



# The rate of muscle wasting in liver transplant recipients on waiting list: post-transplant outcomes and associated serum metabolite patterns

Zachary P. Rokop<sup>1^</sup>, Thomas M. O'Connell<sup>2,3</sup>, Taylor Munsch<sup>1</sup>, Lauren Nephew<sup>4</sup>, Eric Orman<sup>4</sup>, Plamen Mihaylov<sup>1</sup>, Richard S. Mangus<sup>1</sup>, Chandrashekhara Kubal<sup>1</sup>

<sup>1</sup>Division of Transplant Surgery, Department of Surgery, Indiana University School of Medicine, Indianapolis, IN, USA; <sup>2</sup>Department of Otolaryngology, Head & Neck Surgery, Indiana University School of Medicine, Indianapolis, IN, USA; <sup>3</sup>Indiana Center for Musculoskeletal Health, Indiana University School of Medicine, Indianapolis, IN, USA; <sup>4</sup>Division of Gastroenterology and Hepatology, Department of Medicine, Indiana University School of Medicine, Indianapolis, IN, USA

**Contributions:** (I) Conception and design: C Kubal, RS Mangus, TM O'Connell; (II) Administrative support: C Kubal, RS Mangus, TM O'Connell; (III) Provision of study materials or patients: All authors; (IV) Collection and assembly of data: All authors; (V) Data analysis and interpretation: All authors; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

**Correspondence to:** Zachary P. Rokop, MD. Division of Transplant Surgery, Department of Surgery, Indiana University School of Medicine, 545 Barnhill Drive, Emerson Hall 105, Indianapolis, IN 46202, USA. Email: zrokop@indiana.edu.

**Background:** Sarcopenia at the time of liver transplantation (LT) is an established risk factor for mortality following LT. However, most studies in this context have defined sarcopenia by one-time, static measurements. The aims of this study were (I) to determine the impact of the rate of muscle loss in waitlisted LT recipients on post-LT outcomes and (II) to identify patterns of serum metabolites associated with patients with more progressive sarcopenia.

**Methods:** Patients undergoing liver transplant from 2008 to 2018 who received more than one computed tomography (CT) scans within 12 months prior to liver transplant were included (n=61). The psoas muscle index (PMI) was calculated using Slice-O-Matic software and corrected for patient height (m<sup>2</sup>). Patients were classified into two groups based the rate of reduction in PMI—high wasting [HW; change in PMI ( $\Delta$ PMI)  $\leq$ -1%/month] and low wasting (LW;  $\Delta$ PMI  $>$ -1%/month). Pre-transplant serum metabolic profiles were collected using nuclear magnetic resonance (NMR) spectroscopy. Living kidney donor sera was used as healthy controls.

**Results:** Median  $\Delta$ PMI was -2.0%/month in HW and -0.15%/month in LW patients (P<0.001). Post-transplant 1-year mortality was significantly higher in HW patients. There were no significant differences in metabolite concentrations between HW and LW patients. However, perturbations in taurine, sarcosine, betaine and the aromatic amino acids (AAAs), were observed in patients with liver disease as compared to healthy controls. Liver disease was also associated with a decrease in lipoprotein profiles, especially high-density lipoprotein (HDL) particles.

**Conclusions:** In patients undergoing LT, the rate of progression of sarcopenia is a strong prognostic indicator of post-LT death. Serum metabolite profiles were not characteristically unique to HW patients, and most closely resemble derangements associated with chronic liver disease.

**Keywords:** Liver transplantation (LT); sarcopenia; psoas muscle index (PMI); metabolome

Submitted Dec 17, 2023. Accepted for publication May 22, 2024. Published online Aug 22, 2024.

doi: 10.21037/hbsn-23-645

**View this article at:** <https://dx.doi.org/10.21037/hbsn-23-645>

<sup>^</sup> ORCID: 0000-0002-5733-4330.

## Introduction

Sarcopenia is the significant loss of muscle mass as the result of normal aging or an underlying pathologic process (1). A strong link exists between sarcopenia and liver disease, with a prevalence between 40% and 70% in end-stage liver disease (ESLD) patients (2). Within the cirrhotic population, sarcopenia has been associated with increased hospitalizations, disease-related complications, and health care costs (3,4). Additionally, sarcopenia demonstrates significant influence on liver transplantation (LT) outcomes, contributing to increased waitlist mortality, post-transplant mortality, post-transplant infection, and length of post-operative hospitalization (5-7). While standard metrics [Model for End-Stage Liver Disease (MELD) and Child-Pugh scores] are utilized for prognostic and organ allocation purposes, studies have shown that these measures of liver dysfunction do not correlate with sarcopenia (8). Therefore, assessment for sarcopenia is important for identification of patients whose risk may not be adequately captured by conventional means.

Current hypotheses suggest sarcopenia in the cirrhotic population is the result of an imbalance between muscle synthesis and muscle breakdown (9). ESLD promotes a proteolytic state, providing substrate for gluconeogenesis as a way to compensate for impaired glycogen storage of the

cirrhotic liver, and for use in ammonia detoxification (10). As a result, ESLD patients have lower levels of circulating branched-chain amino acids (BCAAs) as compared to healthy controls (11). Liver failure is also associated with an increase in circulating aromatic amino acids (AAAs) due to a decrease in AAA breakdown in the liver (12). The ratio of the concurrent decrease in circulating BCAAs and increase in AAAs is called Fischer's index and is an indicator of liver dysfunction. Muscle breakdown may be further accentuated by increased levels of the pro-inflammatory cytokines tumor necrosis factor (TNF)- $\alpha$ , interleukin (IL)-1, and IL-6, which have been associated with upregulation of the ubiquitin-proteasome pathway (UPP) (13). Additionally, cirrhotic patients have been identified as having lower caloric intake and impaired absorption of fats and vitamins (10). Taken together, these observations suggest that the depletion of building block metabolites necessary for maintenance of skeletal muscle mass may explain the sarcopenia of liver disease. However, the wide range of muscle wasting observed in cirrhotic populations suggests that this process remains poorly understood (7).

The quantification of sarcopenia in patients is most effectively performed through imaging modalities. Dual-energy X-ray absorptiometry (DEXA) has been utilized in numerous studies examining age-related changes in body composition, however this technique is limited by anasarca or ascites commonly seen in liver patients (14). As an alternative, single slice axial computed tomography (CT) techniques have been identified as an accurate measure of skeletal muscle mass, visceral fat, and subcutaneous fat. These measurements have been subsequently validated as predictors of total body composition and nutritional status in the LT population (15-17). Studies performed using these methods have thus far been limited by utilizing one-time measurements of body composition, and few account for changes in lean muscle mass that occur over the course of a patient's liver disease (18). We hypothesized that the rate of muscle wasting influenced post-LT outcomes more so than one time measurement of sarcopenia.

Here, utilizing serial CT scan assessments of sarcopenia, we demonstrate the importance of the rate of muscle wasting to LT outcomes. Additionally, with metabolomic analysis, we examine whether serum metabolites profiles in patients with relatively rapid progression of wasting differed from other patients. We present this article in accordance with the STROBE reporting checklist (available at <https://hbsn.amegroups.com/article/view/10.21037/hbsn-23-645/rc>).

### Highlight box

#### Key findings

- The rate of progression of sarcopenia is a strong prognostic indicator of post-liver transplantation (LT) mortality.
- Serum metabolite profiles of patients with rapid progression of sarcopenia most closely resemble derangements associated with chronic liver disease.

#### What is known and what is new?

- Sarcopenia at the time of LT is an established risk factor for mortality following LT, but most studies in this context have defined sarcopenia by one-time, static measurements.
- The aims of this study were (I) to determine the impact of the rate of muscle loss in waitlisted LT recipients on post-LT outcomes and (II) to identify patterns of serum metabolites associated with patients with more progressive sarcopenia.

#### What is the implication and what should change now?

- Metabolic perturbations identified herein suggest that nutritional interventions aimed at maintaining muscle mass may benefit liver transplant patients irrespective of the rate of sarcopenic progression.

## Methods

### *Patient selection and clinical data collection*

All liver transplant recipients between 2008 and 2018 were retrospectively identified within the prospectively maintained Indiana University Liver Transplant database (n=1,513). From this group, patients who underwent at least two CT scans within a period of 12 months and at least 4 months apart prior to transplantation, were identified (n=129). Of the 129 subjects who met these criteria, 61 patients were selected pursuant to available clinical follow-up. Demographic and clinical outcomes data was obtained from the electronic medical records. Additionally, 10 living-related kidney donor (LKD) patients were identified for use as healthy controls for metabolite analyses. All research was conducted in accordance with both the Declarations of Helsinki (as revised in 2013) and Istanbul. Analysis of patient data for this study was reviewed and approved by the Institutional Review Board of the Indiana University School of Medicine (IRB #1305011413) and individual consent for this retrospective analysis was waived.

### *Psoas muscle measurement and interpretation*

The psoas muscle index (PMI) was utilized as a surrogate for sarcopenia as previously described (15,19). PMI is defined as the cross-sectional area of right and left psoas muscle at the level of L3/L4 relative to the patient's height squared ( $\text{cm}^2/\text{m}^2$ ). To obtain the psoas measurements, patients' CT imaging was reviewed using Slice-O-Matic software. The area of right and left psoas muscles area was measured at the most superior aspect of L3/L4 using the freehand drawing function, and the areas were summed to obtain the total psoas muscle area. Total psoas area was normalized by each patient's height squared to generate the PMI. To minimize human error in CT measurements, each image was analyzed by three individual research assistants who had been trained in interpreting abdominal CT images and Slice-O-Matic software. Research assistants were blinded to patients, their outcomes, and the measurements made by other assistants. Psoas area measurements were compared between reviewers. Any individual measurement that differed by >15% of the mean was re-examined until a consensus was reached. The mean of the three psoas measurements for each patient was used to calculate the PMI (17).

Median rate of progression was  $-0.5\%$  change PMI/

month. There was no difference in post-LT survival when using this cutoff to define median high wasting ( $\text{HW}_{\text{med}}$ ) vs. median low wasting ( $\text{LW}_{\text{med}}$ ) (Figure S1). Therefore, we performed an additional subset analysis of patients with extreme progression of sarcopenia, as defined as >1% decrease per month. Patients with >1% decrease in PMI per month were considered to be very HW (VHW) and patients with <1% decrease/month PMI change were considered to be LW.

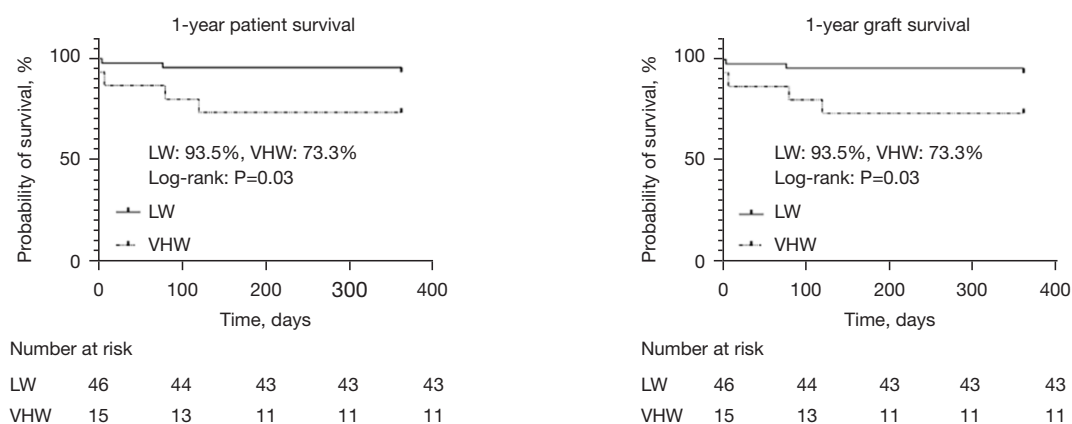
### *Serum nuclear magnetic resonance (NMR) metabolomic analysis*

As part of standard care, serum samples from all recipients and living donors are archived prior to transplant/donation at our institution. Serum from 36 LT recipients was available in sufficient quantity for this analysis. We elected to use the median change in PMI ( $\Delta\text{PMI}$ ) of the cohort ( $-0.5\%$ /month) to delineate between  $\text{HW}_{\text{med}}$  and  $\text{LW}_{\text{med}}$  for metabolomics analysis. The final sample set included 15 patients in the  $\text{HW}_{\text{med}}$  group, 21 in the  $\text{LW}_{\text{med}}$  group, and 10 in the LKD group. We performed a similar subset analysis as well, comparing metabolite profiles in VHW patients against LW and LKD controls.

For NMR-based metabolomics, 100 mL aliquots of serum samples were diluted with 300 mL of deuterated phosphate buffer (pH 7.4). The solution was filtered through a 10 kDa molecular weight cutoff filter to remove proteins and lipoproteins. To the filtrate was added 60 mL of a Chenomx reference solution containing 5.0 mM, 2,2-dimethyl-2-silapentane-5-sulfonate-d<sub>6</sub> (DSS-d<sub>6</sub>) sodium salt as an internal chemical shift and quantitation standard. The total volume of each sample was brought to 600 mL with deuterated phosphate buffer.

NMR data were acquired on a Bruker Avance III 700 MHz NMR spectrometer with a triple-resonance (TXI) probe operating at 25 °C. Spectra were collected with a 1D NOESY pulse sequence covering 12 ppm. The spectra were digitized with 32,768 points during a 3.9-second acquisition time. The mixing time was set to 100 ms and the relaxation delay between scans was set to 2.0 seconds.

The data were processed using the Advanced Chemistry Development Spectrus Processor (version 2016.1, Toronto, Canada). The spectra were zero filled to 65,536 points, apodised using a 0.3 Hz decaying exponential function and fast Fourier transformed. Automated phase correction and linear baseline correction were applied to all samples.



**Figure 1** One-year patient and graft survival curves, comparing VHW and LW cohorts. LW, low wasting; VHW, very high wasting.

Metabolite concentrations were quantified using the Chenomx NMR Suite (version 8.2, Edmonton, Canada). The DSS-d6 was used as a chemical shift and quantification reference for all spectra and was set to a chemical shift of 0.00 and a concentration of 0.5 mM. Quantitative fitting of each spectrum was carried out in batch mode, followed by manual adjustment for some spectra to correct for errors arising from spectral overlap.

### NMR lipoprotein analysis

As the liver plays a major role in lipoprotein metabolism, we measured a range of lipoprotein particle concentrations. Determination of lipoprotein particle subfractions by NMR was assessed using the NMR-based platform at LipoScience (now LabCorp, Morrisville, NC, USA). Lipoprotein particle numbers and sizes were determined using a version of the LP2 algorithm modified for murine plasma (20). A total of 21 lipoprotein subclasses were measured.

### Statistical analysis

Patient demographic and outcomes data is expressed as medians with interquartile ranges (IQRs) unless otherwise specified. Student's *t*-test, analysis of variance (ANOVA), chi-squared, and nonparametric hypothesis testing was employed as relevant. Metabolomics data was analyzed using Viime using ANOVA with Tukey post-hoc analysis (21). Volcano plots were generated in R. All hypotheses were tested at a significance level of 0.05 using two-tailed tests.

## Results

### Patient demographics and clinical outcomes—median $\Delta$ PMI cutoff

In total, 61 patients were identified with >1 CT scan obtained prior to LT. Median time between CT scans was 8.3 months (IQR, 4.7, 11 months). Thirty patients were classified as  $HW_{med}$  and 31 patients as  $LW_{med}$  according to their rate of wasting as compared to the median of the study population. Demographic variables were similar between these two groups (Table S1). Importantly, there was no significant difference in post-LT survival outcomes. The 1-year patient and graft survival rates were statistically similar between  $HW_{med}$  and  $LW_{med}$  groups on Kaplan-Meier survival curve analysis (Figure S1: 86.7% vs. 93.5%,  $P=0.38$ ).

### Patient demographics and clinical outcomes—subgroup analysis ( $\Delta$ PMI <−1%)

While using the median  $\Delta$ PMI as a cutoff resulted in even cohorts, this classification did not identify any significant differences in post-transplant outcomes. We therefore sought to identify a  $\Delta$ PMI cutoff that was associated with a difference in post-LT survival. We found that patients with a decrease in PMI of >1%/month had significantly worse 1-year graft and patient survival rates as compared to the rest of the study population (73.3% vs. 93.5%,  $P=0.03$ ; Figure 1). After identifying this inflection point, we proceeded to perform a subset analysis. A group of 15 patients were classified as VHW and 31 patients as LW using these criteria. Demographic variables are summarized in Table 1. Patients in each group were of similar age, sex, and MELD

**Table 1** Patient demographics comparing patients with very high rates of muscle wasting (>1% decrease in PMI/month) to the rest of the study population

Variables	Wasting group		P value
	VHW (n=15)	LW (n=46)	
Age at Txp (years)	63 [61, 66]	63.5 [61, 66]	0.74
Sex (female)	3 (20.0)	13 (28.3)	0.53
Race (Caucasian)	13 (86.7)	40 (87.0)	
MELD at Txp	22 [21, 29]	21.5 [16.7, 23]	0.08
Pre-LT GFR (mL/min)	>60 [43.3, >60]	>60 [47, >60]	0.72
Etiology liver disease			0.30
NASH	6 (40.0)	18 (39.1)	
Alcoholic	2 (13.3)	12 (26.1)	
Idiopathic	5 (33.3)	14 (30.4)	
Hepatitis C	2 (13.3)	1 (2.2)	
Malignancy	0	1 (2.2)	
History of ascites	14 (93.3)	31 (67.4)	0.04*
History of hepatic encephalopathy	14 (93.3)	31 (67.4)	0.04*
History of diabetes	6 (40.0)	23 (50.0)	0.50
History of tobacco use	11 (73.3)	28 (60.9)	0.38
Waitlist days	26 [6, 72]	68.5 [20.8, 122]	0.03*
HCC on explant pathology	9 (60.0)	17 (37.0)	0.12
BMI at transplant (kg/m <sup>2</sup> )	25.2 [21.7, 27.3]	29.8 [26.4, 34]	0.002*
Initial PMI (cm <sup>2</sup> /m <sup>2</sup> )	6.11 [4.9, 7.8]	6.2 [4.2, 7.5]	0.93
PMI at transplant (cm <sup>2</sup> /m <sup>2</sup> )	4.2 [3.7, 6.3]	6.4 [4.3, 7.5]	0.04*
ΔPMI (% change PMI/month)	-2.0 [-3.4, -1.3]	-0.15 [-0.65, 0.9]	<0.001*
Months between CT scans	9.4 [4.4, 12.3]	7.9 [4.9, 10.7]	0.74
Serum albumin at transplant (g/dL)	2.6 [2.2, 2.9]	3.1 [2.6, 3.4]	0.03*
Serum protein at transplant (g/dL)	5.2 [3.8, 6.0]	6.3 [5.4, 6.8]	0.11

Data are presented as median [range] or n (%). \*, P<0.05. PMI, psoas muscle index; VHW, very high wasting; LW, low wasting; Txp, transplant; MELD, Model for End-Stage Liver Disease; LT, liver transplantation; GFR, glomerular filtration rate; NASH, non-alcoholic steatohepatitis; HCC, hepatocellular carcinoma; BMI, body mass index; ΔPMI, change in PMI; CT, computed tomography.

score at time of transplant. Etiology of liver failure in each group was similar, as was the proportion of patients with hepatocellular carcinoma (HCC) on explant pathology. The proportion of VHW patients with pre-operative ascites or hepatic encephalopathy was significantly greater than in the LW group. Median ΔPMI was -2.0%/month in VHW and -0.15%/month in LW patients (P<0.001). Body mass index (BMI) (25.2 vs. 29.8 kg/m<sup>2</sup>, P=0.002) and PMI (4.2 vs. 6.4 cm<sup>2</sup>/m<sup>2</sup>, P=0.04) at time of transplant were significantly

lower in the HW group. These patients also had significantly lower pre-transplant serum albumin levels as compared to the rest of the study (2.6 vs. 3.1 g/dL, P=0.03). Patients in the HW group spent significantly less time on the waitlist (26 vs. 68.5 days, P=0.03).

Table 2 summarizes post-operative outcomes in this subset analysis. There were no significant differences in rates of early allograft dysfunction, primary nonfunction, or early post-operative complications, as characterized

**Table 2** Post-operative outcomes comparing patients with very high rates of muscle wasting (>1% decrease in PMI/month) to the rest of the study population

Variables	Wasting group		P value
	VHW (n=15)	LW (n=46)	
Peak serum ALT (U/L)	479 [167, 867]	457 [236, 856]	0.73
Peak serum AST (U/L)	563 [254, 1,021]	729 [358, 1,463]	0.37
Early allograft dysfunction	1 (6.7)	10 (21.7)	0.19
Primary nonfunction	0	0	>0.99
Acute kidney injury	6 (40.0)	20 (43.5)	0.81
Clavien-Dindo complication grade			0.34
Grade 1	3 (20.0)	10 (21.7)	
Grade 2	1 (6.7)	8 (17.4)	
Grade 3	4 (26.7)	6 (13.0)	
Grade 4	4 (26.7)	7 (15.2)	
Grade 5	3 (20.0)	2 (4.3)	
Hospitalization duration (days)	9 [7, 13]	9.5 [7, 15.3]	0.81
ICU duration (days)	3 [2, 9]	3 [2, 5.8]	0.98
Days on ventilator	1 [0, 2]	1 [0, 2]	0.78
Reintubation	2 (13.3)	2 (4.3)	0.22
Tracheostomy	1 (6.7)	2 (4.3)	0.72
90-day readmission	7 (46.7)	22 (47.8)	0.94
Discharge disposition	n=12	n=43	0.28
Home	7 (58.3)	32 (74.4)	
Rehab	5 (41.7)	11 (25.6)	
1-year graft survival (%)	73.3	93.5	0.03*
1-year patient survival (%)	73.3	93.5	0.03*

Data are presented as median [range], n (%), or percentage. \*, P<0.05. PMI, psoas muscle index; VHW, very high wasting; LW, low wasting; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ICU, intensive care unit.

according to the Clavien-Dindo classification scheme. Patients in both groups also had similar durations of hospitalization and intensive care unit (ICU) admission. As mentioned above, 1-year patient and graft survival rates in VHW patients were significantly reduced.

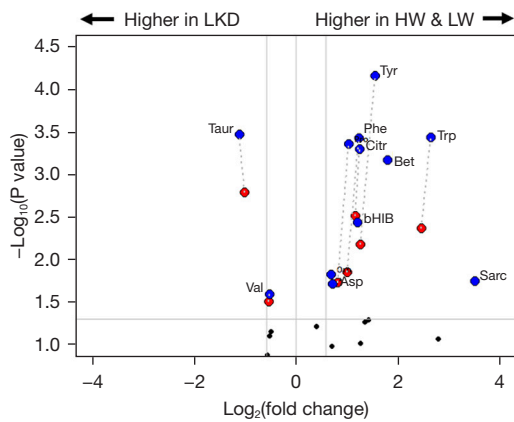
### **Serum metabolomic analysis—median $\Delta$ PMI cutoff**

Serum metabolomics analysis quantified 43 metabolites. Our initial analysis compared metabolite profiles between HW<sub>med</sub> and LW<sub>med</sub> groups, as defined by the median  $\Delta$ PMI cutoff. ANOVA revealed that 12 metabolites demonstrated

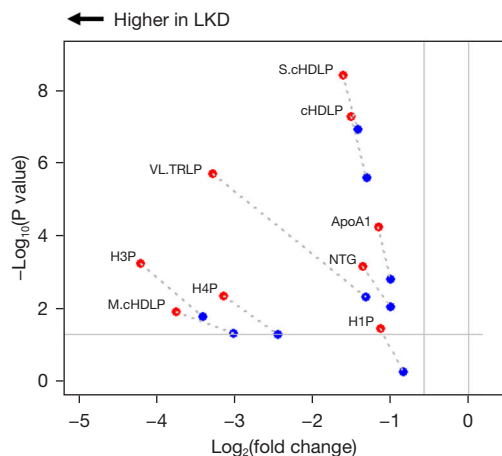
statistically different levels between at least two groups. With the application of Tukey post-hoc testing, it was revealed that all 12 were different between the LW<sub>med</sub> and LKD groups and that seven were significantly different between the HW<sub>med</sub> and LKD groups. No significant differences were observed between the HW<sub>med</sub> and LW<sub>med</sub> group. A table containing the fold changes and significance values between each of the groups is presented in Table S2. The significant differences are presented in the volcano plot in Figure 2.

### **Perturbations to serum amino acids**

The metabolomics platform included measurements for



**Figure 2** Volcano plot of metabolomics data comparing HW<sub>med</sub> (red) and LW<sub>med</sub> (blue) *vs.* LKD. The plot shows the metabolites which have  $-\log_{10}P$  value  $>1.3$ , which is equal to an unadjusted P value of  $<0.05$ . Dashed lines connect the 14 metabolites that were significantly altered in both groups. For clarity, only the blue dots are labeled. LKD, living-related kidney donor; HW, high wasting; LW, low wasting; Tyr, tyrosine; Taur, taurine; Phe, phenylalanine; Trp, tryptophan; Citr, citrate; Bet, betaine; bHIB, b-hydroxyisobutyrate; Asp, aspartate; Sarc, sarcosine; Val, valine; HW<sub>med</sub>, median HW; LW<sub>med</sub>, median LW.



**Figure 3** Volcano plot of lipidomics data comparing HW<sub>med</sub> (red) and LW<sub>med</sub> (blue) *vs.* LKD. The plot shows the metabolites which have  $-\log_{10}P$  value  $>1.3$ , which is equal to an unadjusted P value of  $<0.05$ . Dashed lines connect the lipoprotein measurements that were altered in both the HW and LW groups. For clarity, only the HW<sub>med</sub> (red) dots were labeled. LKD, living-related kidney donor; S.cHDL, small cHDL; cHDL, calibrated high-density lipoprotein particle; VL.TRLP, very large triglyceride-rich lipoprotein; ApoA1, apolipoprotein A1; NTG, triglyceride; H3P, high-density lipoprotein 3; H4P, high-density lipoprotein 4; M.cHDL, medium cHDL; H1P, high-density lipoprotein 1; HW<sub>med</sub>, median HW; HW, high wasting; LW<sub>med</sub>, median LW; LW, low wasting.

17 of the canonical, proteogenic amino acids. Of these, six showed a significant difference in at least one of the liver disease groups. Proline was significantly increased in both the LW<sub>med</sub> and HW<sub>med</sub> groups as compared to healthy controls. Aspartate was increased only in the LW<sub>med</sub> group compared to the LKD group. The three AAAs—phenylalanine, tryptophan, and tyrosine—demonstrated significant increases in both liver disease groups compared with the LKD group. The only amino acid that showed a reduction in the liver disease group was the BCAA valine. Consistent with this finding, there was also a trend towards reduction noted for leucine (LW<sub>med</sub> *vs.* LKD:  $P=0.07$ ; HW<sub>med</sub> *vs.* LKD:  $P=0.08$ ). The most significantly downregulated metabolite was taurine with a more than two-fold reduction in both the HW<sub>med</sub> and LW<sub>med</sub> groups as compared to LKD.

### Perturbations to one-carbon metabolism

Several metabolites associated with one-carbon metabolism showed distinct elevations in the liver disease groups. One-carbon metabolism involves the folate-mediated movement of single carbon units *i.e.*, methyl groups to facilitate a range of metabolic processes. This overall pathway includes the folate cycle, methionine salvage, and purine biosynthesis. Sarcosine showed the largest increase of any of these metabolites with a three-fold increase in the LW<sub>med</sub> group as compared to healthy controls ( $P=0.001$ ). There was also a nearly two-fold increase in the sarcosine levels in the HW<sub>med</sub> group compared to LKD, however this difference was not statistically significant ( $P=0.06$ ). Interestingly, the difference in sarcosine levels between HW<sub>med</sub> and LW<sub>med</sub> patients approached statistical significance ( $P=0.058$ ). Betaine levels were significantly increased in HW<sub>med</sub> and LW<sub>med</sub> patients as compared to LKD. The amino acids tryptophan and methionine are also involved in the one-carbon metabolism pathway. Significant differences were observed between both liver disease groups and the LKDs for tryptophan and methionine.

### Changes in serum lipoprotein profiles

There were significant decreases in seven of the lipoprotein particle concentrations in patients with liver disease compared to healthy controls. Additionally, the concentrations of total triglycerides and apolipoprotein (Apo)A1 were decreased as well. These results are shown in *Figure 3* with raw data supplied in *Table S3*. Only the concentration of the very large triglyceride rich particles showed a difference between the HW<sub>med</sub> and LW<sub>med</sub> groups. The other lipoprotein particle

changes all involved decreases in high-density lipoprotein (HDL) particles as compared to healthy controls. Consistent with this was a significant reduction in ApoA1, the major protein component of HDL particles.

#### *Serum metabolomic analysis—subgroup analysis* ( $\Delta\text{PMI} < -1\%$ )

As we identified significantly worse 1-year survival in VHW patients, we performed an additional analysis of the metabolomics data including a separate group for the VHW. In this analysis, the initial HW group was separated into VHW (n=6) and HW (n=9). Table S4 shows the P values of all of the inter-group comparisons in this new analysis. The list of altered metabolites was nearly identical to the original analysis with the inclusion of one extra metabolite, a-hydroxyisovalerate (aHIV). As shown in Table S2, the P value for aHIV in the first analysis (comparing HW<sub>med</sub> and LW<sub>med</sub>) was 0.0511 and thus very nearly significant.

## Discussion

The aim of this study was two-fold: (I) to establish the clinical relevance that the rate of muscle loss has on post-LT outcomes and (II) to identify patterns in small molecule metabolism unique to patients with rapidly progressive sarcopenia. While multiple studies have demonstrated the negative impact that sarcopenia has on post-LT outcomes (5-8), few have examined the rate at which this muscle loss develops. Single slice, axial CT imaging has been validated as an accurate measure of total body composition, and given the ubiquity of the technology, is an attractive tool for measurement of changes in body composition over time (15-17). Here we demonstrate that patients with more rapid muscle wasting (i.e., a decrease in PMI of >1%/month) had significantly worse 1-year patient and graft survival. Hanai *et al.* examine the relationship between the rate of muscle loss and mortality in cirrhotic patients (18). However, in this study, patients had a median percent change in PMI of -2.2%/year, and no outcomes after LT were considered. Our present study examines a population of patients with much more rapid muscle wasting (median  $\Delta\text{PMI}$  in HW group = -2%/month), and specifically analyzes outcomes following LT. Our findings suggest that dynamic assessment of sarcopenia can identify a high-risk subset of patients who may need prioritization or careful consideration in terms of liver transplant candidacy to avoid futility.

Interestingly, there were no differences in early post-

operative complications that might explain this difference in mortality between VHW and LW in our study. This suggests that the detrimental effects of rapid muscle wasting remain relevant even if patients progress through an uneventful perioperative course. Previous studies into the impact of sarcopenia on post-LT survival outcomes demonstrate that short- (1-year) and long-term (3-year) patient survival is inferior in patients with pre-LT sarcopenia (5). Additionally, pre-operative sarcopenia has been found to persist in the majority of post-LT patients, further supporting this observation (22,23). These studies used static measures of sarcopenia; long-term follow-up in our cohort can identify if similar trends exist in rapid wasters. It should be noted that our overall 1-year survival rate for the entire study population was 89% which is fairly in-line with published outcomes in the United States according to the most recent OPTN annual report (92.8% 1-year survival) (24). This 1-year survival rate improved to 93.5% when VHW patients were removed from the analysis.

The second part of the study focused on the serum metabolite profiles of patients with varying rates of muscle loss. Our goal was to search for metabolic perturbations that are associated with accelerated muscle wasting. A number of metabolomics analyses have been carried out on liver diseases and have identified specific metabolic perturbations associated with non-alcoholic fatty liver disease (NAFLD), cirrhosis, non-alcoholic steatohepatitis (NASH), etc. (25,26). These metabolic profiles have been utilized as a noninvasive screening tool for liver disease such as NASH and hepatic fibrosis (25). In this study we expected that some of these same metabolic perturbations would be observed in the liver disease subjects. Our hypothesis was that rate of muscle wasting in liver disease patients with sarcopenia would also influence the metabolome and that some of these perturbations would be detectable in addition to changes induced by liver disease. Such distinct changes could potentially serve as markers for patients who are at high risk from their sarcopenia for poor outcomes following LT. In our study, we did not identify specific changes in metabolites between HW and LW patients. However, we did find that, while metabolic perturbations generally appeared similar to those found in chronic liver disease, some distinct differences were also identified. These metabolic profiles, discussed ahead, indicated that the muscle metabolic status may indeed be influencing the circulating metabolome, and may suggest therapeutic interventions based on metabolic or nutritional supplementation.

A well-documented metabolic characteristic of liver



cirrhosis is the decrease in BCAAs and an accompanying increase in AAAs (27,28). The reduction in BCAAs occurs primarily in the skeletal muscle, where these amino acids are utilized in reactions that form glutamate from  $\alpha$ -ketoglutarate. The latter is a critical metabolite in the detoxification of ammonia that occurs with liver disease (29). The observed increase in AAA increase is due to the decreased ability of the diseased liver to metabolize these amino acids. A decrease in BCAAs is also observed in age-related muscle wasting (30-32). A potentially confounding factor to quantifying serum BCAAs is dietary intake. As mentioned earlier, cirrhotic patients often have reduced caloric intake. BCAA metabolism is very sensitive to changes in the amount and composition of food ingested, which is highly variable in both healthy and diseased states. Brief periods of starvation can acutely increase the circulating concentrations of BCAA (33) as a result of increased proteolysis and reduced protein synthesis (34). This makes the assessment of a patient's true BCAA status difficult to ascertain. Although recent clinical trials have shown some improvement in muscle mass with BCAA supplementation (35), the major body of evidence in this field remains inconclusive (36). This discrepancy suggests that careful patient selection may be necessary for BCAA supplementation to achieve clinical benefits.

The impact of liver disease on one-carbon metabolism has been well-established (37). The liver is a major site of one-carbon metabolism, which plays a critical role in the transfer of one-carbon units through the folate cycle for remethylation of homocysteine. The metabolite betaine acts as a methyl donor in the remethylation of homocysteine to methionine. This reaction is catalyzed by the enzyme betaine hydroxymethyltransferase (BHMT). Teng *et al.* demonstrated that mice lacking BHMT have elevated liver betaine and homocysteine and rapidly develop HCC (38). The elevated betaine levels in the liver disease groups of our study suggest a similar dysregulation in BHMT. Conversely, in a study of sarcopenia in a cohort of community-dwelling Chinese adults, serum betaine levels were inversely correlated with low lean muscle mass (39). In our study, the betaine levels in the HW group were not different from the LKD. It is possible that the expected increase in betaine from the patient's liver disease is offset in the HW group by a reduction in muscle mass.

Sarcosine also plays a role in the methionine remethylation pathway through the conversion of S-adenosylmethionine to S-adenosylhomocysteine. This reaction is catalyzed by the enzyme glycine-N-methyltransferase (GNMT) which also converts glycine to sarcosine. This significant

elevation in sarcosine in the LW group may be related to a specific alteration to GNMT activity or may be the result of an upstream perturbation in BHMT. Interestingly, there was only a trending increase in the methionine levels in the LW group. As with betaine, the sarcosine levels are significantly increased in the LW group, but not in the HW group. Furthermore, sarcosine levels are very close to being significantly different between HW and LW. Currently, the literature on sarcosine and sarcopenia is rather sparse. A study of circulating amino acids in older persons with physical frailty found an increase in circulating sarcosine associated with physical frailty and sarcopenia (40). A reduction in circulating sarcosine was associated with aging in both rodents and humans (41). Although there were no significant differences in the ages of the HW and LW groups, it is interesting to consider that the HW group may have distinct metabolic features of aging that contribute to the observed sarcosine levels.

Taurine is a non-proteogenic amino acid and is the most abundant free amino acid in skeletal muscle. It is involved in a wide range of functions including the response to oxidative stress and inflammation (42). Taurine levels are maintained in the liver by both endogenous biosynthesis and exogenous transport, and are typically decreased in liver diseases (43). A significant decrease in taurine levels is also observed in age-associated sarcopenia (44,45). Our findings are consistent with this knowledge. It is interesting to note that therapeutic supplementation with taurine has been studied to treat metabolic imbalances in both liver diseases and sarcopenia (46,47) and thus taurine supplementation may be beneficial in liver disease patients irrespective of the degree and rate of sarcopenia.

The liver plays a critical role in lipoprotein metabolism, contributing functions such as lipid synthesis and transport. Patients with cirrhosis often have several defects in lipid metabolism, such as hypocholesterolemia in association with reduced levels of HDL cholesterol (HDL-C), low-density lipoprotein cholesterol (LDL-C), and ApoA1 (48). HDL levels have been shown to decrease with increasing severity of liver disease (40,49). Our results showed a consistent decrease in HDLs as measured by the HDL particle numbers (HDL-P). A similar reduction in HDLs has been observed in a study of sarcopenic patient with chronic obstructive pulmonary disease (COPD) with HDL levels being an independent predictor of sarcopenia in both controls and COPD patients (50).

It should be noted that this study was performed at a single institution and includes a relatively small sample size.

Although our program performed over 1,500 transplants during the study period, only 61 patients qualified. This was mainly a result of the fact that the average waitlist time for a liver transplant at our center is less than 4 months, which means that many patients did not have the requisite >4-month gap between successive CT scans. Due to the scope of this study, a number of social determinants of health (i.e., socioeconomic class, insurance status, access to nutritious foods, etc.) were not accounted for. These may indirectly influence the progression of muscle loss in future transplant recipients and attempts to control for these factors should be made in future studies.

## Conclusions

The rate of progression of sarcopenia in ESLD patients awaiting transplant is a strong prognostic indicator of worse post-transplant survival. Metabolite profiles did not differ between HW and LW patients, but the overall patterns most closely resembled derangements associated with chronic liver disease. While we were unable to identify a “high risk” metabolic profile, metabolic perturbations identified herein suggest that nutritional interventions aimed at maintaining muscle mass may benefit liver transplant patients irrespective of the rate of sarcopenic progression.

## Acknowledgments

We would like to thank Dr. Peter Thompson at the Molecular Education, Technology and Research Innovation Center (METRIC) at NC State University for collection of the NMR data.

*Funding:* None.

## Footnote

*Reporting Checklist:* The authors have completed the STROBE reporting checklist. Available at <https://hbsn.amegroups.com/article/view/10.21037/hbsn-23-645/rc>

*Data Sharing Statement:* Available at <https://hbsn.amegroups.com/article/view/10.21037/hbsn-23-645/dss>

*Peer Review File:* Available at <https://hbsn.amegroups.com/article/view/10.21037/hbsn-23-645/prf>

*Conflicts of Interest:* All authors have completed the ICMJE

uniform disclosure form (available at <https://hbsn.amegroups.com/article/view/10.21037/hbsn-23-645/coif>). The authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All research was conducted in accordance with both the Declarations of Helsinki (as revised in 2013) and Istanbul. Analysis of patient data for this study was reviewed and approved by the Institutional Review Board of the Indiana University School of Medicine (IRB #1305011413) and individual consent for this retrospective analysis was waived.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

## References

1. Fuggle N, Shaw S, Dennison E, et al. Sarcopenia. *Best Pract Res Clin Rheumatol* 2017;31:218-42.
2. Scaglione S, Kliethermes S, Cao G, et al. The Epidemiology of Cirrhosis in the United States: A Population-based Study. *J Clin Gastroenterol* 2015;49:690-6.
3. Amaral TF, Matos LC, Tavares MM, et al. The economic impact of disease-related malnutrition at hospital admission. *Clin Nutr* 2007;26:778-84.
4. Hanai T, Shiraki M, Nishimura K, et al. Sarcopenia impairs prognosis of patients with liver cirrhosis. *Nutrition* 2015;31:193-9.
5. Englesbe MJ, Patel SP, He K, et al. Sarcopenia and mortality after liver transplantation. *J Am Coll Surg* 2010;211:271-8.
6. Montano-Loza AJ, Meza-Junco J, Baracos VE, et al. Severe muscle depletion predicts postoperative length of stay but is not associated with survival after liver transplantation. *Liver Transpl* 2014;20:640-8.
7. Tandon P, Ney M, Irwin I, et al. Severe muscle depletion in patients on the liver transplant wait list: its prevalence

- and independent prognostic value. *Liver Transpl* 2012;18:1209-16.
8. Montano-Loza AJ, Meza-Junco J, Prado CM, et al. Muscle wasting is associated with mortality in patients with cirrhosis. *Clin Gastroenterol Hepatol* 2012;10:166-73, 173.e1.
  9. Sinclair M, Gow PJ, Grossmann M, et al. Review article: sarcopenia in cirrhosis--aetiology, implications and potential therapeutic interventions. *Aliment Pharmacol Ther* 2016;43:765-77.
  10. Dichi JB, Dichi I, Maio R, et al. Whole-body protein turnover in malnourished patients with child class B and C cirrhosis on diets low to high in protein energy. *Nutrition* 2001;17:239-42.
  11. Dam G, Sørensen M, Buhl M, et al. Muscle metabolism and whole blood amino acid profile in patients with liver disease. *Scand J Clin Lab Invest* 2015;75:674-80.
  12. Soeters PB, Fischer JE. Insulin, glucagon, aminoacid imbalance, and hepatic encephalopathy. *Lancet* 1976;2:880-2.
  13. Lin SY, Chen WY, Lee FY, et al. Activation of ubiquitin-proteasome pathway is involved in skeletal muscle wasting in a rat model with biliary cirrhosis: potential role of TNF-alpha. *Am J Physiol Endocrinol Metab* 2005;288:E493-501.
  14. Shaw KA, Srikanth VK, Fryer JL, et al. Dual energy X-ray absorptiometry body composition and aging in a population-based older cohort. *Int J Obes (Lond)* 2007;31:279-84.
  15. Shen W, Punyanitya M, Wang Z, et al. Total body skeletal muscle and adipose tissue volumes: estimation from a single abdominal cross-sectional image. *J Appl Physiol* (1985) 2004;97:2333-8.
  16. Giusto M, Lattanzi B, Albanese C, et al. Sarcopenia in liver cirrhosis: the role of computed tomography scan for the assessment of muscle mass compared with dual-energy X-ray absorptiometry and anthropometry. *Eur J Gastroenterol Hepatol* 2015;27:328-34.
  17. Bush WJ, Davis JP, Maluccio MA, et al. Computed Tomography Measures of Nutrition in Patients With End-Stage Liver Disease Provide a Novel Approach to Characterize Deficits. *Transplant Proc* 2018;50:3501-7.
  18. Hanai T, Shiraki M, Ohnishi S, et al. Rapid skeletal muscle wasting predicts worse survival in patients with liver cirrhosis. *Hepatol Res* 2016;46:743-51.
  19. Braunschweig CA, Sheean PM, Peterson SJ, et al. Exploitation of diagnostic computed tomography scans to assess the impact of nutrition support on body composition changes in respiratory failure patients. *JPEN J Parenter Enteral Nutr* 2014;38:880-5.
  20. Jeyarajah EJ, Cromwell WC, Otvos JD. Lipoprotein particle analysis by nuclear magnetic resonance spectroscopy. *Clin Lab Med* 2006;26:847-70.
  21. Choudhury R, Beezley J, Davis B, et al. Viime: Visualization and Integration of Metabolomics Experiments. *J Open Source Softw* 2020;5:2410.
  22. Lai JC, Segev DL, McCulloch CE, et al. Physical frailty after liver transplantation. *Am J Transplant* 2018;18:1986-94.
  23. Bhanji RA, Takahashi N, Moynagh MR, et al. The evolution and impact of sarcopenia pre- and post-liver transplantation. *Aliment Pharmacol Ther* 2019;49:807-13.
  24. Kwong AJ, Ebel NH, Kim WR, et al. OPTN/SRTR 2021 Annual Data Report: Liver. *Am J Transplant* 2023;23:S178-263.
  25. Caussy C, Ajmera VH, Puri P, et al. Serum metabolites detect the presence of advanced fibrosis in derivation and validation cohorts of patients with non-alcoholic fatty liver disease. *Gut* 2019;68:1884-92.
  26. Wang X, Rao B, Wang H, et al. Serum metabolome alterations in patients with early nonalcoholic fatty liver disease. *Biosci Rep* 2022;42:BSR20220319.
  27. Dasarathy S, Hatzoglou M. Hyperammonemia and proteostasis in cirrhosis. *Curr Opin Clin Nutr Metab Care* 2018;21:30-6.
  28. Dejong CH, van de Poll MC, Soeters PB, et al. Aromatic amino acid metabolism during liver failure. *J Nutr* 2007;137:1579S-1585S; discussion 1597S-1598S.
  29. Holeček M, Kandar R, Sispera L, et al. Acute hyperammonemia activates branched-chain amino acid catabolism and decreases their extracellular concentrations: different sensitivity of red and white muscle. *Amino Acids* 2011;40:575-84.
  30. Lu Y, Karagounis LG, Ng TP, et al. Systemic and Metabolic Signature of Sarcopenia in Community-Dwelling Older Adults. *J Gerontol A Biol Sci Med Sci* 2020;75:309-17.
  31. Meng L, Yang R, Wang D, et al. Specific lysophosphatidylcholine and acylcarnitine related to sarcopenia and its components in older men. *BMC Geriatr* 2022;22:249.
  32. Yamada M, Kimura Y, Ishiyama D, et al. Plasma Amino Acid Concentrations Are Associated with Muscle Function in Older Japanese Women. *J Nutr Health Aging* 2018;22:819-23.
  33. Holeček M. Branched-chain amino acids in health and

- disease: metabolism, alterations in blood plasma, and as supplements. *Nutr Metab (Lond)* 2018;15:33.
34. Holeček M, Mičuda S. Amino acid concentrations and protein metabolism of two types of rat skeletal muscle in postprandial state and after brief starvation. *Physiol Res* 2017;66:959-67.
  35. Siramolpiwat S, Limthanetkul N, Pornthisarn B, et al. Branched-chain amino acids supplementation improves liver frailty index in frail compensated cirrhotic patients: a randomized controlled trial. *BMC Gastroenterol* 2023;23:154.
  36. van Dijk AM, Bruins Slot AS, Portincasa P, et al. Systematic review with meta-analysis: Branched-chain amino acid supplementation in liver disease. *Eur J Clin Invest* 2023;53:e13909.
  37. da Silva RP, Eudy BJ, Deminice R. One-Carbon Metabolism in Fatty Liver Disease and Fibrosis: One-Carbon to Rule Them All. *J Nutr* 2020;150:994-1003.
  38. Teng YW, Mehedint MG, Garrow TA, et al. Deletion of betaine-homocysteine S-methyltransferase in mice perturbs choline and 1-carbon metabolism, resulting in fatty liver and hepatocellular carcinomas. *J Biol Chem* 2011;286:36258-67.
  39. Huang BX, Zhu YY, Tan XY, et al. Serum betaine is inversely associated with low lean mass mainly in men in a Chinese middle-aged and elderly community-dwelling population. *Br J Nutr* 2016;115:2181-8.
  40. Calvani R, Picca A, Marini F, et al. A Distinct Pattern of Circulating Amino Acids Characterizes Older Persons with Physical Frailty and Sarcopenia: Results from the BIOSPHERE Study. *Nutrients* 2018;10:1691.
  41. Walters RO, Arias E, Diaz A, et al. Sarcosine Is Uniquely Modulated by Aging and Dietary Restriction in Rodents and Humans. *Cell Rep* 2018;25:663-676.e6.
  42. Lambert IH, Kristensen DM, Holm JB, et al. Physiological role of taurine--from organism to organelle. *Acta Physiol (Oxf)* 2015;213:191-212.
  43. Miyazaki T, Matsuzaki Y. Taurine and liver diseases: a focus on the heterogeneous protective properties of taurine. *Amino Acids* 2014;46:101-10.
  44. Miyamoto K, Hirayama A, Sato Y, et al. A Metabolomic Profile Predictive of New Osteoporosis or Sarcopenia Development. *Metabolites* 2021;11:278.
  45. Yeung SSY, Zhu ZLY, Kwok T, et al. Serum Amino Acids Patterns and 4-Year Sarcopenia Risk in Community-Dwelling Chinese Older Adults. *Gerontology* 2022;68:736-45.
  46. Barbiera A, Sorrentino S, Lepore E, et al. Taurine Attenuates Catabolic Processes Related to the Onset of Sarcopenia. *Int J Mol Sci* 2020;21:8865.
  47. Zhang Z, Liu D, Yi B, et al. Taurine supplementation reduces oxidative stress and protects the liver in an iron-overload murine model. *Mol Med Rep* 2014;10:2255-62.
  48. Rubiés-Prat J, Masdeu S, Nubiola AR, et al. High-density lipoprotein cholesterol and phospholipids, and apoprotein A in serum of patients with liver disease. *Clin Chem* 1982;28:525-7.
  49. Poynard T, Abella A, Pignon JP, et al. Apolipoprotein AI and alcoholic liver disease. *Hepatology* 1986;6:1391-5.
  50. Abu-Libdeh W, Khrais J, Suwan L, et al. Relation of Plasma High-Density Lipoproteins-Cholesterol with Sarcopenia in Patients with Chronic Obstructive Pulmonary. *Indiana J Respir Care* 2022;11:327-32.

**Cite this article as:** Rokop ZP, O'Connell TM, Munsch T, Nephew L, Orman E, Mihaylov P, Mangus RS, Kubal C. The rate of muscle wasting in liver transplant recipients on waiting list: post-transplant outcomes and associated serum metabolite patterns. *HepatoBiliary Surg Nutr* 2024;13(6):962-973. doi: 10.21037/hbsn-23-645