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SPECIALTY SECTION This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Oncology

RECEIVED 03 August 2022 ACCEPTED 08 September 2022 PUBLISHED 23 September 2022

CITATION

Cherry ABC, Gherardin NA and Sikder HI (2022) Intracellular radar: Understanding $\gamma\delta$ T cell immune surveillance and implications for clinical strategies in oncology. *Front. Oncol.* 12:1011081. doi: 10.3389/fonc.2022.1011081

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Intracellular radar: Understanding $\gamma\delta$ T cell immune surveillance and implications for clinical strategies in oncology

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T cells play a key role in anticancer immunity, with responses mediated through a diversity of $\alpha\beta$ or $\gamma\delta$ T cell receptors. Although $\alpha\beta$ and $\gamma\delta$ T cells stem from common thymic precursors, the development and subsequent biological roles of these two subsets differ considerably. $\gamma\delta$ T cells are an unconventional T cell subset, uniquely poised between the adaptive and innate immune systems, that possess the ability to recognize intracellular disturbances and non-peptide-based antigens to eliminate tumors. These distinctive features of $\gamma\delta$ T cells have led to recent interest in developing $\gamma\delta$ -inspired therapies for treating cancer patients. In this minireview, we explore the biology of $\gamma\delta$ T cells, including how the $\gamma\delta$ T cell immune surveillance system can detect intracellular disturbances, and propose a framework to understand the $\gamma\delta$ T cell-inspired therapeutic strategies entering the clinic today.

KEYWORDS

 $\gamma\delta T,$ immunotherapy, anticancer, intracellular, surveillance, clinical, chimeric antigen receptor, gamma delta ($\gamma\delta)$ T cells

Introduction

The field of cancer immunology has grown around the idea that cancer elimination is a key role of the human immune system (1). Recent discoveries have revealed that T cells play a central role in immune surveillance, with responses mediated through a diversity of $\alpha\beta$ or $\gamma\delta$ T cell receptors (TCRs) (2). Despite sharing common thymic precursors, the development and subsequent biological roles of $\alpha\beta$ and $\gamma\delta$ T cells differ considerably (2). While $\alpha\beta$ T cell activation typically induces adaptive and long-lasting responses, $\gamma\delta$ T cells can respond rapidly and kill tumor cells directly (2, 3). In order to evade the host immune response, tumors have evolved diverse escape mechanisms, such as overexpression of programmed death ligand-1 (PD-L1) or immune exclusion, which are employed following malignant transformation (4–7). This convergent evolution of tumors toward a few immune escape mechanisms across a wide variety of cancer types underscores the central role of immune surveillance in preventing tumor growth, and has enabled the broad efficacy of immune checkpoint inhibitors (8).

Disturbances in intracellular processes, including metabolism, lipid composition and processing, nucleic acid packaging, glycans, biomechanical properties, and endoplasmic reticulum stress are key characteristics of tumor cells (8–13). Non-protein cellular signals of cancer have been recognized for over a century, since the metabolic shift from mitochondrial respiration to aerobic glycolysis was identified in the 1920s by Otto Warburg and colleagues as a fundamental trait of cancer (14). While "conventional" $\alpha\beta$ T cells detect tumor-derived peptide antigens to identify malignant cells, a heterogeneous family of "unconventional" T cells recognizes non-peptide-based antigens to facilitate tumor recognition (15).

 $\gamma\delta$ T cells are one such unconventional T cell subset, uniquely poised to have both adaptive and innate immune functions (16). Like $\alpha\beta$ T cells, $\gamma\delta$ T cells express a TCR on their cell surface that associates with the CD3 complex (16). During thymic development, successful rearrangement of a $\gamma\delta$ TCR drives commitment to the $\gamma\delta$ lineage *via* a different pathway than that of $\alpha\beta$ T cells, resulting in the innate-like phenotypic features that differentiate $\gamma\delta$ T cells from their $\alpha\beta$ counterparts (3). Despite the discovery of $\gamma\delta$ T cells over three decades ago, the mechanisms by which $\gamma\delta$ T cells recognize antigens remain poorly understood (17). Nonetheless, an emerging paradigm suggests that $\gamma\delta$ TCRs recognize and respond to modulation in cell surface "sentry" molecules indicative of intracellular stress (7, 18). The unique features of $\gamma\delta$ T cells have led to widespread interest in exploring their potential application in the development of novel therapeutics for cancer (3).

In this minireview we summarize the current understanding of how the human $\gamma\delta$ T cell immune surveillance system can detect intracellular disturbances and explore new therapeutic approaches undergoing preclinical and clinical development.

The science of $\gamma \delta$ T cells: Intracellular radar

Despite decades of research, our understanding of the mechanisms by which the immune system detects intracellular disturbances is still incomplete (17). However, recent research suggests that $\gamma\delta$ T cells monitor intracellular health *via* detection of transmembrane sentry proteins (18). When a cell's sentry proteins indicate intracellular stress, $\gamma\delta$ T cells can promptly deliver a cytotoxic response to eliminate the stressed cell, as well as release cytokines to orchestrate a broader immune response (3, 17).

Human $\gamma\delta$ T cells are themselves heterogeneous, and are broadly categorized into two major subsets based on their distinct antigen-recognition and phenotypic features, described according to the antigen-recognition domain of their TCR- δ chains: V δ 2+ and

non-V δ 2 (19, 20). The latter consists primarily of V δ 1+ cells, although other less well-characterized cells expressing VS3-VS8 also exist (20, 21). T cells carrying the V δ 2+ chain are more common in circulation, and respond to intracellular accumulation of tumor-associated metabolic byproducts by interacting with a family of sentry proteins called butyrophilins (BTNs) (22, 23). Non-VS2 cells, on the other hand, respond to a variety of stressassociated sentry proteins, the most well-characterized of which are major histocompatibility complex (MHC) class I-like proteins (19, 20, 24). Each $\gamma\delta$ T cell subset also possesses unique tissue localization and homing capabilities, as evidenced by V\delta2+ cells tending to be more enriched in blood, versus non-V δ 2 cells, which are often found enriched in tissues such as the gut (25, 26). Here, we provide an overview of these two distinct groups of yo T cells, including what is known about intracellular stress responses and how $\gamma\delta$ TCRs are able to detect these changes (Figure 1).

Phosphoantigens, butyrophilins, and V\delta2+ $\gamma\delta$ T cells

Most V δ 2+ $\gamma\delta$ T cells express an invariant TCR, imbuing responsiveness to the intracellular presence of low molecular weight, phosphorylated adducts or "phosphoantigens" (pAgs) (27). pAgs are found in healthy cells at low levels as part of the mevalonate synthesis pathway, but elevated levels signal cellular dysregulation (28). For example, highly potent pAgs such as (E)-4-Hydroxy-3methyl-2-but-2-enyl pyrophosphate are generated as byproducts of microbial isoprenoid synthesis *via* the methylerythritol phosphate pathway, and thus represent a molecular signature of microbial infection (29). V δ 2+ $\gamma\delta$ T cells also respond to mammalian pAgs of lower potency, such as the isoprenoid precursor isopentyl pyrophosphate, that accumulate from an overactive mevalonate metabolic pathway during cancer development (30). Accordingly, V δ 2+ $\gamma\delta$ T cells act as immune sentinels, monitoring intracellular stress *via* a conserved molecular pattern.

The precise mechanism by which pAgs are recognized by the $\gamma\delta$ TCR has been a conundrum for the field. Recent studies, however, have revealed that the process is dependent on two members of the BTN protein family, BTN3A1 and BTN2A1 (31, 32). Here, pAgs bind to the intracellular domain of BTN3A1, resulting in an undefined process of extracellular remodeling to reveal an antigenic target for the TCR, which includes direct interactions with BTN2A1 (25, 31, 32). This process is a distinct mode of antigen-sensing compared to $\alpha\beta$ TCR recognition of peptide-MHC, although the molecular details are yet to be revealed (27).

Other intracellular stressors, sentry proteins, and non-V δ 2 $\gamma\delta$ T cells

Like V δ 2+ cells, non-V δ 2 $\gamma\delta$ T cells exhibit both antimicrobial and antitumor potential at a population level; however, the antigens



recognized by non-V δ 2 cells are less well understood (19, 20). Unlike their invariant V δ 2+ counterparts, non-V δ 2 $\gamma\delta$ T cells exhibit extensive TCR diversity *via* TCR- γ chain pairing and more extensive complementarity-determining region 3 variation, and thus have highly variable antigen-recognition potential (19, 20). Despite a paucity of literature on non-V δ 2 antigen identification, the antigens described to date appear to be an array of molecules indicative of various forms of intracellular stress (18).

One major group of antigens consists of monomorphic MHC Ilike molecules, including the CD1 family and the MHC-related protein 1 (MR1) (24, 33-38). CD1 sentry proteins are involved in the presentation of lipid antigens to various subsets of V δ 1+ $\gamma\delta$ T cells (33-38). MR1 proteins present microbial vitamin-B metabolite antigens, as well as undefined tumor-associated antigens to various subsets of non-V δ 2 $\gamma\delta$ T cells (24, 39–42). While research on CD1/ MR1-y T cell interactions continues, early evidence indicates that subsets of $\gamma\delta$ T cells also respond to a plethora of other intracellular stress signals through sentry binding partners, such as the endothelial cell protein C receptor (EPCR), MHC class I chainrelated protein A, and UL16-binding protein, which are MHC Ilike, and annexin A2, EphA2, and phycoerythrin, most of which still remain poorly understood (43-48). Taken together, it appears that non-V $\delta 2 \gamma \delta$ T cells interact with a wide range of sentry proteins to detect a broad array of intracellular stresses.

The mode of $\gamma\delta$ TCR recognition of MHC I-like molecules appears more varied than $\alpha\beta$ TCRs, which conform to a

relatively rigid docking mode atop peptide-MHC complexes (49). In contrast, $\gamma\delta$ TCRs appear to dock across the entire antigen-presenting platform, and in some cases, adopt highly unusual modes of recognition (41, 43). For example, some MR1-restricted TCRs dock with MR1 underneath the antigen-binding cleft, while others such as EPCR-restricted clones are able to recognize the underside of the antigen-binding platform (41, 43). Thus, while these sentries are bona fide antigen-presenting molecules, and many $\gamma\delta$ TCRs exhibit antigen dependence, other TCRs seem to bind in an antigen-independent manner (50).

Role in anticancer immunosurveillance

The research described here has shed light on how sentry proteins monitor for intracellular stress and convey that information to $\gamma\delta$ T cells. While it remains an open question whether certain parts of these systems respond preferentially to infection, cancer, or other stressors, there is abundant evidence demonstrating that $\gamma\delta$ T cells play an active role in cancer surveillance. This includes observation of $\gamma\delta$ T cell enrichment across a broad range of tumor types, *in vitro* evidence of direct V δ 2+ T cell tumor cell killing in a BTN-dependent manner, and an analysis of gene expression signatures from 18,000 tumors that demonstrated the $\gamma\delta$ T cell signature as the most favorable for patient survival among 22 immune cell subtypes (51). $\gamma\delta$ T

cell tumor recognition and elimination is also supported by the expression of receptors associated with natural killer cell activity, including NKG2D, DNAM-1, and other natural cytotoxicity receptors such as NKp30, NKp44, and NKp46.

While the evidence is clear that $\gamma\delta$ T cells play an active role against tumors, an important unanswered question is how tumors evade $\gamma\delta$ T cell recognition and destruction. More broadly, identifying the mechanisms by which tumors evade immunological surveillance is a rapidly evolving area of research, and disentangling the separate contributions of the $\alpha\beta$ vs. $\gamma\delta$ T cell surveillance systems remains difficult (52). There are a number of mechanisms by which tumor cells directly or indirectly suppress T cell immune responses, many of which likely apply to both the $\alpha\beta$ and $\gamma\delta$ T cell compartments (53). In particular, expression of immune checkpoints such as PD-L1 have been successfully targeted with cancer immunotherapies (8). Additionally, cancer immunoediting of surface-expressed molecules involved in antigen presentation to $\gamma\delta$ T cells has been observed, which could represent another mechanism by which tumors evade recognition by $\gamma\delta$ T cells (54–56). Finally, in some environments, $\gamma\delta$ T cells could also play an immunosuppressive role, which may contribute to tumor immune evasion (57).

Overall, these advances show that $\gamma\delta$ T cells monitor the intracellular well-being of tissues *via* a network of sentry proteins. When a cell experiences intracellular distress, $\gamma\delta$ T cells can directly kill the cell and/or recruit other arms of the immune system to respond. While $\gamma\delta$ T cells are involved in routine anticancer immunosurveillance, ongoing research is focused on improving our understanding of the underlying mechanisms.

Discussion: Clinical strategies and progress to date

There is strong biological rationale for harnessing the $\gamma\delta$ T cell surveillance system to enhance antitumor immunity. Despite unprecedented clinical success with checkpoint inhibitor therapies over the past decade, many patients do not respond and most will eventually relapse (58). While other $\alpha\beta$ T cell-based approaches are being developed to address these limitations, such as engineered chimeric antigen receptor (CAR)-T cells, there is increasing interest among investigators to develop novel $\gamma\delta$ T cell-based therapeutics with superior activity to fight against human cancers (7). However, it is yet unknown how $\gamma\delta$ T cell-inspired therapies will compare to other existing and emerging types of cell therapies.

A broad interventional toolkit is investigating the antitumor activity of $\gamma\delta$ T cell surveillance, including cell therapy, protein engineering, and direct modulation by monoclonal antibody (mAb) or small molecules (Figure 2). At present, there is no aligned vision for how $\gamma\delta$ T cell biology can best be leveraged to fight cancer, and thus researchers are manipulating various aspects of the immunological surveillance system in an effort to maximize antitumor activity (59). In the following section, we propose a framework for understanding the $\gamma\delta$ T cell-inspired therapeutic strategies in clinics today.

Innate action: $\gamma\delta$ T cells kill tumor cells (strategies 1 & 2)

 $\gamma\delta$ T cells can directly kill tumor cells with innate-like cytotoxicity. In this way, therapeutic approaches leveraging the ability of $\gamma\delta$ T cells to kill tumors directly are similar to CAR-NK approaches, which direct natural killer cells' innate cytotoxicity against cancer (60). Innate action therapies seek to either recruit endogenous $\gamma\delta$ T cells to the tumor (Strategy 1) or expand and engineer $\gamma\delta$ T cells *ex vivo* to deliver as therapy (Strategy 2).

Strategy 1: Recruit endogenous $\gamma\delta$ T cells

A strategy involving endogenous $\gamma\delta$ T cells is complicated by the fact that the patient's immune system has already allowed the tumor to grow. To overcome this challenge, researchers are working on ways to reinvigorate the endogenous $\gamma\delta$ T cell antitumor response. For example, researchers at ImCheck are attempting to increase signaling from the sentry protein using a mAb activating BTN3A (61). Other examples include Lava Therapeutics and Shattuck Labs, both of which are developing bispecific proteins designed to bring $\gamma\delta$ T cells within close proximity to tumors (62). Two divergent strategies are being employed to attract the $\gamma\delta$ T cells (1): Lava Therapeutics has developed an anti-V γ 9V δ 2 nanobody arm; and (2) Shattuck Labs is utilizing a piece of V γ 9V δ 2's natural ligand, BTN, as part of its heterodimer (62, 63). If these approaches prove successful, they will have the distinct advantage of being off-the-shelf, protein-based therapeutics, which are easier to manufacture and deliver than cell therapies. Of note, these examples rely on V δ 2+ cells, rather than V δ 1+ cells, and thus assuming that BTNbased signaling will provide maximal antitumor efficacy, but not considering other types of intracellular stress.

Strategy 2: Expand and engineer $\gamma \delta$ T cells *ex vivo* to deliver as therapy

Another innate strategy involves adding a CAR or TCR to the surface of a $\gamma\delta$ T cell body for increased specificity against cancer cells. One advantage of this approach is that CARs or TCRs that have been clinically validated as cell therapies can be brought into the $\gamma\delta$ T cell paradigm. For example, Adicet is collecting and expanding V δ 1+ T cells from donors, then adding a CAR construct targeted against CD20 with a 4-1BB costimulatory domain to these V δ 1+ cells (64). Another company, In8bio, is seeking to harness $\gamma\delta$ T cells' natural ability to kill stressed cells by including a chemoresistance gene in its cell therapy, enabling chemotherapy to be administered without damaging the $\gamma\delta$ -based CAR-T therapy (65). Most companies employing Strategy 2 are developing allogeneic cell therapies, leveraging $\gamma\delta$ T cells' natural



 $\gamma\delta$ T Cell-Inspired Clinical Strategies. (A) $\gamma\delta$ T cell biology may be employed to eliminate tumors by four different strategies. In strategy 1, endogenous $\gamma\delta$ T cells kill the tumor. In strategy 2, exogenous engineered $\gamma\delta$ T cell therapies kill the tumor. In strategy 3, $\gamma\delta$ T cell receptor biology is used to recruit an endogenous $\alpha\beta$ T cell adaptive response to stressed tumor cells. In strategy 4, $\gamma\delta$ T cell receptor biology is used as a targeting strategy for traditional $\alpha\beta$ CAR-T cells to identify and attack stressed tumor cells. In each of these strategies, researchers can either employ validated protein constructs (e.g., CARs using GP3C – 4-1BB – CD3 ζ from Adicet) or engineer innovative protein structures (e.g., fusion protein chains of CD19 – Fc – BTN3A1/BTN2A1 from Shattuck Labs) to maximize anti-tumor activity. (B) Researchers and companies leading investigation into each clinical strategy.

avoidance of graft vs. host disease, which can generate hundreds or thousands of doses in a single production run. However, the long-term success of these approaches rests on a crucial question: Will cell therapies using $\gamma\delta$ T cells demonstrate the same *in vivo* expansion, persistence, and long-term activity that other CAR-T therapies offer? Currently, this question remains unanswered. The success of many CAR-T products available today is attributable to the adaptive immune behavior of the carrier $\alpha\beta$ T cell, and whether the more innate-like $\gamma\delta$ T cell will deliver the same results has yet to be seen.

Adaptive action: $\alpha\beta$ T cells kill tumor cells (strategies 3 & 4)

Instead of using $\gamma\delta$ T cells directly, some investigators are seeking to exploit their ability to detect and respond to intracellular stress. As a field, cell therapy has remained highly dependent on tumor cell-specific antigens; however, with $\gamma\delta$ TCR biology, investigators are seeking to link the innate and adaptive immune systems, using sentries' stress signals as an "antigen" to trigger adaptive immune activity. These adaptive

action therapies can rely on either activation of endogenous $\alpha\beta$ T cell responses against stressed cells (Strategy 3) or an $\alpha\beta$ CAR-T therapy built using a $\gamma\delta$ -inspired CAR or TCR (Strategy 4).

Strategy 3: Activation of endogenous $\alpha\beta$ T cells against stressed cells

CD3 bispecific proteins exert antitumor effects by binding to a cancer-specific antigen at the same time they bind to CD3, a molecule expressed on both CD8+ and CD4+ T cells. This binding event allows the T cells to directly kill the cancer cell, overcoming usual antigen presentation and MHC matching processes. Researchers from the University of Utrecht have developed a bispecific protein that includes both a CD3binding arm and V γ 9V δ 2 TCR (66). As previously discussed, V δ 2+ TCRs bind to activated BTN2A1 protein sentries, bringing CD8+ and CD4+ T cells within close proximity to stressed tumor cells. Akin to Strategy 1, this approach relies entirely on the V γ 9V δ 2/BTN axis, rather than including V δ 1+ cells, and if successful it could offer logistical and manufacturing advantages.

Strategy 4: $\alpha\beta$ CAR-T therapy built using a $\gamma\delta\text{-}$ inspired CAR or TCR

The most nascent strategy in development is the addition of $\gamma\delta$ TCRs to $\alpha\beta$ T cells. In theory, this approach could merge the powerful, long-lasting benefit of traditional CAR-T therapy with $\gamma\delta$ T cells' ability to detect intracellular stress in a non-MHCrestricted manner. However, investigators must address at least three key challenges before this strategy can become a reality: 1) Identify a $\gamma\delta$ TCR sequence that can reliably recognize a stressed sentry protein, whether V δ 2+ or V δ 1+. Sequencing of tumor-reactive $\gamma\delta$ TCRs is still in early stages, and it is not yet known whether a single TCR would be broadly effective or if different tumors would require different TCRs. The specificity of the TCR for stressed vs. non-stressed cells will also be crucial, as this will provide a benchmark for the degree of off-target activity. Early work in this area was conducted by Wei He and colleagues, who added tumor-specific $\gamma\delta$ TCRs to $\alpha\beta$ T cells and observed a reduction in tumor growth in a mouse model (67). 2) Determine which parts of the chimeric receptor should be derived from $\gamma\delta$ T cells vs. traditional CAR/TCR molecules, and how these components impact the therapy's behavior in vivo. For example, Gadeta is developing $\alpha\beta$ T cells equipped with $\gamma\delta$ TCRs to detect stress signals from sentry proteins (68). However, Eureka Therapeutics is using a hybrid CAR receptor that includes $\gamma\delta$ TCR sequence on the intracellular portion of the CAR, while the extracellular portion is a traditional anti-CD19 (69). The impact of receptor designs on safety, including cytokine release syndrome, should also be determined. 3) Finally, $\alpha\beta$ -based therapies will initially be limited to the autologous setting, until an extra step is introduced to genetically modify the source material. Accordingly, both $\gamma\delta$ inspired $\alpha\beta$ CAR-T cells in the clinic today are autologous.

Conclusion and remaining questions

 $\gamma\delta$ T cell-based therapies represent an emerging class of anticancer treatments, with a growing pipeline of investigational agents in varied stages of clinical development. In this minireview, we explored the biology of $\gamma\delta$ cells and have proposed a framework for understanding the $\gamma\delta$ T cellinspired therapeutic strategies currently in use. Despite recent progress, there remains a need for more comprehensive understanding of $\gamma\delta$ immune surveillance, as the success of clinical strategies depends on several key questions that remain unanswered. These questions include: What protein sequence will be the most effective bait to recruit endogenous $\gamma\delta$ T cells? Will cell therapies using $\gamma\delta$ T cell biology demonstrate the same expansion and long-term activity that traditional CAR-T therapies have shown? Is there a $\gamma\delta$ TCR sequence that would consistently recognize stressed cells across patients, and if so, how can it be identified? Can $\gamma\delta$ T cell biology be used to design a traditional CAR-T therapy that activates the adaptive immune system against stressed cells? The answers to these questions will determine which $\gamma \delta$ T cell-inspired clinical strategy will be more effective in the fight against cancer.

Author contributions

AC and NG drafted the manuscript and designed the figures. HS discussed and reviewed the final manuscript. All authors contributed to the article and approved the submitted version.

Funding

The authors received no financial support for the research or authorship of this article. NAG was supported by an Australian Research Council Discovery Early Career Award Fellowship (DE210100705). Publication was supported by Axiom Healthcare Strategies, LLS (Princeton, NJ).

Acknowledgments

Medical writing and editorial support was provided by Caleb Rans, of Rans Medical Communications (Vancouver, Canada) and was funded by Axiom Healthcare Strategies.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Finn OJ. A believer's overview of cancer immunosurveillance and immunotherapy. J Immunol (2018) 200:385–91. doi: 10.4049/jimmunol.1701302

2. Chien YH, Meyer C, Bonneville M. γδ T cells: first line of defense and beyond. Annu Rev Immunol (2014) 32:121–55. doi: 10.1146/annurev-immunol-032713-120216

3. Gustafsson K, Herrmann T, Dieli F. Editorial: Understanding gamma delta T cell multifunctionality - towards immunotherapeutic applications. *Front Immunol* (2020) 11:921. doi: 10.3389/fimmu.2020.00921

4. Cha JH, Chan LC, Li CW, Hsu JL, Hung MC. Mechanisms controlling PD-L1 expression in cancer. *Mol Cell* (2019) 76(3):359-70. doi: 10.1016/j.molcel.2019.09.030

5. Pai SI, Cesano A, Marincola FM. The paradox of cancer immune exclusion: Immune oncology next frontier. *Cancer Treat Res* (2020) 180:173–95. doi: 10.1007/ 978-3-030-38862-1_6

6. Baugh R, Khalique H, Seymour LW. Convergent evolution by cancer and viruses in evading the NKG2D immune response. *Cancers (Basel)* (2020) 12 (12):3827. doi: 10.3390/cancers12123827

7. Sebestyen Z, Prinz I, Déchanet-Merville J, Silva-Santos B, Kuball J. Translating gammadelta ($\gamma\delta$) T cells and their receptors into cancer cell therapies. *Nat Rev Drug Discovery* (2020) 19(3):169–84. doi: 10.1038/s41573-019-0038-z

8. Robert C. A decade of immune-checkpoint inhibitors in cancer therapy. *Nat Commun* (2020) 11:3801. doi: 10.1038/s41467-020-17670-y

9. DeBerardinis RJ. Is cancer a disease of abnormal cellular metabolism? new angles on an old idea. *Genet Med* (2008) 10(11):767-77. doi: 10.1097/GIM.0b013e31818b0d9b

10. Zhang F, Du G. Dysregulated lipid metabolism in cancer. *World J Biol Chem* (2012) 3(8):167–74. doi: 10.4331/wjbc.v3.i8.167

11. Wang Z, Chen K, Jia Y, Chuang JC, Sun X, Lin YH, et al. Dual ARID1A/ ARID1B loss leads to rapid carcinogenesis and disruptive redistribution of BAF complexes. *Nat Cancer* (2020) 1(9):909–22. doi: 10.1038/s43018-020-00109-0

12. Mereiter S, Balmaña M, Campos D, Gomes J, Reis CA. Glycosylation in the era of cancer-targeted therapy: Where are we heading? *Cancer Cell* (2019) 36(1):6–16. doi: 10.1016/j.ccell.2019.06.006

13. Runel G, Lopez-Ramirez N, Chlasta J, Masse I. Biomechanical properties of cancer cells. *Cells* (2021) 10(4):887. doi: 10.3390/cells10040887

14. Cassim S, Vučetić M, Ždralević M, Pouyssegur J. Warburg and beyond: The power of mitochondrial metabolism to collaborate or replace fermentative glycolysis in cancer. *Cancers (Basel)* (2020) 12(5):1119. doi: 10.3390/ cancers12051119

15. Godfrey DI, Le Nours J, Andrews DM, Uldrich AP, Rossjohn J. Unconventional T cell targets for cancer immunotherapy. *Immunity* (2018) 48 (3):453-73. doi: 10.1016/j.immuni.2018.03.009

16. Pistoia V, Tumino N, Vacca P, Veneziani I, Moretta A, Locatelli F, et al. Human $\gamma\delta$ T-cells: From surface receptors to the therapy of high-risk leukemias. Front Immunol (2018) 9:984. doi: 10.3389/fimmu.2018.00984

17. Simões AE, Di Lorenzo B, Silva-Santos B. Molecular determinants of target cell recognition by human $\gamma\delta$ T cells. Front Immunol (2018) 9:929. doi: 10.3389/fimmu.2018.00929

18. Willcox BE, Willcox CR. $\gamma\delta$ TCR ligands: the quest to solve a 500-million-year-old mystery. Nat Immunol (2019) 20(2):121–8. doi: 10.1038/s41590-018-0304-y

19. Davey MS, Willcox CR, Hunter S, Kasatskaya SA, Remmerswaal EBM, Salim M, et al. The human V δ 2+ T-cell compartment comprises distinct innate-like V γ 9+ and adaptive V γ 9- subsets. *Nat Commun* (2018) 9(1):1760. doi: 10.1038/s41467-018-04076-0

20. Davey MS, Willcox CR, Hunter S, Oo YH, Willcox BE. V $\delta2+$ T cells-two subsets for the price of one. Front Immunol (2018) 9:2106: doi: 10.3389/fimmu.2018.02106

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21. Biradar S, Lotze MT, Mailliard RB. The unknown unknowns: Recovering gamma-delta T cells for control of human immunodeficiency virus (HIV). *Viruses* (2020) 12(12):1455. doi: 10.3390/v12121455

22. Laplagne C, Ligat L, Foote J, Lopez F, Fournié JJ, Laurent C, et al. Self-activation of V γ 9V δ 2 T cells by exogenous phosphoantigens involves TCR and butyrophilins. *Cell Mol Immunol* (2021) 18(8):1861–70. doi: 10.1038/s41423-021-00720-w

23. Rigau M, Ostrouska S, Fulford TS, Johnson DN, Woods K, Ruan Z, et al. Butyrophilin 2A1 is essential for phosphoantigen reactivity by $\gamma\delta$ T cells. *Science* (2020) 367(6478):eaay5516. doi: 10.1126/science.aay5516

24. Kjer-Nielsen L, Patel O, Corbett AJ, Le Nours J, Meehan B, Liu L, et al. MR1 presents microbial vitamin b metabolites to MAIT cells. *Nature* (2012) 491 (7426):717–23. doi: 10.1038/nature11605

25. Parker CM, Groh V, Band H, Porcelli SA, Morita C, Fabbi M, et al. Evidence for extrathymic changes in the T cell receptor gamma/delta repertoire. *J Exp Med* (1990) 171(5):1597–612. doi: 10.1084/jem.171.5.1597

26. Groh VE, Porcelli S, Fabbi M, Lanier LL, Picker LJ, Anderson T, et al. Human lymphocytes bearing T cell receptor gamma/delta are phenotypically diverse and evenly distributed throughout the lymphoid system. *J Exp Med* (1989) 169(4):1277–94. doi: 10.1084/jem.169.4.1277

27. Uldrich AP, Rigau M, Godfrey DI. Immune recognition of phosphoantigenbutyrophilin molecular complexes by $\gamma\delta$ T cells. *Immunol Rev* (2020) 298(1):74–83. doi: 10.1111/imr.12923

28. Gogoi D, Chiplunkar SV. Targeting gamma delta T cells for cancer immunotherapy: Bench to bedside. *Indian J Med Res* (2013) 138(5):755-61.

29. Hintz M, Reichenberg A, Altincicek B, Bahr U, Gschwind RM, Kollas AK, et al. Identification of (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate as a major activator for human gammadelta T cells in escherichia coli. *FEBS Lett* (2001) 509(2):317–22. doi: 10.1016/s0014-5793(01)03191-x

30. Tanaka Y, Morita CT, Tanaka Y, Nieves E, Brenner MB, Bloom BR. Natural and synthetic non-peptide antigens recognized by human gamma delta T cells. *Nature* (1995) 375(6527):155–8. doi: 10.1038/375155a0

31. Harly C, Guillaume Y, Nedellec S, Peigné CM, Mönkkönen H, Mönkkönen J, et al. Key implication of CD277/butyrophilin-3 (BTN3A) in cellular stress sensing by a major human $\gamma\delta$ T-cell subset. *Blood* (2012) 120(11):2269–79. doi: 10.1182/blood-2012-05-430470

 Sandstrom A, Peigné CM, Léger A, Crooks JE, Konczak F, Gesnel MC, et al. The intracellular B30.2 domain of butyrophilin 3A1 binds phosphoantigens to mediate activation of human Vγ9Vδ2 T cells. *Immunity* (2014) 40(4):490–500. doi: 10.1016/j.immuni.2014.03.003

33. Uldrich AP, Le Nours J, Pellicci DG, Gherardin NA, McPherson KG, Lim RT, et al. CD1d-lipid antigen recognition by the $\gamma\delta$ TCR. *Nat Immunol* (2013) 14 (11):1137–45. doi: 10.1038/ni.2713

34. Bai L, Picard D, Anderson B, Chaudhary V, Luoma A, Jabri B, et al. The majority of CD1d-sulfatide-specific T cells in human blood use a semiinvariant V δ 1 TCR. *Eur J Immunol* (2012) 42(9):2505–10. doi: 10.1002/eji.201242531

35. Porcelli S, Brenner MB, Greenstein JL, Balk SP, Terhorst C, Bleicher PA. Recognition of cluster of differentiation 1 antigens by human CD4-CD8-cytolytic T lymphocytes. *Nature* (1989) 341(6241):447–50. doi: 10.1038/341447a0

36. Spada FM, Grant EP, Peters PJ, Sugita M, Melián A, Leslie DS, et al. Selfrecognition of CD1 by gamma/delta T cells: Implications for innate immunity. *J Exp Med* (2000) 191(6):937-48. doi: 10.1084/jem.191.6.937

 Reijneveld JF, Ocampo TA, Shahine A, Gully BS, Vantourout P, Hayday AC, et al. Human γδ T cells recognize CD1b by two distinct mechanisms. *PNAS* (2020) 117(37):22944–52. doi: 10.1073/pnas.2010545117

38. Wegrecki M, Ocampo TA, Gunasinghe SD, von Borstel A, Tin SY, Reijneveld JF, et al. Atypical sideways recognition of CD1a by autoreactive $\gamma\delta$ T cell receptors. *Nat Commun* (2022) 13(1):3872. doi: 10.1038/s41467-022-31443-9

39. Crowther MD, Dolton G, Legut M, Caillaud ME, Lloyd A, Attaf M, et al. Genome-wide CRISPR-Cas9 screening reveals ubiquitous T cell cancer targeting

via the monomorphic MHC class I-related protein MR1. Nat Immunol (2020) 21 (2):178–85. doi: 10.1038/s41590-019-0578-8

40. Lepore M, Kalinichenko A, Calogero S, Kumar P, Paleja B, Schmaler M, et al. Functionally diverse human T cells recognize non-microbial antigens presented by MR1. *Elife* (2017) 6:e24476. doi: 10.7554/eLife.24476

41. Le Nours J, Gherardin NA, Ramarathinam SH, Awad W, Wiede F, Gully BS, et al. A class of $\gamma\delta$ T cell receptors recognize the underside of the antigen-presenting molecule MR1. *Science* (2019) 366(6472):1522–7. doi: 10.1126/science.aav3900

42. Rice MT, von Borstel A, Chevour P, Awad W, Howson LJ, Littler DR, et al. Recognition of the antigen-presenting molecule MR1 by a V δ 3+ $\gamma\delta$ T cell receptor. PNAS (2021) 118(49):e2110288118. doi: 10.1073/pnas.2110288118

43. Willcox CR, Pitard V, Netzer S, Couzi L, Salim M, Silberzahn T, et al. Cytomegalovirus and tumor stress surveillance by binding of a human $\gamma\delta$ T cell antigen receptor to endothelial protein c receptor. *Nat Immunol* (2012) 13(9):872–79. doi: 10.1038/ni.2394

44. Salih HR, Rammensee HG, Steinle A. Cutting edge: Down-regulation of MICA on human tumors by proteolytic shedding. *J Immunol* (2002) 169(8):4098–102. doi: 10.4049/jimmunol.169.8.4098

45. Kong Y, Cao W, Xi X, Ma C, Cui L, He W. The NKG2D ligand ULBP4 binds to TCRgamma9/delta2 and induces cytotoxicity to tumor cells through both TCRgammadelta and NKG2D. *Blood* (2009) 114(2):310–7. doi: 10.1182/blood-2008-12-196287

46. Marlin R, Pappalardo A, Kaminski H, Willcox CR, Pitard V, Netzer S, et al. Sensing of cell stress by human $\gamma\delta$ TCR-dependent recognition of annexin A2. PNAS (2017) 114(12):3163–68. doi: 10.1073/pnas.1621052114

47. Harly C, Joyce SP, Domblides C, Bachelet T, Pitard V, Mannat C, et al. Human $\gamma\delta$ T cell sensing of AMPK-dependent metabolic tumor reprogramming through TCR recognition of EphA2. *Sci Immunol* (2021) 6(61):eaba9010. doi: 10.1126/sciimmunol.aba9010

48. Zeng X, Wei YL, Huang J, Newell EW, Yu H, Kidd BA, et al. $\gamma\delta$ T cells recognize a microbial encoded b cell antigen to initiate a rapid antigen-specific interleukin-17 response. *Immunity* (2012) 37(3):524–34. doi: 10.1016/j.immuni.2012.06.011

49. Rossjohn J, Gras S, Miles JJ, Turner SJ, Godfrey DI, McCluskey J. T Cell antigen receptor recognition of antigen-presenting molecules. *Annu Rev Immunol* (2015) 33:169–200. doi: 10.1146/annurev-immunol-032414-112334

50. Gully BS, Rossjohn J, Davey MS. Our evolving understanding of the role of the $\gamma\delta$ T cell receptor in $\gamma\delta$ T cell mediated immunity. *Biochem Soc Trans* (2021) 49 (5):1985–95. doi: 10.1042/BST20200890

51. Gentles AJ, Newman AM, Liu CL, Bratman SV, Feng W, Kim D, et al. The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nat Med* (2015) 21(8):938–45. doi: 10.1038/nm.3909

52. Beatty GL, Gladney WL. Immune escape mechanisms as a guide for cancer immunotherapy. *Clin Cancer Res* (2015) 21(4):687–92. doi: 10.1158/1078-0432.CCR-14-1860

53. Labani-Motlagh A, Ashja-Mahdavi M, Loskog A. The tumor microenvironment: A milieu hindering and obstructing antitumor immune responses. *Front Immunol* (2020) 11:940. doi: 10.3389/fimmu.2020.00940

54. Dhatchinamoorthy K, Colbert JD, Rock KL. Cancer immune evasion through loss of MHC class I antigen presentation. *Front Immunol* (2021) 12:636568. doi: 10.3389/fimmu.2021.636568

55. Minchom A, Yuan W, Crespo M, Gurel B, Figueiredo I, Wotherspoon A, et al. Molecular and immunological features of a prolonged exceptional responder with malignant pleural mesothelioma treated initially and rechallenged with

pembrolizumab. J Immunother Cancer (2020) 8(1):e000713. doi: 10.1136/jitc-2020-000713

56. Hix LM, Shi YH, Brutkiewicz RR, Stein PL, Wang CR, Zhang M. CD1dexpressing breast cancer cells modulate NKT cell-mediated antitumor immunity in a murine model of breast cancer metastasis. *PloS One* (2011) 6(6):e20702. doi: 10.1371/journal.pone.0020702

57. Kabelitz D, Serrano R, Kouakanou L, Peters C, Kalyan S. Cancer immunotherapy with $\gamma\delta$ T cells: Many paths ahead of us. Cell Mol Immunol (2020) 17(9):925–39. doi: 10.1038/s41423-020-0504-x

58. Hegde A, Andreev-Drakhlin AY, Roszik J, Huang L, Liu S, Hess K, et al. Responsiveness to immune checkpoint inhibitors versus other systemic therapies in RET-aberrant malignancies. *ESMO Open* (2020) 5(5):e000799. doi: 10.1136/ esmoopen-2020-000799

59. Li Y, Li G, Zhang J, Wu X, Chen X. The dual roles of human $\gamma\delta$ T cells: Antitumor or tumor-promoting. Front Immunol (2021) 11:619954. doi: 10.3389/fimmu.2020.619954

60. Zhang C, Hu Y, Shi C. Targeting natural killer cells for tumor immunotherapy. Front Immunol (2020) 11:60. doi: 10.3389/fimmu.2020.00060

61. De Gassart A, Le KS, Brune P, Agaugué S, Sims J, Goubard A, et al. Development of ICT01, a first-in-class, anti-BTN3A antibody for activating V γ 9V δ 2 T cell-mediated antitumor immune response. *Sci Transl Med* (2021) 13 (616):eabj0835. doi: 10.1126/scitranslmed.abj0835

62. Lava Therapeutics. Utrecht And Philadelphia: GlobeNewswire; 2022 may 17. press release, LAVA therapeutics provides business update and reports first quarter financial results. Available at: https://ir.lavatherapeutics.com/news-release-details/lava-therapeutics-provides-business-update-and-reports-first-0.

63. Shattuck Labs. Austin And Durham: GlobeNewswire; 2022 may 12. press release, shattuck labs reports first quarter 2022 financial results and recent business highlights. Available at: https://ir.shattucklabs.com/investors/news-events/press-releases/news-details/2022/Shattuck-Labs-Reports-First-Quarter-2022-Financial-Results-and-Recent-Business-Highlights-05-12-2022/default.aspx.

64. Bartkowiak T, Curran MA. 4-1BB agonists: Multi-potent potentiators of tumor immunity. *Front Oncol* (2015) 5:117. doi: 10.3389/fonc.2015.00117

65. IN8bio. New York: GlobeNewswire; 2022 July 21. press release, newly issued European patent broadens IN8bio's drug resistant immunotherapy (DRI) platform. Available at: https://www.globenewswire.com/en/news-release/2022/07/21/ 2483478/0/en/Newly-Issued-European-Patent-Broadens-IN8bio-s-Drug-Resistant-Immunotherapy-DRI-Platform.html.

66. van Diest E, Hernández López P, Meringa AD, Vyborova A, Karaiskaki F, Heijhuurs S, et al. Gamma delta TCR anti-CD3 bispecific molecules (GABs) as novel immunotherapeutic compounds. J Immunother Cancer (2021) 9(11): e003850. doi: 10.1136/jitc-2021-003850

67. He K, You H, Li Y, Cui L, Zhang J, He W. TCR γ 4 δ 1-engineered $\alpha\beta$ T cells exhibit effective antitumor activity. *Mol Med* (2016) 22:519–29. doi: 10.2119/molmed.2016.00023

68. Drent E, Bisso A, Baardman S, Verweij D, Salcedo E, Coomans C, et al. Targeting solid tumors with GDT002, a first-in-class võTCR-based T cell therapy. In: Proceedings of the American Association for Cancer Research Annual Meeting 2022, Apr 8-13, Vol. 2022. Philadelphia (PA): AACR. Available at: https://gadeta. nl/gadeta/assets/Abstract4559_Gadeta.pdf.

69. Xu Y, Yang Z, Horan LH, Zhang P, Liu L, Zimdahl B, et al. A novel antibody-TCR (AbTCR) platform combines fab-based antigen recognition with gamma/delta-TCR signaling to facilitate T-cell cytotoxicity with low cytokine release. *Cell Discovery* (2018) 4:62. doi: 10.1038/s41421-018-0066-6