

Procalcitonin may not be a differential diagnostic marker for bacterial infection in febrile patients with chronic gouty arthritis

Journal of International Medical Research

2018, Vol. 46(10) 4197–4206

© The Author(s) 2018

Article reuse guidelines:

sagepub.com/journals-permissions

DOI: 10.1177/0300060518791093

journals.sagepub.com/home/imr



Jing Zhang^{1,*}, Cheng Zhao^{2,*}, Tong Wu¹,
Jiang Su¹, Xiaodan Wu¹, Jian Liu¹, Jing Zhu¹
and Bin Zhou¹

Abstract

Objective: This study aimed to examine the diagnostic value of serum procalcitonin (PCT) levels for identifying bacterial infection in febrile patients with chronic gouty arthritis.

Methods: Sixty-six febrile patients with chronic gouty arthritis were divided into non-bacterial infection ($n = 45$) and bacterial infection groups ($n = 21$). PCT levels were measured by an immunoassay. Other laboratory parameters, including the erythrocyte sedimentation rate (ESR), C-reactive protein (CRP), white blood cells (WBCs), and the neutrophil ratio were extracted from medical records. Receiver-operating characteristic curves were used to evaluate diagnostic values and accuracy.

Results: Serum PCT levels, the ESR, CRP levels, WBC count, and neutrophil ratio were not different between the groups. To assess the ability of PCT to discriminate bacterial infection in febrile patients with chronic gouty arthritis (cut-off value: 0.5 ng/mL), the sensitivity and specificity of PCT were 22.2% and 61.5%, respectively. The area under the curve (AUC) of serum PCT levels was 0.526. The AUCs of related inflammatory indicators were 0.530 for the ESR, 0.635 for CRP, 0.577 for the WBC count, and 0.712 for the neutrophil ratio.

Conclusion: Serum PCT levels may not be a good biomarker for bacterial infection in febrile patients with chronic gouty arthritis.

¹Department of Rheumatology and Immunology, Sichuan Academy of Medical Sciences & Sichuan Provincial People's Hospital, Chengdu, China

²Department of Rheumatology and Immunology, the Affiliated Drum Tower Hospital of Nanjing University Medical School, Nanjing, China

*These authors contributed equally to this work.

Corresponding author:

Tong Wu, Department of Rheumatology and Immunology, Sichuan Academy of Medical Sciences & Sichuan Provincial People's Hospital. No. 32, Section 2, West 1st Ring Road, Chengdu 610072, Sichuan Province, China.
Email: tongwu09@163.com



Keywords

Bacterial infection, biomarker, chronic gouty arthritis, procalcitonin, inflammation, fever

Date received: 3 April 2018; accepted: 5 July 2018

Introduction

Clinical manifestation of gouty arthritis (GA) is a metabolic disorder and shows an inflammatory response to deposition of monosodium urate crystals in joints, bones, and various tissues. GA generally presents as acute and chronic arthritis, which affects the first metatarsal joint, followed by other multiple joints, tophi, urolithiasis, and renal disease.¹ Patients with GA show inflammatory clinical symptoms, joint swelling, and stiffness over inflamed joints, and even systematic fever during a period of an acute attack. Findings of leukocytosis and an increase in the erythrocyte sedimentation rate (ESR) and serum C-reactive protein (CRP) levels in GA are similar to those of infectious diseases.² Although a positive bacterial culture is commonly regarded as the gold standard in diagnosing infections, it is time-consuming and has a low positive rate.³ Therefore, finding an effective method to promptly identify bacterial infection in febrile patients with chronic GA is important.

Procalcitonin (PCT) is the precursor of calcitonin, which is secreted by C cells of the thyroid gland and produced by hydrolysis of specific proteins in cells. In systemic inflammation, particularly in bacterial infections, PCT is released from various forms of pro-inflammatory cytokines, such as interleukin-6 (IL-6), tumor necrosis factor- α (TNF- α), and interleukin-1 (IL-1), indirectly and directly from microbial toxin.⁴ PCT levels rapidly increase in the first 2 to 4 hours after infection and then reach peak levels in serum within 6 to 24 hours.^{4,5} PCT levels then decrease as infection is controlled (normal

condition: < 0.05 ng/mL).⁶ Therefore, PCT is generally regarded as a helpful biomarker in detecting bacterial infections. Most previous studies have shown that inflammatory markers for infectious arthritis, such as serum PCT, are significantly elevated.⁷⁻⁹ A previous study reported that PCT levels in serum and synovial fluid are a sensitive biomarker for differentiating septic arthritis from non-infectious arthritis, such as rheumatoid arthritis, osteoarthritis, and GA.⁷

However, no studies have demonstrated whether PCT levels can show bacterial infection in patients with chronic GA who experience an acute attack. Therefore, this study aimed to evaluate the ability of PCT to diagnosis bacterial infection and guide prompt antibiotic therapy in febrile patients with chronic GA.

Materials and methods

Patients

This was a single-center, retrospective, observational study. All of the included patients were diagnosed with chronic tophaceous gout and fever (axillary temperature $> 37.4^{\circ}\text{C}$) and were admitted between November 2011 and January 2014 at the Department of Rheumatology and Immunology, Sichuan Academy of Medical Sciences & Sichuan Provincial People's Hospital in Chengdu, China. The protocol for this investigation was approved by Sichuan Academy of Medical Sciences & Sichuan Provincial People's Hospital Ethics Committee. This research was conducted in compliance with the Helsinki Agreement.

Written informed consent was obtained from each participant.

Patients with chronic GA and tophi were diagnosed in accordance with the 1977 American College of Rheumatology criteria.¹⁰ According to the criteria of bacterial infection, patients with chronic GA were divided into two groups as follows: the bacterial infection group (n=21) and the non-bacterial infection group (n=45). Inclusion criteria of patients with bacterial infection were as follows: (1) patients showed symptoms of chronic GA; (2) a pathogen was detected, with positive results for sputum culture, blood culture, secretion, and joint fluid culture; and (3) bacterial infection was diagnosed. Exclusion criteria were as follows: (1) patients with non-bacterial infections; and (2) uncertain causes with fever.

Clinical and laboratory assessment

Clinical and laboratory data of patients with chronic GA were retrospectively collected from medical records at the time of sampling serum PCT levels, including measurement of CRP levels, the ESR, the white blood cell (WBC) count, and the neutrophil ratio. Serum PCT levels were measured by using a chemiluminescent enzyme immunoassay (VIDAS; BioMerieux SA, Marcy l'Etoile, France). PCT levels of 0.25 to 0.5 ng/mL were likely to be a bacterial infection and those ≥ 0.5 ng/mL were very likely to be a bacterial infection. Therefore, PCT levels ≥ 0.5 ng/mL were considered as a positive result according to Schuetz et al.¹¹ The WBC count ($> 10.0 \times 10^9/L$ as a positive result) and the neutrophil ratio ($>70\%$ as a positive result) were estimated by using the Automatic Blood Cell Analyzer (SYSMEX XE-2100; Sysmex, Kobe, Japan). CRP levels > 8 mg/L were considered as positive and measured by the Nephelometer (Olympus AU5421 automatic biochemistry analyzer, Olympus, Tokyo, Japan).

An ESR > 20 mm/h was considered as positive by using Wintrobe's method.¹²

Statistical analysis

Continuous variables are presented as mean \pm standard deviation. The variables tested in the study are shown as descriptive statistics. We tested the normality of data distribution with the Kolmogorov–Smirnov test and the distribution of the data was considered normal. Significance testing was carried out using the Student's t-test and one-way analysis of variance was used with the Bonferroni post hoc test. The ability of distinguishing between bacterial infection and non-bacterial infection was evaluated by using receiver-operating characteristic (ROC) analysis. ROC curves were analyzed by using MedCalc software version 12 (MedCalc, Ostend, Belgium), and the sensitivity and specificity values of the optimal cut-off point were also calculated. The area under the curve (AUC) was calculated to evaluate diagnostic accuracy. In all analyses, $p < 0.05$ was considered to be statistically significant. All statistical analysis was performed by using SPSS Statistics version 16 (SPSS Inc., Chicago, IL, USA).

Results

Patients' characteristics

The demographic and clinical characteristics of the included patients are shown in Table 1. A total of 66 consecutive patients were studied. The mean ages of patients in the bacterial infection and non-bacterial infection groups were 60.2 ± 17.1 years and 56.2 ± 13.5 years, respectively. The mean disease duration of patients with tophaceous gout and bacterial infection was 14.8 ± 9.7 years and that in patients without infection was 12.4 ± 8.7 years. In patients with bacterial infection,

Table 1. Demographic and laboratory characteristics of enrolled patients

Parameters	Bacterial infection group (n = 21)	Non-bacterial infection group (n = 45)
Age, mean \pm SD (years)	60.2 \pm 17.1	56.2 \pm 13.5
Course of disease, mean \pm SD (years)	14.8 \pm 9.7	12.4 \pm 8.7
Type of infection		
Pulmonary	11 (52.4)	N/A
Joint	2 (9.5)	N/A
Skin soft tissue	8 (38.1)	N/A
Renal insufficiency	1 (4.8)	12 (26.7)
Hepatic failure	0	2 (4.4)
Glucocorticoid use	11 (52.4)*	7 (15.6)
Ruptured tophi	9 (42.9)*	4 (8.9)
Hypertension	12 (57.1)	24 (53.3)
Hyperlipidemia	1 (4.8)	0
Diabetes mellitus	6 (28.6)	9 (20)
Coronary heart disease	2 (9.5)	1 (2.2)

Values are mean \pm SD or n (%). Ruptured tophi refers to tophus, which is a symptom of chronic gouty arthritis. SD: standard deviation; N/A: not available. * $p < 0.05$

Table 2. Clinical information of patients with bacterial infection

Parameters	Type of infection	Type of bacteria	Patients (n)	Percentage
Infection	Pulmonary infection	<i>Streptococcus pneumoniae</i>	5	42.6
		<i>Klebsiella pneumoniae</i>	2	
		<i>Haemophilus influenzae</i>	1	
		<i>Streptococcus hemolyticus</i>	1	
	Skin soft tissue infection	<i>Staphylococcus aureus</i>	8	38.1
	Pulmonary combined skin Soft tissue infection	<i>Klebsiella pneumoniae</i>	1	1.9
		<i>Staphylococcus aureus</i>		
Joint infection	<i>Streptococcus pneumoniae</i>	2	9.5	
Arthritis infection	<i>Staphylococcus aureus</i>	1	1.9	
Culture site	Department of Rheumatology and Immunology, Sichuan Academy of Medical Sciences & Sichuan Provincial People's Hospital			
Disease duration of infection	7 to 10 days before admission			

the most common type of infection was pulmonary infection, followed by skin soft tissue infection, and joint infection (Table 2). Patients in the bacterial infection group had a significantly higher rate of glucocorticoid use and more ruptured tophi than did patients in the non-bacterial infection group (both $p < 0.05$).

Serum PCT levels and related inflammatory markers in patients in the bacterial infection and non-bacterial infection groups

Serum PCT levels and related indices of inflammation were measured in patients in the two groups. Patients in the bacterial

infection group had similar PCT levels to those in the non-bacterial infection group (Table 3). A total of 28.6% of patients in the bacterial group had PCT-positive results and 46.7% in the non-bacterial group had PCT-positive results. There were no significant differences in the ESR, WBC count, and the neutrophil ratio between the two groups. CRP levels tended to be higher in patients in the bacterial group than in those in the non-bacterial group ($p = 0.05$).

Ability of PCT and related inflammatory markers to distinguish infection in febrile patients with chronic GA

ROC curves of serum PCT levels and related inflammatory indicators were used for discriminating infection in febrile patients with chronic GA. With a cut-off value of 0.5 ng/mL, the sensitivity and specificity of serum PCT levels were the highest

(22.2% and 61.5%, respectively) (Table 4). The positive likelihood ratio and negative likelihood ratio were 0.577 and 1.265, respectively. The AUC of PCT was 0.526 (95% confidence interval [CI], 0.399–0.650). The AUCs of related inflammatory indicators were 0.530 for ESR (95% CI, 0.403–0.654), 0.635 for CRP (95% CI, 0.508–0.751), 0.577 for the WBC count (95% CI, 0.449–0.697), and 0.712 for the neutrophil ratio (95% CI, 0.588–0.817) (Table 4, Figure 1). Figure 2 shows scatter plots for each biomarker (PCT, ESR, CRP, WBC count, and neutrophil ratio).

When serum PCT levels > 0.5 ng/mL were regarded as a positive result, the positive predictive value and negative predictive value in patients with chronic tophaceous gout were 21.2% and 62.9%, respectively. The positive predictive values of CRP, the ESR, the neutrophil ratio, and the WBC count were 31.3%, 35.6%, 35.2%, and 34.9%, respectively.

Table 3. Related inflammatory indices for detecting bacterial infection in patients with chronic gouty arthritis

Parameters	Bacterial infection group (n = 21)	Non-bacterial infection group (n = 45)	p value
PCT (ng/mL)			0.5
Mean	0.72 ± 1.10	0.72 ± 0.81	
Maximum	3.86	3.48	
Positive rate, % (n)	28.6 (6/21)	46.7 (21/45)	
ESR (mm/h)			0.08
Mean	59.1 ± 32	48.3 ± 28	
Positive rate, % (n)	81 (17/21)	75.6 (34/45)	
CRP (mg/L)			0.05
Mean	124.32 ± 85	91.1 ± 72.4	
Positive rate, % (n)	95.2 (20/21)	95.6 (43/45)	
WBC count ($\times 10^9/L$)			0.13
Mean	14.3 ± 5.61	12.9 ± 4.2	
Positive rate, % (n)	71.4 (15/21)	62.2 (28/45)	
Neutrophil ratio (%)			0.08
Mean	84.8 ± 5.85	82.3 ± 6.8	
Positive rate, % (n)	100 (21/21)	84.4 (38/45)	

Values are mean ± standard deviation or % (n). PCT: procalcitonin; CRP: C-reactive protein; WBC: white blood cell; ESR: erythrocyte sedimentation rate

Table 4. Areas under the receiver-operating characteristic curve for PCT, and related inflammatory indices for detecting bacterial infection in patients with chronic gouty arthritis

Biomarkers	AUC (95% CI)	Cut-off value	Sensitivity (%)	Specificity (%)
PCT, ng/mL	0.526 (0.399–0.650)	0.5	22.2	61.5
ESR, mm/h	0.530 (0.403–0.654)	20	90.4	22.2
CRP, mg/L	0.635 (0.508–0.751)	8	95.2	2.2
WBC count, $\times 10^9/L$	0.577 (0.449–0.697)	10	71.4	37.8
Neutrophils, %	0.712 (0.588–0.817)	0.70	100	15.6

AUC: area under the curve; PCT: procalcitonin; CI: confidence interval; ESR: erythrocyte sedimentation rate; CRP: C-reactive protein; WBC: white blood cell

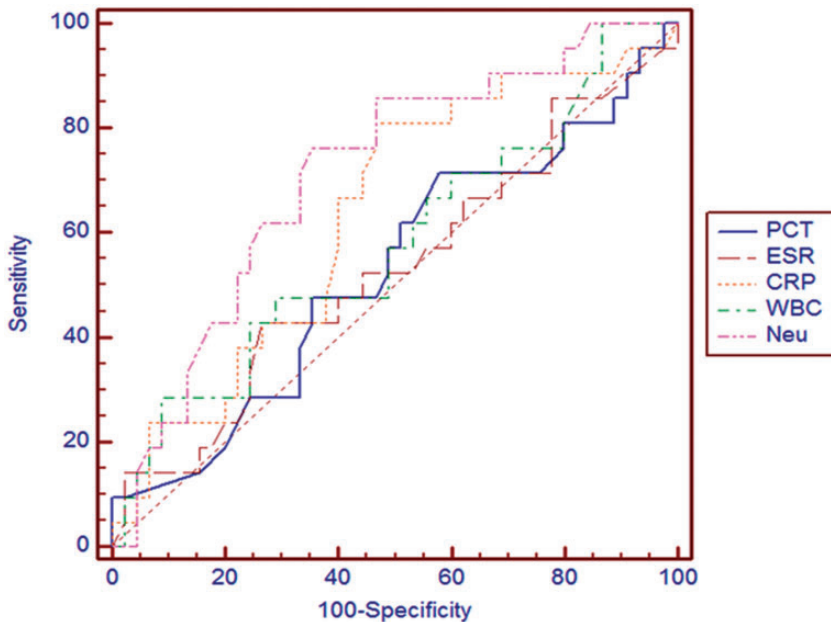


Figure 1. Receiver-operating characteristic curves for PCT, the ESR, CRP, WBCs, and the percentage of neutrophils for diagnosis of infection in patients with chronic gouty arthritis

PCT: procalcitonin; ESR: erythrocyte sedimentation rate; CRP: C-reactive protein; WBCs: white blood cells

Discussion

Many recent studies have reported that PCT is highly accurate in diagnosing bacterial infections, especially bacterial meningitis, septic shock, bacteremia, and pyelonephritis.^{13,14} PCT levels are closely related to the severity of infection and clinical prognosis, suggesting that PCT can be a sensitive indicator of early detection.¹⁵

Studies have shown that PCT can be used for differential diagnosis of non-infectious arthritis, such as rheumatoid arthritis, and other non-infectious arthritis and bacterial infections.^{16–18} However, no studies have examined the significance of PCT in clinical manifestation of gout, which is similar to clinical manifestations of bacterial infections and arthritis. To the best of our knowledge, there has been little research

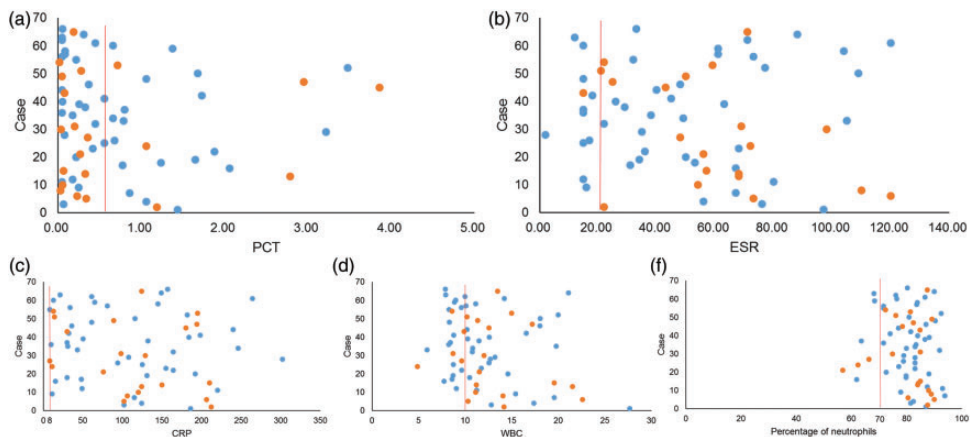


Figure 2. Scatter plots for each biomarker of PCT, the ESR, CRP, WBCs, and the percentage of neutrophils for diagnosis of infection in patients with chronic gouty arthritis. Orange circles indicate infectious patients and blue circles indicate non-infectious patients. (a) Scatter plot for PCT levels; (b) scatter plot for the ESR; (c) scatter plot for CRP levels; (d) scatter plot for WBCs; (e) scatter plot for the percentage of neutrophils
PCT: procalcitonin; ESR: erythrocyte sedimentation rate; CRP: C-reactive protein; WBCs: white blood cells

on the utility of PCT to identify gout with bacterial infections and crystal arthritis. One of the reasons for this lack of research could be because the incidence of bacterial infections is relatively low. Another reason may be that gout with recurrent bacterial infections is rare, and thus it has not attracted sufficient attention. Therefore, using PCT as a biomarker to monitor infectious diseases by these features could be advantageous.

In patients with acute exacerbation of chronic GA, bacterial infections should be identified early. Many previous studies have demonstrated that PCT is a useful marker of bacterial infections in sepsis/septic shock, and this is more superior to conventional bacterial culture.^{13,19} Additionally, PCT appears to be a highly sensitive and specific marker in differentiation between septic and non-septic arthritis.^{16,20} In this study, we assessed whether PCT levels could be a reliable biomarker in differential diagnosis between patients with bacterial infection and those with non-bacterial infection with chronic GA. We found that serum

PCT levels in patients in the bacterial infection group were not significantly higher than those in patients in the non-bacterial infection group (Table 3). Additionally, 21 of 45 patients in the non-bacterial infection group showed PCT-positive results, 12 of whom had chronic renal insufficiency, 2 had hepatic insufficiency, and 1 had renal transplantation. A previous study reported the isolated presence of calcitonin precursors in patients with benign liver diseases (such as hepatitis) or with hepatocellular carcinoma.²¹ Therefore, we believe that renal or hepatic insufficiency might influence PCT levels to some extent. Liu et al.⁸ reported that serum PCT levels were higher in patients with GA than in healthy controls without infection. This finding can be explained by various pro-inflammatory cytokines, such as IL-6, TNF- α , and particularly IL-1 β , which have important roles in inflammation caused by an acute gout attack. However, an inflammatory response has also been reported in patients with autoimmune diseases,²² such as adult-onset Still's disease,²³ granulomatosis with

polyangiitis (Wegener's granulomatosis),²⁴ and Good pasture's syndrome.²⁵ These autoimmune diseases often cause a sepsis-like high fever, which is called systemic inflammatory response syndrome. Some reports have indicated that massive production of inflammatory cytokines, such as TNF and IL-6, in such sepsis-like conditions may stimulate production of PCT.²⁶⁻²⁸ Additionally, a previous study suggested that PCT levels were not elevated in patients with local infections, such as wound infection or cellulitis.⁹ Therefore, patients with chronic GA who experience an acute attack with fever might have non-infectious systemic inflammatory response syndrome, and most of the inflammatory cytokines are released into the blood. However, larger studies are needed to prove this hypothesis.

Schuetz et al.¹¹ suggested that a bacterial infection may be present when a PCT value is between 0.25 ng/mL and 0.5 ng/mL in patients. Furthermore, a cut-off level of PCT > 0.5 ng/mL suggested bacterial infection and needed immediate antibiotic therapy. Therefore, PCT levels \geq 0.5 ng/mL were considered as a positive signal for investigating its diagnostic value in our study. We found a low sensitivity of 22.2% and a low specificity of 61.5% for differentiating bacterial infection in patients with chronic GA and fever with a cut-off PCT value of 0.5 ng/mL. Furthermore, the positive predictive value for chronic tophaceous gout was 21.2%. Therefore, we showed that PCT measurement may be not a highly specific marker for detecting bacterial infections in ROC curve analysis, which is inconsistent with previous studies.¹¹ Additionally, glucocorticoids can elevate the WBC count, but the effects of glucocorticoids on PCT levels are unclear. In our study, 11 patients who used glucocorticoids were enrolled in the bacterial infection group and seven were in the non-bacterial infection group. However, almost

half of the patients presented with elevated PCT levels in those who used glucocorticoids. Therefore, the effects of glucocorticoids on PCT levels should be investigated in the future.

Our study showed that there were no other significant differences in other conventional inflammatory markers, such as CRP levels, and the ESR, neutrophil ratio, and WBC count, between the two groups. The positive predictive value was also low for these variables (31.3%, 35.6%, 35.2% and 34.9%, respectively), which may have been due to the inflammatory reaction in gout. The WBC count may also have been related to the effects of glucocorticoids because the bacterial infection group had a significantly higher rate of glucocorticoid use than did the non-bacterial infection group.

Our study suggests that serum PCT levels may not be a good biomarker for bacterial infection in febrile patients with chronic GA. However, because the incidence of chronic GA is low, and the number of patients with chronic GA associated with fever is even fewer, the number of patients enrolled in this study was small. Further studies with a larger number of samples are required to verify our findings.

Declaration of conflicting interest

The authors declare that there is no conflict of interest.

Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

References

1. Richette P and Bardin T. Gout. *Lancet* 2010; 375: 318-328.
2. Talebi-Taher M, Shirani F, Nikanjam N, et al. Septic versus inflammatory arthritis:

- discriminating the ability of serum inflammatory markers. *Rheumatol Int* 2013; 33: 319–324.
3. Mathews CJ, Weston VC, Jones A, et al. Bacterial septic arthritis in adults. *Lancet* 2010; 375: 846–855.
 4. Mehanic S and Baljic R. The importance of serum procalcitonin in diagnosis and treatment of serious bacterial infections and sepsis. *Mater Sociomed* 2013; 25: 277–281.
 5. Chan T and Gu F. Early diagnosis of sepsis using serum biomarkers. *Expert Rev Mol Diagn* 2011; 11: 487–496.
 6. Schneider HG and Lam QT. Procalcitonin for the clinical laboratory: a review. *Pathology* 2007; 39: 383–390.
 7. Wang C, Zhong D, Liao Q, et al. Procalcitonin levels in fresh serum and fresh synovial fluid for the differential diagnosis of knee septic arthritis from rheumatoid arthritis, osteoarthritis and gouty arthritis. *Exp Ther Med* 2014; 8: 1075–1080.
 8. Liu W, Sigdel KR, Wang Y, et al. High level serum procalcitonin associated gouty arthritis susceptibility: from a southern Chinese Han population. *PLoS One* 2015; 10: e0132855.
 9. Saeed K, Ahmad N and Dryden M. The value of procalcitonin measurement in localized skin and skin structure infection, diabetic foot infections, septic arthritis and osteomyelitis. *Expert Rev Mol Diagn* 2014; 14: 47–54.
 10. Wallace SL, Robinson H, Masi AT, et al. Preliminary criteria for the classification of the acute arthritis of primary gout. *Arthritis Rheum* 1977; 20: 895–900.
 11. Schuetz P, Christ-Crain M, Wolbers M, et al. Procalcitonin guided antibiotic therapy and hospitalization in patients with lower respiratory tract infections: a prospective, multicenter, randomized controlled trial. *BMC Health Serv Res* 2007; 7: 102.
 12. Wintrobe MM, Landsberg JW. A standardized technique for the blood sedimentation test. *Am J Med Sci* 1935; 189: 102–114.
 13. Aikawa N, Fujishima S, Endo S, et al. Multicenter prospective study of procalcitonin as an indicator of sepsis. *J Infect Chemother* 2005; 11: 152–159.
 14. Butbul-Aviel Y, Koren A, Halevy R, et al. Procalcitonin as a diagnostic aid in osteomyelitis and septic arthritis. *Pediatr Emerg Care* 2005; 21: 828–832.
 15. Lind L, Bucht E and Ljunghall S. Pronounced elevation in circulating calcitonin in critical care patients is related to the severity of illness and survival. *Intensive Care Med* 1995; 21: 63–66.
 16. Hugel T, Schuetz P, Mueller B, et al. Serum procalcitonin for discrimination between septic and non-septic arthritis. *Clin Exp Rheumatol* 2008; 26: 453–456.
 17. Kuuliala A, Takala A, Siitonen S, et al. Cellular and humoral markers of systemic inflammation in acute reactive arthritis and early rheumatoid arthritis. *Scand J Rheumatol* 2004; 33: 13–18.
 18. Martinot M, Sordet C, Soubrier M, et al. Diagnostic value of serum and synovial procalcitonin in acute arthritis: a prospective study of 42 patients. *Clin Exp Rheumatol* 2005; 23: 303–310.
 19. Nylen ES, Whang KT, Snider RH, Jr., et al. Mortality is increased by procalcitonin and decreased by an antiserum reactive to procalcitonin in experimental sepsis. *Crit Care Med* 1998; 26: 1001–1006.
 20. Shaikh MM, Hermans LE and van Laar JM. Is serum procalcitonin measurement a useful addition to a rheumatologist's repertoire? A review of its diagnostic role in systemic inflammatory diseases and joint infections. *Rheumatology (Oxford)* 2015; 54: 231–240.
 21. Ghillani PP, Motte P, Troalen F, et al. Identification and measurement of calcitonin precursors in serum of patients with malignant diseases. *Cancer Res* 1989; 49: 6845–6851.
 22. Delevaux I, Andre M, Colombier M, et al. Can procalcitonin measurement help in differentiating between bacterial infection and other kinds of inflammatory processes? *Ann Rheum Dis* 2003; 62: 337–340.
 23. Scire CA, Cavagna L, Perotti C, et al. Diagnostic value of procalcitonin measurement in febrile patients with systemic autoimmune diseases. *Clin Exp Rheumatol* 2006; 24: 123–128.

24. Zycinska K, Wardyn KA, Zielonka TM, et al. Procalcitonin as an indicator of systemic response to infection in active pulmonary Wegener's granulomatosis. *J Physiol Pharmacol* 2008; 59(Suppl 6): 839–844.
25. Morath C, Sis J, Haensch GM, et al. Procalcitonin as marker of infection in patients with Goodpasture's syndrome is misleading. *Nephrol Dial Transplant* 2007; 22: 2701–2704.
26. Chen DY, Lan JL, Lin FJ, et al. Proinflammatory cytokine profiles in sera and pathological tissues of patients with active untreated adult onset still's disease. *J Rheumatol* 2004; 31: 2189–2198.
27. Lamprecht P, Kumanovics G, Mueller A, et al. Elevated monocytic IL-12 and TNF-alpha production in Wegener's granulomatosis is normalized by cyclophosphamide and corticosteroid therapy. *Clin Exp Immunol* 2002; 128: 181–186.
28. Rau M, Schiller M, Krienke S, et al. Clinical manifestations but not cytokine profiles differentiate adult-onset still's disease and sepsis. *J Rheumatol* 2010; 37: 2369–2376.