

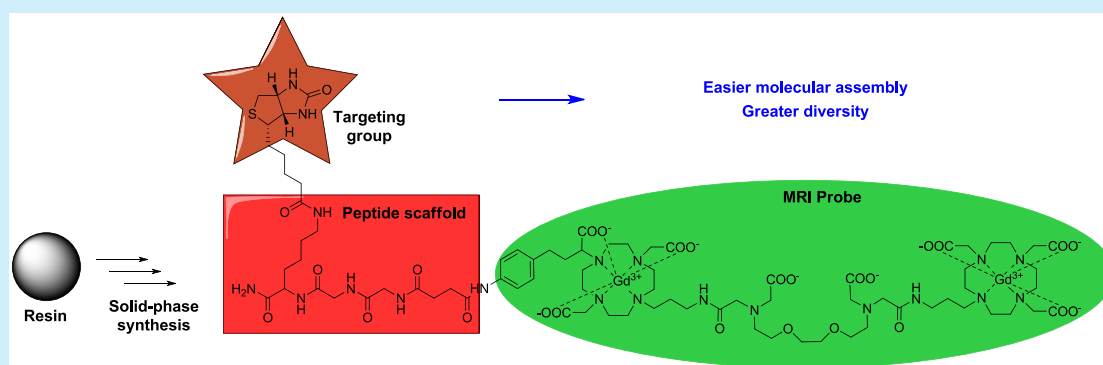
## Solid-Phase-Supported Approach for the Preparation of Bioresponsive and Multifunctional MRI Probes

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### S Supporting Information



**ABSTRACT:** The development of bifunctional imaging probes can often be challenging with difficult and time-consuming solution phase chemistry protocols and purification techniques. A solid phase synthetic protocol was therefore utilized to produce a functionalized derivative of a potent bismacrocyclic calcium-responsive contrast agent for magnetic resonance imaging. Through a convenient building block approach, the applicability of this methodology in the preparation and simple future development of multifunctional imaging probes was demonstrated.

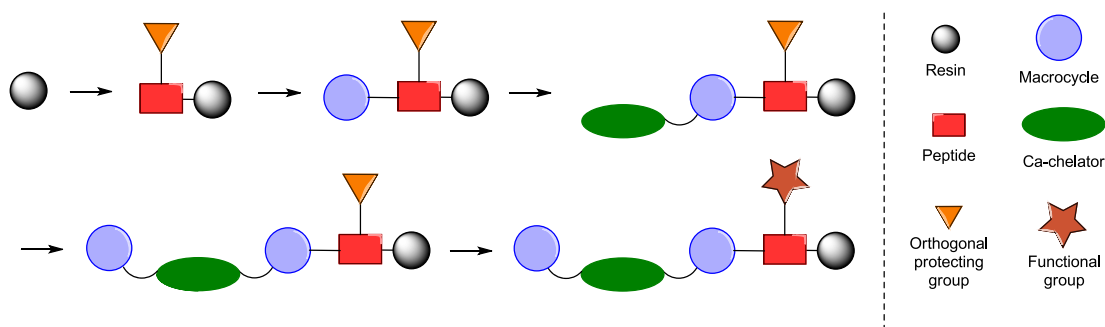
Bifunctional chelators based on polyazamacrocycles are often utilized in the development of imaging probes.<sup>1</sup> The macrocyclic component acts as a metal chelator while the second function can be exploited for further synthetic modifications, conjugation to biomolecules, targeting, or the detection of specific analytes. As such, they are heavily used in various molecular imaging techniques, such as positron emission tomography (PET), single photon emission computed tomography (SPECT), or magnetic resonance imaging (MRI). Among these, MRI is a preferred technique utilized to obtain three-dimensional anatomical images of soft tissue with excellent spatial resolution. Moreover, the use of MRI contrast agents increases its specificity, while expanding the scope and utilization potential. For instance, its functional variant, which aims to observe processes on the molecular level (molecular fMRI), has progressively gained in importance lately,<sup>2</sup> as its application aids the visualization of biochemical events in a dynamic manner. Subsequently, understanding various biological and pathological states becomes a challenging but highly rewarding task using this technique and the appropriate biomarkers. Thus, the application of bioresponsive or “smart” contrast agents (SCAs) in the development of functional MRI has become an increasingly investigated research area over recent years.<sup>3–5</sup> In order to observe alterations that are

coupled to the execution of particular biological processes, dedicated SCAs are employed, as they are capable of displaying a change in their MR behavior upon a physical or chemical change in their microenvironment. To achieve this, SCAs, which mostly consist of bifunctional systems, have been developed to detect enzymatic activity, pH changes, or various metal ion concentrations. However, the development of such systems includes arduous synthetic procedures. Moreover, limitations in synthetic modifications or further addition of functionalities to provide more diverse derivatives frequently lead to complications and challenges that cause limitations in structural diversity, hence the broader utilization of SCAs.

To this end, solid-phase synthetic (SPS) methods have been frequently used as an alternative to conventional solution-phase techniques, eventually finding its application in the development of MRI probes.<sup>6–11</sup> Introduced by Bruce Merrifield in the 1960s, SPS has been typically used in the area of peptide synthesis, as it allows for the building of a molecule in a stepwise fashion on an insoluble solid support.<sup>12–14</sup>

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**Figure 1.** Schematic representation of the synthetic approach followed in this study.

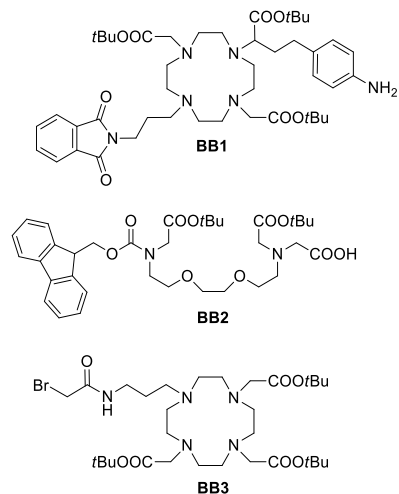
Significantly, the use of an insoluble solid support gave significant advantages over standard solution-phase chemistry techniques,<sup>15</sup> specifically: (i) the ability to drive reactions to completion through the use of excess reagents; (ii) the minimization of physical losses; (iii) quick washing steps after each reaction instead of tedious solution-phase workups and purification; (iv) to allow for the synthesis of more complex molecules and (v) the possibility to fully automate the synthetic procedure.

Such advantages could be useful for circumventing tedious and demanding preparation procedures of MRI chelates, as mentioned above. Yet, the potential of SPS has not been fully exploited in this field, especially on the preparation of SCAs. Being bifunctional chelators by default, SCAs are very suitable targets for convenient techniques such as SPS. Moreover, the establishment of a reliable SPS methodology to produce SCAs can enable additional modifications toward the incorporation of further functionalities (e.g., targeting vector, second probe suitable for additional imaging modality, etc.), thus expanding the scope of SCA use.

Hence, aiming to access more diverse bioresponsive probes, we embarked on developing an SPS synthetic protocol which would enable the production of such probes in a simple and convenient manner. Of particular importance was the preparation of a bismacrocylic SCA, which generates a strong response in the presence of  $\text{Ca}^{2+}$ , and in cellular model systems.<sup>16,17</sup> The previously developed solution-phase procedure allowed for its convenient preparation; however, the symmetric nature did not allow for the incorporation of additional functionalities without damaging the response toward  $\text{Ca}^{2+}$ .

We have therefore established a novel SPS approach based on the Fmoc-SPPS strategy<sup>15</sup> to develop the desired multifunctional bismacrocylic probe. The approach consisted of five stages (Figure 1): (1) the synthesis of a peptide unit capable of providing an orthogonal functionality; (2–4) the building of the bismacrocylic component with a series of smaller building blocks (BB1–BB3, Figure 2); (5) the incorporation of a functional molecule to the existing SCA. Beside the commercially available amino acids necessary for step 1, the exploration of this SPS methodology required the preparation of specific building blocks in solution prior to their introduction in steps 2–4.

The building blocks BB1–3 were selected as the best candidates for the assembly of the bismacrocylic system. Importantly, these BBs included protecting groups that could be selectively deprotected and were compatible with the resin used. Furthermore, through the use of BB2 with the Fmoc protecting group, we were able to employ a strategy of



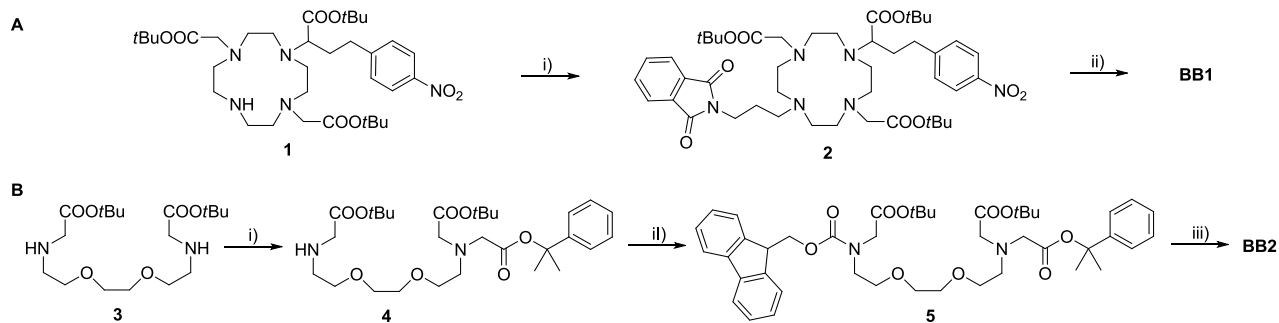
**Figure 2.** Chemical structures of the building blocks used.

alkylation in the final building block assembly step, which is a rather unconventional approach to SPS (*vide infra*).

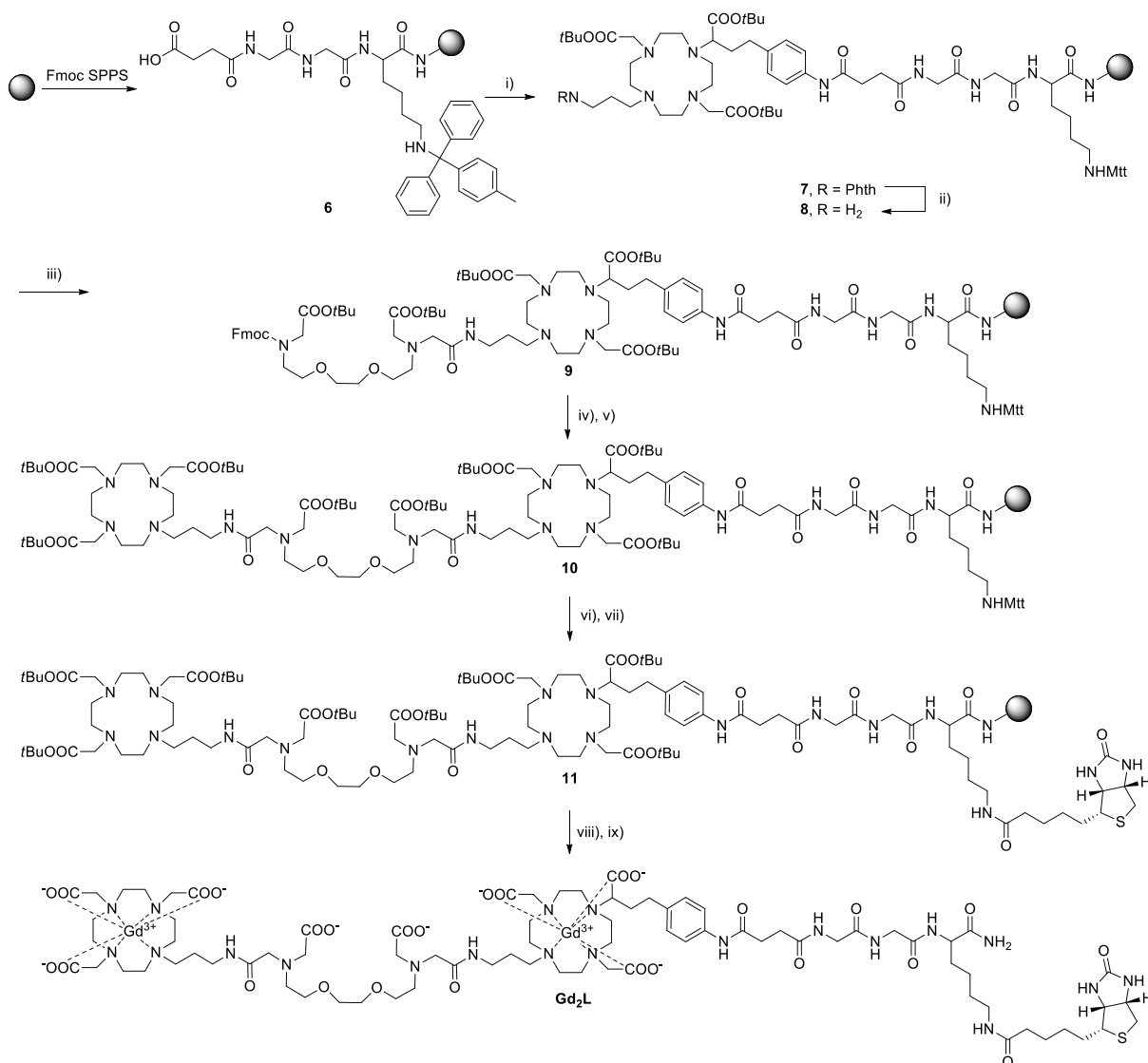
BB1 and BB2 were prepared in a few straightforward synthetic steps (Scheme 1), whereas BB3 was synthesized in accordance with a reported literature procedure.<sup>16</sup> The starting material for the synthesis of BB1 was secondary amine **1** (Scheme 1A). Its N-alkylation with *N*-(3-bromopropyl)-phthalimide gave **2**, which was reduced to give the BB1. Hence, this approach resulted in a DO2A-based macrocyclic component with one functional group (an aromatic amine) available for coupling to the peptide scaffold and another (phthalimide-protected amine) which can be selectively deprotected and used for further synthetic transformations.

The design of the  $\text{Ca}^{2+}$  chelator BB2, derived from ethylene glycol tetraacetic acid (EGTA), required differentiation from the “standard” derivatives routinely used in our group. Typically, the EGTA-derived block used for the synthesis of this type of SCAs contains benzyl esters as orthogonal protecting groups, which are incompatible with SPS due to the use of a heterogeneous catalyst under a reducing hydrogen atmosphere. Thus, the alternative derivative bears an Fmoc protecting group and a phenyl isopropanol ester. The conditions of deprotection for this acid labile ester were orthogonal to that of Fmoc, *tert*-butyl esters, and the resin.

The synthesis of BB2 started via the monoalkylation of secondary diamine **3** with 2-phenylpropan-2-yl 2-bromoacetate in acetonitrile to give **4** (Scheme 1B). Acylation of **4** with FmocCl afforded **5**. To yield the desired BB2, cleavage of the phenylisopropanol ester was carried out in a solution of 3% TFA in dichloromethane.

Scheme 1. Synthetic Route towards Building Blocks BB1 and BB2<sup>a</sup>

<sup>a</sup>(A) (i) *N*-(3-Bromopropyl)phthalamide, K<sub>2</sub>CO<sub>3</sub>, MeCN, 16 h, 70 °C, 66%; (ii) H<sub>2</sub>, Pd/C (20%wt), EtOH, rt, 78%. (B) (i) 2-Phenylpropan-2-yl 2-bromoacetate, DCC, DMAP, MeCN, 2 days, rt, 44%; (ii) FmocCl, Na<sub>2</sub>CO<sub>3</sub>, dioxane/water, 16 h, rt, 70%; (iii) 3% TFA/CH<sub>2</sub>Cl<sub>2</sub>, 1 h, rt, quant.

Scheme 2. Synthesis of Gd<sub>2</sub>L<sup>a</sup>

<sup>a</sup>(i) **BB1**, HATU, DIPEA, DMF; (ii) ethylenediamine, *i*PrOH; (iii) **BB2**, HATU, DIPEA, DMF; (iv) 40% piperidine/DMF; (v) **BB3**, DIPEA, DMF; (vi) TFA/triisopropylsilane/CH<sub>2</sub>Cl<sub>2</sub> (3:3:94), 4 × 2 min; (vii) biotin, HATU, HOBT, DIPEA, DMF; (viii) TFA/triisopropylsilane/H<sub>2</sub>O (95:2.5:2.5); (ix) GdCl<sub>3</sub>·6H<sub>2</sub>O, pH 7.

Once the BBs were prepared, the assembly of the multifunctional SCA was conducted using standard conjugation procedures (Scheme 2).<sup>15</sup> Initially, we prepared the

peptide sequence Lys-Gly-Gly between the rink-amide resin and the rest of the SCA to allow for the inclusion of an additional functional molecule at the final stage of conjugate

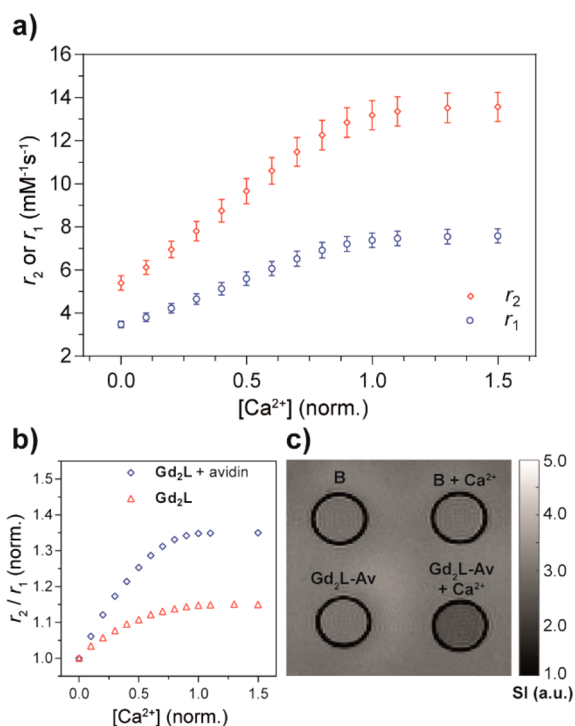
preparation (*vide infra*). Consequently, Lys bearing a 4-methyltrityl (Mtt) orthogonal protection of the  $\omega$ -NH<sub>2</sub> was used as the first amino acid in the SPS protocol.

Furthermore, reaction of the terminal amine of the Lys-Gly-Gly backbone with succinic anhydride provided **6**, which acted as a spacer unit and as a method to convert the N-terminal functionality to a carboxylic acid suitable for conjugation to **BB1**. Using the more reactive HATU coupling agent, the coupling of **BB1** to the peptidyl resin was achieved to give **7**. Here, phthalimide deprotection of **7** was carried out with 10 equiv of ethylene diamine in isopropanol at room temperature to yield amine **8**. Interestingly, phthalimide is an amino protecting group that is not frequently employed in SPS. If utilized, the most popular deprotection conditions involve hydrazine.<sup>18</sup> On the other hand, ethylene diamine is one attractive alternative due to its less harsh nature, increased reactivity, and overall safer use.<sup>19</sup> Procedures describing this method typically employ an excess of ethylene diamine in butanol or isopropanol at reflux.<sup>19,20</sup> Here we demonstrate the use of ethylene diamine as an efficient reagent for phthalimide deprotection on solid phase at room temperature, proceeding smoothly to provide conversion to the amino function. Importantly, the mild deprotection conditions described here can allow for increased flexibility in terms of future preparations involving hydrazine sensitive protecting groups.

Conjugation of amine **8** to **BB2** was performed using HATU to afford **9**. Subsequent Fmoc deprotection of **9** was carried out with 40% piperidine in DMF. The resulting amine was *N*-alkylated with **BB3** to give **10**. In total, 4.8 equiv of **BB3** were added in two separate portions to drive the reaction to completion. Finally, with the SCA assembled on the resin, the incorporation of a functional molecule could be carried out. For this proof of concept study, we selected biotin, as its interaction with avidin is well characterized.<sup>21</sup> However, in principle this site could be reserved for a number of various functional molecules such as antibodies, targeting moieties, or functional dyes depending on the desired application for the specific SCA.

Prior to biotin coupling, the orthogonal Mtt protecting group of lysine was removed with a solution of TFA/triisopropylsilane/dichloromethane (3:3:94) to afford the primary amine. This was then coupled to biotin with HATU yielding **11**. Cleavage of the final biotinylated-SCA, **11**, from the resin was achieved with the cleavage cocktail TFA/triisopropylsilane/H<sub>2</sub>O (95:2.5:2.5) to yield ligand **L**, which was purified by reversed-phase HPLC. Subsequent complexation with Gd<sup>3+</sup> was carried out in H<sub>2</sub>O at room temperature and neutral pH to afford the final SCA Gd<sub>2</sub>L with an overall yield of 24% across the whole synthetic procedure.

To assess the potency of the multifunctional SCA toward Ca<sup>2+</sup>, its relaxometric characterization was performed with proton longitudinal and transverse relaxometric titrations. The response observed for Gd<sub>2</sub>L was excellent and even higher than the previously reported bismacrocylic analogue.<sup>16</sup> The relaxometric titrations show an increase of longitudinal  $r_1$  relaxivity upon the addition of Ca<sup>2+</sup> from 3.47 to 7.58 mM<sup>-1</sup> s<sup>-1</sup>, which is almost a 118% enhancement relative to the initial value in the absence of Ca<sup>2+</sup>. Concurrently, the transverse  $r_2$  relaxivity increased from 5.40 to 13.57 mM<sup>-1</sup> s<sup>-1</sup>, or around 150% (Figure 3a). The obtained results indicate that the effective response of the SCA has been retained despite the change in molecular design. Moreover, the intrinsic property of Gd<sub>2</sub>L to trigger concurrent and massive changes on two



**Figure 3.** Relaxometric and MRI phantom experiments for Gd<sub>2</sub>L and Gd<sub>2</sub>L + avidin with Ca<sup>2+</sup> ([Gd<sup>3+</sup>] = 1 mM, pH 7.4, 50 mM HEPES, 25 °C, 7 T). (a) Relaxometric titration of Gd<sub>2</sub>L with Ca<sup>2+</sup>; the  $r_1$  and  $r_2$  values are represented as means  $\pm$  SD of three independent measurements. (b) Ratio of  $r_2$  over  $r_1$  from relaxometric titrations of Gd<sub>2</sub>L and Gd<sub>2</sub>L + avidin with Ca<sup>2+</sup>. (c)  $T_2/T_1$  ratiometric MR phantom images of the buffer (B) or Gd<sub>2</sub>L + avidin (Gd<sub>2</sub>L-Av) sample tubes in presence and absence of Ca<sup>2+</sup> (1 equiv).

paramagnetic Gd<sup>3+</sup> ions upon interaction with a single Ca<sup>2+</sup> ion is very advantageous, and could now be more frequently exploited with this new preparation methodology.

With the inclusion of biotin as a functional moiety, we also proceeded to demonstrate its binding with avidin. The number of biotin binding sites available for Gd<sub>2</sub>L was determined through an HABA assay revealing three sites (Figure S1 in Supporting Information).<sup>22</sup> Following this assessment, 3 equiv of Gd<sub>2</sub>L were added to avidin in HEPES buffer (50 mM) and incubated for 2 h. Relaxometric titrations and  $T_2/T_1$ -weighted ratiometric MR images were recorded showing an increase in  $r_1$  and  $r_2$  upon Ca<sup>2+</sup> addition and consequently a change in MRI signal (Figures 3b,c and S2 in Supporting Information). The enhancement in  $r_1$  was lower than on Gd<sub>2</sub>L alone, which was previously observed after the attachment of “small-size” SCAs to nanosized systems.<sup>23</sup> Furthermore, the unexplored interactions of Gd<sub>2</sub>L with the protein surface could also play a role. On the other hand, the values recorded for  $r_2$  were typical for nanosized-type SCAs, indicating the possibility to use the Gd<sub>2</sub>L avidin conjugate in combination with the fast  $T_2/T_1$ -weighted imaging protocol.<sup>24</sup>

In summary, we have developed a solid-phase synthetic protocol for the preparation of functionalized and bioresponsive bismacrocylic SCAs. We diversified our building block synthesis to incorporate SPS-friendly functional groups such as phthalimide, which we successfully deprotected using mild conditions. Furthermore, through the use of a peptide backbone, significant flexibility can be achieved in terms of designing a molecule which can be highly specific, biocompat-

ible and possess a range of functional molecules. The results described here offer an exciting range of possibilities for the development of various bifunctional bioresponsive probes ranging from multifunctional molecules to multimeric species.

## ■ ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: [10.1021/acs.orglett.9b01341](https://doi.org/10.1021/acs.orglett.9b01341).

Synthetic procedures for preparation of **BB1**, **BB2**, and **Gd<sub>2</sub>L**; purification procedure for **Gd<sub>2</sub>L**; HABA assay; additional relaxometric titration experiments; parameters for MRI phantom measurements; NMR and HR mass spectra; LC-MS and HPLC data (PDF)

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### Notes

The authors declare no competing financial interest.

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