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Establishing a Framework for the Clinical Translation of Germline Findings in Precision Oncology

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Abstract

Inherited genetic variation has important implications for cancer screening, early diagnosis, and disease prognosis. A role for germline variation has also been described in shaping the molecular landscape, immune response, microenvironment, and treatment response of individual tumors. However, there is a lack of consensus on the handling and analysis of germline information that extends beyond known or suspected cancer susceptibility in large-scale cancer genomics initiatives. As part of the Personalized OncoGenomics program in British Columbia, we performed whole-genome and transcriptome sequencing in paired tumor and normal tissues from advanced cancer patients to characterize the molecular tumor landscape and identify putative targets for therapy. Overall, our experience supports a multidisciplinary and integrative approach to germline data management. This includes a need for broader definitions and standardized recommendations regarding primary and secondary germline findings in precision oncology. Here, we propose a framework for identifying, evaluating, and returning germline variants of potential clinical significance that may have indications for health management beyond cancer risk reduction or prevention in patients and their families.

Characterizing hereditary genetic variation in high- and moderate-penetrance cancer predisposition genes may have implications for cascade carrier testing, cancer risk-reduction, and screening interventions in individuals at risk of having inherited a causal germline variant. Individual variability in disease prognosis and treatment response also occurs within

the context of heterogeneous genetic backgrounds, including rare variants associated with cancer susceptibility and common polymorphisms in other cancer-related genes (1–4). However, standards for the clinical translation of genetic information relevant to cancer susceptibility, pathogenesis, prognosis, and treatment, as well as secondary or incidental

genetic information unrelated to cancer, are inconsistent across cancer genomics programs (5). This may result from varying regional policies regarding the return of research results, clinician preference, or genetic literacy.

The Personalized OncoGenomics (POG) program is a precision medicine initiative in British Columbia (BC), Canada, that was established to identify clinically actionable molecular events in adult metastatic cancer patients and pediatric patients with poor prognosis cancers (NCT02155621). Wholegenome sequencing (WGS) and RNA sequencing of fresh, frozen tumor biopsies performed with WGS of paired normal tissues has helped identify somatic alterations and inherited genetic variants that shape tumor progression (6-9). This and similar projects, such as the Cancer Genome Atlas and International Cancer Genome Consortium, have provided important resources for understanding the morbid human genome (10, 11). At the same time, the rate of data production has far outpaced the development of evidence-based guidelines for managing germline findings. Here, we advocate for a broader understanding of what defines primary germline findings in oncology and propose a framework for identifying, evaluating, and reporting research germline findings within the clinical infrastructure of a publicly funded provincial health authority.

Early Years of the POG Program

Since its establishment in 2012, the POG program has continued to develop effective, collaborative, and transparent data management and reporting strategies (6). These have allowed the accommodation of advancements in technology, computational pipelines, therapeutic developments, and biological and clinical knowledge. Before standard recommendations for variant interpretation were published by the American College of Medical Genetics and Genomics (ACMG) and Association for Molecular Pathology in 2015, germline data were assessed on an ad hoc basis to identify high-penetrance variants relevant to inherited cancer susceptibility or treatment (12). Any research findings were discussed with treating oncologists as part of the POG tumor board, but this approach was not broadly or consistently actionable in a maturing program. This was complicated by conflicting interpretations of variant pathogenicity and/or differing opinions regarding the return of results, indicating the need for a standardized procedure for germline assessment. Given the ethical challenges of germline data analysis within the oncology setting, the POG Ethics and Germline Working Group was created in 2014. Using autonomy, beneficence, nonmaleficence, and justice as guiding principles, the mandate of the Ethics and Germline Working Group includes addressing ongoing issues related to the management and clinical translation of germline findings. The group meets monthly and consists of a multidisciplinary team of oncologists, medical and molecular geneticists, pathologists, bioinformaticians and other scientists, an ethicist, and lawyers.

Transparency and Standardization of Informed Consent

This POG program is approved by the University of British Columbia Research Ethics Committee, and written informed consent or assent is obtained for all patients involved in this research. The Ethics and Germline Working Group reached a consensus that informed consent or assent should include by

default the reporting of any clinically actionable cancer-related germline genetic information for both adults and children with cancer. This mandate keeps with the primary goal of the POG program to characterize the complete genomic architecture of an individual cancer while maintaining data transparency and allowing opportunities for clinical translation. Germline findings of clinical significance related to cancer risk are currently returned to the research participant, designee, or next of kin by the patient's oncologist, and a referral is made to the provincial BC Cancer Hereditary Cancer Program (HCP) for genetic counseling and clinical variant confirmation. For both tumor-only and tumor-normal sequencing, an appropriate consent procedure includes a detailed review of family history and pretest counseling regarding the potential risks and benefits of germline findings. This is critical for ensuring ethical justification of studies involving the analysis of hereditary genetic variation.

An opt-in procedure for the return of germline findings of clinical significance unrelated to cancer has been adopted by the POG program, consistent with recommendations based on patient and public preference studies (13). Although these findings are not routinely sought in cancer genomics analyses, 97.9% of participants enrolled in the program between 2012 and July 2019 (n = 993) opted for the return of incidental germline findings unrelated to cancer. These observations suggest that the level of potential risk in learning about inherited genetic variation, including risks related to cancer susceptibility, disease carrier status, or paternity, is acceptable to patients and has not resulted in a decline in participation in the program. The potential for learning about genetic cancer predisposition in particular should be an essential part of the patients' education and communicated to them at the time of diagnosis.

Germline Variant Curation, Validation, and Return

Pathogenic and likely pathogenic germline variants in moderate- to high-penetrance cancer predisposition genes underlie 5%-10% of all cancers, with a prevalence of up to 20% in certain cancer types (14, 15). In collaboration with HCP and the Cancer Genetics and Genomics Laboratory (CGL), the Ethics and Germline Working Group developed standard procedures for germline variant prioritization and evaluation (Figures 1 and 2). Genome-wide variant calling is performed in parallel pipelines for small variants, including single nucleotide variants and small insertions and deletions, and structural variants (SVs), including copy number variants and balanced genomic rearrangements (Figure 1). SV calling through short-read-based nextgeneration sequencing in particular has inherent technical and computational challenges because of low-complexity sequences in the human reference genome. Therefore, using multiple computational tools that employ complementary variant calling methods is preferred to improve the sensitivity of SV detection. Following gene- and function-based filtering, candidate germline variants are manually reviewed in a genome browser to flag putative technical or sequence artifacts prior to clinical review.

To allow rapid return of results for variants with established clinical actionability, a tiered list of cancer susceptibility genes is analyzed according to the following guidelines. Known pathogenic and likely pathogenic variants in curated variant databases, such as ClinVar and local CGL database, and novel predicted loss of function variants (eg, deletion, frameshift,

Figure 1. Approach for germline variant calling, annotation, and filtering in tumor-normal whole-genome sequencing. In the Personalized OncoGenomics program, parallel pipelines are implemented for the analysis of small variants, including single nucleotide variants (SNVs) and small insertions and deletions (indels), and structural variants (SVs). Low-complexity regions, strong GC bias, and repetitive elements limit the accuracy of SV calling through short-read (50-300 bp) sequencing. Consequently, complementary read depth-, flanking read-, split read-, and contig-based computational approaches are incorporated to increase the sensitivity of SV detection. COSMIC = Catalog of Somatic Mutations in Cancer; MAVIS = Merging, Annotation, Validation, and Illustration of Structural Variants.

Germline variants with known or putative clinical significance are prioritized by clinical annotation, functional effect prediction, and population frequency in 98 cancer predisposition genes. All candidate variants are reviewed in a genome browser to flag possible technical artifacts, and this information is included in an integrated germline report along with relevant tumor data.

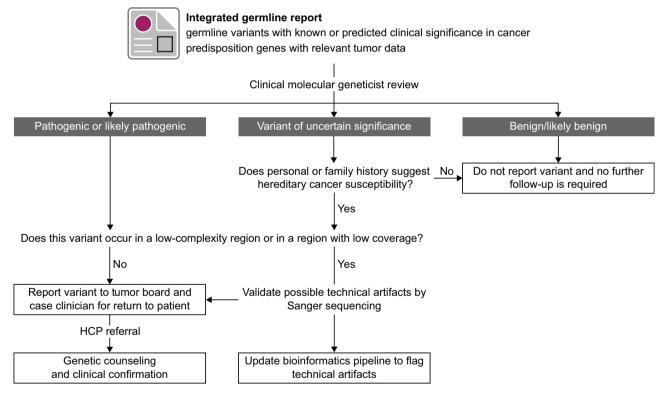


Figure 2. Standard procedure for the review, reporting, and clinical translation of germline variants in the Personalized OncoGenomics program. Given an integrated germline analysis report, a clinical molecular geneticist at the Cancer Genetics and Genomics Laboratory (CGL) curates all germline variants with known or potential clinical significance in cancer predisposition genes undergoing prioritized review. The Ethics and Germline Working Group determine, by consensus, final recommendations for variant reporting and whether a referral to the Hereditary Cancer Program (HCP) should be made for patient counseling and clinical genetic testing. In the absence of functional evidence supporting pathogenicity, variants of uncertain significance are disclosed only when the patient's personal or family history is suggestive of hereditary cancer susceptibility. Clinically actionable variants that occur in areas of the reference genome flagged as low-complexity or repetitive regions will be validated at CGL prior to return of results. If false-positive variants are identified, an updated bioinformatics pipeline is implemented to flag these variants in future cases.

nonsense, canonical splice site, and stop loss) are prioritized for curation in known cancer predisposition genes (Supplementary Table 1, available online). For several genes associated with

high-penetrance syndromes, variants of uncertain significance (VUS) and rare variants in coding and splice regions are further prioritized for review. Variant classification is performed by a

clinical molecular geneticist from CGL and certified by the Canadian College of Medical Genetics and Genomics or ACMG according to updated guidelines from the ACMG, Association for Molecular Pathology, and Clinical Genome Resource. This approach integrates clinical expertise and laboratory protocols in variant classification and confirmatory genetic testing, respectively, allowing seamless translation for patients referred to HCP (Figure 2). Final recommendations for return of information and referral for genetic counseling and clinical testing are made based on consensus among the Ethics and Germline Working Group. In exceptional cases, germline variants may be subject to expedited review by a core expert panel, including a clinical molecular geneticist, medical geneticist, and oncologist, to allow immediate return of information and clinical referral (Supplementary Table 2, available online). Clinical genetics expertise and group consultation are thus integral parts of germline assessment in the POG program to ensure reliability, consistency, and transparency.

Evolving variant curation guidelines indicate the need for dynamic computational pipelines that integrate updated variant information from population and clinical databases and encourage efficient retrospective analysis (16, 17). To aid in largescale variant identification and classification, special consideration should also be given to variants with founder effects, specific modes of disease inheritance, and variants with low to moderate cancer risk. Founder mutations in common cancer predisposition genes such as BRCA1, BRCA2, CHEK2, and MUTYH should be excluded from global allele frequency thresholds used in automated variant filtering. For example, more than 1% of patients in the POG program are carriers for pathogenic MUTYH variants, reflecting a strong representation of individuals of East and Southeast Asian descent in BC (9, 18). In such cases, we recommend developing highly curated internal databases with known pathogenic and likely pathogenic germline variants to reduce the incidence of false-negative findings. Furthermore, variants in Mendelian disease genes underlying cancer predisposition syndromes inherited in autosomal recessive or X-linked recessive patterns, such as MUTYH-associated polyposis and dyskeratosis congenita, respectively, also require exceptional committee review. The position of the Ethics and Germline Working Group is to evaluate the risks and benefits of returning carrier status for autosomal recessive cancer susceptibility genes on a gene- and case-specific basis. Finally, identification of low- to moderate-penetrance cancer predisposition variants may present an additional challenge because of limited evidence-based guidelines regarding effective clinical management and cancer risk reduction strategies. Personal and family medical history should be carefully considered in these cases to determine the appropriateness of variant disclosure for cancer susceptibility.

For individuals with phenotypic indications of highpenetrance cancer predisposition syndromes and uninformative clinical genetic testing, evaluation of tumor WGS and RNA sequencing may improve genetic diagnosis through the resolution of potential splicing variants, noncoding variants in regulatory regions, or structural variants (9, 19). Tumor data may also inform possible roles for VUS or autosomal recessive gene variants in pathogenesis, recently demonstrated in a patient with biallelic variants in MUTYH (p.Gly286Glu and p.Ser346Ser) with tumor evidence supporting global deficiency in base excision repair (7–9). The specific types of molecular data that could be considered in whole-genome and transcriptome studies include genome-wide mutation burden, simple somatic mutations and

mutational signatures, copy number alterations, loss of heterozygosity, structural variants and structural variant signatures, expression outliers, expression-based classifications, and alternative splicing (Supplementary Table 3, available online). However, recommendations for incorporating tumor data into variant classification are still evolving, and recent guidelines recommend cautious variant interpretation based on this evidence alone without corresponding data from in vitro or in vivo functional studies (20). In the POG program, VUS are not reclassified on the basis of tumor data alone without ClinVar classifications or published phenotypic and functional evidence supporting pathogenicity. For patients with relevant personal or family history, VUS may be returned to the treating clinician with a recommendation for referral to HCP if the individual is eligible for publicly funded index genetic testing. The Ethics and Germline Working Group does not support clinical decision making based on the presence of VUS alone and stresses the importance of consultation with core members of a clinical genetics team to ensure appropriate clinical follow-up when indicated.

Implications for Germline Genetic Variation Beyond Cancer Susceptibility

Both germline variation and somatic alterations have potential clinical significance in precision oncology, and integrated analysis of tumor-normal sequencing may identify roles for germline variation that extend beyond cancer susceptibility. Therefore, our framework for germline data management defines primary germline findings as any variants with relevance to tumor biology or treatment. This includes variants with implications for estimating cancer risk, determining the landscape of somatic alterations or tumor evolution, modifying the immune response and microenvironment, and predicting overall response or adverse reactions to treatment (Figure 3). This broad definition is based on findings from large tumor-sequencing projects that have allowed functional characterization of inherited genetic variation other than rare coding variants in known disease genes. These include a

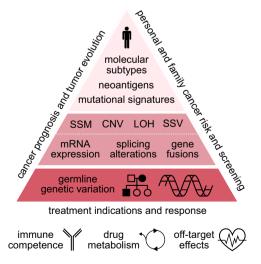


Figure 3. Extending the clinical significance of integrated molecular analysis of tumor and normal tissues beyond cancer susceptibility. CNV = copy number $variants; \ LOH = loss\ of\ heterozygosity; \ mRNA = messenger\ RNA; \ SSM = simple$ somatic mutations; SSV = somatic structural variants.

number of recent studies that have identified roles for common and noncoding variants in tumorigenesis that may ultimately guide treatment interventions or development of targeted therapeutics (21-24). Identifying germline variation with potential tumor relevance is thus an important part of characterizing the complex molecular architecture of individual tumors and may have immediate or future clinical implications.

Framework for the Clinical Translation of **Germline Findings**

Based on our experience during 7 years of the POG program, we developed an integrated schema for the management of germline genetic information. We propose a nonmutually exclusive categorical framework for evaluating clinically actionable germline variants that aims to identify inherited cancer susceptibility, characterize treatment indications, and provide opportunities for the discovery of novel genetic associations (Table 1). This includes pathogenic and likely pathogenic variants in known cancer predisposition genes with defined cancer risk estimates and screening recommendations and may include VUS in cases where the patient's phenotype supports pathogenicity. In the POG program, known and novel pathogenic and likely pathogenic variants in known cancer predisposition genes are reviewed by the Ethics and Germline Working Group, and these are then disclosed to the tumor board and case oncologist with a general recommendation for return to the patient with a referral to HCP for follow-up. Secondary germline findings in the 59 genes defined by the ACMG should consistently be returned to patients if informed consent for return of such findings was given at the time of study enrollment (25, 26).

With an inclusive definition of primary germline findings in the context of cancer genomics, pharmacogenomic variants with known drug associations, polymorphic alleles with putative immune response associations, variants in genes with potential tumor relevance based on histological or molecular characteristics, and other variants with potential clinical relevance as requested by the case oncologist could also be analyzed. The Ethics and Germline Working Group supports routine reporting of germline variants predictive of immune response, such as human leukocyte antigen class I genotypes, but recommends caution in reporting pharmacogenomic variants with limited or conflicting scientific evidence that may prevent use of important supportive medications (27). Clinicians and scientists should refer to public curated databases such as PharmGKB for updated variant reviews and clinical guidelines (28). In practice, a curated variant database relevant for chemotherapy and other cancer therapy with established pharmacogenomic associations, such as gastrointestinal and bone marrow toxicity in carriers of the UGT1A1*28polymorphism, should be prioritized during routine germline analysis unless otherwise requested by the case clinician (29).

Pharmacogenomic variants will be an important aspect of cancer treatment in the era of precision medicine. The potential benefits of profiling pharmacogenomic variants as part of the POG program were demonstrated in a patient with a gastrointestinal stromal tumor and prolonged QT interval during a high-dose course of imatinib. Further genomic profiling was performed at the request of the case clinician, and a common polymorphism in SCN5A (p.His558Arg) that has been previously reported as a genetic modifier in cardiac arrhythmia syndromes was identified (30, 31). Metabolic and cardiovascular gene variants that are known to modulate drug metabolism or mediate

adverse events in response to treatment have immediate clinical utility in treatment selection. Thus, prior knowledge of variants that can impact treatment choice may provide indications for or against the use of certain drugs. In the case described previously, this may have indicated an avoidance of specific tyrosine kinase inhibitors associated with prolonged QT intervals (32, 33).

Novel relationships between genetic alterations and histological or molecular phenotypes could also be investigated through an agnostic analysis of germline variation. Mutation and expression comparisons using public datasets are invaluable tools for characterizing the individual tumor genome and in validating putative molecular associations. However, these must be interpreted cautiously given expected differences between sample handling and preparation protocols, sequencing chemistries and platforms, and computational pipelines. With a secondary goal of discovery, deleterious germline variants in known cancer genes defined in the Cancer Gene Census of the Catalog of Somatic Mutations in Cancer or genes in other disease-related pathways could be prioritized but not limiting when investigating novel associations with outlier tumor phenotypes (Supplementary Table 4, available online) (34, 35). It should be noted that germline findings with unconfirmed implications for disease pathogenesis, prognosis, or treatment are purely the result of research and, because of their uncertain nature, are strongly discouraged from being returned to the patient unless indicated by the tumor board for treatment indications. If germline findings are determined to be of clinical importance, notice should be given to the case clinician to ensure banking of a clinical DNA sample and patient and/or family referral for genetic counseling and variant confirmation. Disclosure of germline variants to the POG tumor board and case clinician is encouraged in cases with established implications for clinical management, including for cancer risk reduction, screening, or treatment indications. In the future, larger studies and agnostic analyses may uncover additional roles for these variants in disease pathogenesis that may be predictive or prognostic of overall survival and treatment outcomes.

Hereditary cancer susceptibility is commonly observed in unselected cohorts of cancer patients who do not meet current clinical testing guidelines (36, 37). In precision oncology, the utility of exploring inherited genetic variation is achieved through the implementation of cancer prevention and screening strategies and by the use of targeted therapies. However, personnel and financial resources in jurisdictions with universal health care may be a growing barrier to service accessibility as genetic testing becomes more common and additional germline variants with clinical significance are discovered. These issues were strongly considered by the Ethics and Germline Working Group in assessing the benefits of disclosure for certain variants, but these guidelines must be re-evaluated moving forward to help inform cost-effective, patient-centered care. Based on our experience in the POG program, genome-wide analysis of inherited genetic variation should allow for the examination of normal and disease-causing variation that may affect tumor evolution, response to therapy, and immune function; predict adverse events; and/or be relevant to noncancer disease risk that may be meaningful to patients and their care.

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Table 1. Framework for the return of germline findings in precision oncology

| Tier | Clinical indication | Recommended categories for consideration | Examples | PEWG recommendations for disclosure |
|-----------|-------------------------------------|---|---|--|
| Primary | Cancer susceptibility | Known or novel pathogenic and likely pathogenic variants associated with moderate- to high-penetrance cancer susceptibility ^a | CHEK2 c.1100delC (p.Thr367Metfs) | Return to patient with referral to HCP |
| | | Known or novel pathogenic and likely pathogenic variants associated with low-penetrance cancer susceptibility ^a | CHEK2 c.470T>C (p.lle157Thr) | Return to patient with referral to HCP on a caseby-case basis |
| | | Autosomal or X-linked carrier status for known or novel pathogenic and likely pathogenic var- iants associated with low-, moderate- or high- penetrance cancer susceptibility* | ERCC5 c.529-1G>A | Return to patient with referral to HCP on a caseby-case basis |
| | | VUS in cancer predisposition genes with clinical and molecular indications for pathogenicity | MUTYH c.996G>A (p.Ser332Ser) | Return to patient with referral to HCP on a caseby-case basis |
| | Disease pathogenesis, prog- | Known or novel variants in cancer predisposition | Biallelic loss through combined germline | Return to patient with referral to HCP on a case- |
| | nosis, or treatment | genes understood to contribute to tumor phe- | and somatic aberrations: | by-case basis and if clinical significance has |
| | | notype and evolution | NTHL1 carrier status MUTYH carrier status | been established |
| | | Deleterious variants in known or novel cancer-related genes | Deletion polymorphisms in APOBEC3A and APOBEC3B | Return to patient if clinical significance has been established |
| | | Pharmacogenomic variants with established or potential cancer treatment associations | UGT1A1*28 | Return to patient if clinical significance has been established |
| | | Alleles associated with immune response | HLA class I genotypes | Return to patient if clinical significance has been established |
| | | Variants in gene(s) reviewed at the request of the case clinician given patient consent | SCN5A c.1673A>G (p.His558Arg) | Return to patient if clinical significance has been established |
| Secondary | Secondary Mendelian disease risk or | Deleterious variants in genes without known | Pathogenic and likely pathogenic variants | Return to patient with referral to medical genetics |
| | carrier status | implications for cancer prevention, cancer screening, or treatment | in Mendelian disease genes defined by the ACMG | program if patient preference for noncancer-re- lated information is indicated in consent |
| | | | | |

*Implications for cancer susceptibility should be considered in the context of variant zygosity and the typical mode of inheritance observed for a given gene. In particular, this includes variants in autosomal recessive genes or in X-linked genes conferring recessive disease risk that may have differing indications for XY, XX, and XO individuals. ACMG = American College of Medical Genetics and Genomics; HCP = Hereditary Cancer Program; HLA = human leukocyte antigen; PEWG = Personalized OncoGenomics Ethics and Germline Working Group; VUS = variants of uncertain significance.

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References

- Copson ER, Maishman TC, Tapper WJ, et al Articles germline BRCA mutation and outcome in young-onset breast cancer (POSH): a prospective cohort study. Lancet Oncol. 2018;19(2):169–180.
- Robson M, Im S-A, Senkus E, et al Olaparib for metastatic breast cancer in patients with a germline BRCA mutation. N Engl J Med. 2017;377(6):523–533.
- Ryu JS, Hong YC, Han HS, et al Association between polymorphisms of ERCC1 and XPD and survival in non-small-cell lung cancer patients treated with cisplatin combination chemotherapy. Lung Cancer. 2004;44(3):311–316.
- Musolino A, Naldi N, Bortesi B, et al Immunoglobulin G fragment C receptor polymorphisms and clinical efficacy of trastuzumab-based therapy in patients with HER-2/neu-positive metastatic breast cancer. J Clin Oncol. 2008; 26(11):1789–1796.
- Mandelker D, Zhang L. The emerging significance of secondary germline testing in cancer genomics. J Pathol. 2018;244(5):610–615.
- Laskin J, Jones S, Aparicio S, et al Lessons learned from the application of whole-genome analysis to the treatment of patients with advanced cancers. Cold Spring Harb Mol Case Stud. 2015;1(1):a000570.
- Zhao EY, Shen Y, Pleasance E, et al Homologous recombination deficiency and platinum-based therapy outcomes in advanced breast cancer. Clin Cancer Res. 2017;23(24):7521–7530.
- Wong H-L, Yang KC, Shen Y, et al Molecular characterization of metastatic pancreatic neuroendocrine tumors (PNETs) using whole-genome and transcriptome sequencing. Cold Spring Harb Mol Case Stud. 2018;4(1):a002329.
- 9. Thibodeau ML, Zhao EY, Reisle C, et al Base excision repair deficiency signatures implicate germline and somatic MUTYH aberrations in pancreatic

- ductal adenocarcinoma and breast cancer oncogenesis. Cold Spring Harb Mol Case Stud. 2019;5(2):a003681.
- Weinstein JN, Collisson EA, Mills GB, et al The Cancer Genome Atlas Pan-Cancer Analysis Project. Nat Genet. 2013;45(10):1113–1120.
- Hudson TJ, Anderson W, Artez A, et al International network of cancer genome projects. Nature. 2010;464(7291):993–998.
- 12. Richards S, Aziz N, Bale S, et al Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015;17(5):405–423.
- Regier DA, Peacock SJ, Pataky R, et al Societal preferences for the return of incidental findings from clinical genomic sequencing: a discrete-choice experiment. CMAJ. 2015;187(6):E190–E197.
- Rahman N. Realizing the promise of cancer predisposition genes. Nature. 2014;505(7483):302–308.
- 15. Huang K, Mashl RJ, Wu Y, et al Pathogenic germline variants in 10,389 adult cancers. Cell. 2018;173(2):355–370.e14.
- Landrum MJ, Lee JM, Benson M, et al ClinVar: improving access to variant interpretations and supporting evidence. Nucleic Acids Res. 2018;46(D1): D1062–D1067.
- Li Q, Wang K. InterVar: clinical interpretation of genetic variants by the 2015 ACMG-AMP guidelines. Am J Hum Genet. 2017;100(2):267–280.
- Statistics Canada. British Columbia [Province] and Canada [Country] (table). Census Profile. 2016 Census. Statistics Canada Catalogue no. 98-316-X2016001. Ottawa. Released November 29, 2017. https://www12.statcan.gc. ca/census-recensement/2016/dp-pd/prof/index.cfm?Lang=E. Accessed June 14, 2020.
- Cummings BB, Marshall JL, Tukiainen T, et al Improving genetic diagnosis in Mendelian disease with transcriptome sequencing Genotype-Tissue Expression Consortium; 2017. http://stm.sciencemag.org.ezproxy.library. ubc.ca/content/scitransmed/9/386/eaal5209.full.pdf. Accessed September 9, 2017.
- Walsh MF, Ritter DI, Kesserwan C, et al Integrating somatic variant data and biomarkers for germline variant classification in cancer predisposition genes. Hum Mutat. 2018;39(11):1542–1552. doi: 10.1002/humu.23640
- Middlebrooks CD, Banday AR, Matsuda K, et al Association of germline variants in the APOBEC3 region with cancer risk and enrichment with APOBEC-signature mutations in tumors. Nat Genet. 2016;48(11):1330–1338.
- 22. Carter H, Marty R, Hofree M, et al Interaction landscape of inherited polymorphisms with somatic events in cancer. Cancer Discov. 2017;7(4):410–423.
- Lim YW, Chen-Harris H, Mayba O, et al Germline genetic polymorphisms influence tumor gene expression and immune cell infiltration. Proc Natl Acad Sci U S A. 2018;115(50):E11701–E11710.
- Lin P-C, Yeh Y-M, Wu P-Y, Hsu K-F, Chang J-Y, Shen M-R. Germline susceptibility variants impact clinical outcome and therapeutic strategies for stage. Sci Rep. 2019;9(1):3931.
- Green RC, Berg JS, Grody WW, et al ACMG recommendations for reporting of incidental findings in clinical exome and genome sequencing. Genet Med. 2013;15(7):565–574.
- Kalia SS, Adelman K, Bale SJ, et al Recommendations for reporting of secondary findings in clinical exome and genome sequencing, 2016 update (ACMG SF v2.0): a policy statement of the American College of Medical Genetics and Genomics. Genet Med. 2017;19(2):249–255.
- Chowell D, Morris LGT, Grigg CM, et al Patient HLA class I genotype influences cancer response to checkpoint blockade immunotherapy. Science. 2018; 359(6375):582–587.
- Whirl-Carrillo M, McDonagh EM, Hebert JM, et al Pharmacogenomics knowledge for personalized medicine. Clin Pharmacol Ther. 2012;92(4):414

 –417.
- Iyer L, Das S, Janisch L, et al UGT1A1*28 polymorphism as a determinant of irinotecan disposition and toxicity. Pharmacogenomics J. 2002;2(1):43–47.
- Ye B, Valdivia CR, Ackerman MJ, Makielski JC. A common human SCN5A polymorphism modifies expression of an arrhythmia causing mutation. Physiol Genomics. 2003;12(3):187–193.
- Matsumura H, Nakano Y, Ochi H, et al H558R, a common SCN5A polymorphism, modifies the clinical phenotype of Brugada syndrome by modulating DNA methylation of SCN5A promoters. J Biomed Sci. 2017;24(1):91.
- 32. Strevel EL, Ing DJ, Siu LL. Molecularly targeted oncology therapeutics and prolongation of the QT interval. J Clin Oncol. 2007;25(22):3362–3371.
- Kloth JSL, Pagani A, Verboom MC, et al Incidence and relevance of QTcinterval prolongation caused by tyrosine kinase inhibitors. Br J Cancer. 2015; 112(6):1011–1016.
- Futreal PA, Coin L, Marshall M, et al A census of human cancer genes. Nat Rev Cancer. 2004;4(3):177–183.
- Tate JG, Bamford S, Jubb HC, et al COSMIC: the Catalogue of Somatic Mutations in Cancer. Nucleic Acids Res. 2019;47(D1):D941–D947.
- Schrader KA, Cheng DT, Joseph V, et al Germline variants in targeted tumor sequencing using matched normal DNA. JAMA Oncol. 2016;2(1):104–111.
- Mandelker D, Zhang L, Kemel Y, et al Mutation detection in patients with advanced cancer by universal sequencing of cancer-related genes in tumor and normal DNA vs guideline-based germline testing. JAMA. 2017;318(9):825.