator of chemokine CCL20 production, which drives the migration of cytotoxic CD8+ T cells from systemic circulation into peripheral tissues [15, 16]. In mouse models, the migration of CD8+ T cells to the skin results in melanocyte loss [17, 18]. Additionally, IL-17 stimulates endothelial cells to express E- and P-selectins and the adhesion molecules ICAM-1 and VCAM-1, resulting in increased neutrophil migration [19].

Nevertheless, the role of IL-17A in vitiligo remains poorly understood. In this cross-sectional study, we evaluated the relationship between serum levels of IL-17A and the clinicopathological characteristics of Vietnamese individuals with nonsegmental vitiligo.

2. Materials and Methods

2.1. Study Subjects. We recruited 52 nonsegmental vitiligo patients (Group 1), including patients with focal acrofacial, mucosal, generalized, and universal vitiligo. Subjects who were treated with systemic or topical corticosteroids, or who received any other systemic immunosuppressive therapy in the preceding 2 months of recruitment, were excluded. Pregnant/lactating patients, or patients with a history of Hashimoto thyroiditis, Graves' disease, insulin-dependent diabetes, Addison's disease, alopecia areata, psoriasis, rheumatoid arthritis, or dysfunctional thyroid, were also excluded. We also enrolled 50 age- and sex-matched healthy individuals (Group 2) who did not have any criteria above as well as acute or chronic diseases. In Group 1, we recorded age, sex, duration of the disease, age of onset, family history, Koebner phenomenon, clinical features of the lesions, body area involvement (in quartile percentiles), and disease activity by using the vitiligo disease activity (VIDA) score [20]. The study was approved by IRB of University of Medicine and Pharmacy at Ho Chi Minh City (IRB number: 424/DHYD-HDDD).

2.2. Measurement of Serum IL-17A Levels by ELISA. Serum samples were collected from study subjects. Serum IL-17A levels were measured using an ELISA with a Human IL-17A ELISA Kit (ANOGEN Inc, Ontario, Canada) following the manufacturer's instructions.

2.3. Statistical Analysis. Data are expressed as mean \pm standard deviation (SD), median and range, or prevalence. Mean values were compared using Mann-Whitney U (for two groups) or the Kruskal-Wallis test (for three or more groups). Spearman's rank correlation coefficient was used to investigate the relationship between clinical parameters. *P* values less than 0.05 were considered statistically significant. Statistical analyses were performed using R packages (RStudio Desktop, version 1.2 for Windows, RStudio Inc., Boston, USA).

3. Results

3.1. Clinical Characteristics of the Study Subjects. The clinical characteristics of the study subjects are shown in Table 1. Group 1 included 18 males (34.62%) and 34 females (65.38%), with a mean age of 32.71 ± 14.92 (range, 6–61 years). Group 2 included 18 males (36%) and 32 females (64%).

3.2. Serum Interleukin-17A Level in Patients and Controls. The serum IL-17A levels of Group 1 (median 3.09 (2.16–15.03) pg/mL) were significantly higher than those of Group 2 (median 2.66 (1.86–4.80) pg/mL) ($P=0.018$). In addition, serum IL-17A levels of patients with progressive vitiligo (median 4.35 (2.78–32.26) pg/mL) were significantly higher than those with stable vitiligo (median 2.26 (1.94–6.32) pg/mL, $P=0.014$) as well as healthy controls (median 2.66 (1.85–4.79) pg/mL, $P=0.002$). However, there were no significant differences in serum IL-17A levels between the patients with stable disease and healthy controls (Figure 1).

3.3. Association of Serum IL-17A Levels with Clinical Characteristics. Vitiligo patients with leukotrichia had significantly higher serum levels of IL-17A than those without leukotrichia (8.27 (2.78–122.62) pg/mL vs 2.99 (2.02–10.38) pg/mL, $P<0.05$). Nevertheless, serum IL-17A levels were not associated with sex, family history of vitiligo, affected body surface area, or lesional characteristics (including trichrome, confetti-like, perifollicular hyperpigmentation, Koebner phenomenon, and nevus halo) (Table 2).

In the subgroup analysis of active vitiligo, we also could not find the association between affected body surface area and the concentration of IL-17A ($r=-0.15$, $P=0.38$). Furthermore, serum IL-17A levels were negatively correlated with age ($r=-0.39$, $P=0.004$) and age of onset ($r=-0.33$, $P=0.016$) in vitiligo patients but not in healthy controls (Table 3 and Figure 2).

4. Discussion

The pathophysiology of vitiligo remains understudied. Changes in humoral and cellular immunity are considered important causes of melanocyte destruction and subsequent vitiligo. Consistently, vitiligo is associated with autoimmune endocrinopathies [2]. Furthermore, the presence of autoantibodies against melanocyte antigens, including tyrosinase and tyrosinase-related protein (TYRP) 1 and 2, in the sera of vitiligo patients suggests an important role of humoral immunity in vitiligo pathophysiology [21]. In addition, high numbers of T-cell in the lesional margins in inflammatory vitiligo indicate the role of cellular immunity in vitiligo [22]. Besides the Th1 response and subsequent TNF- α and IFN- γ production, Th17 cells and IL-17A have recently been implicated in the pathogenesis of vitiligo. IL-17A targets multiple cell types, including fibroblasts, endothelial cells, epithelial cells, keratinocytes, and macrophages. Kotobuki et al. [23] investigated the biological effects of IL-17A and

TABLE 1: Clinical characteristics of study subjects.

	Group 1 (n = 52)	Group 2 (n = 50)	P value
Age (mean ± SD)	32.71 ± 14.92	36.34 ± 10.34	0.22
Male, n (%)	18 (34.62%)	18(36%)	0.88
Age of onset, n (%)			
≤30 years	29 (55.77%)	NA	NA
>30 years	23 (44.23%)		
Duration of disease, n (%)			
<5 years	40 (76.92%)	NA	NA
5–10 years	8 (15.39%)		
>10 years	4 (7.69%)		
Family history of vitiligo, n (%)	11 (21.15%)	NA	NA
Affected body surface area, n (%)			
<3%	32 (61.54%)	NA	NA
3–10%	13 (25.00%)		
>10%	7 (13.46%)		
Triggers, n (%)			
Trauma	11 (21.15%)	NA	NA
Psychological stress	19 (36.54%)		
Sunburn	6 (11.54%)		
Pregnancy	6 (11.54%)		
Vitiligo activity, n (%)		NA	NA
Progressive			
VIDA +4	23 (44.23%)		
VIDA +3	12 (23.08%)		
Stable			
VIDA +2	7 (13.46%)		
VIDA +1	8 (15.39%)		
VIDA 0	0 (0.00%)		
VIDA -1	2 (3.84%)		
Clinical variants, n (%)		NA	NA
Focal	11 (21.15%)		
Acro/acrofacial	4 (7.69%)		
Mucosa	1 (1.93%)		
Generalized	36 (69.23%)		
Universal	0 (0.00%)		
Lesion characteristics, n (%)		NA	NA
Trichrome	31 (59.62%)		
Confetti-like	10 (19.23%)		
Perifollicular hyperpigmentation	22 (42.31%)		
Leukotrichia	14 (26.92%)		
Koebner phenomenon	12 (23.08%)		
Nevus halo	2 (3.85%)		

NA, not available.

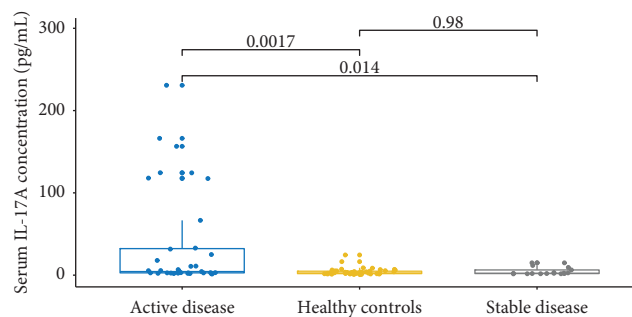


FIGURE 1: Serum levels of IL-17A in active and stable disease and healthy controls.

TABLE 2: The association of serum IL-17A levels with clinical characteristics of vitiligo patients.

Clinical characteristics	N	Serum IL-17 level (pg/mL)	P value
Gender			
Females	34	3.12 (2.16–15.03)	0.81
Males	18	2.78 (2.11–10.08)	
Ages			
≤30 years	21	10.68 (3.09–32.91)	0.003
>30 years	31	2.78 (1.96–5.34)	
Vitiligo activity			
Stable	17	2.26 (1.94–6.32)	0.013
Progressive	35	4.35 (2.78–32.26)	
Family history of vitiligo			
Yes	11	2.26 (1.96–49.10)	0.69
No	41	3.09 (2.47–11.86)	
Leukotrichia			
Yes	14	8.27 (2.78–122.62)	0.04
No	38	2.99 (2.02–10.38)	
Affected body surface area			
<3%	32	3.08 (2.24–12.02)	0.82
3–10%	13	4.35 (2.42–11.86)	
>10%	7	2.78 (1.78–16.00)	

P values were obtained by the Mann–Whitney U test. Data were shown as median (25th–75th).

TABLE 3: The correlations of serum IL-17A levels with ages and age of onset in the study groups.

	Group 1 (n = 52)	Group 2 (n = 50)
Age	$r = -0.39, P = 0.004$	$r = 0.27, P = 0.06$
Age of onset	$r = -0.33, P = 0.016$	NA

NA, not available.

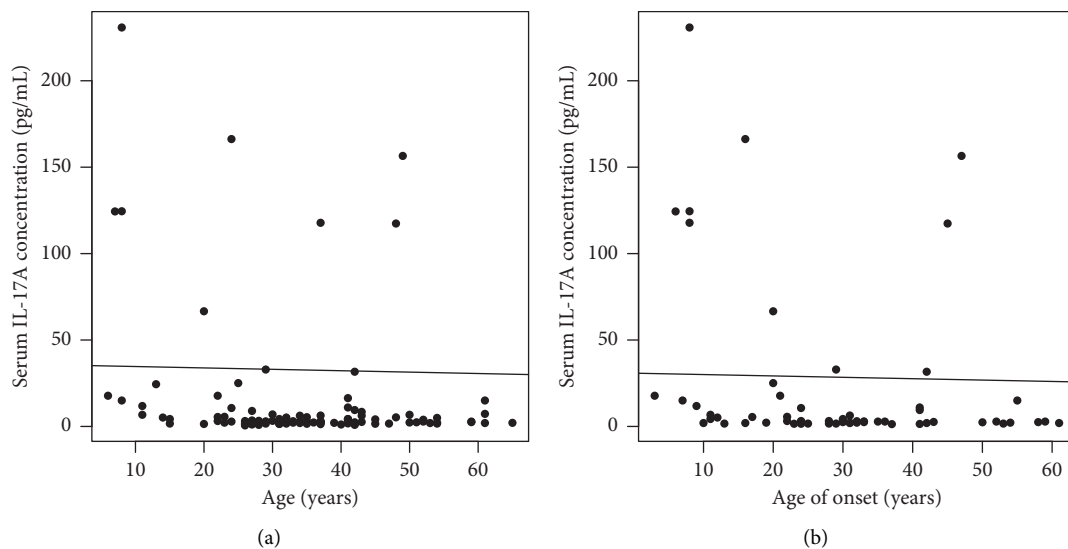


FIGURE 2: The correlations of serum IL-17A levels with age (a) and age of onset (b) in vitiligo patients.

found that expression of MITF, an important transcriptional regulator of melanogenesis, and its downstream genes was reduced by more than 10% in melanocytes treated with IL-17. In addition, melanin production from IL-17-treated melanocytes decreased by approximately 30%, accompanied by profound morphological changes. In addition, IL-17 synergizes with various inflammatory mediators, including

IL-1 α , IL-6, and TNF- α [24], to inhibit melanocyte proliferation [25].

In this study, we found that serum IL-17A level was significantly increased in patients with nonsegmental vitiligo compared to healthy controls, confirming the findings of studies [26–30]. We also found that serum IL-17A levels were negatively correlated with age in patients with vitiligo

but not in healthy controls. Previously, Kosar Hedayat et al. [31] demonstrated that vitiligo patients under 30 years had a lower quality of life than those aged above 30 years. Interestingly, mental stress is a trigger for vitiligo in 50–65% of vitiligo patients [32, 33], likely due to aggravation of innate and adaptive immunity [34]. Additionally, chronic stress polarizes immune response towards Th17 rather than Th2 response [35, 36]. Consequently, we hypothesized that the increased levels in younger vitiligo patients may be due to mental stress. However, further investigations are required to elucidate the relationship between IL-17A levels and age.

We also found a negative correlation between IL-17A levels and age of onset in vitiligo patients. This finding is in agreement with the findings of Basak et al. [25] but contradict those of Zhou et al. [37]. This suggests that IL-17A may contribute to the immune response in early-onset vitiligo. In addition, Arcos-Burgos et al. [38] demonstrated an association of early onset vitiligo with HLA-DR4, which is involved in CD4+ T cell activation and subsequent production of proinflammatory cytokines, including IL-17 and IFN- γ [39]. More studies are needed to understand the role of IL-17A in early onset vitiligo.

A correlation between serum IL-17A levels and body surface involvement has been reported in previous studies [25, 26, 29]. However, we found no significant association between these two factors, consistent with Tembhe et al. [28] and Zhou et al. [37]. Consistent with previous findings [28, 37, 40, 41], we found that IL-17A levels were correlated with progressive vitiligo, suggesting that IL-17A might be involved in vitiligo induction and progression. Zhen et al. [41] reported that while serum IL-17A levels were significant higher in the patients with nonsegmental vitiligo, there were no significant differences in serum levels of IFN- γ , IL-4, and TGF- β 1. Furthermore, in this study, the numbers of peripheral CD4+IL-17A+ Th17 cells were significantly higher in vitiligo patients compared to healthy volunteers; however, no differences were observed in the numbers of peripheral CD4+IL-4+ Th2 or regulatory T cells. These findings suggest that IL-17 may be crucial in the onset and progression of autoimmune nonsegmental vitiligo. However, regarding the biomarkers of disease activity, circulating cytokines (IL-1 β , IL-17, IFN- γ , and TGF- β), autoantibodies, oxidative stress markers, soluble CDs (sCD25 and sCD27), and chemokines (CXCL9 and CXCL10) remain competing [42].

Follicular melanocytes serve as a melanocyte reservoir, facilitating repigmentation in vitiligo. The presence of leukotrichia in white patches may be a sign of refractory vitiligo [43], and untreated leukotrichia could severely impact the quality of life [44]. The understanding of follicular vitiligo pathogenesis and the relationship between IL-17A and leukotrichia, in particular, is limited. In the present study, we found that nonsegmental vitiligo patients with leukotrichia had significantly higher serum levels of IL-17A than patients without leukotrichia. Gan et al. [45] reported the presence of perifollicular lymphocytes in the infundibular region of the hair follicles, while melanocytes were absent in the basal layer of the epidermis and the hair follicle. These findings suggest that both CD8+ and CD4+ T cells could attack hair follicle melanocytes [46]. In addition, Rongioletti et al. [47]

reported a repigmentation of follicular melanocytes in psoriasis patients after treatment with secukinumab, supporting the role of IL-17A in the pathogenesis of follicular melanocyte destruction. Further studies are warranted to understand the role of IL-17 in the pathogenesis of vitiligo.

This study had some limitations that need to be addressed. First of all, the concentration of IL-17A was only measured in the serum, but not in the skin or follicles of the vitiligo patients. Thus, further researches implemented at the lesional sites should be conducted. Secondly, the size sample of our study was modest partly due to the uncommon characteristic of the vitiligo. Finally, this was a cross-sectional study in which a causal relationship could not be established.

5. Conclusions

In conclusion, this study provides evidence of that IL-17A may play a crucial role in the induction of vitiligo along with other factors, namely, the genetics, environment, and inflammatory cytokines. In addition, we found that serum levels of IL-17A were significantly associated with leukotrichia, suggesting the role of IL-17A in the destruction of follicular melanocytes.

Data Availability

Due to privacy and ethical concerns, neither the data nor the source of the data can be made available.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

References

- [1] A. Alikhan, L. M. Felsten, M. Daly, and V. Petronic-Rosic, "Vitiligo: a comprehensive overview," *Journal of the American Academy of Dermatology*, vol. 65, no. 3, pp. 473–491, 2011.
- [2] A. Alkhateeb, P. R. Fain, A. Thody, D. C. Bennett, and R. A. Spritz, "Epidemiology of vitiligo and associated autoimmune diseases in Caucasian probands and their families," *Pigment Cell Research*, vol. 16, no. 3, pp. 208–214, 2003.
- [3] C. Krüger and K. U. Schallreuter, "A review of the worldwide prevalence of vitiligo in children/adolescents and adults," *International Journal of Dermatology*, vol. 51, no. 10, pp. 1206–1212, 2012.
- [4] R. E. Boissy and P. Manga, "On the etiology of contact/occupational vitiligo," *Pigment Cell Research*, vol. 17, no. 3, pp. 208–214, 2004.
- [5] M. L. Cucchi, P. Frattini, G. Santagostino, and G. Orecchia, "Higher plasma catecholamine and metabolite levels in the early phase of nonsegmental vitiligo," *Pigment Cell Research*, vol. 13, no. 1, pp. 28–32, 2000.
- [6] S. Garbelli, S. Mantovani, B. Palermo, and C. Giachino, "Melanocyte-specific, cytotoxic T cell responses in vitiligo: the effective variant of melanoma immunity?" *Pigment Cell Research*, vol. 18, no. 4, pp. 234–242, 2005.
- [7] K. Ongenaes, N. Van Geel, and J.-M. Naeyaert, "Evidence for an autoimmune pathogenesis of vitiligo," *Pigment Cell Research*, vol. 16, no. 2, pp. 90–100, 2003.

- [8] R. M. Halder, C. S. Walters, B. A. Johnson, S. G. Chakrabarti, and J. A. Kenney Jr., "Aberrations in T lymphocytes and natural killer cells in vitiligo: a flow cytometric study," *Journal of the American Academy of Dermatology*, vol. 14, no. 5, pp. 733–737, 1986.
- [9] D. P. Antelo, A. L. Filgueira, and J. M. T. Cunha, "Reduction of skin-homing cytotoxic T cells (CD8+·CLA+) in patients with vitiligo," *Photodermatology, Photoimmunology & Photomedicine*, vol. 27, no. 1, pp. 40–44, 2011.
- [10] M. L. Frisoli and J. E. Harris, "Vitiligo: mechanistic insights lead to novel treatments," *Journal of Allergy and Clinical Immunology*, vol. 140, no. 3, pp. 654–662, 2017.
- [11] M. Rashighi and J. E. Harris, "Interfering with the IFN- γ /CXCL10 pathway to develop new targeted treatments for vitiligo," *Annals of Translational Medicine*, vol. 3, no. 21, p. 343, 2015.
- [12] A. B. Blechman, C. E. Cabell, C. H. Weinberger et al., "Aggressive skin cancers occurring in patients treated with the janus kinase inhibitor ruxolitinib," *Journal of Drugs in Dermatology: JDD*, vol. 16, no. 5, pp. 508–511, 2017.
- [13] J.-M. Schröder, H. Gregory, J. Young, and E. Christophers, "Neutrophil-activating proteins in psoriasis," *Journal of Investigative Dermatology*, vol. 98, no. 2, pp. 241–247, 1992.
- [14] I. C. Le Poole, R. M. Van Den Wijngaard, W. Westerhof, and P. K. Das, "Presence of T cells and macrophages in inflammatory vitiligo skin parallels melanocyte disappearance," *Annals of Translational Medicine*, vol. 148, pp. 1219–1228, 1996.
- [15] S. P. Singh, H. H. Zhang, J. F. Foley, M. N. Hedrick, and J. M. Farber, "Human T cells that are able to produce IL-17 express the chemokine receptor CCR6," *The Journal of Immunology*, vol. 180, no. 1, pp. 214–221, 2008.
- [16] N. Martin-Orozco, P. Muranski, Y. Chung et al., "T helper 17 cells promote cytotoxic T cell activation in tumor immunity," *Immunity*, vol. 31, no. 5, pp. 787–798, 2009.
- [17] J. G. Van Den Boorn, D. Konijnenberg, T. A. M. Dellemijn et al., "Autoimmune destruction of skin melanocytes by perilesional T cells from vitiligo patients," *Journal of Investigative Dermatology*, vol. 129, no. 9, pp. 2220–2232, 2009.
- [18] P. Muranski, A. Boni, P. A. Antony et al., "Tumor-specific Th17-polarized cells eradicate large established melanoma," *Blood*, vol. 112, no. 2, pp. 362–373, 2008.
- [19] E. Suzuki, E. D. Mellins, M. E. Gershwin, F. O. Nestle, and I. E. Adamopoulos, "The IL-23/IL-17 axis in psoriatic arthritis," *Autoimmunity Reviews*, vol. 13, no. 4-5, pp. 496–502, 2014.
- [20] F. Aydin, N. Sentürk, B. Şahin, Y. Bek, E. Pancar Yuksel, and A. Turanlı, "A practical method for the estimation of vitiligo surface area: a comparison between the point counting and digital planimetry techniques," *European Journal of Dermatology: EJD*, vol. 17, pp. 30–32, 2007.
- [21] T. Okamoto, S. Fujii, S. K. S. Huang et al., "Anti-tyrosinase-related protein-2 immune response in vitiligo patients and melanoma patients receiving active-specific immunotherapy," *Journal of Investigative Dermatology*, vol. 111, no. 6, pp. 1034–1039, 1998.
- [22] A. Wankowicz-Kalinska, R. M. Van Den Wijngaard, B. J. Tigges et al., "Immunopolarization of CD4+ and CD8+ T cells to Type-1-like is associated with melanocyte loss in human vitiligo," *Lab Invest*, vol. 83, pp. 683–695, 2003.
- [23] Y. Kotobuki, A. Tanemura, L. Yang et al., "Dysregulation of melanocyte function by Th17-related cytokines: significance of Th17 cell infiltration in autoimmune vitiligo vulgaris," *Pigment Cell & Melanoma Research*, vol. 25, no. 2, pp. 219–230, 2012.
- [24] V. B. Swope, Z. Abdel-Malek, L. M. Kassem, and J. J. Nordlund, "Interleukins 1 α and 6 and tumor necrosis factor- α are paracrine inhibitors of human melanocyte proliferation and melanogenesis," *Journal of Investigative Dermatology*, vol. 96, no. 2, pp. 180–185, 1991.
- [25] P. Y. Basak, A. K. Adiloglu, A. M. Ceyhan, T. Tas, and V. B. Akkaya, "The role of helper and regulatory T cells in the pathogenesis of vitiligo," *Journal of the American Academy of Dermatology*, vol. 60, no. 2, pp. 256–260, 2009.
- [26] D. A. Bassiouny and O. Shaker, "Role of interleukin-17 in the pathogenesis of vitiligo," *Clinical and Experimental Dermatology*, vol. 36, no. 3, pp. 292–297, 2011.
- [27] R. Khan, S. Gupta, and A. Sharma, "Circulatory levels of T-cell cytokines (interleukin [IL]-2, IL-4, IL-17, and transforming growth factor-beta) in patients with vitiligo," *Journal of the American Academy of Dermatology*, vol. 66, no. 3, pp. 510–511, 2012.
- [28] M. K. Tembhre, V. K. Sharma, A. Sharma, P. Chattopadhyay, and S. Gupta, "T helper and regulatory T cell cytokine profile in active, stable and narrow band ultraviolet B treated generalized vitiligo," *Clinica Chimica Acta*, vol. 424, pp. 27–32, 2013.
- [29] M. A. Elela, R. A. Hegazy, M. M. Fawzy, L. A. Rashed, and H. Rasheed, "Interleukin 17, interleukin 22 and FoxP3 expression in tissue and serum of non-segmental vitiligo: a case-controlled study on eighty-four patients," *European Journal of Dermatology*, vol. 23, no. 3, pp. 350–355, 2013.
- [30] P. Acharya and M. Mathur, "Interleukin-17 level in patients with vitiligo: a systematic review and meta-analysis," *Australasian Journal of Dermatology*, vol. 61, no. 2, pp. e208–e212, 2020.
- [31] K. Hedayat, M. Karbakhsh, M. Ghiasi et al., "Quality of life in patients with vitiligo: a cross-sectional study based on Vitiligo Quality of Life index (VitiQoL)," *Health and Quality of Life Outcomes*, vol. 14, p. 86, 2016.
- [32] A. Firooz, N. Bouzari, N. Fallah, B. Ghazisaidi, M. R. Firoozabadi, and Y. Dowlati, "What patients with vitiligo believe about their condition," *International Journal of Dermatology*, vol. 43, no. 11, pp. 811–814, 2004.
- [33] L. Manolache and V. Benea, "Stress in patients with alopecia areata and vitiligo," *Journal of the European Academy of Dermatology and Venereology*, vol. 21, no. 7, pp. 921–928, 2007.
- [34] I. Harpaz, S. Abutbul, A. Nemirovsky, R. Gal, H. Cohen, and A. Monsonego, "Chronic exposure to stress predisposes to higher autoimmune susceptibility in C57BL/6 mice: glucocorticoids as a double-edged sword," *European Journal of Immunology*, vol. 43, no. 3, pp. 758–769, 2013.
- [35] T. J. D'zurilla, A. Maydeu-Olivares, and G. L. Kant, "Age and gender differences in social problem-solving ability," *Personality and Individual Differences*, vol. 25, no. 2, pp. 241–252, 1998.
- [36] E. Hamarat, D. Thompson, K. M. Zabrocky, D. Steele, K. B. Matheny, and F. Aysan, "Perceived stress and coping resource availability as predictors of life satisfaction in young, middle-aged, and older adults," *Pigment Cell & Melanoma Research*, vol. 27, pp. 181–196, 2001.
- [37] L. Zhou, Y.-L. Shi, K. Li et al., "Increased circulating Th17 cells and elevated serum levels of TGF-beta and IL-21 are correlated with human non-segmental vitiligo development," *Pigment Cell & Melanoma Research*, vol. 28, no. 3, pp. 324–329, 2015.

- [38] M. Arcos-Burgos, E. Parodi, M. Salgar et al., "Vitiligo: complex segregation and linkage disequilibrium analyses with respect to microsatellite loci spanning the HLA," *Human Genetics*, vol. 110, no. 4, pp. 334–342, 2002.
- [39] A. K. Mangalam, V. Taneja, and C. S. David, "HLA class II molecules influence susceptibility versus protection in inflammatory diseases by determining the cytokine profile," *Journal of Immunology*, vol. 190, pp. 513–518, 2013.
- [40] S. Bhardwaj, S. Rani, N. Srivastava, R. Kumar, and D. Parsad, "Increased systemic and epidermal levels of IL-17A and IL-1 β promotes progression of non-segmental vitiligo," *Cytokine*, vol. 91, pp. 153–161, 2017.
- [41] Y. Zhen, L. Yao, S. Zhong, Y. Song, Y. Cui, and S. Li, "Enhanced Th1 and Th17 responses in peripheral blood in active non-segmental vitiligo," *Archives of Dermatological Research*, vol. 308, no. 10, pp. 703–710, 2016.
- [42] R. Speeckaert, M. Speeckaert, S. De Schepper, and N. Van Geel, "Biomarkers of disease activity in vitiligo: a systematic review," *Autoimmunity Reviews*, vol. 16, no. 9, pp. 937–945, 2017.
- [43] C.-Y. Kim, T.-J. Yoon, and T.-H. Kim, "Epidermal grafting after chemical epilation in the treatment of vitiligo," *Dermatologic Surgery*, vol. 27, no. 10, pp. 855–856, 2001.
- [44] G. Agarwal, "Vitiligo: an under-estimated problem," *Experimental Dermatology*, vol. 15, no. 1, pp. S19–S23, 1998.
- [45] E. Y. Gan, M. Cario-André, C. Pain et al., "Follicular vitiligo: a report of 8 cases," *Journal of the American Academy of Dermatology*, vol. 74, no. 6, pp. 1178–1184, 2016.
- [46] J. E. Harris, "Vitiligo and alopecia areata: apples and oranges?" *Experimental Dermatology*, vol. 22, no. 12, pp. 785–789, 2013.
- [47] F. Rongioletti, C. Mugheddu, and S. Murgia, "Repigmentation and new growth of hairs after anti-interleukin-17 therapy with secukinumab for psoriasis," *JAAD Case Reports*, vol. 4, no. 5, pp. 486–488, 2018.