

RESEARCH ARTICLE

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Exosomal cell-to-cell transmission of alpha synuclein oligomers

Karin M Danzer^{1,3}, Lisa R Kranich¹, Wolfgang P Ruf¹, Ozge Cagsal-Getkin^{1,2}, Ashley R Winslow¹, Liya Zhu¹, Charles R Vanderburg^{1,2} and Pamela J McLean^{1,4*}

Abstract

Background: Aggregation of alpha-synuclein (asyn) and resulting cytotoxicity is a hallmark of sporadic and familial Parkinson's disease (PD) as well as dementia with Lewy bodies, with recent evidence implicating oligomeric and pre-fibrillar forms of asyn as the pathogenic species. Recent *in vitro* studies support the idea of transcellular spread of extracellular, secreted asyn across membranes. The aim of this study is to characterize the transcellular spread of asyn oligomers and determine their extracellular location.

Results: Using a novel protein fragment complementation assay where asyn is fused to non-bioluminescent amino-or carboxy-terminus fragments of humanized Gaussia Luciferase we demonstrate here that asyn oligomers can be found in at least two extracellular fractions: either associated with exosomes or free. Exosome-associated asyn oligomers are more likely to be taken up by recipient cells and can induce more toxicity compared to free asyn oligomers. Specifically, we determine that asyn oligomers are present on both the outside as well as inside of exosomes. Notably, the pathway of secretion of asyn oligomers is strongly influenced by autophagic activity.

Conclusions: Our data suggest that asyn may be secreted via different secretory pathways. We hypothesize that exosome-mediated release of asyn oligomers is a mechanism whereby cells clear toxic asyn oligomers when autophagic mechanisms fail to be sufficient. Preventing the early events in asyn exosomal release and uptake by inducing autophagy may be a novel approach to halt disease spreading in PD and other synucleinopathies.

Keywords: Alpha synuclein, Oligomers, Exosomes, Parkinson's disease, Aggregation, Secretion

Background

Parkinson's disease (PD) is pathologically characterized by alpha-synuclein (α syn) immunopositive intracellular deposits termed Lewy bodies [1]. Gene multiplication of the asyn gene [2,3] and missense mutations [4-6] are linked to familial forms of PD. Together, these data support a role for asyn in the pathogenesis of PD.

Because α syn inclusion body pathology associated with PD occurs in a hierarchical distribution with its epicenter in the brainstem, then extends to the mesolimbic cortex and associated areas [7], Braak et al. have suggested that α syn pathology spreads gradually throughout the neuraxis as PD progresses [8]. However, as yet, the

underlying mechanisms of disease progression in PD remain to be determined.

The main component of Lewy bodies and Lewy neurites are fibrillar aggregates of asyn [1] but a growing body of evidence suggests that prefibrillar oligomers of asyn are key contributors in the progression of Parkinson's disease [9-15].

Until recently asyn was thought to exert its toxic effects intracellularly. However, this concept was challenged when El-Agnaf et al. detected asyn species in human plasma and CSF [12]. Furthermore, Desplats et al. demonstrated that asyn can be directly transmitted from neuronal cells overexpressing asyn to transplanted embryonic stem cells both in tissue culture and in transgenic animals [16]. Concurrently, our group was able to demonstrate that cell produced asyn oligomers are secreted and taken up by neighboring cells where they have detrimental consequences [17]. These results suggest that the pathogenic action of asyn oligomers are

* Correspondence: mclean.pamela@mayo.edu

¹MassGeneral Institute for Neurodegenerative Disease, Department of Neurology, Massachusetts General Hospital, Charlestown, MA, USA

⁴Present address: Department of Neuroscience, Mayo Clinic, 4500 San Pablo Rd, Jacksonville, FL 32224, USA

Full list of author information is available at the end of the article

not limited to the donor cells but can extend into the extracellular space and affect neighboring cells. In support of this hypothesis, recombinant α syn oligomers added to cell culture medium have been shown to be internalized by recipient cells causing either cell death or seeding of α syn [18-22]. The mechanism(s) of α syn transmission from cell to cell that contribute to the spread of α syn pathology remain largely unknown. One intriguing question is how intracellularly generated α syn is released into the extracellular space. A first hint that α syn may be secreted by externalized vesicles that have hallmarks of exosomes was recently provided [23,24].

The aim of this current study is to characterize α syn associated with exosomes and to explore the nature of α syn secretion using a highly sensitive protein complementation assay [25-30]. Moreover, we examine the specific relationship of α syn oligomers with exosomes and find that both intra- and extra-exosomal associated α syn oligomers exist. The importance of intact exosomes for re-uptake of α syn oligomers into neighboring cells and the role of autophagic activity on exosomal secretion of α syn oligomers are also examined.

Results

Alpha-synuclein oligomers are found in exosomes

Increasing evidence suggests that α syn can be released by neurons and neuronal like cells [17,31-33] although extracellular α syn and its pathological relevance are still hotly debated in the field. Recent work from our own group and the elegant study from Desplats et al. suggest that α syn can be transferred from cell to cell and thus may provide an explanation for the spread of α syn pathology in PD patients [16,17]. However, little is known about the mechanism of α syn secretion.

Recently, secretion of α syn in association with membrane vesicles, identified as exosomes based on their composition and biophysical properties, has been described [23,24]. However, the specific α syn species (monomers vs pathogenic oligomers) secreted with exosomes and the location of α syn remains to be determined.

To investigate whether oligomeric species of α syn are present in the exosome enriched fractions we employed a bioluminescent protein-fragment complementation assay [25,26,30]. In this strategy, α syn was fused to non-bioluminescent amino-terminal (S1) or carboxy-terminal fragments (S2) of *Gaussia princeps* luciferase [28] that can reconstitute when brought together by α syn- α syn interactions [25], thus providing a readout of α syn oligomerization (Additional file 1: Figure S1A). The same principle of protein complementation with fluorescent venus YFP was used generating the fluorescent protein-fragment complementation pair V1S or SV2 whereby N-terminal half of Venus YFP is fused to the N-terminus of α syn (V1S) and C-terminal half of Venus

YFP is fused to the C-terminus of α syn (SV2) [25] (Additional file 1: Figure S1B).

Human H4 neuroglioma cells were co-transfected with S1 and S2 that reconstitute luciferase activity upon α syn oligomerization. Exosomes were isolated from conditioned media (CM) of H4 cells using an established sub-cellular fractionation methodology [34,35] and the exosomal pellet was analyzed for luciferase activity that is indicative of α syn oligomers. Interestingly, we witnessed a large increase in luciferase activity in the exosomal fraction derived from H4 cells transfected with S1 and S2 compared to exosomes from mock transfected cells (Figure 1A), suggesting that α syn, and specifically α syn oligomers are present in the exosomal fraction. To exclude the possibility that N- or C-terminal fragments of human Gaussia Luciferase can interfere with protein sorting in exosomes, our results were verified in exosomes isolated from human H4 cells transfected with untagged wild-type (wt) α syn using a human α syn ELISA. Significant levels of α syn were present in the exosomal fraction from wt α syn and S1/S2 transfected H4 cells compared to exosomes derived from empty vector (mock) transfected cells (Figure 1B).

We extended these observations to primary cortical neurons where α syn oligomers were also found in the exosomal fraction isolated from primary neurons co-transduced with adeno-associated virus (AAV) encoding S1 (AAV-S1) and S2 (AAV-S2) as demonstrated by a significant increase in luciferase activity compared to exosomes isolated from naive neurons (Figure 1C). In accord with the experiments performed in H4 cells, we also confirmed the presence of α syn in exosomes derived from primary neurons infected with a variety of different AAV constructs encoding either α syn-ires-GFP, AAV-S1 and AAV-S2 or α syn-venusYFP fluorescent protein-fragment complementation pair (AAV8-V1S or AAV8-SV2) (Figure 1D) using an α syn ELISA. Taken together, our data provide evidence that α syn oligomers are present in the exosomal fractions from both neurons and non-neuronal cells.

Characterization of exosomes

To confirm the presence of exosomes, fractions from both primary neurons and H4 cells were subjected to SDS-PAGE and immunoblotting. All exosomal fractions were found to be immunopositive for the exosome-specific proteins alix and flotillin, whereas the 'exosome-free' supernatant was immuno-negative for alix and flotillin (Figure 2A).

Moreover, exosome-enriched fractions isolated from CM of H4 cells transfected with the α syn complementation pair S1 and S2 were also analyzed using electron microscopy and demonstrated the distinctive vesicular morphological structures characteristic of exosomes

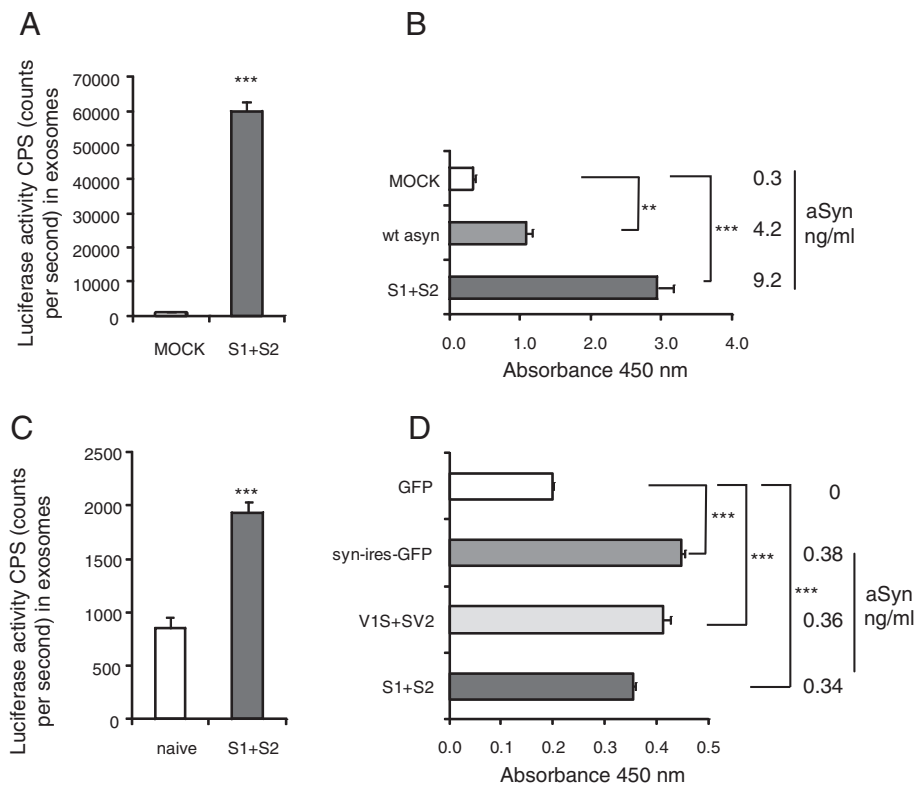


Figure 1 Extracellular α Syn oligomers are associated with exosomes: (A) Exosomal fractions from human H4 cells transfected with asyn complementation pair S1 and S2 contain high amounts of asyn oligomers, analyzed with a luciferase assay. Signal corresponds to 1.25x100mm dishes. $n=6$, unpaired t test with Welch's correction, $*p < 0.001$ (B) asyn ELISA analysis from exosomal fractions from human H4 cells transfected with S1/S2, wt asyn or Mock. Signal corresponds to 1x100 mm dish. $n=4$, unpaired t test with Welch's correction, $***p < 0.001$ (C) Exosomal fractions from primary neurons infected with asyn complementation pair S1 and S2 contain high amounts of asyn oligomers, analyzed with a luciferase assay, signal corresponds to 1x60 mm dish, $n=4$, unpaired t test with Welch's correction, $***p < 0.001$ (D) asyn ELISA analysis from exosomal fractions from primary neurons infected with AAV expressing GFP, asyn -ires-GFP, V1S/SV2 and S1/S2. Values correspond to 1x60 mm dish, $n=4$, Tukey's Multiple Comparison Test, $***p < 0.001$.**

(Figure 2B). Immuno-electron microscopy with an antibody against the exosomal marker CD63 [36], confirmed characteristic exosomal vesicles typically 60–100 nm in size in exosome enriched fractions from CM of primary neurons co-transduced with AAV expressing the asyn complementation pair V1S or SV2 (Figure 2C).

Because microRNAs (miRs) have been found in exosomes [37], miR profiling is a powerful tool to definitively characterize exosomes. Exosome fractions from both S1/S2 transfected H4 cells and primary neurons transduced with AAV- α syn-ires-GFP were found to contain a large number of miRs that have previously been reported to be present in exosomes (see Tables 1 and 2). Of interest, we did not detect miR-7, which has been previously identified as a negative regulator of α syn expression [66].

Localization of α syn oligomers in the extracellular space

Cytosolic proteins can be secreted from cells via at least two distinct pathways which include exocytosis and

fusion of multi-vesicular bodies with the plasma membrane to release exosomes. Defining the localization of α syn in the extracellular space will provide insight into the mechanisms and pathways involved in α syn release. To examine the localization of α syn oligomers in the extracellular space we first digested exosome-enriched fractions containing α syn S1/S2 oligomers with 0.25% trypsin. Interestingly, trypsin digestion significantly reduced luciferase activity in the exosome fraction by 62% (Figure 3A) suggesting the presence of α syn oligomers either on the external surface of the exosomes or outside of exosomes. However, trypsin treatment did not eliminate luciferase activity to background levels completely, indicating that α syn oligomers must exist in the lumen of the exosomes that are insensitive to the trypsin treatment. To further clarify, we cotreated exosome fractions with trypsin and the detergent saponin (0.1%). The presence of saponin results in exosome-membrane permeabilization. Notably, we saw a complete elimination of luciferase activity in the presence of trypsin and

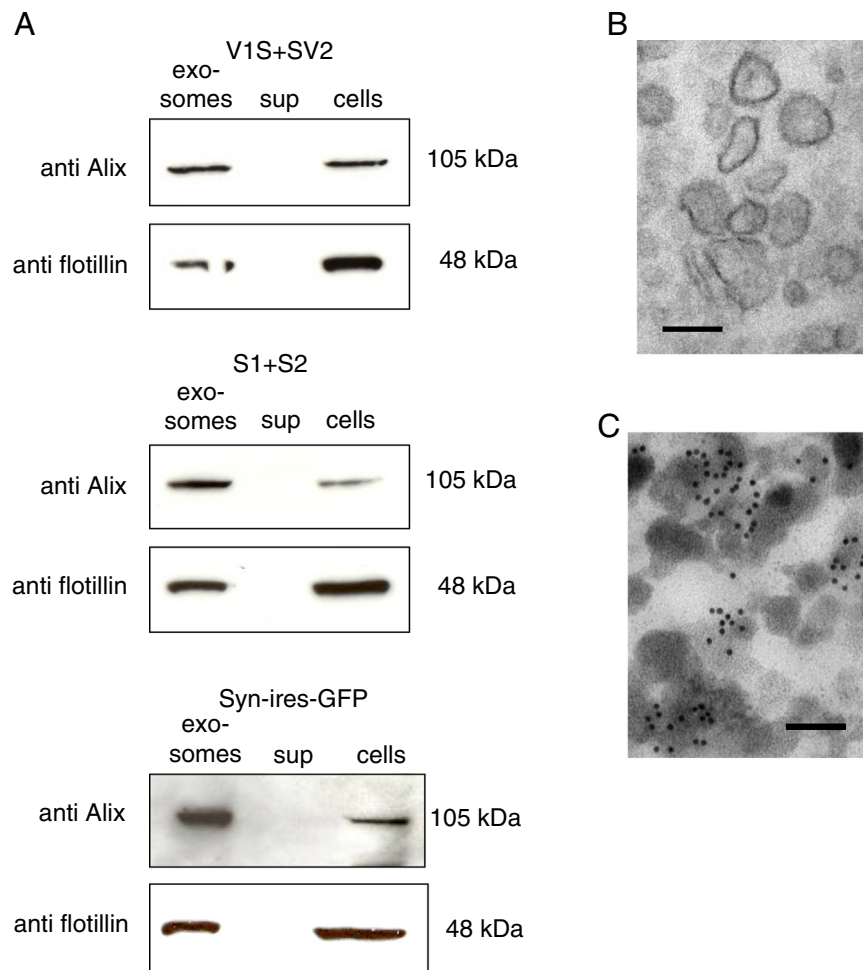


Figure 2 Characterization of exosomes: (A) Exosomal pellets derived from primary neurons infected with AAV-V1S/SV2, AAV-S1/S2, AAV- α syn-ires-GFP were resuspended in 1xPBS and analyzed by Western blotting using exosome specific antibodies anti Alix and anti flotillin. Also total cell lysates and exosome free supernatant (sup) were loaded as positive and negative controls, respectively. (B) Exosomes from S1/S2 transfected H4 cells were prepared as described before. After fixation vesicles were negatively stained with 2% uranyl acetate and observed under electron microscopy. Scale bar, 100 nm. (C) Exosomes from primary neurons infected with AAV-V1S/SV2 were negatively stained with 2% uranyl acetate and immunolabeled with CD63 antibody as exosomal marker. Scale bar, 100 nm.

saponin (Figure 3A). Treatment with saponin alone slightly increased luciferase activity compared to untreated control exosomes, although it was not a significant increase. This could be due to enhanced substrate availability to luminal α syn oligomers. The same experimental paradigm was tested on the "exosome-free" supernatant fraction. As expected, trypsin eliminated all luciferase activity from "free" α syn oligomers in the supernatant fraction in the presence or absence of saponin (Figure 3B). These data confirm the absence of exosomes from the supernatant fraction and verify that the experimental paradigm is sufficient to digest all available α syn oligomers.

To confirm our results on the localization of α syn oligomers inside/outside exosomes we examined samples

prepared under the same experimental conditions using dot blot immunoblotting. Probing with Syn-1 antibody showed that exosome free α syn oligomers (sup) were completely digested by trypsin independent of saponin treatment (lane 1, sup, Figure 3C). In contrast, Syn-1 signal was not completely eliminated when exosome fractions were treated with trypsin (lane 2, exo, Figure 3C). Only the combination of trypsin and saponin resulted in a complete digestion of α syn oligomers and a consequent abolishment of α syn immunostaining in exosome fractions. Probing with an antibody against the exosomal marker CD63, which is known to be located solely on the outside of exosomes, shows reactivity only in the exosome fractions not treated with trypsin and no reactivity at all in supernatant-associated α syn oligomers.

Table 1 MicroRNA profiling of alpha-synuclein containing exosomes from H4 cell culture media

miRNAs	function	remarks	Key citations
7b	Inhibition of cell cycle progression and growth of melanoma cells	present in exosomes	[38,39]
7i	regulatory responses to microbial infection	present in exosomes	[40,41]
18b	exocytosis, angiogenesis, hematopoiesis, tumorigenesis	present in exosomes	[40]
182	Repression of tumor suppressors	present in exosomes	[39,42]
184	Block of NFAT1 protein expression	present in exosomes	[40,43]
363*	-		[44,45]
373*	Involvement in tumor migration and invasion	present in exosomes	[39,46]
375	exocytosis, angiogenesis, hematopoiesis, tumorigenesis	present in exosomes	[39,40]
376*	-		[47,48]
424	controlling the macrophage differentiation program	present in exosomes	[39,49]
431	-	CNS specific miRNA	[47,50]
455	-		[51]
487a	Role in angiogenesis and cell aging		[52]
492	-		[44]
494	regulation of PTEN expression and functions as a micro-oncogene in carcinogenesis	present in exosomes	[39,53]
518c	-		[44,54]

Dot blots were also performed on fractions prepared from CM of cells transfected with wt untagged α syn. As expected, trypsin treatment resulted in a reduction in Syn-1 signal in exosome associated α syn oligomers but

only the combination of trypsin and saponin resulted in a complete digestion and abolishment of Syn-1 signal. Together, the data indicate that α syn oligomers are located on the inside and outside of exosomes.

Table 2 MicroRNA profiling of alpha-synuclein containing exosomes from primary cortical neuron media

miRNAs	function	remarks	Key citations
27a	activating the expression of P-glycoprotein	present in exosomes	[39] [55]
28	inhibitor of thrombopoietin receptor translation	present in exosomes	[39] [56]
34a	suppression of cell proliferation through modulation of the E2F signaling pathway	-	[57]
106b	modulation of TGF β signaling in tumors	present in exosomes	[39] [58]
184	Block of NFAT1 protein expression	present in exosomes	[40,43]
185	Induction of cell cycle arrest in lung cancer cell lines	present in exosomes	[40,59]
192	affects cellular proliferation through the p53-miRNA circuit	present in exosomes	[39] [60]
199b	Regulation of protein phosphatase 2A inhibitor) in human choriocarcinoma	-	[61]
302b*	regulatory mechanism in tuning stem cell properties	-	[62]
373*	Involvement in tumor migration and invasion	present in exosomes	[39,46]
422b	-	present in exosomes	[40]
431	-	CNS specific miRNA	[47,50]
448	-	-	[63]
455	-	-	[51]
487a	Role in angiogenesis and cell aging	-	[52]
491	decreases cell viability by induction of apoptosis	-	[64]
493-3p	-	-	[54]
518*	Predicted to be involved in Huntington's disease	-	[54,65]
522	-	present in exosomes	[39]
526a	-	present in exosomes	[39]
526b	-		

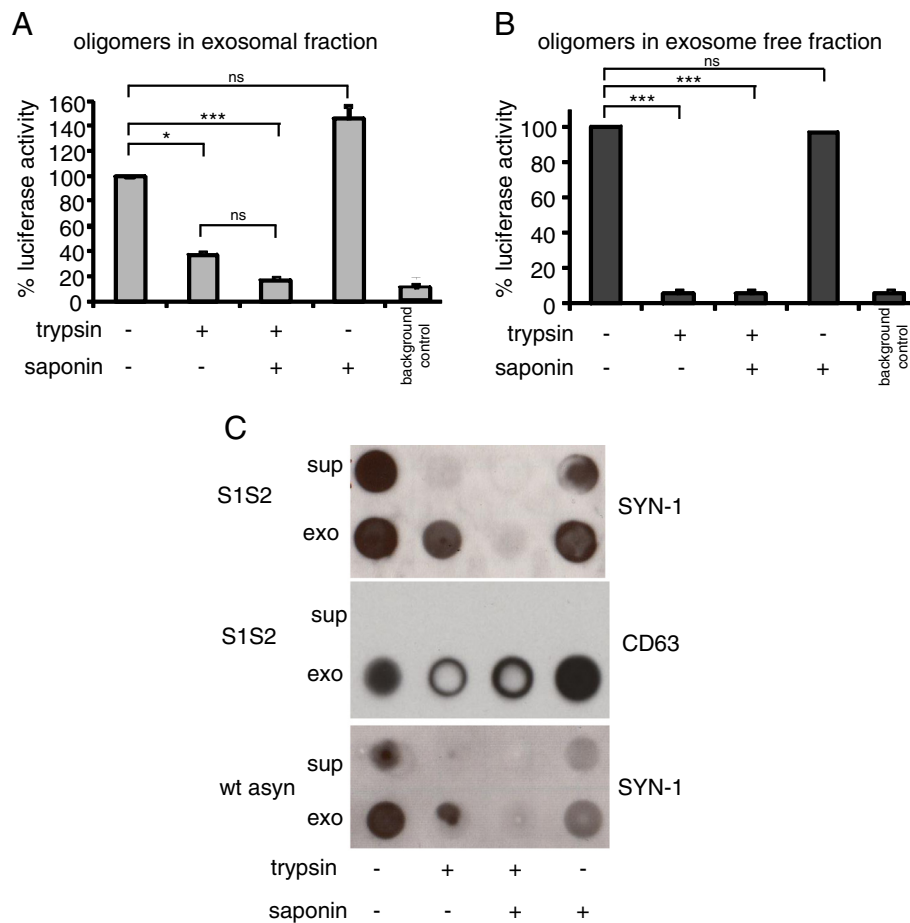


Figure 3 Localization of asyn oligomers within exosomes: (A) Luciferase assay with exosomes derived from human H4 cells transfected with S1/S2 or 0.1% saponin/or 0.1% saponin. (B) Luciferase assay with exosome-free supernatants after asyn oligomer digestion using 0.25% trypsin and/or 0.1% saponin. (C) Dot blot approach with exosomal fraction (exo) and exosome free supernatant (sup) from H4 cells transfected with S1/S2 after asyn oligomer digestion using 0.25% trypsin and/or 0.1% saponin. Probing with syn-1 antibody shows only a complete digestion of asyn oligomers when trypsin and saponin are used concurrently. Signal of exosomal marker CD63 is abolished by addition of solely trypsin.

Exosome-associated asyn oligomers are more prone to internalization than exosome-free asyn oligomers

It has been reported that recombinant asyn or asyn oligomers can be internalized by cells and result in various cellular effects [18-20,33,67]. Furthermore, we and others have shown that cell produced asyn oligomers can be secreted and taken up by proliferating cells and primary neurons [17,23]. To investigate if exosomes are required for the internalization of asyn oligomers, we exposed naive H4 cells to exosome-associated asyn oligomers or exosome-free supernatant containing asyn oligomers derived from S1/S2 transfected H4 cells for 3 days. Concurrently, exosomes or exosome-free supernatant from "mock" transfected cells were added to naive H4 cells. Interestingly, we found that exosome associated asyn oligomers are more prone to being taken up than exosome free asyn oligomers (Figure 4A). To control for the variable amounts of

asyn in each exosome or supernatant preparation added to cells, the luciferase signal detected in the recipient cells was normalized back to the initial luciferase counts added to the naive cells. Data analysed in this way revealed a 2.4 fold increase in uptake of exosome-associated asyn oligomers compared to exosome-free asyn oligomers (Figure 4B).

Recombinant oligomers as well as physiologically secreted asyn oligomers can cause cell death when applied to culture medium of different cell lines and primary neurons [18-20,23,33,67]. To determine if exosome-associated asyn oligomers confer more cytotoxicity compared to exosome-free asyn oligomers, we applied exosome-enriched fractions or exosome-free fractions derived from S1/S2 or MOCK transfected H4 cells to naive proliferating H4 cells and found an increase in Caspase 3/7 activation conferred by exosome associated asyn oligomers (Figure 4C). To ensure the

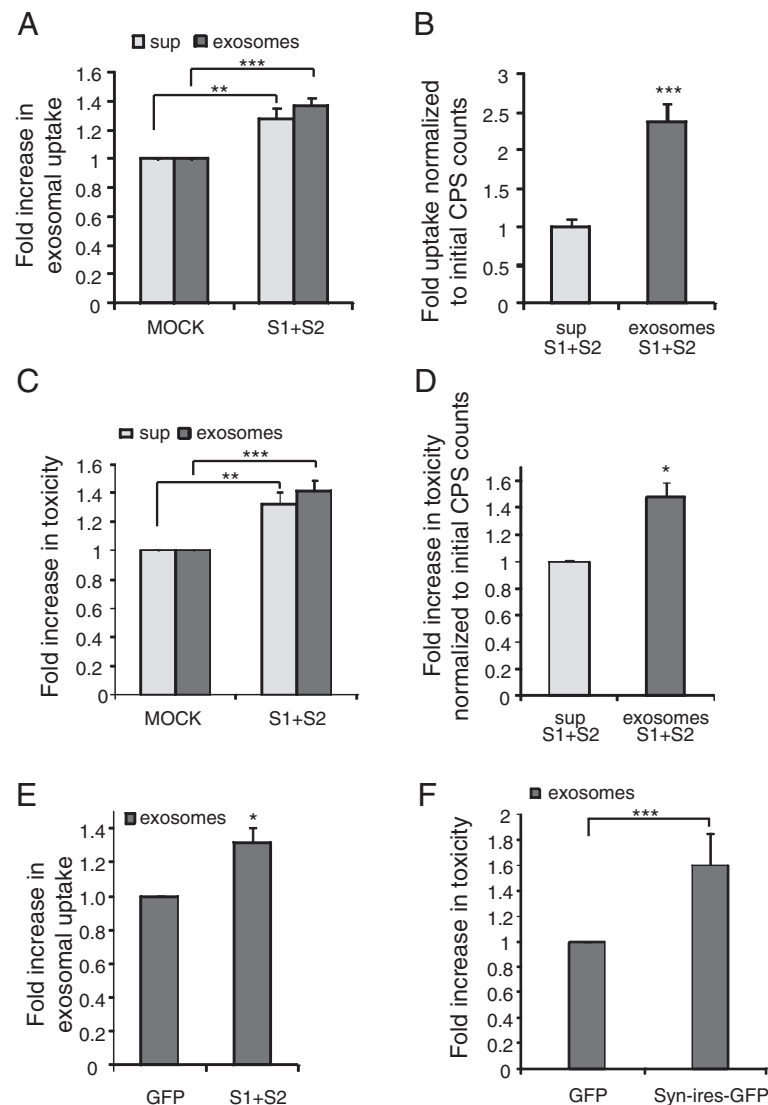


Figure 4 Exosome-associated asyn oligomers are preferentially internalized and more toxic than exosome-free asyn oligomers. (A) Exosomal fractions and exosome-free supernatants (sup) from human H4 cells transfected with S1/S2 or mock were applied to naïve H4 cells and incubated for 3 days. Uptake of asyn oligomers into recipient cells was measured by luciferase assay. $n = 9$, unpaired t test with Welch's correction, $***p < 0.001$. **(B)** Naïve H4 cells treated with exosome-associated S1/S2 oligomers and exosome-free S1/S2 oligomers (sup) were assayed for luciferase activity. Luciferase signal was normalized to the amount of asyn oligomers in the input, $n = 9$, unpaired t test with Welch's correction, $***p < 0.001$. **(C)** H4 cells treated with exosome-associated asyn fractions conferred greater toxicity on naïve H4 cells than exosome-free asyn (sup $n = 5$, unpaired t test with Welch's correction, $***p < 0.001$) **(D)** Naïve H4 cells treated with exosome-associated asyn oligomers and exosome free asyn oligomers (sup) were assayed for toxicity. Level of toxicity was normalized to the luciferase activity in each input $n = 5$, Mann Whitney test, $*p < 0.05$. **(E)** Exosomal fractions derived from primary neurons infected with AAV-S1/S2 or AAV-GFP were applied to naïve neurons and incubated for 3 days. Uptake of asyn oligomers into recipient neurons was measured performing a luciferase assay on recipient cells. $n = 3$, one-sample t-test, $*p < 0.05$. **(F)** Toxicity assay of naïve primary neurons treated with exosome-enriched fractions and exosome-free fractions (sup) derived from either AAV-asyn-ires-GFP or AAV-GFP infected primary neurons. $n = 9$, one sample t-test, $***p < 0.001$.

same amount of asyn oligomers in each fraction, the level of Caspase 3/7 activation was normalized to the amount of asyn oligomers prior to the addition to naïve cells. Interestingly, a significant 1.5-fold increase in Caspase3/7 activation and resulting apoptosis induction from exosome-associated asyn oligomers compared to exosome-free asyn oligomers was detected (Figure 4D).

In accordance with our data for human H4 cells we confirmed that exosome-associated asyn oligomers could also be taken-up by naïve primary neurons (Figure 4E) and induce apoptosis as characterized by an increase in caspase3/7 activity (Figure 4F). Unfortunately, due to high levels of non-specific background bioluminescence from B-27 supplement in our neuronal

cell culture medium, we were unable to assess the internalization of exosome-free asyn oligomers by primary neurons.

Exosomes need to be intact to be internalized

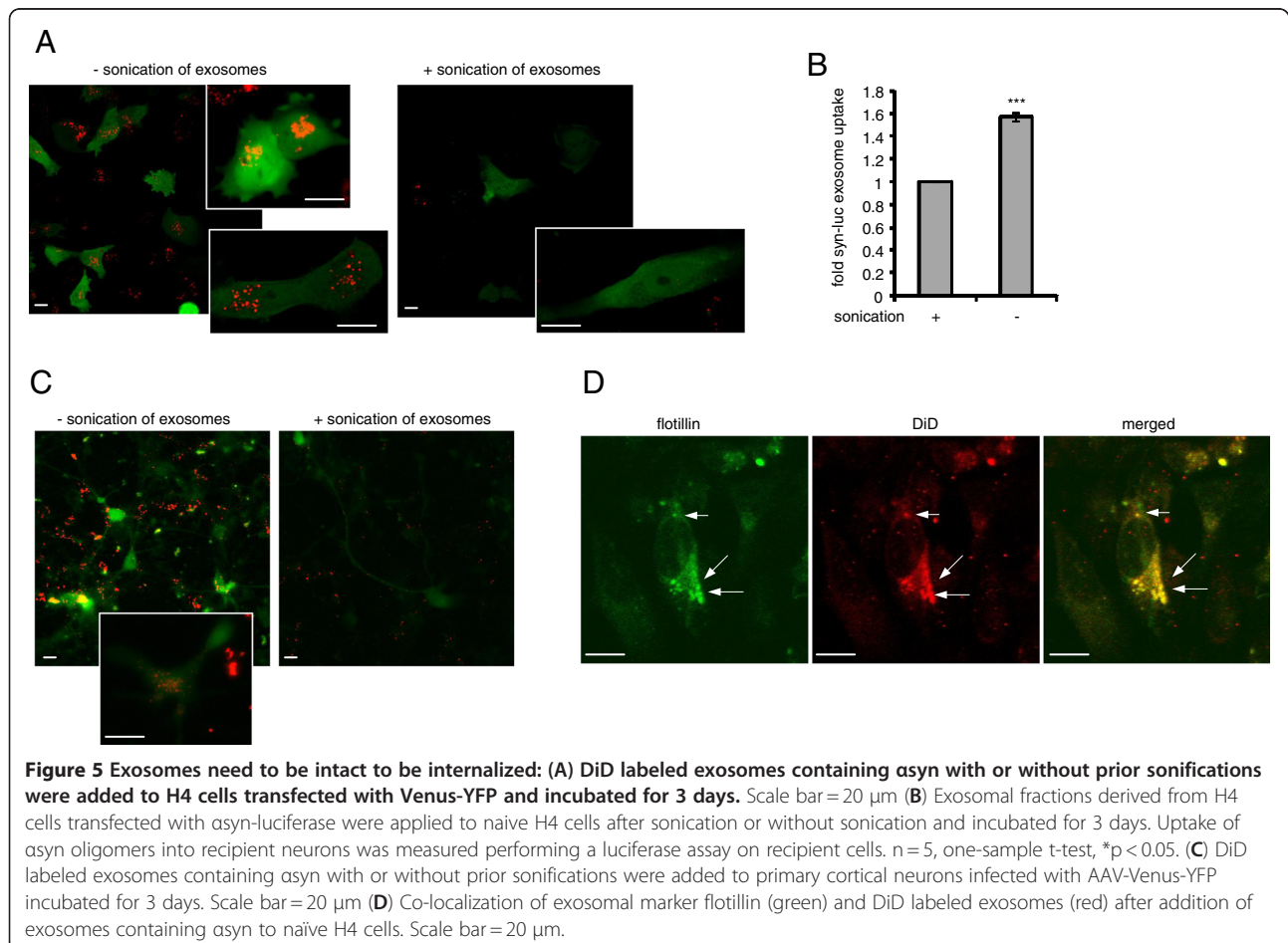
Because our data suggest that exosome-associated asyn may be preferentially taken up by neighboring cells, we next asked whether exosomes need to be intact for uptake to occur. To explore this question, we labeled purified exosome-enriched fractions derived from S1/S2 transfected H4 cells with the membrane dye DiD [68]. To delineate the morphology of H4 cells or primary neurons, we transfected cells with venus-YFP prior to exosome addition resulting in a subpopulation of H4 cells or primary neurons that could be identified via green fluorescence. As expected when labeled exosomes were exogenously added to H4 cells or primary neurons in culture, we observed a rapid uptake of labeled exosomes into the cytosol of cells. To investigate whether membrane integrity is important for uptake, exosomes were subjected to sonication which is known to disrupt lipid bilayer integrity [69-71]. Sonication of exosomes prevented the uptake of exosomes by recipient cells

(Figure 5A). These results were confirmed by measuring luciferase activity inside naive cells following incubation with intact or sonicated exosome fractions from syn-luc transfected H4 cells. Where exosomes were sonicated, significantly less luciferase activity was detected inside the naive cells (Figure 5B) although sonication itself had no effect on luciferase activity of exosomal fractions. This uptake was not unique to proliferating cells as we were also able to confirm the uptake of labeled exosomes into primary cortical neurons (Figure 5C). These results demonstrate that intact exosomes promote exosomal uptake.

To confirm that the labeled microvesicles being observed by confocal microscopy were actually exosomes, we immunostained cells with the exosomal marker flotillin and found that the flotillin immunoreactivity colocalized with the DiD labeled exosomes (Figure 5D).

Autophagy regulates asyn secretion

Exosomes are derived from multivesicular bodies (MVBs), which are endocytic organelles generated by membrane invagination [72-74]. Proteins that are designated for lysosomal degradation are sequestered by



MVBs, however, an alternative destination of MVBs is their exocytic fusion with the plasma membrane leading to the release of intraluminal vesicles (ILVs; i.e., exosome) into the extracellular environment. Interestingly, induction of autophagy can markedly increase the interaction of MVBs and autophagosomes and thereby block exosome secretion [75].

We detected a higher luciferase activity extracellularly (Figure 6A) which may suggest that more oligomers are secreted. However, it's also possible that coelentraxine is more sensitive to extracellular α syn oligomers although it is cell permeable. Nevertheless, given that we detected α syn oligomers in the exosomal fraction of cells and observed that α syn oligomer secretion could be modulated by autophagic activity (Figure 6A) we asked whether autophagy would be a release pathway for exosome-associated α syn oligomers.

Following guidelines for assays monitoring autophagy [76] we measured the levels of LC3.II (ratio with β -actin) and p62 (ratio with α -tubulin) in S1/S2 transfected H4 cells treated with rapamycin, bafilomycin A1 and DMSO. Levels of LC3.II and p62 significantly increased when H4 cells were treated with bafilomycin A1 compared to DMSO controls, indicating an abundance of autophagosomes due to reduced downstream fusion with lysosomes. The opposite effect was observed when treated with rapamycin (Figure 6 B, C). We speculated that an increased pool of autophagosomes could be the basis for an increased exosomes secretion. Exosomes were isolated from conditioned media of S1/S2 transfected H4 cells treated with bafilomycin A1, rapamycin or DMSO, and luciferase activity was monitored. A significant increase in luciferase activity in the exosomal fraction from cells that were treated with bafilomycin A1 compared to DMSO control (Figure 6D) was observed which suggests that inhibition of the fusion of the autophagosome with the lysosome by bafilomycin A1 provides an increased pool autophagosomes which enhances exosomal release. Likewise, treatment with the autophagy enhancer rapamycin showed a decreased α syn oligomer signal in the exosomal fraction as measured by luciferase activity compared with DMSO treatment (Figure 6D), suggesting that enhanced lysosomal activity results in effective α syn oligomer degradation and less secretion. We extended these findings to primary neurons and observed an increase in the α syn oligomer signal in the exosomal fraction from neurons treated with bafilomycin A1 and a decrease in the α syn oligomer signal with rapamycin treatment in the exosomal fraction compared to DMSO control (Figure 6E), although these effects did not reach statistical significance ($p = 0.17$), probably due to the fact that the yield of exosomes are significantly less from primary neuronal preparations resulting in barely detectable luciferase and an decrease in the

signal/noise ratio. Together, these experiments indicate autophagy can be a specific release pathway for secretion of α syn oligomers.

Discussion

Multivesicular bodies (MVBs) and their intraluminal vesicles (ILVs) are involved in the sequestration of proteins destined for degradation in lysosomes. However, MVBs can also fuse with the plasma membrane leading to the release of 50-90 nm ILVs into the extracellular milieu, which are then called exosomes [77,78]. Exosome secretion can therefore be used by cells, including neurons and astrocytes, to clear molecules originally destined for lysosomal degradation [77]. Recently, exosomes have been suggested to play a role in neurodegeneration: Exosomes from prion infected cells have been demonstrated to be efficient initiators of prion propagation in uninfected recipient cells and more importantly, to produce prion disease when inoculated into mice [34,79]. Also the beta-amyloid peptide has been found to be secreted from cells in association with exosomes [80]. Ghidoni et al. suggested that exosomes could be the "Trojan horses" of neurodegeneration; a mechanism underlying the death of cells by shipping toxic agents in exosomes from cell to cell [81]. In our study we identified α syn oligomers to be present in exosomes and found that exosome-associated α syn oligomers are more toxic to neighboring cells than exosome-free α syn oligomers. In contrast to the study of Hasegawa et al. we found that α syn oligomers are present in both the exosomal pellet and the exosome free supernatant from the conditioned media of α syn overexpressing cells whereas Hasegawa et al. recovered α syn mainly from the supernatant fraction [82]. One possible explanation for this discrepancy is the different cellular models and resulting levels of sensitivity. The use of a highly sensitive luciferase protein complementation assay, allows the detection of minimal amounts of protein compared to Western blot analyses.

To our knowledge, this is the first report of α syn oligomers in the exosomal fraction of primary neurons or neuronal cells. Specifically, we have conclusively shown that α syn oligomers can be found outside exosomes, presumably on the outer surface of exosomes. The existence of α syn oligomers outside exosomes has been demonstrated in two ways. First, any external α syn has been digested using trypsin which resulted in a significant decrease in luciferase counts indicative for α syn oligomers. Second, trypsin digestion also led to a dramatic decrease of α syn signal in a Dot blot approach. There are several possible reasons for the existence of α syn oligomers on the external surface of exosomes (Figure 7). First, lipid raft components were found on the membrane surface of secreted exosomes [83]. α Syn has been

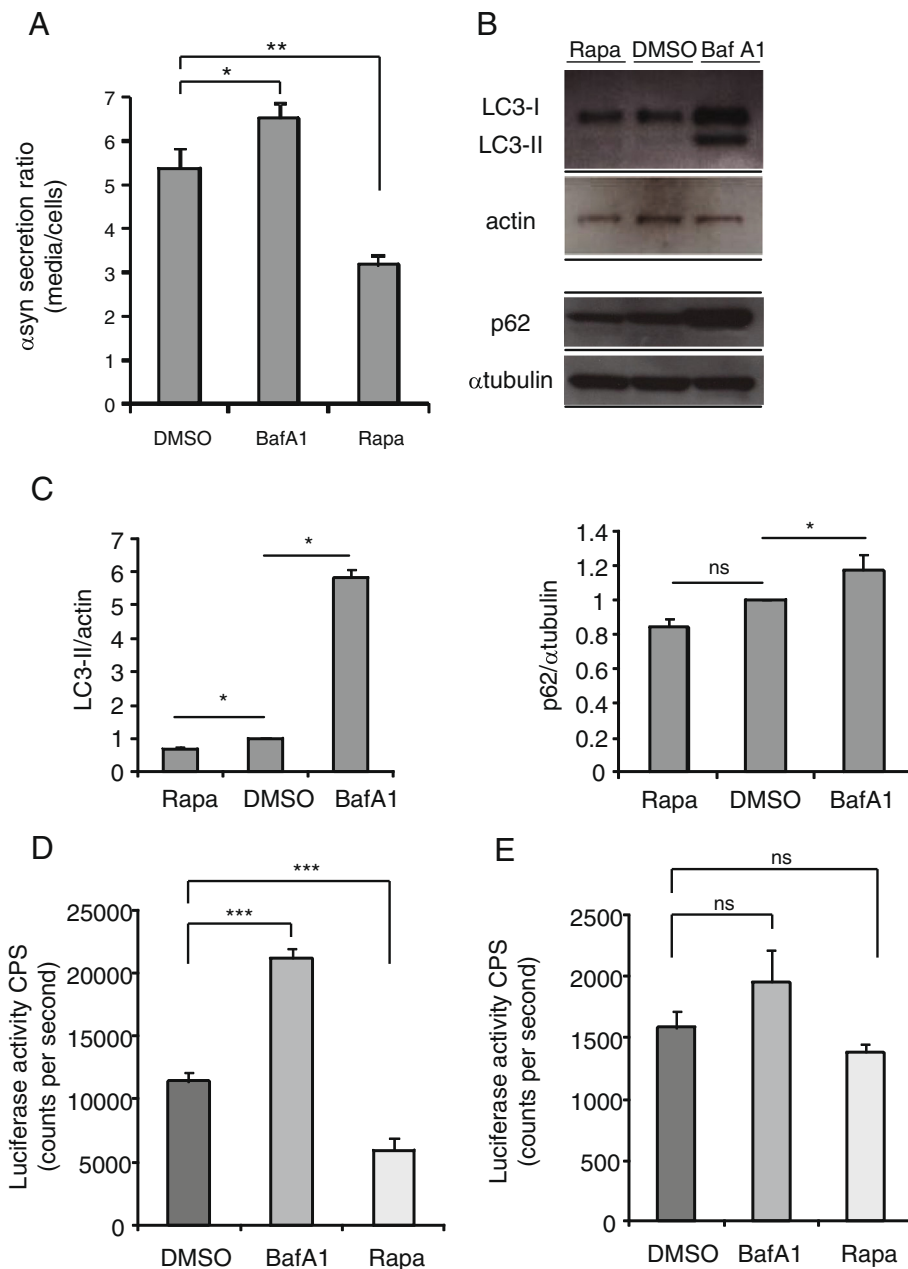
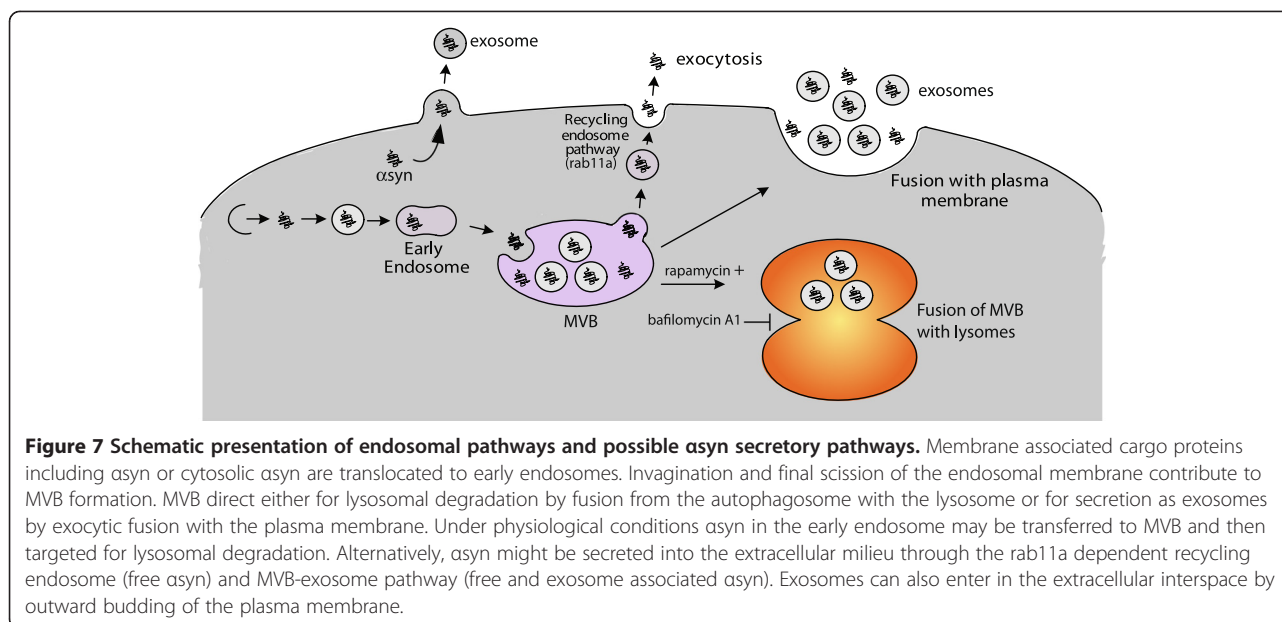


Figure 6 αSyn oligomer secretion is modulated by autophagic activity. (A) Human H4 cells were transfected with asyn complementation pair S1/S2 and treated with DMSO, rapamycin or bafilomycin A1. After 48 h cells CM was assayed for luciferase activity and a ratio of luciferase activity in media compared to cells was expressed. n = 4, One way ANOVA, Bonferroni's Multiple Comparison Test, **p < 0.001. (B) Immunoblot levels of autophagosome marker LC3.II and macrophagy substrate p62 in H4 cells treated with autophagy inhibitor Bafilomycin A1, autophagy inducer rapamycin or DMSO. (C) Densitometric analysis of immunoblots probed with LC3.II normalized to β-actin [n = 3 (Baf A1) p < 0.0001, t-test; n = 3 (rapa), p = 0.0021, t-test] and p62 normalized α-tubulin [n = 3 (Baf A1) p = 0.022, t-test; n = 3 (rapa), p = 0.0022, t-test] (D) Exosomal fractions from H4 cells transfected with S1/S2 and treated with DMSO, rapamycin or bafilomycin A1 were assayed for luciferase activity. n = 4, One way ANOVA, Bonferroni's Multiple Comparison Test, ***p < 0.001 (E) Exosomal fractions from primary neurons infected with AAV-S1/S2 and treated with DMSO, rapamycin or bafilomycin A1 were assayed for luciferase activity. n = 4, One way ANOVA, Bonferroni's Multiple Comparison Test, p = 0.17, ns = non-significant.



shown to interact with lipid rafts and artificial membranes [84,85] and a recent report demonstrates that α syn can penetrate in the outer leaflet of a bilayer [86].

We also found α syn present in the inside of exosomes. Cytosolic proteins like α syn can end up in ILVs because plasma membrane invagination occurs during endocytosis and results in the formation of early endosomes encapsulating a significant amount of cytosol (Figure 7). Endosomal membranes further invaginate to form ILVs, which give endosomes their multi-vesicular appearance. When MVBs fuse with the plasma membrane they can then be released as exosomes containing the encapsulated cytosolic proteins like α syn [87]. Why α syn oligomers are encapsulated in MVB's and released as exosomes is not understood. Possible explanations include first, that α syn oligomers in exosomes could represent the species that are most harmful to cells and is thus targeted for release. Second, Fang et al. have proposed that proteins which exhibit higher order oligomerization and are associated with the plasma membrane, are preferentially sorted into exosomes [88]. A third explanation for the presence of α syn oligomers in exosomes could be that exosomes provide an environment that is favorable for the oligomerization process. Indeed, lipid-mediated oligomerization seems to be important in amyloid formation and polyunsaturated fatty acids have been shown to trigger multimerization of recombinant α syn [89].

We also found α syn oligomers in the exosome-free fraction. One explanation for the presence of exosome-free α syn oligomers could be that the the exosomal membrane undergoes extracellular degradation via proteases or lipases that would lead to the release of proteins

from the exosomal lumen to the extracellular environment during the fractionation process [90,91]. Another explanation could be that α syn oligomers might become unstable and lose their affinity for lipid membranes after being released from the cell, due to changes in the pH value or ionic strength in the extracellular space. In fact, only a small fraction of A β peptide is found associated with exosomes [80], which supports our observations. A third explanation for the secretion of soluble, non-exosomal α syn oligomers could be that there is an exosome-independent pathway of α syn secretion (Figure 7) maybe through the Rab11a-dependent recycling endosomal pathway [82,92], however, further in depth studies will be needed to determine if this is the case. Supporting evidence for the presence of α syn in the exosomal fraction also comes from the recent studies of Emmanouilidou et al. and Alvarez-Erviti [23,24]. Taken together with the previously published studies, our findings support the "Trojan horse" hypothesis [81].

Our data make a case for exosomal transfer of α syn from cell to cell and could represent a key mechanism in the spread of α syn aggregates between neurons in the brain. Indeed, exosomes are biologically active vesicles that are thought to be important for intercellular communication [93,94]. Valadi et al. recently reported that exosomes also contain both mRNA and microRNA, which can be delivered to neighboring cells and be functional in the recipient cell [40]. Exosomes can interact with recipient cells in different ways, including endocytosis, fusion with the plasma membrane, receptor-ligand-binding or attachment [78,95-97].

Critical to our understanding of the toxic effects of secreted α syn oligomers on neighboring cells is the

identification of the toxic species. Our data support the possibility that both exosome-associated α syn oligomers and exosome-free α syn oligomers can confer toxicity on neighboring cells. We found that exosome-associated α syn oligomers are more likely to be taken up by neighboring cells, although we also observe the uptake of free α syn oligomers. The same holds true in terms of toxicity: exosome-associated α syn oligomers are more toxic to neighboring cells compared to free α syn oligomers. It is tempting to speculate that the more α syn oligomers that are taken up by recipient cells, the greater the toxicity. However, it remains to be determined whether α syn oligomers exert their toxic effects after being taken up by recipient cells by impacting cellular homeostasis [98,99] or if extracellular α syn oligomers exert their effect at the cell membrane.

Autophagy can function as a protective mechanism in cells and is particularly crucial in the aging brain and in neurodegeneration where aggregated proteins accumulate [100]. It is now thought that α syn can be degraded by either the proteasome or by autophagy and both macroautophagy and CMA have been reported to contribute to α syn degradation [101-104]. In this study, we demonstrate that secretion of α syn oligomers is increased when lysosomal activity is blocked by Baf A1. Baf A1 inhibits the fusion of the autophagosome with the lysosome by inhibiting vacuolar type H(+)-ATPase [105], thereby inhibiting lysosomal activity. We speculate that by blocking the major degradation pathway for α syn oligomers, the cells use secretion as an alternative pathway to eliminate harmful α syn oligomeric species. By contrast, we did not detect a significant effect of proteasomal inhibition with MG132 on the secretion of α syn oligomers. These results support a hypothesis where autophagy is the major route for degradation of α syn oligomers which are then targeted to the plasma membrane to be cleared by secretion as an alternative route upon failure of this pathway. This assumption is also supported by the fact that rapamycin decreased α syn secretion by enhancing autophagy and thereby triggering intracellular degradation of α syn oligomers. Our results are also in line with the recent work from Emmanouilidiou et al., who did not observe an effect of proteasome inhibitor on levels of extracellular α syn, but found a profound increase in the levels of secreted α syn when the lysosomal pathway was blocked by methylamine [23]. Our study specifically investigates the regulation of secretion of oligomeric α syn upon autophagy inhibition or activation, supporting and significantly augmenting the published study. The fact that we observed more α syn oligomers in the exosomal fraction after inhibition with BafA1 raises the possibility that α syn oligomer containing vesicles (presumably ILVs) originally destined for lysosomal degradation, were re-directed to the plasma

membrane and released as exosomes. This hypothesis requires an interaction between exosomal and autophagic pathways. Indeed, a recent study by Fader et al. demonstrated that induction of autophagy markedly increased the interaction of MVBs and autophagosomes and concurrently blocked exosome secretion, suggesting that MVBs are directed to the autophagic pathway with a consequent inhibition in exosome release [75].

In conclusion, we demonstrate that α syn oligomers can be found in different extracellular fractions in association with exosomes or as exosome-free oligomers. α Syn oligomers associated with exosomes are more toxic to recipient cells compared to free α syn oligomers. The toxic mechanisms of α syn oligomers spreading from cell to cell described here in cell culture could resemble events explaining the spread of α syn pathology that has been observed in human post-mortem brains [8]. Additional studies are needed to verify exosome-associated α syn oligomers and exosomal release in the brains of PD patients. Preventing the early events in exosomal release and uptake by inducing autophagy might be a novel approach for the development of effective drugs for the treatment of PD and other synucleinopathies.

Conclusions

These data demonstrate that oligomeric forms of α syn can be found in multiple extracellular fractions: associated with exosomes and free. Exosome-associated α syn oligomers are more likely to be taken up by recipient cells and can induce more toxicity compared to free α syn oligomers. In addition, we determined α syn oligomers to be present both on the outside of exosomes as well as inside of exosomes. Notably, the pathway of secretion of α syn oligomers is strongly influenced by autophagic activity. Preventing the early events in α syn exosomal release and uptake by inducing autophagy may be a novel approach to halt disease spreading in PD and other synucleinopathies.

Methods

Plasmid generation

Fusion constructs α syn-hGLuc1 (S1), α syn-hGLuc2 (S2) and Venus1- α syn (V1S), α syn-Venus2 (SV2) were generated by subcloning α syn into Not1/Cla1 sites of humanized Gaussia Luciferase and VenusYFP constructs provided by Dr. Stephen Michnick of University of Montreal [25,28].

AAV vectors construction and production

The viral vectors rAAV-CBA-WPRE, rAAV-CBA-IRES-EGFP and rAAV-CBA-SYNUCLEIN-IRES-EGFP were described previously [29]. rAAV-CBA-SYNUCLEIN-LUC1-WPRE (AAV-S1) and rAAV-CBA-SYNUCLEIN-LUC2-WPRE (AAV-S2) were constructed as follows:

α syn -hGLuc1 (S1) and α syn -hGLuc2 (S2) were subcloned into Not1/Nhe1 sites of AAV-CBA-WPRE vector. rAAV-CBA- VENUS1-SYNUCLEIN-WPRE (AAV-V1S) and rAAV-CBA-SYNUCLEIN-VENUS2-WPRE (AAV-SV2) were constructed as follows: the fragments Venus1-Synuclein and Synuclein-Venus2 was inserted into the EcoRV and NheI sites of the pAAV-CBA-WPRE vector. Recombinant adeno-associated type 2/8 was generated by tripartite transfection (AAV-rep/cap expression plasmid, adenovirus miniplasmid and AAV vector plasmid) into 293A cells and purified by iodixanol gradient followed by Q sepharose column chromatography (Harvard Gene Therapy Initiative, Harvard Medical School). The purified virus was dialyzed against PBS, concentrated by Amicon spin column, and tittered by dot blot hybridization. Final titers for virus were for AAV-S1 1.5E13 gc/ml, AAV-S2 1.3E13 gc/ml, V1S 8.3E12 gc/ml and SV2 8.7E12 gc/ml.

Human α synuclein ELISA

Alpha synuclein concentration was quantified using human α syn specific ELISA (#KHB0061, Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions. Absorbance is read at 450 nm. The absorbance is directly proportional to the concentration of α syn present in the original specimen. α Syn concentration was determined by plotting sample absorbances against standards using Graph Pad Prism fitting software (four parameter algorithm).

Cell culture and transfections

Unless otherwise stated, human H4 neuroglioma cells (HTB-148 - ATCC, Manassas, VA, USA) were maintained in OPTI-MEM medium supplemented with 10% fetal bovine serum (both from Invitrogen) and incubated at 37°C. Cells were plated 24 hours prior to transfection, growing to 80–90% confluency prior to transfection. Transfection was performed using Superfect (Qiagen, Chatsworth, CA, USA) using equimolar ratios of plasmids according to the manufacturer's instructions. Conditioned media was collected 48 hours post-transfection and centrifuged for 5 min at 3000 g to eliminate floating cells before being used.

Gaussia luciferase protein-fragment complementation assay

Fusion constructs α syn hGLuc1 (S1) and α syn -hGLuc2 (S2) were generated as described previously [25]. S1 and S2 were transfected into H4 cells in a 96-well plate format as described above. 48 h after transfection, culture media was transferred to a new 96 well plate (Costar, Corning, NY, USA). Cells were washed with PBS and replaced with serum- and phenol-red free media. Luciferase activity from protein complementation was

measured for conditioned media and live cells in an automated plate reader at 480 nm following the injection of the cell permeable substrate, coelenterazine (20 μ M) (Prolume Ltd, Pinetop, AZ) with a signal integration time of 2 seconds.

Primary cortical cell culture

Primary cortical neurons were prepared from cerebral cortices of E14-16 mouse embryos. Cortices were dissected from embryonic brain and the meninges were removed. Cortices were dissociated by titration at RT and cells were resuspended in Neurobasal (NB) (Gibco) medium supplemented with 10% fetal bovine serum, 2 mM Glutamax, 100 U/mL penicillin, and 100 μ g/mL streptomycin and plated at a density of 3.84×10^4 cells/well on 96w plates (Corning, NY, USA), 60 mm dishes at a density of 3.6×10^6 cells/dish (Costar, Corning, NY, USA) coated with 20 μ g/mL poly-D-lysine (Sigma-Aldrich, St. Louis, MO, USA). After 2 h medium was changed into NB/B-27 [NB medium containing 2% (v/v) B-27 supplement], 100 U/mL penicillin, 100 μ g/mL streptomycin, and 2 mM/L glutamine. Cells were maintained at 37°C in 5% CO₂ in a humidified incubator. Medium was changed every third day. Neurons were grown for 4–5 days in vitro (DIV) before infected with AAV8-S1/AAV8-S2, AAV8-V1S/AAV8-SV2, AAV8-syn-ires-GFP or AAV8-GFP. Infections were carried out as followed: 6 μ l rAAV2/8 expressing eGFP (1.3E13gc/ml) per 60 mm dish, 6 μ l rAAV2/8 expressing α syn(wt)-ires-GFP (1.1E13gc/ml) per 60 mm dish and 3 μ l rAAV2/8 expressing V1S (8.3E12gc/ml) together with 3 μ l rAAV2/8 expressing SV2 (8.7E12gc/ml) per 60 mm dish, as well as 3 μ l rAAV2/8 expressing S1 (1.5E13gc/ml) together with 3 μ l rAAV2/8 expressing S2 (1.3E13gc/ml) per 60 mm dish.

Exosome isolation

Exosomes from Human H4 cells and primary neurons were prepared as described earlier [34,35] with minor modifications. Briefly, conditioned medium was collected and spun for 5 min at 500xg to remove floating cells. The supernatants were then sequentially centrifuged at 300xg (10 min) and 2x 200xg (10 min) at 4°C each. Then supernatants were filtered through a 0.45 μ m (Whatmann, Florham Park, NJ) and then 0.22 μ m (Millipore, Carrigtowhill, Cork, Ireland) filter, and centrifuged for 30 minutes at 10,000xg (2X) at 4°C. After ultracentrifugation at 100,000xg for 70 min at 4°C, exosomal pellet was then resuspended in 1xPBS for Western Blotting, electron microscopy or luciferase assay or resuspended culture medium for cell treatments.

Exosome depleted medium was prepared as described above, except after ultracentrifugation at 100,000xg for

70 min at 4°C, exosome free supernatant was filtered through a 0.22 µm filter before used in cell culture.

Digestion of exosomes

Exosome associated or exosome free α syn oligomers were digested by addition of 0.25% trypsin (Invitrogen, Carlsbad, CA, USA) and/or 0.1% saponin and incubated for 20 min at 37°C. After complete digestion samples were analyzed in luciferase assay or Dot blot approach.

Labeling of exosomes

The exosomes were labelled using DiD (Biotium, Hayward, CA USA) according to manufacturer's instructions in a 1:200 dilution. Briefly, after the final spin in exosome purification exosomal pellet was resuspended in 1 ml DiD solution and incubated for 5 minutes. After ultracentrifugation at 100,000xg for 70 min at 4°C the exosomal pellet was washed in 1xPBS centrifuged again for 90 min at 150 000 g to remove free dye. Then the pellet is resuspended finally as exosome fraction.

Dot blot

Exosomes or exosome free supernatant was collected as described previously. 100ul of each condition was applied to nitrocellulose membrane (pore size 0.22 µm, Whatman Protran, Sanford, ME, USA) placed in a Dot blot apparatus (Schleicher & Schuell Minifold-I Dot-Blot System, Whatman, Sanford, ME, USA,) and incubated at RT for 1 h. Samples were filtered through the membrane by gentle vacuum and developed using conditions as described previously (21). Briefly, the membrane was blocked with 10% non-fat dried milk in Tris-buffered saline (TBS, Sigma-Aldrich, St. Louis, MO, USA) containing 0.01% Tween 20 (TBS-T), at room temperature for 1 h. After three washes with TBS-T, the membrane was incubated with anti- Syn-1 antibody (1:1000; BD transduction, Franklin Lakes, NJ, USA) or monoclonal anti-CD63 antibody (1:500, BD Transduction) overnight at 4 C with gentle agitation. The membranes were then washed three times for 5 min with TBS-T, incubated with horseradish peroxidase conjugated anti mouse IgG (Jackson Immuno Research Laboratories, Baltimore, PA, USA) diluted 1:2000 in 5% non-fat dried milk in Tris-buffered saline (TBS) containing 0.01% Tween 20 (TBS-T) and incubated for 1 hour at room temperature. The blots were washed three times with TBS-T and developed with Pierce ECL chemiluminescence kit from Thermo Scientific (Rockford, IL, USA).

Pharmacological treatments in vitro

H4 cells were plated into 96 well plates or 60 mm dishes (Costar, Corning, NY, USA) and transfected as described above. Transfection mix was incubated for 2 h according to manufacturer's protocol, then media was replaced by

fresh culture media containing 0.4 µg/ml rapamycin (Sigma Aldrich, St. Louis, MO) or DMSO (Sigma Aldrich, St. Louis, MO) and incubated for 48 h. 200nM Bafilomycin A1 (Merck KG, Darmstadt, Germany) was added to the culture medium 20 h before harvesting the medium. Conditioned medium was collected for luciferase assay or exosomal isolations. To ensure that pharmacological treatments result in a true increase in the secretion ratio of α syn oligomers and not simply a matter of more available α syn oligomers in the cytoplasm, we calculated the ratio of secreted α syn oligomers in the conditioned medium to intracellular α syn oligomers.

Primary neurons were plated into 60 mm dishes (Costar, Corning, NY, USA) and infected as described above. DMSO and 0.1 µg/ml rapamycin were added to the culture medium right after infection, whereas 100nM bafilomycin A1 was added after 3 days expression and incubated for 20 h. Conditioned medium was then collected to perform exosomal isolations.

Exosomal uptake experiments

Conditioned media from naïve H4 cells or naïve primary neurons was replaced by exosome containing culture media. After 3–4 days incubation cells were washed twice with 1xPBS and then assayed for luciferase activity.

Toxicity assay

Toxicity was analyzed 3–4 days after exosome application by measuring the activity of Caspases 3 and 7 using a fluorometric substrate Z-DEVD-Rhodamine 110 (Apo-ONE homogeneous Caspase-3/7 assay #G7790, Promega, Madison, WI) according to the manufacturer's protocol.

Western blotting

Primary cortical neurons were scraped from 60 mm dishes and washed by centrifugation and resuspension in cold PBS. The cells were resuspended in 1x PBS containing protease inhibitors (protease inhibitor cocktail tablet 1 tablet/10 mL (Roche Diagnostics) sheared by passing through a 27-gauge 1 ml syringe 4–6 times and centrifuged for 5 min at 13,000 g. Lysates or exosomal samples were resolved by electrophoresis on a 4–12 % Bis-Tris gradient gel (NuPAGE Novex Bis-Tris Gel, Invitrogen, Carlsbad, CA, USA) according to manufacturer's instructions using NuPAGE MOPS buffer. After transfer to nitrocellulose membrane (Protran, Schleicher and Schuell, Whatman GmbH, Dassel, Germany) membranes were blocked in either 5% milk in TBS-T or Li-Cor blocking buffer (LI-COR, Lincoln, NE, USA) for 1 hour at room temperature. Membranes were then incubated with primary antibodies (mouse anti-Alix,

1:500, BD Transduction; mouse anti-flotillin: monoclonal, 1:500, BD Transduction) overnight at 4 °C. After three 5–10 min TBS-T washes, membranes were incubated at room temperature for 1 hour with either IR-labeled secondary antibodies (IR800 goat anti-mouse, 1:2000, Rockland Immunochemicals, PA, USA) or HRP-conjugated secondary antibodies (1:2000). After three 5–10 min TBS-T washes, immunoblots were analyzed using either the Odyssey Infrared imaging system (Licor, Lincoln, NE, USA) or the ECL chemiluminescent detection system (Amersham/GE HealthCare, Buckinghamshire, UK).

Cell imaging and immunofluorescence staining

All images were acquired using a 20x Plan Achromat lens (Carl Zeiss), 25x APO-Plan NEOFLU lens (Carl Zeiss) or Zeiss 63x 1.2 NA C-APO-Plan NEOFLU water immersion lens (Carl Zeiss), mounted on the microscope described before. H4 cells or cortical neurons were washed three times with phosphate-buffered saline (PBS) following 30 min incubation in a fixation solution containing 4% paraformaldehyde in PBS. After washing, the cells were permeabilized and unspecific binding sites were blocked using 0.05% Saponin and 1% bovine serum albumin in PBS followed by another washing step. The primary rabbit antibody against flotillin (1:500, BD Transduction) was added for 1 h at RT, followed by another washing step and incubation with the secondary antibody (anti-mouse antibody labeled with Alexa-Fluor 488; Invitrogen) for 1 h at RT.

Electron microscopy

An exosome pellet from either human H4 cells or primary neurons was prepared by centrifugation as described above and then resuspended in 20 µl of cold Karnovsky's EM fixative (2% formaldehyde and 2.5% glutaraldehyde in 0.1 M Sodium Cacodylate buffer, pH 7.4). Ultrathin sections from LR white embedded samples were picked up from the knife with a loop, dipped in a 2:1 mixture of 2.3 M sucrose and 2% methylcellulose, and adsorbed to the surface of a formvar/carbon coated copper grid. Grids are placed on 2% gelatin in a small petri dish and stored in the fridge until immunogold labeling. This was accomplished by washing grids in PBS and then either treating with CD63 antibody (BD Transduction) followed by 10 nm gold labelled secondary antibodies (Sigma Aldrich, St. Louis, MO) or processing without immunolabelling. These exosome containing grids were then post-fixed with 2.5% glutaraldehyde, washed and contrasted with 2% methyl cellulose and 3% aqueous uranyl acetate. Samples were examined and photographed with a JEOL 1200EX electron microscope.

microRNA profiling and data analysis

For nucleic acid analysis, the entire exosome pellet is gently resuspended in 20 µl of 1xPBS. Any cellular/ribosomal RNAs that may exist in the extra-exosomal solution (53) are eliminated by adding 8 µg of RNase T1/A (Fermentas) to the 20ul preparation and incubating for 10 minutes at 37°C. Four hundred units of Superase-in RNase (Ambion) inhibitor are then added to inactivate the RNases and the sample is held at 25°C for 10 minutes. The entire mixture is then dissolved in 60 µl of miRNA extraction buffer (Arcturus), incubated at 42°C for 30 minutes, and stored at –80°C prior further processing. In order to generate amplified sense RNA from small quantities (<1 ng) of purified miRNAs, we used the NCode miRNA Amplification System (Invitrogen) according to the manufacturer's instructions. miRNA expression profiles were generated by adding 250 ng of this amplified miRNA to the FlexMiR miRNA assay from Luminex Corporation (Austin, Texas) and running on a Luminex FlexMAP 3D system according to the manufacturer's instructions.

Statistical analysis

Statistical analyses were carried out using the program GraphPad Prism, Version 4.0. Values in the figures are expressed as means ± SEM.

Additional file 1

Additional file 1: Schematic representation of the asyn protein fragment complementation constructs. (A) Nonbioluminescent halves of humanized *Gussia luciferase* are fused to asyn monomers (B) Non fluorescent halves of *Venus-YFP* are fused to asyn monomers.

Abbreviations

PD: Parkinson's disease; asyn: Alpha synuclein; CM: Conditioned media; V1S: Venus YFP fused to the N-terminus of asyn; SV2: C-terminal half of Venus YFP fused to the C-terminus of asyn; S1: asyn fused to amino-terminal fragment of *Gussia princeps luciferase*; S2: asyn fused to carboxy-terminal fragment of *Gussia princeps luciferase*; MVBs: Multivesicular bodies; CSF: Cerebrospinal fluid.

Competing interests

The author declares that they have no competing interests.

Authors' contributions

KMD, LRK, WPR, OCG, ARW, LZ and CRV performed the experiments. KMD and PJM and analyzed the results, designed the study, and wrote the manuscript. All authors read and approved the final manuscript.

Acknowledgements

This work was supported by NIH NS063963 to PJM. We thank Harvard Medical School EM core for electron microscopy assistance.

Author details

¹MassGeneral Institute for Neurodegenerative Disease, Department of Neurology, Massachusetts General Hospital, Charlestown, MA, USA.

²Advanced Tissue Resource Center, Harvard NeuroDiscovery Center, Harvard Medical School, Boston, MA, USA. ³Present address: Neurology Department, University of Ulm, Ulm, Germany. ⁴Present address: Department of Neuroscience, Mayo Clinic, 4500 San Pablo Rd, Jacksonville, FL 32224, USA.

Received: 25 April 2012 Accepted: 14 August 2012
Published: 24 August 2012

References

- Goedert M: **Alpha-synuclein and neurodegenerative diseases.** *Nat Rev Neurosci* 2001, **2**(7):492–501.
- Chartier-Harlin MC, Kachergus J, Roumier C, Mouroux V, Douay X, Lincoln S, Leveque C, Larvor L, Andrieux J, Hulihan M, et al: **Alpha-synuclein locus duplication as a cause of familial Parkinson's disease.** *Lancet* 2004, **364**(9440):1167–1169.
- Singleton AB, Farrer M, Johnson J, Singleton A, Hague S, Kachergus J, Hulihan M, Peuralinna T, Dutra A, Nussbaum R, et al: **alpha-Synuclein locus triplication causes Parkinson's disease.** *Science* 2003, **302**(5646):841.
- Krüger R, Kuhn W, Müller T, Woitalla D, Graeber M, Kosel S, Przuntek H, Epplen JT, Schols L, Riess O: **Ala30Pro mutation in the gene encoding α -synuclein in Parkinson's disease.** *Nat Genet* 1998, **18**:106–108.
- Polymeropoulos MH, Lavedan C, Leroy E, Idle SE, Dehejia A, Dutra A, Pike B, Root H, Rubenstein J, Boyer R, et al: **Mutation in the α -Synuclein gene identified in families with Parkinson's disease.** *Science* 1997, **276**:2045–2047.
- Zarranz JJ, Alegre J, Gomez-Esteban JC, Lezcano E, Ros R, Ampuero I, Vidal L, Hoenicka J, Rodriguez O, Atares B, et al: **The new mutation, E46K, of alpha-synuclein causes Parkinson and Lewy body dementia.** *Ann Neurol* 2004, **55**(2):164–173.
- Irizarry MC, Growdon W, Gomez-Isla T, Newell K, George JM, Clayton DF, Hyman BT: **Nigral and cortical Lewy bodies and dystrophic nigral neurites in Parkinson's disease and cortical Lewy body disease contain alpha-synuclein immunoreactivity.** *J Neuropathol Exp Neurol* 1998, **57**(4):334–337.
- Braak H, Del Tredici K, Rub U, de Vos RA, Jansen Steur EN, Braak E: **Staging of brain pathology related to sporadic Parkinson's disease.** *Neurobiology of aging* 2003, **24**(2):197–211.
- Auluck PK, Chan HYE, Trojanowski JQ, Lee VM-Y, Bonini NM: **Chaperone suppression of α -synuclein toxicity in a *Drosophila* model for Parkinson's disease.** *Science* 2002, **295**:865–868.
- Bodner RA, Outeiro TF, Altmann S, Maxwell MM, Cho SH, Hyman BT, McLean PJ, Young AB, Housman DE, Kazantsev AG: **Pharmacological promotion of inclusion formation: a therapeutic approach for Huntington's and Parkinson's diseases.** *Proc Natl Acad Sci U S A* 2006, **103**(11):4246–4251.
- Bucciantini M, Calloni G, Chiti F, Formigli L, Nosi D, Dobson CM, Stefani M: **Prefibrillar amyloid protein aggregates share common features of cytotoxicity.** *J Biol Chem* 2004, **279**(30):31374–31382.
- El-Agnaf OM, Salem SA, Paleologou KE, Curran MD, Gibson MJ, Court JA, Schlossmacher MG, Allsop D: **Detection of oligomeric forms of alpha-synuclein protein in human plasma as a potential biomarker for Parkinson's disease.** *FASEB J* 2006, **20**(3):419–425.
- Kayed R, Head E, Thompson JL, McIntire TM, Milton SC, Cotman CW, Glabe CG: **Common structure of soluble amyloid oligomers implies common mechanism of pathogenesis.** *Science* 2003, **300**(5618):486–489.
- Lashuel HA, Petre BM, Wall J, Simon M, Nowak RJ, Walz T, Lansbury PT Jr: **Alpha-synuclein, especially the Parkinson's disease-associated mutants, forms pore-like annular and tubular protofibrils.** *J Mol Biol* 2002, **322**(5):1089–1102.
- Masliah E, Rockenstein E, Veinbergs I, Mallory M, Hashimoto M, Takeda A, Sagara Y, Sisk A, Mucke L: **Dopaminergic loss and inclusion body formation in α -synuclein mice: Implications for neurodegenerative disorders.** *Science* 2000, **287**:1265–1269.
- Desplats P, Lee HJ, Bae EJ, Patrick C, Rockenstein E, Crews L, Spencer B, Masliah E, Lee SJ: **Inclusion formation and neuronal cell death through neuron-to-neuron transmission of alpha-synuclein.** *Proc Natl Acad Sci U S A* 2009, **106**(31):13010–13015.
- Danzer KM, Ruf WP, Putcha P, Joyner D, Hashimoto T, Glabe C, Hyman BT, McLean PJ: **Heat Shock Protein 70 modulates toxic extracellular alpha-synuclein oligomers and rescues trans-synaptic toxicity.** *FASEB J* 2010, In Press.
- Ahn KJ, Paik SR, Chung KC, Kim J: **Amino acid sequence motifs and mechanistic features of the membrane translocation of alpha-synuclein.** *J Neurochem* 2006, **97**(1):265–279.
- Danzer KM, Haasen D, Karow AR, Moussaoud S, Habeck M, Giese A, Kretzschmar H, Hengger B, Kostka M: **Different species of alpha-synuclein oligomers induce calcium influx and seeding.** *J Neurosci* 2007, **27**(34):9220–9232.
- Danzer KM, Krebs SK, Wolff M, Birk G, Hengger B: **Seeding induced by alpha-synuclein oligomers provides evidence for spreading of alpha-synuclein pathology.** *J Neurochem* 2009, **111**(1):192–201.
- Lee HJ, Suk JE, Bae EJ, Lee JH, Paik SR, Lee SJ: **Assembly-dependent endocytosis and clearance of extracellular alpha-synuclein.** *Int J Biochem Cell Biol* 2008, **40**(9):1835–1849.
- Luk KC, Song C, O'Brien P, Stieber A, Branch JR, Brunden KR, Trojanowski JQ, Lee VM: **Exogenous alpha-synuclein fibrils seed the formation of Lewy body-like intracellular inclusions in cultured cells.** *Proc Natl Acad Sci U S A* 2009, **106**(47):20051–20056.
- Emmanouilidou E, Melachroinou K, Roumeliotis T, Garbis SD, Ntzouni M, Margaritis LH, Stefanis L, Vekrellis K: **Cell-produced alpha-synuclein is secreted in a calcium-dependent manner by exosomes and impacts neuronal survival.** *J Neurosci*, **30**(20):6838–6851.
- Alvarez-Erviti L, Seow Y, Schapira AH, Gardiner C, Sargent IL, Wood MJ, Cooper JM: **Lysosomal dysfunction increases exosome-mediated alpha-synuclein release and transmission.** *Neurobiol Dis* 2011, **42**(3):360–367.
- Outeiro TF, Putcha P, Tetzlaff JE, Spoelgen R, Koker M, Carvalho F, Hyman BT, McLean PJ: **Formation of toxic oligomeric alpha-synuclein species in living cells.** *PLoS One* 2008, **3**(4):e1867.
- Putcha P, Danzer KM, Kranich LR, Scott A, Silinski M, Mabbett S, Hicks CD, Veal JM, Steed PM, Hyman BT, et al: **Brain permeable small molecule inhibitors of Hsp90 prevent alpha-synuclein oligomer formation and rescue alpha-synuclein-induced toxicity.** *J Pharmacol Exp Ther* 2009, in press.
- Remy I, Michnick SW: **A cDNA library functional screening strategy based on fluorescent protein complementation assays to identify novel components of signaling pathways.** *Methods* 2004, **32**(4):381–388.
- Remy I, Michnick SW: **A highly sensitive protein-protein interaction assay based on *Gussia* luciferase.** *Nat Methods* 2006, **3**(12):977–979.
- St Martin JL, Klucken J, Outeiro TF, Nguyen P, Keller-McGandy C, Cantuti-Castelvetri I, Grammatopoulos TN, Standaert DG, Hyman BT, McLean PJ: **Dopaminergic neuron loss and up-regulation of chaperone protein mRNA induced by targeted over-expression of alpha-synuclein in mouse substantia nigra.** *J Neurochem* 2007, **100**(6):1449–1457.
- Tetzlaff JE, Putcha P, Outeiro TF, Ivanov A, Berezovska O, Hyman BT, McLean PJ: **CHIP targets toxic alpha-synuclein oligomers for degradation.** *J Biol Chem* 2008, **283**(26):17962–17968.
- Lee HJ, Patel S, Lee SJ: **Intravesicular localization and exocytosis of alpha-synuclein and its aggregates.** *J Neurosci* 2005, **25**(25):6016–6024.
- Mollenhauer B, Cullen V, Kahn I, Krastins B, Outeiro TF, Pepivani I, Ng J, Schulz-Schaeffer W, Kretzschmar HA, McLean PJ, et al: **Direct quantification of CSF alpha-synuclein by ELISA and first cross-sectional study in patients with neurodegeneration.** *Exp Neurol* 2008, **213**(2):315–325.
- Sung JY, Kim J, Paik SR, Park JH, Ahn YS, Chung KC: **Induction of neuronal cell death by Rab5A-dependent endocytosis of alpha-synuclein.** *J Biol Chem* 2001, **276**(29):27441–27448.
- Fevrier B, Vilette D, Archer F, Loew D, Faigle W, Vidal M, Laude H, Raposo G: **Cells release prions in association with exosomes.** *Proc Natl Acad Sci U S A* 2004, **101**(26):9683–9688.
- Thery C, Amigorena S, Raposo G, Clayton A: **Isolation and characterization of exosomes from cell culture supernatants and biological fluids.** *Curr Protoc Cell Biol* 2006, **Chapter 3**:Unit 3 22.
- Escola JM, Kleijmeer MJ, Stoorvogel W, Griffith JM, Yoshie O, Geuze HJ: **Selective enrichment of tetraspan proteins on the internal vesicles of multivesicular endosomes and on exosomes secreted by human B-lymphocytes.** *J Biol Chem* 1998, **273**(32):20121–20127.
- Simpson RJ, Lim JW, Moritz RL, Mathivanan S: **Exosomes: proteomic insights and diagnostic potential.** *Expert Rev Proteomics* 2009, **6**(3):267–283.
- Schultz J, Lorenz P, Gross G, Ibrahim S, Kunz M: **MicroRNA let-7b targets important cell cycle molecules in malignant melanoma cells and interferes with anchorage-independent growth.** *Cell Res* 2008, **18**(5):549–557.
- Taylor DD, Gercel-Taylor C: **MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer.** *Gynecol Oncol* 2008, **110**(1):13–21.
- Valadi H, Ekstrom K, Bossios A, Sjostrand M, Lee JJ, Lotvall JO: **Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells.** *Nat Cell Biol* 2007, **9**(6):654–659.

41. Chen XM, Splinter PL, O'Hara SP, LaRusso NF: **A cellular micro-RNA, let-7i, regulates Toll-like receptor 4 expression and contributes to cholangiocyte immune responses against *Cryptosporidium parvum* infection.** *J Biol Chem* 2007, **282**(39):28929–28938.
42. Segura MF, Hanniford D, Menendez S, Reavie L, Zou X, Alvarez-Diaz S, Zakrzewski J, Blochin E, Rose A, Bogunovic D, et al: **Aberrant miR-182 expression promotes melanoma metastasis by repressing FOXO3 and microphthalmia-associated transcription factor.** *Proc Natl Acad Sci U S A* 2009, **106**(6):1814–1819.
43. Weitzel RP, Lesniewski ML, Haviernik P, Kadereit S, Leahy P, Greco NJ, Laughlin MJ: **microRNA 184 regulates expression of NFAT1 in umbilical cord blood CD4+ T cells.** *Blood* 2009, **113**(26):6648–6657.
44. Bentwich I, Avniel A, Karov Y, Aharonov R, Gilad S, Barad O, Barzilai A, Einat P, Einav U, Meiri E, et al: **Identification of hundreds of conserved and nonconserved human microRNAs.** *Nat Genet* 2005, **37**(7):766–770.
45. Sewer A, Paul N, Landgraf P, Aravin A, Pfeffer S, Brownstein MJ, Tuschl T, van Nimwegen E, Zavolan M: **Identification of clustered microRNAs using an ab initio prediction method.** *BMC Bioinforma* 2005, **6**:267.
46. Huang Q, Gumireddy K, Schrier M, le Sage C, Nagel R, Nair S, Egan DA, Li A, Huang G, Klein-Szanto AJ, et al: **The microRNAs miR-373 and miR-520c promote tumour invasion and metastasis.** *Nat Cell Biol* 2008, **10**(2):202–210.
47. Altuvia Y, Landgraf P, Lithwick G, Elefant N, Pfeffer S, Aravin A, Brownstein MJ, Tuschl T, Margalit H: **Clustering and conservation patterns of human microRNAs.** *Nucleic Acids Res* 2005, **33**(8):2697–2706.
48. Kawahara Y, Zinshteyn B, Sethupathy P, Iizasa H, Hatzigeorgiou AG, Nishikura K: **Redirection of silencing targets by adenosine-to-inosine editing of miRNAs.** *Science* 2007, **315**(5815):1137–1140.
49. Rosa A, Ballarino M, Sorrentino A, Sthandier O, De Angelis FG, Marchioni M, Masella B, Guarini A, Fatica A, Peschle C, et al: **The interplay between the master transcription factor PU.1 and miR-424 regulates human monocyte/macrophage differentiation.** *Proc Natl Acad Sci U S A* 2007, **104**(50):19849–19854.
50. Wheeler G, Ntounia-Fousara S, Granda B, Rathjen T, Dalmay T: **Identification of new central nervous system specific mouse microRNAs.** *FEBS Lett* 2006, **580**(9):2195–2200.
51. Ujifuku K, Mitsutake N, Takakura S, Matsue M, Saenko V, Suzuki K, Hayashi K, Matsuo T, Kamada K, Nagata I, et al: **miR-195, miR-455-3p and miR-10a(*) are implicated in acquired temozolomide resistance in glioblastoma multiforme cells.** *Cancer Lett* 2010, **296**(2):241–248.
52. Goodarzi H, Elemento O, Tavazoie S: **Revealing global regulatory perturbations across human cancers.** *Mol Cell* 2009, **36**(5):900–911.
53. Liu L, Jiang Y, Zhang H, Greenlee AR, Han Z: **Overexpressed miR-494 down-regulates PTEN gene expression in cells transformed by anti-benzo(a)pyrene-trans-7,8-dihydrodiol-9,10-epoxide.** *Life Sci*, **86**(5–6):192–198.
54. Landgraf P, Rusu M, Sheridan R, Sewer A, Iovino N, Aravin A, Pfeffer S, Rice A, Kamphorst AO, Landthaler M, et al: **A mammalian microRNA expression atlas based on small RNA library sequencing.** *Cell* 2007, **129**(7):1401–1414.
55. Zhu H, Wu H, Liu X, Evans BR, Medina DJ, Liu CG, Yang JM: **Role of MicroRNA miR-27a and miR-451 in the regulation of MDR1/P-glycoprotein expression in human cancer cells.** *Biochem Pharmacol* 2008, **76**(5):582–588.
56. Girardot M, Pecquet C, Boukour S, Knoops L, Ferrant A, Vainchenker W, Giraudier S, Constantinescu SN: **miR-28 is a thrombopoietin receptor targeting microRNA detected in a fraction of myeloproliferative neoplasm patient platelets.** *Blood* 2010, **116**(3):437–445.
57. Tazawa H, Tsuchiya N, Izumiya M, Nakagama H: **Tumor-suppressive miR-34a induces senescence-like growth arrest through modulation of the E2F pathway in human colon cancer cells.** *Proc Natl Acad Sci U S A* 2007, **104**(39):15472–15477.
58. Petrocca F, Vecchione A, Croce CM: **Emerging role of miR-106b-25/miR-17-92 clusters in the control of transforming growth factor beta signaling.** *Cancer Res* 2008, **68**(20):8191–8194.
59. Takahashi Y, Forrest AR, Maeno E, Hashimoto T, Daub CO, Yasuda J: **MiR-107 and MiR-185 can induce cell cycle arrest in human non small cell lung cancer cell lines.** *PLoS One* 2009, **4**(8):e6677.
60. Song B, Wang Y, Kudo K, Gavin EJ, Xi Y, Ju J: **miR-192 Regulates dihydrofolate reductase and cellular proliferation through the p53-microRNA circuit.** *Clin Cancer Res* 2008, **14**(24):8080–8086.
61. Chao A, Tsai CL, Wei PC, Hsueh S, Chao AS, Wang CJ, Tsai CN, Lee YS, Wang TH, Lai CH: **Decreased expression of microRNA-199b increases protein levels of SET (protein phosphatase 2A inhibitor) in human choriocarcinoma.** *Cancer Lett*, **291**(1):99–107.
62. Barroso-delJesus A, Romero-Lopez C, Lucena-Aguilar G, Melen GJ, Sanchez L, Ligerio G, Berzal-Herranz A, Menendez P: **Embryonic stem cell-specific miR302-367 cluster: human gene structure and functional characterization of its core promoter.** *Mol Cell Biol* 2008, **28**(21):6609–6619.
63. Xie X, Lu J, Kulbokas EJ, Golub TR, Mootha V, Lindblad-Toh K, Lander ES, Kellis M: **Systematic discovery of regulatory motifs in human promoters and 3' UTRs by comparison of several mammals.** *Nature* 2005, **434**(7031):338–345.
64. Nakano H, Miyazawa T, Kinoshita K, Yamada Y, Yoshida T: **Functional screening identifies a microRNA, miR-491 that induces apoptosis by targeting Bcl-X(L) in colorectal cancer cells.** *Int J Cancer* 2009, **127**(5):1072–1080.
65. Roshan R, Ghosh T, Scaria V, Pillai B: **MicroRNAs: novel therapeutic targets in neurodegenerative diseases.** *Drug Discov Today* 2009, **14**(23–24):1123–1129.
66. Junn E, Lee KW, Jeong BS, Chan TW, Im JY, Mouradian MM: **Repression of alpha-synuclein expression and toxicity by microRNA-7.** *Proc Natl Acad Sci U S A* 2009, **106**(31):13052–13057.
67. Zhang W, Wang T, Pei Z, Miller DS, Wu X, Block ML, Wilson B, Zhou Y, Hong JS, Zhang J: **Aggregated alpha-synuclein activates microglia: a process leading to disease progression in Parkinson's disease.** *FASEB J* 2005, **19**(6):533–542.
68. Tian T, Wang Y, Wang H, Zhu Z, Xiao Z: **Visualizing of the cellular uptake and intracellular trafficking of exosomes by live-cell microscopy.** *J Cell Biochem* 2010, **111**(2):488–496.
69. Stoeckl L, Funk A, Kopitzki A, Brandenburg B, Oess S, Will H, Sirma H, Hildt E: **Identification of a structural motif crucial for infectivity of hepatitis B viruses.** *Proc Natl Acad Sci U S A* 2006, **103**(17):6730–6734.
70. Keller S, Ridinger J, Rupp AK, Janssen JW, Altevogt P: **Body fluid derived exosomes as a novel template for clinical diagnostics.** *Journal of translational medicine* 2011, **9**:86.
71. Papahadjopoulos D, Miller N: **Phospholipid model membranes. I. Structural characteristics of hydrated liquid crystals.** *Biochim Biophys Acta* 1967, **135**(4):624–638.
72. Denzer K, Kleijmeer MJ, Heijnen HF, Stoorvogel W, Geuze HJ: **Exosome: from internal vesicle of the multivesicular body to intercellular signaling device.** *J Cell Sci* 2000, **113**(Pt 19):3365–3374.
73. Johnstone RM: **Exosomes biological significance: A concise review.** *Blood cells, molecules & diseases* 2006, **36**(2):315–321.
74. Mathivanan S, Ji H, Simpson RJ: **Exosomes: extracellular organelles important in intercellular communication.** *Journal of proteomics* 2010, **73**(10):1907–1920.
75. Fader CM, Sanchez D, Furlan M, Colombo MI: **Induction of autophagy promotes fusion of multivesicular bodies with autophagic vacuoles in k562 cells.** *Traffic* 2008, **9**(2):230–250.
76. Klionsky DJ, Abeliovich H, Agostinis P, Agrawal DK, Aliev G, Askew DS, Baba M, Baehrecke EH, Bahr BA, Ballabio A, et al: **Guidelines for the use and interpretation of assays for monitoring autophagy in higher eukaryotes.** *Autophagy* 2008, **4**(2):151–175.
77. Faure J, Lachenal G, Court M, Hirrlinger J, Chatellard-Causse C, Blot B, Grange J, Schoehn G, Goldberg Y, Boyer V, et al: **Exosomes are released by cultured cortical neurones.** *Mol Cell Neurosci* 2006, **31**(4):642–648.
78. van Niel G, Porto-Carreiro I, Simoes S, Raposo G: **Exosomes: a common pathway for a specialized function.** *J Biochem* 2006, **140**(1):13–21.
79. Vella LJ, Sharples RA, Lawson VA, Masters CL, Cappai R, Hill AF: **Packaging of prions into exosomes is associated with a novel pathway of PrP processing.** *J Pathol* 2007, **211**(5):582–590.
80. Rajendran L, Honsho M, Zahn TR, Keller P, Geiger KD, Verkade P, Simons K: **Alzheimer's disease beta-amyloid peptides are released in association with exosomes.** *Proc Natl Acad Sci U S A* 2006, **103**(30):11172–11177.
81. Ghidoni R, Benussi L, Binetti G: **Exosomes: the Trojan horses of neurodegeneration.** *Med Hypotheses* 2008, **70**(6):1226–1227.
82. Hasegawa T, Konno M, Baba T, Sugeno N, Kikuchi A, Kobayashi M, Miura E, Tanaka N, Tamai K, Furukawa K, et al: **The AAA-ATPase VPS4 regulates extracellular secretion and lysosomal targeting of alpha-synuclein.** *PLoS One* 2011, **6**(12):e29460.

83. de Gassart A, Geminard C, Fevrier B, Raposo G, Vidal M: **Lipid raft-associated protein sorting in exosomes.** *Blood* 2003, **102**(13):4336–4344.
84. Fortin DL, Troyer MD, Nakamura K, Kubo S, Anthony MD, Edwards RH: **Lipid rafts mediate the synaptic localization of alpha-synuclein.** *J Neurosci* 2004, **24**(30):6715–6723.
85. Kubo S, Nemani VM, Chalkley RJ, Anthony MD, Hattori N, Mizuno Y, Edwards RH, Fortin DL: **A combinatorial code for the interaction of alpha-synuclein with membranes.** *J Biol Chem* 2005, **280**(36):31664–31672.
86. Pfefferkorn CM, Heinrich F, Sodt AJ, Maltsev AS, Pastor RW, Lee JC: **Depth of alpha-Synuclein in a Bilayer Determined by Fluorescence, Neutron Reflectometry, and Computation.** *Biophys J* 2012, **102**(3):613–621.
87. Aguzzi A, Rajendran L: **The transcellular spread of cytosolic amyloids, prions, and prionoids.** *Neuron* 2009, **64**(6):783–790.
88. Fang Y, Wu N, Gan X, Yan W, Morrell JC, Gould SJ: **Higher-order oligomerization targets plasma membrane proteins and HIV gag to exosomes.** *PLoS Biol* 2007, **5**(6):e158.
89. Perrin RJ, Woods WS, Clayton DF, George JM: **Exposure to long chain polyunsaturated fatty acids triggers rapid multimerization of synucleins.** *J Biol Chem* 2001, **276**(45):41958–41962.
90. Hughes RC: **Secretion of the galectin family of mammalian carbohydrate-binding proteins.** *Biochim Biophys Acta* 1999, **1473**(1):172–185.
91. Mehul B, Hughes RC: **Plasma membrane targeting, vesicular budding and release of galectin 3 from the cytoplasm of mammalian cells during secretion.** *J Cell Sci* 1997, **110**(Pt 10):1169–1178.
92. Liu J, Zhang JP, Shi M, Quinn T, Bradner J, Beyer R, Chen S, Zhang J: **Rab11a and HSP90 regulate recycling of extracellular alpha-synuclein.** *J Neurosci* 2009, **29**(5):1480–1485.
93. Fevrier B, Raposo G: **Exosomes: endosomal-derived vesicles shipping extracellular messages.** *Curr Opin Cell Biol* 2004, **16**(4):415–421.
94. Ratajczak J, Wysoczynski M, Hayek F, Janowska-Wieczorek A, Ratajczak MZ: **Membrane-derived microvesicles: important and underappreciated mediators of cell-to-cell communication.** *Leukemia* 2006, **20**(9):1487–1495.
95. Keller S, Sanderson MP, Stoeck A, Altevogt P: **Exosomes: from biogenesis and secretion to biological function.** *Immunol Lett* 2006, **107**(2):102–108.
96. Thery C, Zitvogel L, Amigorena S: **Exosomes: composition, biogenesis and function.** *Nat Rev Immunol* 2002, **2**(8):569–579.
97. Vincent JP, Magee T: **Argosomes: membrane fragments on the run.** *Trends Cell Biol* 2002, **12**(2):57–60.
98. Inouye H, Kirschner DA: **Alzheimer's beta-amyloid: insights into fibril formation and structure from Congo red binding.** *Subcell Biochem* 2005, **38**:203–224.
99. Sanchez I, Mahlke C, Yuan J: **Pivotal role of oligomerization in expanded polyglutamine neurodegenerative disorders.** *Nature* 2003, **421**(6921):373–379.
100. Rubinsztein DC, Gestwicki JE, Murphy LO, Klionsky DJ: **Potential therapeutic applications of autophagy.** *Nat Rev Drug Discov* 2007, **6**(4):304–312.
101. Bandyopadhyay U, Cuervo AM: **Chaperone-mediated autophagy in aging and neurodegeneration: lessons from alpha-synuclein.** *Exp Gerontol* 2007, **42**(1–2):120–128.
102. Bennett MC, Bishop JF, Leng Y, Chock PB, Chase TN, Mouradian MM: **Degradation of alpha-synuclein by the proteasome.** *J Biol Chem* 1999, **274**(48):33855–33858.
103. Vogiatzi T, Xilouri M, Vekrellis K, Stefanis L: **Wild type alpha-synuclein is degraded by chaperone-mediated autophagy and macroautophagy in neuronal cells.** *J Biol Chem* 2008, **283**(35):23542–23556.
104. Webb JL, Ravikumar B, Atkins J, Skepper JN, Rubinsztein DC: **Alpha-Synuclein is degraded by both autophagy and the proteasome.** *J Biol Chem* 2003, **278**(27):25009–25013.
105. Yamamoto A, Tagawa Y, Yoshimori T, Moriyama Y, Masaki R, Tashiro Y: **Bafilomycin A1 prevents maturation of autophagic vacuoles by inhibiting fusion between autophagosomes and lysosomes in rat hepatoma cell line, H-4-II-E cells.** *Cell Struct Funct* 1998, **23**(1):33–42.

doi:10.1186/1750-1326-7-42

Cite this article as: Danzer et al.: Exosomal cell-to-cell transmission of alpha synuclein oligomers. *Molecular Neurodegeneration* 2012 **7**:42.

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