Inhibition of the PI3K/AKT/mTOR pathway in pancreatic cancer: is it a worthwhile endeavor?

Al Jarroudi Ouissam (D, Chibani Hind, Brahmi Sami Aziz and Afqir Said

Abstract: Pancreatic cancer (PC) is an aggressive disease that is challenging to treat and is associated with a high mortality rate. The most common type of PC is pancreatic ductal adenocarcinoma (PDAC), and the existing treatment options are insufficient for PDAC patients. Due to the complexity and heterogeneity of PDAC, personalized medicine is necessary for effectively treating this illness. To achieve this, it is essential to understand the mechanism of PDAC carcinogenesis. Targeted therapies are a promising strategy to improve patient outcomes. Aberrant activation of the phosphatidylinositol 3-kinase/protein kinase B/ mammalian target of rapamycin (PI3K/AKT/mTOR) signaling pathway plays a crucial role in PC pathogenesis, from initiation to progression. This review provides a comprehensive overview of the current state of knowledge regarding the PI3K pathway in PDAC, summarizes clinical data on PI3K pathway inhibition in PDAC, and explores potential effective combinations that are a promising direction requiring further investigation in PDAC.

Keywords: clinical trials, genetic alterations, pancreatic ductal adenocarcinoma, PI3K/AKT/ mTOR pathway, targeted therapy

Received: 19 April 2024; revised manuscript accepted: 3 September 2024.

Introduction

Pancreatic cancer (PC) is one of the leading causes of cancer-related deaths worldwide, ranking 14th in incidence (2.6% of all cancers) and 7th in mortality (4.7% of all cancer-related deaths).¹ Pancreatic ductal adenocarcinoma (PDAC) is the most common histological type of PC, representing 90% of all PCs.² The pathology of this aggressive cancer is multifactorial and involves both modifiable risk factors (such as smoking, alcohol, and dietary factors) and non-modifiable factors (including gender, age, and genetic factors).^{3,4}

PDAC represents a challenging neoplasm characterized by its limited therapeutic options, poor prognosis, and dismal survival.⁵ Although surgery is the only curative approach for PC patients, it is usually impossible to perform optimal surgery due to the insidious onset of the disease, which results in most patients being diagnosed at an advanced stage.⁶ Cytotoxic chemotherapy is the only systemic treatment currently recommended for PC patients. Olaparib, a PARP inhibitor, is the only targeted therapy that has been indicated for PC as a maintenance treatment for patients with metastatic disease who have a BRCA germline mutation.⁷ However, PDAC is considered an immunologically cold tumor, which explains the inefficacy of immune checkpoint inhibitors, such as PD1/PDL1 and CTLA-4 inhibitors, in this aggressive malignancy.^{8,9} Currently, PDAC management is of utmost importance for a better and more extensive understanding of PC biology and the subsequent identification of promising molecular targets.

According to recent reports, several genetic changes have been detected in PDAC. In particular, the activation of the proto-oncogene Kristen rat sarcoma viral oncogene homolog (KRAS) has been observed in approximately 90% of cases,

Ther Adv Med Oncol

2024, Vol. 16: 1–21 DOI: 10.1177/ 17588359241284911

© The Author(s), 2024. Article reuse guidelines: sagepub.com/journalspermissions

Correspondence to: Al Jarroudi Ouissam Department of Medical Oncology, Mohammed VI University Hospital, Oujda, Morocco Faculty of Medicine and Pharmacy, Mohammed Ist University, Oujda, Morocco Aljarroudi.ouissam@ gmail.com Chibani Hind

Brahmi Sami Aziz Afgir Said

Department of Medical Oncology, Mohammed VI University Hospital, Oujda, Morocco

Faculty of Medicine and Pharmacy, Mohammed Ist University, Oujda, Morocco

1

and the inactivation of tumor suppressor genes, such as CDKN2A, SMAD4, and TP53, has been implicated in the tumors' proliferation and progression.¹⁰ The activating KRAS mutation is the main oncogenic driver mutation included in the characteristic hallmarks, which occurs early in the PDAC pathogenesis process. This mutation leads to constitutive activation of downstream signaling pathways (RAS/RAF/MAPK) and phosphatidylinositol 3-kinase/protein kinase B/mammalian target of rapamycin (PI3K/AKT/mTOR).11 The PI3K signaling pathway is commonly dysregulated in PDAC cells and plays a crucial role in promoting tumor growth, survival, and metastasis.12 Therefore, targeting the PI3K/AKT/mTOR cascade may be a potential therapeutic strategy for PDAC.

The objective of this narrative review is to provide a comprehensive overview of the current understanding of the PI3K pathway in PDAC, summarize the clinical data on PI3K pathway inhibition in PDAC, and investigate the potential of targeting the PI3K/AKT/mTOR cascade as a therapeutic strategy for this highly aggressive neoplasm.

Overview of the PI3K/AKT/mTOR pathway in PDAC

Signaling pathways are intricate networks that maintain crucial biological functions in all cell types. In tumors, these pathways are often disrupted, leading to deregulated cellular signaling implicated in various molecular hallmarks of cancer.¹³ The PI3K/AKT/mTOR pathway is a key signaling pathway that is essential for cellular processes such as proliferation, survival, motility, and metabolism.¹⁴. Research has shown that the PI3K pathway, which is frequently activated in various types of cancer, including PC, plays a pivotal role in tumorigenesis and progression.¹⁵ Deregulation of the PI3K pathway is frequently observed in 50% of all cancer patients and in 60% of PDAC patients.¹⁶

PI3K enzymes are a group of lipid kinases that catalyze the phosphatidylinositols' phosphorylation of the 3'-hydroxyl group. These enzymes are grouped into three distinct complexes (classes 1, 2, and 3) based on their structure and function.¹⁷ Class I PI3Ks, which are composed of two subclasses (IA and IB), are the only group of PI3Ks implicated in human cancer. They are activated by various stimuli, such as receptor tyrosine kinases (RTKs), Ras, or G-protein-coupled

receptors¹⁸ (Figure 1). The composition of class IA PI3Ks consists of a regulatory p85 (with three isoforms: $p85\alpha$, $p85\beta$, and $p55\gamma$) and a catalytic p110 (with three isoforms: p110a, p110b, and p1108) subunits.¹⁹ Upon activation, PI3K enzymes stimulate the conversion of phosphatidvlinsotide-4,5-biphosphate (PIP2) to phosphatidylinsotide-3,4,5-triphosphate (PIP3). PIP3 functions as a second messenger, activating downstream effectors involved in the phosphoinositide-dependent kinase 1 (PDK1)-mediated phosphorylation of the serine-threonine kinase AKT at threonine site 308.20 AKT activation leads to the phosphorylation of downstream components, including transcription factors, antiapoptotic proteins, and cell-cycle-related proteins. AKT, also known as protein kinase B (PKB), plays a crucial role in cell survival and growth. AKT activity is increased in PC due to its hyperphosphorylation or gene amplification.²¹ Finally, mTOR is the third effector player in the PI3K signaling pathway. The role of mTOR is to regulate cell growth and apoptosis through two distinct complexes: mTORC1 (rapamycin-sensitive) and mTORC2 (rapamycin-insensitive).^{22,23} mTORC1 is one of the major downstream effectors of AKT. However, mTORC2 is involved in the AKT phosphorylation on the serine residue-473. AKT activation is due to its phosphorylation mediated by PDK1 and mTORC2 on the threonine and serine sites, respectively. Thus, increased AKT activity phosphorylates the tuberous sclerosis complex 2 (TSC2) and subsequently activates the formation of the TSC2-TSC1 complex, which is directly responsible for mTORC1 activation. This complex can also bind mTORC2, leading to the promotion of its activity.^{18,24}

Mutations in PI3K pathway genes and other associated regulated pathway genes are responsible for PDAC tumorigenesis. KRAS oncogenic driver mutations (75%-90% of PDAC) are the primary molecular aberration that activates the PI3K downstream signaling pathway in PDAC.^{25,26} Studies have demonstrated that the PI3K pathway is activated in human PDAC and KRAS-driven mouse models of PC.27 KRASactivating mutations lead to overexpression of PI3KCA, which triggers acinar-to-ductal metaplasia (ADM) reprogramming through PDK1, ultimately resulting in PDAC formation.²⁸ The KRAS/PI3K axis plays a key role in mediating ADM, PDAC formation, and maintenance. The enhanced ducts formed from acinar cells further develop pancreatic intraepithelial neoplasia





(PanIN) lesions in in vivo models.²⁹ Therefore, understanding the mechanisms and cellular functions of the PI3K pathway is crucial for the development of effective cancer therapies. Research has demonstrated that blocking the p110a PI3K isoform can prevent preneoplastic transformation induced by KRAS in animal models.³⁰ PIK3CA gene mutations, occurring in 3%-5% of PC patients, have been associated with an increase in ADM and PanIN in a mouse model.³¹ Reactive oxygen species (ROS) are an important determinant of PC pathogenesis. ROS production is regulated by oncogenic KRAS-driven metabolic and signaling alterations. Once activated, ROS mediate the production of NADPH oxidases (NOX), which subsequently regulate ROS pro-survival activities through sustained apoptosis suppression.³² Additionally, AKT has a direct role in the activation of NOX proteins.33 Constitutive activation of the PI3K-effector AKT is a marker of the aggressiveness of PC. AKT2 amplification has been reported in 10%-32% of PDAC cases and contributes to the malignant phenotype.34

Recent analysis of the human PDAC genome and genetic research on mice has revealed that PTEN gene deletion is frequently observed in PC tissues, activating the NF- κ B and downstream cytokine pathways, which is linked to modifying the tumor microenvironment (TME) in PDAC.³⁵ When considered together, it becomes clear that the PI3K/AKT/PTEN signaling loop is a crucial signaling hub that is modified during the onset and course of PDAC.

PI3K has a crucial function in the regulation of immune cells and the coordination of interactions between cancer cells and immune cells. Additionally, it regulates the metabolic attributes of tumor cells and the TME's ability to control tumor growth and survival.³⁶ PDAC is a rare tumor with a higher proportion of stromal cells than tumor cells, and treatment options are limited due to the thick desmoplasia, which creates an acidic, hypoxic, and drug-impermeable TME.³⁷ The pancreatic TME comprises stromal cells, endothelial cells, immune cells, cancerassociated fibroblasts, and the extracellular

matrix. The dynamic nature of the TME is influenced by numerous signaling molecules released during tumor and accessory cell interactions.38 The relationship between stromal cells and PC cells modifies intracellular signaling and metabolic pathways.³⁹ Cancer-associated fibroblasts secrete substances like collagen, proteoglycans, glycoproteins, and extracellular matrix constituents, enhancing cancer cell migration and proliferation while shielding them from chemotherapy drugs. The PI3K pathway is involved in the migration and proliferation of cancer-associated fibroblasts.⁴⁰ Moreover, PI3K signaling activity is crucial for PC progression, controlling macrophage motility, adhesion, metabolism, polarization, and survival.⁴¹ Neutrophils, attracted to the stromal compartment and tumor cells by PC cells, are linked to poor prognosis and tumor development in patients with PDAC. These cells influence the migration of tumor and immune cells, accelerating cancer development through DNA damage caused by ROS.42 The PI3K signaling pathway controls neutrophils' phagocytosis, adhesion, proliferation, survival, and chemotaxis.43 The pancreatic TME also contains mast cells, which are associated with a worse PC patient survival rate.44 The PI3K pathway is a crucial regulator of mast cell development, differentiation, chemotaxis, degranulation, and cytokine production,45 as well as natural killer (NK) cell function.³⁴ NK cells are effector lymphocytic cells that are part of the innate immune system and are involved in the recognition and killing of tumor cells.46 In PC, the activity of NK cells is reduced, and the decreased expression of activating killer receptors correlates with tumor progression in PC patients. By inhibiting the PI3K signaling pathway in NK cells, their function in immune surveillance can be impaired, preventing their degranulation activity and reducing NK cell-mediated lysis of tumors.47,48 Adaptive immune responses are mostly suppressed in PC due to immunoediting, development of immune escape and immune resistance mechanisms.⁴⁹ The PI3K pathway plays a crucial role in T-cell activation, survival, and proliferation,⁵⁰ as well as the expression of the enzyme Bruton tyrosine kinase (BTK), which is essential for suppressing the growth of PC cells.⁵¹ Finally, the PI3K cascade acts as a key regulator of angiogenesis, controlling the proliferation, survival, and maturation of endothelial cells.52 Taken together, these studies suggest that the PI3K signaling pathway is critical for regulating pancreatic tumor biology.

Impact of targeting PI3K pathway in preclinical models of PDAC

Activation of the PI3K signaling pathway is mainly involved in the pathogenesis of PC, suggesting its potential as a promising therapeutic target for this disease. Several preclinical studies have evaluated the effects of inhibiting the PI3K cascade in PC.53 In human PC cell lines, two distinct PI3K inhibitors (wortmannin and LY294002) were found to induce apoptosis in a dose-dependent manner.⁵⁴ In a separate study, LY294002, a specific inhibitor of the PI3K/AKT pathway, simultaneously enhanced the efficacy of nonsteroidal anti-inflammatory drugs in human PC cells by inhibiting cell cycle progression.⁵⁵ Although using PI3K pathway inhibitors as monotherapy appears to be a promising but incomplete therapeutic approach for PC, subsequent studies have investigated the potential of combining these inhibitors with other therapies to optimize their efficacy.²⁸ The combinatorial approach has been explored in several human and mouse models of PC. For instance, MK-2206, a pan-AKT inhibitor, was combined with a cyclindependent kinase (CDK) inhibitor (MK-7965, dinaciclib) in preclinical PC models using orthotopic and subcutaneous patient-derived human PC xenografts, emphasizing the antitumor effect of this combination.⁵⁶ Double inhibition of the PI3K signaling pathway at two different sites seems to be a promising therapeutic option for PC. The combination of MK-2206 (an AKT inhibitor) with buparlisib (a PI3K inhibitor) was investigated in 10 PDAC cell lines, showing promising antiproliferative effects.⁵⁷ Another interesting combination of dual inhibition of PI3K and mTOR (NVP-BEZ235) was assessed in a PC orthotopic mouse xenograft model, which showed a significant decrease in tumor growth.⁵⁸ Recently, a natural compound that inhibits both PI3K and mTOR (urolithin A) was shown to have antiproliferative and proapoptotic effects in tumor xenografts and genetically engineered mouse models in vivo.59

Recently, it has been observed that oncogenic KRAS activates multiple signaling networks, particularly the ERK/MAPK and PI3K/AKT/mTOR pathways, in KRAS-dependent PC. The crosstalk between these two primaries signaling pathways is clinically relevant and forms the basis for the development of combination therapies for PDAC. Inhibition of ERK represents a potential therapeutic strategy. PI3K, on the other hand, regulates the sensitivity of ERK inhibitors. Therefore,

targeting both the ERK and PI3K signaling pathways simultaneously may provide a beneficial therapeutic approach for PC.60 Combining the inhibition of both the MAPK and PI3K pathways has demonstrated synergistic antitumor activity and a reduction in resistance in human PC cell lines.⁶¹ Dual inhibition of PI3K (ZSTK474) and RAS/MEK (RO5126766) inhibitors decreased the viability of human PC cell lines.⁶² Additionally, the combination of a MEK inhibitor (AZD6244) and a PI3K inhibitor (BKM120 or GDC-0941) increased tumor cell apoptosis in 46 human PDAC cell lines.63 Furthermore, combining PI3K (MK-2206) and MAPK (trametinib) inhibitors with chemotherapy (nab paclitaxel and gemcitabine) in preclinical models of PDAC (PDAC cell-derived subcutaneous and PDAC patientderived subcutaneous xenografts studies) demonstrated additive efficacy in reducing cell proliferation and inducing apoptosis.⁶⁴ Preclinical data on mutant KRAS-driven genetically engineered PC mouse models have shown an incremental benefit of the antiproliferative activity using the combination of targeted therapy (MEK and/or PI3K inhibitors) and chemotherapy (gemcitabine) compared to single-agent chemotherapy.65 Interestingly, in vitro screens have confirmed the resistance of PDAC preclinical models to monotherapies, emphasizing the need for combination drug therapies targeting multiple pathways mainly involved in PC. Epidermal growth factor receptor (EGFR) is frequently overexpressed in PC and is associated with a poor prognosis.^{66,67} Therefore, blocking EGFR signaling led to an inhibition of tumor growth and metastasis in human PC xenografts.⁶⁸ Regrettably, erlotinib's applicability for advanced PC is limited due to its restricted clinical response. Therefore, it is essential to investigate new predictive biomarkers that can identify patients with PC who would specifically benefit from erlotinib therapy and to explore potential new combination therapies.69

According to recent reports, the sensitivity of cell lines to the growth inhibition caused by EGFR inhibitors is influenced by the downregulation of activity in the PI3K-AKT pathway.⁷⁰ There are various mechanisms that can lead to EGFRindependent activity of the AKT pathway, such as the presence of other growth factor receptors such as insulin-like growth factor 1 receptor (IGF-1R) and fibroblast growth factor receptor (FGFR), specific mutations like constitutively activating PI3K mutations, or a lack of PTEN activity.⁷¹ It is important to note that multiple inputs can regulate this pathway, so inhibiting EGFR alone may not be sufficient for effectively inhibiting all tumor cells, emphasizing the need for multifaceted intervention. In fact, the combination of sirolimus and erlotinib has shown synergistic inhibitory activity on tumor growth in both cell culture and in vivo xenograft models.72 In addition, the PI3K pathway, as previously demonstrated, holds significant importance in the development of tumors and metastasis in PDAC. Furthermore, it also enables tumors to evade immune surveillance and suppress immune responses.73 These findings offer a compelling rationale for examining the PI3K/AKT/mTOR signaling cascade in clinical settings for patients with PDAC. The PI3K/AKT/mTOR signaling pathway is of great importance in the development of PC. Several inhibitors targeting this pathway have been investigated in early-phase clinical trials for patients with advanced PC, displaying promising outcomes with certain agents.74 However, due to the limited efficacy of single agents in targeting this pathway, the development of multidrug and multitarget combination approaches appears to be a more appealing and effective strategy for treating this aggressive disease.75 Therefore, further investigation is needed for the following combination therapies, which show promise for clinical application: MAPK, Sonic Hedgehog pathways, or pan-histone deacetylase inhibitors in combination with PI3K/ AKT/mTOR inhibitors.76,77

Contemporary clinical trials in PDAC targeting PI3K signaling axis

Based on preclinical data, the PI3K pathway appears to be a relevant target for PDAC management. Inhibition of this pathway can be accomplished at various points, and several drug classes that disrupt the PI3K/AKT/mTOR cascade have been evaluated in clinical research to improve PDAC treatment outcomes.⁷⁸ Notably, recent developments have led to the creation of targeted treatments that selectively inhibit PI3K, AKT, and mTOR as single agents or in combination with other therapies to enhance response rates and overcome resistance mechanisms in PDAC (Table 1).

PI3K inhibitors

Utilizing small molecules that specifically inhibit PI3K and its subsequent downstream signaling pathway offers hope for improving the management and prognosis of PDAC. Several tyrosine kinase inhibitors of PI3K have been explored in clinical trials for PDAC patients.⁷⁹

Table 1. Clinical trials assessing PI3K/AKT/mTOR pathway inhibitors in PDAC studies.

Drug	Target	Population	N	Treatment regimen	Phase	Status	References
Alpelisib (BYL719)	Oral class Ι α-specific PI3K inhibitor	<i>PIK3CA</i> -altered advanced solid tumors	134	Alpelisib monotherapy	I	Completed	80
Alpelisib (BYL719)	Oral class I α-specific PI3K inhibitor	Locally advanced and metastatic PC	15	BYL719 in combination with gemcitabine and nab- paclitaxel	I	Completed (the study was closed prematurely due to slow accrual)	82
Buparlisib (BKM120)	Oral pan-PI3K inhibitor	Advanced solid tumors	113	Buparlisib and trametinib (MEK inhibitor) combination	lb	Completed	84
Buparlisib (BKM120)	Oral pan-PI3K inhibitor	Advanced solid tumors with RAS/RAF alterations	89	Buparlisib in combination with Binimetinib (MEK1/2 inhibitor)	lb	Completed	135
Buparlisib (BKM120)	Oral pan-PI3K inhibitor	Advanced refractory solid tumors	17	BKM120 plus chemotherapy (mF0LF0X6)	I	Completed	85
Sonolisib (PX- 866)	Oral PI3K inhibitor	Advanced solid tumors	48	PX-866 as single agent	I	Completed	86
Copanlisib (BAY 80-6946)	Intravenous pan-class I PI3K inhibitor	Advanced solid tumors and non-Hodgkin's lymphomas	57	Copanlisib monotherapy	I	Completed	87
Pictilisib (GDC-0941)	Oral pan-class I PI3K inhibitor	Advanced solid tumors	60	Pictilisib (GDC-0941) as single agent	I	Completed	88
Pictilisib (GDC-0941)	Pan-PI3K inhibitor	Advanced solid tumors	178	Cobimetinib (MEK) and pictilisib combination therapy	I	Terminated	89
Perifosine (KRX041)	AKT inhibitor	Unreserctable locally advanced or metastatic PC	10	Perifosine monotherapy	II	Terminated for unacceptable adverse events	91
Perifosine (KRX041)	AKT inhibitor	Advanced PC	19	Perifosine as second line therapy	II	Completed	92
Afusertib (GSK2110183)	AKT inhibitor	Solid tumors and multiple myeloma	335	MEK inhibitor GSK1120212 in combination with the AKT inhibitor GSK2110183	1/11	Completed	94
Uprosertib (GSK2141795)	Oral AKT inhibitor	Advanced solid tumors	126	Uprosertib in combination with the oral MEK1/MEK2 inhibitor trametinib	I	Completed	95

(Continued)

Table 1. (Continued)

Drug	Target	Population	N	Treatment regimen	Phase	Status	References
Oleandrin (PBI-05204)	Oleander- derived inhibitor of AKT, FGF2, NF- κB, and p70s6k	Advanced solid tumors	46	PBI-05204 as monotherapy	I	Completed	100
Oleandrin (PBI-05204)	Oleander- derived inhibitor of AKT, FGF2, NF- κB, and p70s6k	Refractory metastatic PC	42	PBI-05204 as single agent	II	Completed	101
MK 2206	AKT inhibitor	Advanced PC	39	MK 2206 in combination with Dinaciclib (cyclin- dependent kinase inhibitor)	I	Completed	97
MK 2206	AKT inhibitor	Advanced treatment- refractory solid cancers	62	MK 2206 in combination with MEK 1/2 inhibitor (Selumetinib, AZD6244)	I	Completed	96
MK 2206	AKT inhibitor	Metastatic PC refractory to Gemcitabine- based therapy	137	MK 2206— Selumetinib combination compared to modified Folfox	II	Completed	98
LY2780301	Dual p70 S6 kinase and AKT inhibitor	Advanced tumors	80	Single-agent LY2780301	I	Completed	93
TAS0612	RSK, AKT, and S6K inhibitor	Advanced or metastatic solid tumors	100	TAS0612 as single agent	I	Recruiting (estimated study completion date: July 2027)	104
Sirolimus (AY- 22989)	Oral mTOR inhibitor	Advanced cancers	57	Sirolimus in combination with Sorafenib or Sunitinib	Ι	Completed	106
Sirolimus (AY- 22989)	Oral mTOR inhibitor	Advanced, gemcitabine- resistant PC	31	Sirolimus as monotherapy	I	Completed	105
Sirolimus (AY- 22989)	Oral mTOR inhibitor	Metastatic PC	22	Metformin with or without sirolimus as maintenance therapy after induction chemotherapy	lb	Completed	108

(Continued)

THERAPEUTIC ADVANCES in

Medical Oncology

Table 1. (Continued)

Drug	Target	Population	N	Treatment regimen	Phase	Status	References
Nab-sirolimus	Intravenous mTOR inhibitor	Locally advanced or metastatic solid tumors and moderate hepatic impairment or normal hepatic function	28	Nab sirolimus as single agent	I	Recruiting (estimated study completion date: April 2025)	110
Nab-sirolimus	Intravenous mTOR inhibitor	Malignant solid tumors with pathogenic alterations in TSC1/TSC2 genes	120	Nab sirolimus as single-agent	II	Recruiting (estimated study completion date: December 2025)	111
Nab-sirolimus	Intravenous mTOR inhibitor	Advanced solid tumors and non-small cell lung cancer with a KRAS G12C mutation	79	Nab sirolimus in Combination with Adagrasib	1/11	Recruiting (estimated study completion date: June 2026)	112
Everolimus (RAD001)	Oral mTOR inhibitor	Gemcitabine- refractory metastatic PC	33	RAD001	II	Completed	113
Everolimus (RAD001)	Oral mTOR inhibitor	Advanced PC	31	Capecitabine and Everolimus combination treatment as first- and second-line therapy in	II	Completed	114
Everolimus (RAD001)	Oral mTOR inhibitor	Advanced PV	43	Everolimus in combination with Cetuximab and Capecitabine	1/11	Completed	115
Everolimus (RAD001)	Oral mTOR inhibitor	Recurrent or refractory advanced solid tumors	100	Combination of three inhibitors Trametinib, Everolimus, and Lenvatinib	II	Recruiting (estimated study completion date: January 2027)	116
Everolimus (RAD001)	Oral mTOR inhibitor	Solid tumors or multiple myeloma	104	VS-6766 (R05126766) (a dual RAF/MEK inhibitor) alone and in combination with Everolimus (RAF/ MEK)	I	Recruiting (estimated study completion date: May 2024)	117

(Continued)

Table 1. (Continued)

Drug	Target	Population	N	Treatment regimen	Phase	Status	References
Temsirolimus (CCI-779)	Intravenous mTOR inhibitor	Advanced PC	5	Temsirolimus as single agent	II	Terminated (the study was closed to accrual due to significant adverse effects resulting from study treatment)	118
Temsirolimus (CCI-779)	Intravenous mTOR inhibitor	Advanced or metastatic PC	30	Temsirolimus in combination xith Gemcitabine	1/11	Completed	119
Ridaforolimus (MK-8669)	MTOR inhibitor	Refractory advanced tumors	17	Ridaforolimus in combination with Bevacizumab	I	Completed	120
Vistusertib (AZD2014)	MTORC1/2 inhibitor	Advanced solid tumors	56	AZD2014 in monotherapy	Ι	Completed	121
Voxtalisib (SAR245409)	Dual inhibitor of pan-PI3K and mTORC1/2	Advanced solid cancers	83	Voxtalisib as monotherapy	I	Completed	124
Voxtalisib (SAR245409)	Dual inhibitor of pan-PI3K and mTORC1/2	Advanced solid cancers	146	Voxtalisib in combination with MEK inhibitor	lb	Completed	125
Gedatolisib (PF05212384)	Dual inhibitor of PI3K and mTOR	Advanced solid tumors	81	Gedatolisib in combination with Irinotecan or MEK inhibitor	I	Completed	128
Gedatolisib (PF05212384)	Dual inhibitor of PI3K and mTOR	Advanced squamous cell lung, pancreatic, head and neck, and other solid tumors	96	Gedatolisib in combination with a cycline dependent kinase 4/6 (Palbociclib, PD- 0332991)	I	Recruiting (estimated study completion date: January 2026)	129
Omipalisib (GSK458)	Inhibitor of pan-PI3K and mTOR	Advanced solid cancers	170	Omipalisib as single agent	I	Completed	127

KRAS, Kristen rat sarcoma viral oncogene homolog; PC, pancreatic cancer; PDAC, pancreatic ductal adenocarcinoma; PI3K/AKT/mTOR, phosphatidylinositol 3-kinase/protein kinase B/mammalian target of rapamycin; TSC2, tuberous sclerosis complex 2.

Alpelisib (BYL719) is one of the first PI3K inhibitors to be tested in various malignancies. This PI3K α -selective inhibitor was explored in a first-in-human phase I trial (NCT01219699) in 134 patients with PIK3CA-altered advanced

solid tumors, demonstrating a safe profile and encouraging preliminary antitumor activity.⁸⁰ On April 5, 2022, alpelisib received accelerated Food and Drug Administration (FDA) approval as a treatment for adult and pediatric patients with PIK3CA-related overgrowth spectrum (PROS).⁸¹ Results from an in vitro study of alpelisib in PC human and murine samples led to its assessment in a phase I clinical trial (NCT02155088) in combination with chemotherapy (gemcitabine and nab paclitaxel) in 15 patients with advanced and metastatic PC. The median progression-free survival (mPFS) was 5.36 months (1.6–10 months), and the median overall survival (mOS) was 8.74 months (3.8–21.2 months). The most common adverse events (AEs) were hyperglycemia, anemia, and a decreased neutrophil count.⁸²

Due to the high prevalence of resistance to targeted therapies administered as single agents, clinical investigations have been conducted to explore combinatorial approaches. As previously mentioned, the MAPK and PI3K signaling pathways are interconnected and play a crucial role in tumor initiation and progression, as well as in drug resistance mechanisms. Inhibition of one pathway can lead to compensatory activation of the other, making dual inhibition a promising strategy to overcome resistance to current therapies.83 In an open-label phase Ib trial (NCT01155453), 113 patients with select solid tumors were administered the pan-PI3K inhibitor buparlisib (also known as BKM120) in combination with an oral MEK1/2 inhibitor (trametinib). The most frequently reported AEs were gastrointestinal and dermatologic. Although the combination showed promising activity in KRAS-mutant ovarian cancers, minimal activity was observed in PDAC patients.84 In a phase Ib, multicenter, open-label, and basket trial (NCT01363232), 89 patients with advanced solid tumors harboring RAS/RAF alterations were evaluated for the effectiveness of buparlisib when combined with a highly potent MEK1/2 inhibitor (binimetinib). Out of these, 6 patients (7%) had PC, and at the data cutoff, 84 patients (94.4%) had discontinued study treatment due to disease progression (48.3%) and AEs (36.0%). Among the 69 patients (77.5%) who were evaluable for tumor response based on RECIST version 1.1, the combined therapy was not as effective as anticipated, with the exception of KRAS/NRAS/BRAF-mutant ovarian cancer, where significant responses were observed (partial response in 5/18 patients). Several factors may have contributed to the reduced effectiveness, including the frequent dose reductions resulting from the combination's toxicities and the consequent suboptimal pathway modulation. Additionally, the genomic

context, whether truncal, shared, or private, may explain the significant differences in responses between the cancers, despite them having common driver alterations. Finally, epigenetic factors can dictate therapeutic vulnerabilities, requiring different doses of targeted therapy to achieve sufficient pathway inhibition.85 Based on these results, further exploration of this combination may be warranted to define a better-tolerated dose and/or schedule, such as alternative scheduling with noncontinuous/pulsatile dosing of either agent, which could be explored further in RAS/ BRAF-mutant tumors. The findings may inform the design of future combination therapy trials in patients with tumors harboring mutations in the PI3K and MAPK pathways. In addition, buparlisib was tested in combination with chemotherapy (mFOLFOX6) in a single-center phase I clinical trial (NCT01571024) involving 17 patients with refractory solid tumors. This combination resulted in an unconfirmed partial response in one patient and stable disease in three patients. Common AEs included neutropenia, fatigue, leukopenia, hyperglycemia, and thrombocytopenia. The increasing toxicity observed in this combination does not encourage further exploration of this treatment in gastrointestinal cancers.85 Sonolisib or PX-866 is an oral, irreversible PI3K inhibitor that was assessed in a multicenter, openlabel, and dose-escalation phase I study. This trial was the first to test PX-866 in humans and showed promising activity in 84 patients with advanced solid malignancies.86 Copanlisib (BAY 80-6946), an intravenous pan-class I PI3K inhibitor, was evaluated in a phase I dose escalation study (NCT00962611) to assess its safety and pharmacokinetics in patients with advanced solid tumors and hematological cancers. This agent exhibited a safe profile and a preliminary antitumor effect.87 Based on the promising results of preclinical models with pictilisib (GDC-0941), a phase I dose escalation trial was conducted in 16 patients with solid tumors, providing impetus for further investigation of this pan-class I PI3K inhibitor.88 However, a phase Ib trial found that combining pictilisib with a MEK inhibitor (cobimetinib) did not result in higher efficacy in patients with advanced solid cancers.⁸⁹ Recently, the impact of taselisib, a selective inhibitor of PI3K, on PDAC cell proliferation was investigated. The study demonstrated a correlation between increasing taselisib concentrations and a reduction in PDAC cell viability. In addition, transcriptomic data from PDAC samples were utilized in this study to develop a risk score based

on the expression profiles of dysregulated PI3K/ AKT/mTOR pathway-related genes and machine learning, which showed a significant increase in overall survival time among patients in the lowrisk group. Hence, the PI3K/AKT/mTOR pathway could be harnessed to better predict individual risk scores.⁹⁰

AKT inhibitors

AKT signaling plays a critical role in activating antiapoptotic pathways that enable cancer cells to survive. According to preclinical evidence, targeting AKT presents a promising strategy for the treatment of PC. Unfortunately, several AKT inhibitors failed to show efficacy in clinical trials. Perifosine, also known as KRX041, is an example of an AKT inhibitor that demonstrated promising results in preclinical studies but did not prove beneficial in two phase II clinical trials for patients with advanced PC.^{91,92} LY2780301, another AKT inhibitor, was assessed as a single agent in a first-in-human phase I study in patients with advanced malignancies, showing minimal antitumor activity.⁹³

Based on biological concepts, the PI3K/AKT/ mTOR and RAF/MEK/ERK pathways are intriguing aspects of human cancer therapy. Accumulated studies on these two pathways will provide new hope for cancer patients. Inhibiting the PI3K/AKT/mTOR and MAPK/MEK/ERK pathways could have a synergistic effect. Afuresertib (GSK2110183) is an AKT inhibitor that was evaluated in combination with a MEK inhibitor (trametinib) in a phase I/II clinical trial (NCT01476137) in 20 patients with solid tumors and multiple myeloma. The purpose of this study was to determine safety and response. This trial concluded that the combination was poorly tolerated using a continuous daily dose schedule.94 Uprosertib (GSK2141795) is another oral AKT inhibitor that failed to show clinical benefit with poor tolerance in advanced solid tumors, including PC, in combination with an oral MEK 1/2 inhibitor (trametinib) in a phase I trial.95 MK-2206 was assessed in combination with an oral, selective MEK 1/2 inhibitor (selumetinib) in a phase I trial (NCT01021748), which involved 62 patients with advanced treatment-refractory solid malignancies and demonstrated a promising therapeutic combination option in KRAS-driven cancers.96 Despite promising results in preclinical trials, including combination approaches with a CDK inhibitor,

MK-2206 failed to show clinical benefit in a phase I study with dinaciclib (a CDK inhibitor) for patients with advanced PC.97 In addition, this combination failed to meet its primary endpoint of improving overall survival compared to modified FOLFOX in patients with metastatic PC that had failed prior gemcitabine-based therapy in a phase II clinical trial (SWOG S1115, NCT01658943).98 Furthermore, oleandrin, a novel plant compound derived from Nerium oleander, has been widely used as a cardiac glycoside and exhibits antitumor activity through the inhibition of phosphorylated AKT expression and downregulation of mTOR in orthotopic models of human PC.99 In a phase I clinical trial, oleandrin demonstrated a safe profile in heavily pretreated patients with advanced solid tumors.¹⁰⁰ However, a phase II, single-arm trial in 42 patients with metastatic PC failed to achieve its primary endpoint of achieving 50% patient survival at 4.5 months.¹⁰¹ Currently, the evaluation of novel and potent kinase inhibitors with multitargets, such as TAS0612, which inhibits RSK, AKT, and S6K, is being conducted.¹⁰² RSK is a serine threonine kinase activated by upstream kinases of the PI3K and MAPK pathways and is implicated in various resistance mechanisms associated with single inhibition of the PI3K cascade.103 TAS0612 is currently being explored in an ongoing phase I clinical trial (NCT04586270) in patients with advanced or metastatic malignancies.¹⁰⁴ The findings underscore the necessity of delving deeper into the intricate molecular mechanisms and complex signaling pathways in order to provide new therapeutic perspectives for clinicians treating PC patients. Despite the development of numerous AKT inhibitors, only a few have shown promise in preclinical studies for PC, and even fewer have been tested in clinical research. Due to their toxicity and lack of efficacy, these inhibitors are not yet approved for use in patients. Therefore, it is imperative to determine the actual impact of these drugs in a clinical setting. Thus, additional research focusing on reducing toxicity and enhancing the effectiveness of AKTinhibitor combinations is essential.

mTOR inhibitors

Targeting mTOR has been widely investigated in the field of PC. mTOR is a serine/threonine protein kinase that is involved in tumor growth through the formation of two protein complexes, mTORC1 and mTORC2. Several mTOR inhibitors have been evaluated in clinical settings for the treatment of PC.

Sirolimus, also known as rapamycin or AY-22989, has demonstrated promising results in preclinical studies through the inhibition of mTOR. However, in a clinical setting, this agent was investigated as a single therapy in patients with advanced PC who were refractory to gemcitabine, with marginal outcomes indicating that sirolimus monotherapy is ineffective in the treatment of PC.¹⁰⁵ As a result, researchers sought to combine it with other drugs. In a phase I trial, sirolimus was tested in combination with potent oral antiangiogenic kinase inhibitors (sorafenib or sunitinib) in several advanced malignancies, including PC cases, demonstrating the feasibility and safety of such a combination.¹⁰⁶ Another combinatorial strategy that has been investigated in the context of PC involves the combination of sirolimus and metformin. This approach was supported by preclinical evidence in PC cell lines, as it led to a synergistic effect due to enhanced inhibition of the mTOR pathway.¹⁰⁷ Metformin, which is an antidiabetic medication, was found to inhibit mTORC1. In a randomized, open-label phase Ib trial (NCT02048384), metformin was administered either alone or in combination with rapamycin as maintenance therapy to advanced PC patients who had previously received chemotherapy. A total of 22 patients were treated according to the protocol and were randomly assigned in a 1:1 ratio to either metformin alone (Arm A) or metformin plus rapamycin (Arm B). The results demonstrated that both treatment options had a favorable safety profile and promising outcomes, with longer-than-expected progression free survival (PFS) and overall survival (OS) in this population with poor prognosis. The median PFS for the entire cohort, Arm A, and Arm B was 3.5 (95% confidence interval (CI): 2.9-9.2), 4.0 (95% CI: 2.9 to not reached (NR)), and 3.0 months (95% CI: 2.8 to NR), respectively, and the median OS was 13.2 (95% CI: 7.8 to NR), 14.8 (95% CI: 6.2 to NR), and 9.7 months (95% CI: 9.0 to NR), for the entire cohort, Arm A, and Arm B, respectively. The study findings support further investigation of these agents in the maintenance setting. However, this trial was not designed to detect differences in clinical activity between the treatment arms. In patients with metastatic PDAC who achieve a response to chemotherapy, the administration of metformin with or without rapamycin was found to be well-tolerated and associated with better-than-expected OS. Further studies are required to prospectively

evaluate the role of these agents compared to maintenance chemotherapy or observation-only approaches.¹⁰⁸ Nab-sirolimus, a novel and potent mTOR inhibitor, has exhibited a higher tumor inhibitory effect and more effective mTOR target suppression in preclinical models as compared to oral mTOR inhibitors.¹⁰⁹ Currently, nab-sirolimus is being investigated in three ongoing clinical trials, including as a single agent in locally advanced or metastatic cancers (NCT05661461),¹¹⁰ in selected patients with tumors harboring pathogenic alterations in TSC1/2 genes (PRECISION-1, NCT05103358),¹¹¹ and in combination with adagrasib in tumors with KRAS G12C mutations (KRYSTAL-19, NCT05840510).¹¹² Everolimus (RAD001) is another oral mTOR inhibitor that has been studied in a phase II multi-institutional and single-arm trial. This trial involved 33 patients with advanced PC who had previously been unresponsive to gemcitabine. The results of the trial suggest that everolimus may have limited effectiveness when used alone and may be more effective when used in combination with other treatments.¹¹³ In a separate phase II clinical trial, the combination of everolimus and chemotherapy (capecitabine) was analyzed in 31 patients with advanced PC. The results of this trial indicated that the combination was feasible and showed moderate effectiveness, with a disease control rate of 38%, an mPFS of 3.6 months, and an mOS of 8.9 months.¹¹⁴ An open-label, multicenter, nonrandomized phase I/II trial (NCT01077986) was conducted to evaluate a triple drug regimen comprising everolimus, cetuximab, and capecitabine in 43 patients with advanced PC. However, the study demonstrated the impracticality of this combination due to severe AEs.¹¹⁵ Everolimus is also under exploration in an ongoing phase II clinical trial (NCT04803318) in combination with trametinib and lenvatinib for the treatment of recurrent or refractory advanced solid malignancies.¹¹⁶ Another phase I trial (NCT02407509) is ongoing to explore the combination of everolimus with a dual inhibitor of RAF/MEK (VS-6766) in patients with advanced solid malignancies.¹¹⁷ Moreover, in addition to the two mTOR inhibitors previously mentioned, two other inhibitors were investigated in the context of PC. The first of these is temsirolimus, an intravenous mTOR inhibitor that was tested in a phase II clinical trial (NCT0075647) involving advanced PC patients, but the results were not positive.¹¹⁸ Consequently, a combination of temsirolimus and gemcitabine was studied in a phase I/II trial (ACTRN12611000643976) for PC patients.

Although the combination was found to be safe, it did not demonstrate significant efficacy.¹¹⁹ The second agent is ridaforolimus (MK-8669), which was combined with bevacizumab in a phase I study (NCT00781846) involving 17 patients with refractory advanced malignancies, 2 of whom had PC. This combination showed promising outcomes in this trial.¹²⁰ Recently, a novel mTORC1 and 2 inhibitor known as vistusertib (AZD2014) was explored in a first-in-human phase I trial, suggesting activity against heavily pretreated patients with advanced solid cancers. This suggests that further investigation into mTORC inhibitors may be merited.¹²¹

Dual inhibitors of PI3K/mT0R pathway

The management of PDAC may benefit from a dual inhibition of PI3K and mTOR, as suggested by preclinical data demonstrating potential efficacy in PC models.^{122,123}

Voxtalisib (SAR245409) is a dual inhibitor of pan-PI3K and mTORC1 and 2 that has been investigated in a first-in-human phase I trial involving 83 patients with advanced solid tumors. In this trial, voxtalisib was found to be safe, with disease stability serving as the best response in 48% of the patients analyzed.¹²⁴ Additionally, voxtalisib was studied in combination with an oral MEK inhibitor (pimasertib) in a dose-escalation and expansion phase Ib study (NCT01390818), which included 146 patients with advanced solid cancers (7 of whom had PC). However, this combination was poorly tolerated and showed a lack of clinical antitumor activity.¹²⁵ Omipalisib (GSK458) is a potent oral inhibitor of pan-PI3K and mTOR.126 One hundred seventy patients with refractory advanced cancers (including two patients with PC) were enrolled in a dose-escalation, singleagent, and phase I study to receive omipalisib to assess its pharmacokinetics, pharmacodynamics, efficacy, and safety.¹²⁷ On the other side, gedatolisib (PF05212384) is another agent that inhibits the PI3K and mTOR pathways and was evaluated in advanced solid malignancies. In an open-label, multicenter, four-arm, dose-escalation phase I trial (NCT01347866), PF05212384 was administered in combination with chemotherapy (irinotecan) or an MEK inhibitor (PD-0325901). The results of this trial indicated the need for further investigation of gedatolisib in advanced solid tumors.¹²⁸ In a phase I clinical trial (NCT03065062) that is currently

underway, the investigational drug gedatolisib is being tested in combination with a CDK4/6 inhibitor (palbociclib) in patients with advanced squamous cell lung, pancreatic, head and neck, and other solid malignancies. The study's findings are anticipated to be released in January 2026.¹²⁹

Mechanisms of resistance to PI3K/AKT/ mTOR inhibitors

Although PI3K/AKT/mTOR pathway-targeted agents have shown promising preclinical activity in PC, acquired and intrinsic resistance have significantly hindered their clinical effectiveness. To develop alternative clinical strategies, it is crucial to understand the biochemical mechanisms that lead to resistance.74 PC cells can overcome downregulation of the PI3K pathway induced by targeted agents by activating alternate signaling pathways or maintaining or reestablishing the targeted pathway.¹³⁰ Resistance to targeted agents can be attributed to several mechanisms, including secondary target mutations, parallel signaling pathways, and amplification of downstream modifications within the same pathway.¹³¹

Upregulation of the PI3K/AKT/mTOR pathway due to PI3K inhibition leads to drug resistance, involving the feedback loop of RTKs, growth factors, and transcription factors like forkhead box (FOXO). FOXO suppresses RTKs or adaptors that activate the PI3K pathway, such as EGFR, IGF-1R, EGFR, FGFR, and insulin receptor (IR). By inhibiting PI3K/AKT signaling, FOXO phosphorylation is suppressed, leading to FOXOdependent repression of RTKs and derepression of downstream molecules. This results in the activation of multiple RTKs and partial maintenance of PIP3, which ultimately leads to cell proliferation and survival.¹³²

The PI3K pathway is crucial for nutrient uptake and cell survival. PC cells exhibit the "Warburg effect," which refers to increased glucose uptake and glycolytic conversion to lactate. This effect is promoted by the constitutive activation of downstream AKT signaling, which enhances glucose uptake through the GLUT family of transporters. Apart from glucose transporter guidance, it has been demonstrated that the PI3K-AKT pathway regulates multiple nodes of the glycolytic cascade, thereby ensuring the continual metabolic requirements of the cell.¹³³ As previously indicated, growth factors that activate cell surface receptors, such as EGFR, FGFR, IR, and IGFR, control proliferation, migration, metabolism, and cell survival. Upon growth factor stimulation, these receptors recruit and phosphorylate insulin receptor substrate-1 (IRS-1) adaptor molecules to activate the PI3K pathway. Furthermore, the interactions between these receptors and their signaling cascades can overlap to conform to synergistic responses, where, for instance, the induction of cell cycle progression also promotes an influx in energy synthesis by altering metabolic activity.¹³⁴ Taken together, these intriguing findings demonstrate the complexity of these intrinsic feedback loops to effectively downregulate PI3K signaling. Notably, combination therapies that target PI3K, AKT, or mTOR alongside various RTK molecules effectively combat this response in preclinical studies. However, there remain limitations in the clinical setting due to primarily negative adverse effects experienced by the patients, as well as a lack of selectivity when it comes to available drugs that target RTKs specifically.

Recently, emerging evidence has demonstrated the potential role in changes in cellular plasticity and PI3K inhibitor resistance. The downregulation of several RTKs involved in PI3K signaling may result in the generation of genetically independent transcriptional programs, which lead to "drug-tolerant" cell populations. In contrast to primary drug resistance, this state allows tumor cells to survive temporarily but not proliferate during treatment. However, these drug-tolerant populations can evade initial drug therapy but do not possess the genetic mechanisms required for tumor progression. They can serve as a reservoir of tumorigenic slow-cycling cells from which secondary genetic mechanisms of acquired resistance can develop. This protective measure has been associated with a phenotypic switch, commonly referred to as cell plasticity, whereby epithelial tumors progress to a more mesenchymal state. This epithelialmesenchymal transition has long been linked to chemoresistance. More importantly, this drugrefractory state is reversible upon drug withdrawal, highlighting the absence of genetic mutations driving this drug-tolerant state.130

Furthermore, mucins (MUC), which are highmolecular-weight glycoproteins, are notably overexpressed in PC. Of these, MUC1, a type I transmembrane glycoprotein, modifies the

aggressiveness of PDAC by impacting metabolism and signaling through the PI3K pathway. MUC1 influences the expression and activation of several RTKs, including PDGFR, EGFR, and c-Met, to enhance cellular activities such as proliferation, dissemination, and survival. Specifically, MUC1 affects EGFR's nuclear localization, which in turn influences how EGFR interacts with transcriptionally active promoter regions. Additionally, MUC1 controls the expression of genes that confer multidrug resistance in PC cells through AKTdependent and independent mechanisms. Considering the MUC-PI3K signaling regulatory axis as a potential treatment approach for PC may be beneficial, as it can also control MUC1-mediated resistance to chemotherapy and radiation.34

In general, the PI3K pathway is highly influenced by intrinsic adaptive responses that re-establish pathway activation following treatment, leading to inadequate pathway inhibition and tumor progression. This not only emphasizes the need for a deeper understanding of the complexities of PI3K signaling but also strongly supports the use of specific combinations to overcome these mechanisms of resistance.

Conclusion and prospect

Exploring the fundamental mechanisms responsible for the development of PDAC is a promising avenue for discovering targeted therapies. The PI3K/AKT/mTOR signaling pathway is a critical hub that plays a significant role in the initiation and progression of PC. It also has a crucial regulatory function in the PDAC TME, which is composed of various cellular components. Despite the development of several inhibitors that target the PI3K/AKT/mTOR pathway, the main challenge remains drug resistance, which results in suboptimal responses when these agents are used as single therapies. Results from early-phase clinical trials have demonstrated that targeting a single component of the PI3K/AKT/mTOR loop is insufficient for effectively treating this genetically diverse type of cancer. The necessity for additional research in order to determine the optimal combination of treatments that will lead to improved outcomes for patients with PDAC is evident. Furthermore, the preclinical and earlyphase clinical trial data supporting the use of downstream inhibition of the PI3K/AKT/mTOR pathway (mTORC1/2 inhibitors) in PDAC

treatment presents a promising option, providing a rationale for the development of dual mTORC1/2 inhibitors as combination therapies for PDAC patients. To improve personalized management for PDAC patients, an increase in the number of clinical trials examining novel multitarget drugs and rational therapeutic combinations is recommended.

It is interesting to note that the urgent need for effective treatment methods and extending the lifespan of PDAC patients requires immediate investigation. To this end, recent advancements in genomics, transcriptomics, and artificial intelligence tools should be utilized in PC field studies. The development of risk prediction models for PDAC is essential for identifying high-risk individuals and incorporating them into individual screening programs. Additionally, innovative approaches are required to detect PC at earlier stages and prevent its occurrence. Finally, the use of in vivo research models such as human organoid models holds promise for enhancing understanding of PC pathogenesis and evaluating the efficacy and mechanisms of resistance of potential drugs and combination-based therapies in PC cells.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication Not applicable.

Author contributions Al Jarroudi Ouissam: Conceptualization; Visualization; Writing – original draft.

Chibani Hind: Writing – review & editing.

Brahmi Sami Aziz: Validation; Writing – review & editing.

Afqir Said: Supervision.

Acknowledgments

We thank Dr El Bairi Khalid for providing inspiration and guidance in the conception and design of this work.

Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

Competing interests

The authors declare that there is no conflict of interest.

Availability of data and materials Not applicable.

ORCID iD

Al Jarroudi Ouissam D https://orcid.org/0000-0001-9946-921X

References

- Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2021; 71(3): 209–249.
- 2. Taherian M, Wang H and Wang H. Pancreatic ductal adenocarcinoma: molecular pathology and predictive biomarkers. *Cells* 2022; 11(19): 3068.
- 3. Klein AP. Pancreatic cancer epidemiology: understanding the role of lifestyle and inherited risk factors. *Nat Rev Gastroenterol Hepatol* 2021; 18(7): 493–502.
- 4. Cai J, Chen H, Lu M, et al. Advances in the epidemiology of pancreatic cancer: trends, risk factors, screening, and prognosis. *Cancer Lett* 2021; 520: 1–11.
- Zhao Z and Liu W. Pancreatic cancer: a review of risk factors, diagnosis, and treatment. *Technol Cancer Res Treat* 2020; 19: 1533033820962117.
- Huang L, Jansen L, Balavarca Y, et al. Resection of pancreatic cancer in Europe and USA: an international large-scale study highlighting large variations. *Gut* 2019; 68(1): 130–139.
- Chi J, Chung SY, Prasad S, et al. The role of olaparib in metastatic pancreatic cancer. *Cancer Med* J 2021; 4(3): 89–91.
- Mukherji R, Debnath D, Hartley ML, et al. The role of immunotherapy in pancreatic cancer. *Curr Oncol* 2022; 29(10): 6864–6892.
- 9. Laface C, Memeo R, Maselli FM, et al. Immunotherapy and pancreatic cancer: a lost challenge? *Life* 2023; 13(7): 1482.
- Baer R, Cintas C, Therville N, et al. Implication of PI3K/Akt pathway in pancreatic cancer: when PI3K isoforms matter? *Adv Biol Regul* 2015; 59: 19–35.
- Conway JR, Herrmann D, Evans TJ, et al. Combating pancreatic cancer with PI3K pathway

inhibitors in the era of personalised medicine. *Gut* 2019; 68(4): 742–758.

- 12. Lee JE, Woo MG, Jung KH, et al. Combination therapy of the active KRAS-targeting antibody in Ras37 and a PI3K inhibitor in pancreatic cancer. *Biomol Ther* 2022; 30(3): 274–283.
- Fares J, Fares MY, Khachfe HH, et al. Molecular principles of metastasis: a hallmark of cancer revisited. *Signal Transduct Target Ther* 2020; 5(1): 1–17.
- 14. Cantley LC. The phosphoinositide 3-kinase pathway. *Science* 2002; 296(5573): 1655–1657.
- 15. Elsayed M and Abdelrahim M. The latest advancement in pancreatic ductal adenocarcinoma therapy: a review article for the latest guidelines and novel therapies. *Biomedicines* 2021; 9(4): 389.
- Schild C, Wirth M, Reichert M, et al. PI3K signaling maintains c-myc expression to regulate transcription of E2F1 in pancreatic cancer cells. *Mol Carcinog* 2009; 48(12): 1149–1158.
- Yuan T and Cantley L. PI3K pathway alterations in cancer: variations on a theme. *Oncogene* 2008; 27(41): 5497–5510.
- Courtney KD, Corcoran RB and Engelman JA. The PI3K pathway as drug target in human cancer. *J Clin Oncol* 2010; 28(6): 1075–1083.
- Engelman JA, Luo J and Cantley LC. The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism. *Nat Rev Genet* 2006; 7(8): 606–619.
- Sarbassov DD, Guertin DA, Ali SM, et al. Phosphorylation and regulation of Akt/PKB by the rictor-mTOR complex. *Science* 2005; 307(5712): 1098–1101.
- Schlieman MG, Fahy BN, Ramsamooj R, et al. Incidence, mechanism and prognostic value of activated AKT in pancreas cancer. *Br J Cancer* 2003; 89(11): 2110–2115.
- 22. Dan HC, Cooper MJ, Cogswell PC, et al. Akt-dependent regulation of NF-{kappa}B is controlled by mTOR and Raptor in association with IKK. *Genes Dev* 2008; 22(11): 1490–1500.
- Dan HC, Antonia RJ and Baldwin AS. PI3K/Akt promotes feedforward mTORC2 activation through IKKα. Oncotarget 2016; 7(16): 21064–21075.
- 24. Huang J and Manning BD. A complex interplay between Akt, TSC2, and the two mTOR complexes. *Biochem Soc Trans* 2009; 37(Pt 1): 217–222.
- 25. Thibault B, Ramos-Delgado F, Pons-Tostivint E, et al. Pancreatic cancer intrinsic PI3Kα activity

accelerates metastasis and rewires macrophage component. *EMBO Mol Med* 2021; 13(7): e13502.

- Wang JY, Lian ST, Chen YF, et al. Unique K-ras mutational pattern in pancreatic adenocarcinoma from Taiwanese patients. *Cancer Lett* 2002; 180(2): 153–158.
- 27. Eser S, Reiff N, Messer M, et al. Selective requirement of PI3K/PDK1 signaling for Kras oncogene-driven pancreatic cell plasticity and cancer. *Cancer Cell* 2013; 23(3): 406–420.
- Mehra S, Deshpande N and Nagathihalli N. Targeting PI3K pathway in pancreatic ductal adenocarcinoma: rationale and progress. *Cancers* 2021; 13(17): 4434.
- Baer R, Cintas C, Dufresne M, et al. Pancreatic cell plasticity and cancer initiation induced by oncogenic Kras is completely dependent on wildtype PI 3-kinase p110α. *Genes Dev* 2014; 28(23): 2621.
- Kennedy AL, Morton JP, Manoharan I, et al. Activation of the PIK3CA/AKT pathway suppresses senescence induced by an activated RAS oncogene to promote tumorigenesis. *Mol Cell* 2011; 42(1): 36–49.
- Payne SN, Maher ME, Tran NH, et al. PIK3CA mutations can initiate pancreatic tumorigenesis and are targetable with PI3K inhibitors. *Oncogenesis* 2015; 4(10): e169.
- Storz P. KRas, ROS and the initiation of pancreatic cancer. *Small GTPases* 2016; 8(1): 38–42.
- Edderkaoui M, Nitsche C, Zheng L, et al. NADPH oxidase activation in pancreatic cancer cells is mediated through Akt-dependent up-regulation of p22phox. *β Biol Chem* 2011; 286(10): 7779–7787.
- 34. Murthy D, Attri KS and Singh PK. Phosphoinositide 3-kinase signaling pathway in pancreatic ductal adenocarcinoma progression, pathogenesis, and therapeutics. *Front Physiol* 2018; 9: 335.
- 35. Ying H, Elpek KG, Vinjamoori A, et al. PTEN is a major tumor suppressor in pancreatic ductal adenocarcinoma and regulates an NF-κB-cytokine network. *Cancer Discov* 2011; 1(2): 158–169.
- 36. Rucki AA, Foley K, Zhang P, et al. Heterogeneous stromal signaling within the tumor microenvironment controls the metastasis of pancreatic cancer. *Cancer Res* 2017; 77(1): 41–52.
- 37. Pandol S, Edderkaoui M, Gukovsky I, et al. Desmoplasia of pancreatic ductal

adenocarcinoma. *Clin Gastroenterol Hepatol* 2009; 7(11 Suppl): S44–S47.

- Feig C, Gopinathan A, Neesse A, et al. The pancreas cancer microenvironment. *Clin Cancer Res* 2012; 18(16): 4266–4276.
- Tape CJ, Ling S, Dimitriadi M, et al. Oncogenic KRAS regulates tumor cell signaling via stromal reciprocation. *Cell* 2016; 165(4): 910–920.
- 40. Bussard KM, Mutkus L, Stumpf K, et al. Tumor-associated stromal cells as key contributors to the tumor microenvironment. *Breast Cancer Res* 2016; 18(1): 84.
- 41. Covarrubias AJ, Aksoylar HI, Yu J, et al. AktmTORC1 signaling regulates Acly to integrate metabolic input to control of macrophage activation. *eLife* 2016; 5: e11612.
- Ino Y, Yamazaki-Itoh R, Shimada K, et al. Immune cell infiltration as an indicator of the immune microenvironment of pancreatic cancer. *Br J Cancer* 2013; 108(4): 914.
- Pinho V, Russo RC, Russo RC, et al. Tissue- and stimulus-dependent role of phosphatidylinositol 3-kinase isoforms for neutrophil recruitment induced by chemoattractants in vivo. *J Immunol* 2007; 179(11): 7891–7898.
- 44. Strouch MJ, Cheon EC, Salabat MR, et al. Crosstalk between mast cells and pancreatic cancer cells contributes to pancreatic tumor progression. *Clin Cancer Res* 2010; 16(8): 2257– 2265.
- Kim MS, Rådinger M and Gilfillan AM. The multiple roles of phosphoinositide 3-kinase in mast cell biology. *Trends Immunol* 2008; 29(10): 493–501.
- Cerwenka A and Lanier LL. Natural killer cell memory in infection, inflammation and cancer. *Nat Rev Immunol* 2016; 16(2): 112–123.
- Jiang K, Zhong B, Gilvary DL, et al. Pivotal role of phosphoinositide-3 kinase in regulation of cytotoxicity in natural killer cells. *Nat Immunol* 2000; 1(5): 419–425.
- Peng YP, Xi CH, Zhu Y, et al. Altered expression of CD226 and CD96 on natural killer cells in patients with pancreatic cancer. *Oncotarget* 2016; 7(41): 66586–66594.
- Gajewski TF, Schreiber H and Fu YX. Innate and adaptive immune cells in the tumor microenvironment. *Nat Immunol* 2013; 14(10): 1014–1022.
- Putz EM, Prchal-Murphy M, Simma OA, et al. PI3Kδ is essential for tumor clearance mediated by cytotoxic T lymphocytes. *PLoS One* 2012; 7(7): e40852.

- Gunderson AJ, Kaneda MM, Tsujikawa T, et al. Bruton tyrosine kinase-dependent immune cell cross-talk drives pancreas cancer. *Cancer Discov* 2016; 6(3): 270–285.
- 52. Yamazaki M, Nakamura K, Mizukami Y, et al. Sonic hedgehog derived from human pancreatic cancer cells augments angiogenic function of endothelial progenitor cells. *Cancer Sci* 2008; 99(6): 1131–1138.
- 53. Thomas AG and Awasthi N. Targeted therapy for pancreatic cancer: lessons learned and future opportunities. *Dig Med Res* 2021; 4: 1–16.
- 54. Bondar VM, Sweeney-Gotsch B, Andreeff M, et al. Inhibition of the phosphatidylinositol 3'-kinase-AKT pathway induces apoptosis in pancreatic carcinoma cells in vitro and in vivo. *Mol Cancer Ther* 2002; 1(12): 989–997.
- 55. Yip-Schneider MT, Wiesenauer CA and Schmidt CM. Inhibition of the phosphatidylinositol 3'-kinase signaling pathway increases the responsiveness of pancreatic carcinoma cells to sulindac. *J Gastrointest Surg* 2003; 7(3): 354–363.
- 56. Hu C, Dadon T, Chenna V, et al. Combined inhibition of cyclin-dependent kinases (dinaciclib) and AKT (MK-2206) blocks pancreatic tumor growth and metastases in patient-derived xenograft models. *Mol Cancer Ther* 2015; 14(7): 1532–1539.
- 57. Ma Y, Sender S, Sekora A, et al. The inhibitory response to PI3K/AKT pathway inhibitors MK-2206 and buparlisib is related to genetic differences in pancreatic ductal adenocarcinoma cell lines. *Int J Mol Sci* 2022; 23(8): 4295.
- Cao P, Maira SM, García-Echeverría C, et al. Activity of a novel, dual PI3-kinase/mTor inhibitor NVP-BEZ235 against primary human pancreatic cancers grown as orthotopic xenografts. Br J Cancer 2009; 100(8): 1267–1276.
- Totiger TM, Srinivasan S, Jala VR, et al. Urolithin A, a novel natural compound to target PI3K/AKT/mTOR pathway in pancreatic cancer. *Mol Cancer Ther* 2019; 18(2): 301–311.
- Hayes TK, Neel NF, Hu C, et al. Long-term ERK inhibition in KRAS-mutant pancreatic cancer is associated with MYC degradation and senescence-like growth suppression. *Cancer Cell* 2016; 29(1): 75–89.
- 61. Wong MH, Xue A, Baxter RC, et al. Upstream and downstream co-inhibition of mitogenactivated protein kinase and PI3K/Akt/mTOR pathways in pancreatic ductal adenocarcinoma. *Neoplasia* 2016; 18(7): 425–435.
- 62. Van Dort ME, Galbán S, Wang H, et al. Dual inhibition of allosteric mitogen-activated protein

kinase (MEK) and phosphatidylinositol 3-kinase (PI3K) oncogenic targets with a bifunctional inhibitor. *Bioorg Med Chem* 2015; 23(7): 1386–1394.

- Alagesan B, Contino G, Guimaraes AR, et al. Combined MEK and PI3K inhibition in a mouse model of pancreatic cancer. *Clin Cancer Res* 2015; 21(2): 396–404.
- 64. Awasthi N, Kronenberger D, Stefaniak A, et al. Dual inhibition of the PI3K and MAPK pathways enhances nab-paclitaxel/gemcitabine chemotherapy response in preclinical models of pancreatic cancer. *Cancer Lett* 2019; 459: 41–49.
- 65. Junttila MR, Devasthali V, Cheng JH, et al. Modeling targeted inhibition of MEK and PI3 kinase in human pancreatic cancer. *Mol Cancer Ther* 2015; 14(1): 40–47.
- 66. Grapa CM, Mocan T, Gonciar D, et al. Epidermal growth factor receptor and its role in pancreatic cancer treatment mediated by nanoparticles. *Int J Nanomedicine* 2019; 14: 9693–9706.
- Shi X, Wang M, Zhang Y, et al. Hypoxia activated HGF expression in pancreatic stellate cells confers resistance of pancreatic cancer cells to EGFR inhibition. *EBioMedicine* 2022; 86: 104352.
- Bruns CJ, Solorzano CC, Harbison MT, et al. Blockade of the epidermal growth factor receptor signaling by a novel tyrosine kinase inhibitor leads to apoptosis of endothelial cells and therapy of human pancreatic carcinoma. *Cancer Res* 2000; 60(11): 2926–2935.
- 69. Ge W, Wang Y, Quan M, et al. Activation of the PI3K/AKT signaling pathway by ARNTL2 enhances cellular glycolysis and sensitizes pancreatic adenocarcinoma to erlotinib. *Mol Cancer* 2024; 23(1): 48.
- 70. Buck E, Eyzaguirre A, Haley JD, et al. Inactivation of Akt by the epidermal growth factor receptor inhibitor erlotinib is mediated by HER-3 in pancreatic and colorectal tumor cell lines and contributes to erlotinib sensitivity. *Mol Cancer Ther* 2006; 5(8): 2051–2059.
- Hennessy BT, Smith DL, Ram PT, et al. Exploiting the PI3K/AKT pathway for cancer drug discovery. *Nat Rev Drug Discov* 2005; 4(12): 988–1004.
- 72. Buck E, Eyzaguirre A, Brown E, et al. Rapamycin synergizes with the epidermal growth factor receptor inhibitor erlotinib in non-small-cell lung, pancreatic, colon, and breast tumors. *Mol Cancer Ther* 2006; 5(11): 2676–2684.

- Dituri F, Mazzocca A, Giannelli G, et al. PI3K functions in cancer progression, anticancer immunity and immune evasion by tumors. *Clin Dev Immunol* 2011; 2011: 947858.
- 74. Glaviano A, Foo ASC, Lam HY, et al. PI3K/ AKT/mTOR signaling transduction pathway and targeted therapies in cancer. *Mol Cancer* 2023; 22: 138.
- 75. Peng Y, Wang Y, Zhou C, et al. PI3K/ Akt/mTOR pathway and its role in cancer therapeutics: are we making headway? *Front Oncol* 2022; 12: 819128.
- 76. Sharma N, Nanta R, Sharma J, et al. PI3K/AKT/ mTOR and sonic hedgehog pathways cooperate together to inhibit human pancreatic cancer stem cell characteristics and tumor growth. *Oncotarget* 2015; 6(31): 32039–32060.
- 77. Venkannagari S, Fiskus W, Peth K, et al. Superior efficacy of co-treatment with dual PI3K/ mTOR inhibitor NVP-BEZ235 and pan-histone deacetylase inhibitor against human pancreatic cancer. Oncotarget 2012; 3(11): 1416–1427.
- Stanciu S, Ionita-Radu F, Stefani C, et al. Targeting PI3K/AKT/mTOR signaling pathway in pancreatic cancer: from molecular to clinical aspects. *Int J Mol Sci* 2022; 23(17): 10132.
- 79. Mortazavi M, Moosavi F, Martini M, et al. Prospects of targeting PI3K/AKT/mTOR pathway in pancreatic cancer. *Crit Rev Oncol Hematol* 2022; 176: 103749.
- Juric D, Rodon J, Tabernero J, et al. Phosphatidylinositol 3-kinase α-selective inhibition with alpelisib (BYL719) in PIK3CAaltered solid tumors: results from the firstin-human study. J Clin Oncol 2018; 36(13): 1291–1299.
- Singh S, Bradford D, Li X, et al. FDA approval summary: alpelisib for PIK3CA-related overgrowth spectrum. *Clin Cancer Res* 2024; 30(1): 23–28.
- Soares HP, Al-Toubah TE, Kim RD, et al. Final report: a phase I trial of BYL719 in combination with gemcitabine and nab-paclitaxel in locally advanced and metastatic pancreatic cancer. *J Clin* Oncol 2018; 36(4 Suppl): 398.
- 83. Shimizu T, Tolcher AW, Papadopoulos KP, et al. The clinical effect of the dual-targeting strategy involving PI3K/AKT/mTOR and RAS/MEK/ ERK pathways in patients with advanced cancer. *Clin Cancer Res* 2012; 18(8): 2316–2325.
- 84. Bedard PL, Tabernero J, Janku F, et al. A phase Ib dose-escalation study of the oral pan-PI3K inhibitor buparlisib (BKM120) in combination

with the oral MEK1/2 inhibitor trametinib (GSK1120212) in patients with selected advanced solid tumors. *Clin Cancer Res* 2015; 21(4): 730–738.

- 85. McRee AJ, Sanoff HK, Carlson C, et al. A phase I trial of mFOLFOX6 combined with the oral PI3K inhibitor BKM120 in patients with advanced refractory solid tumors. *Invest New Drugs* 2015; 33(6): 1225–1231.
- Hong DS, Bowles DW, Falchook GS, et al. A multicenter phase I trial of PX-866, an oral irreversible phosphatidylinositol 3-kinase inhibitor, in patients with advanced solid tumors. *Clin Cancer Res* 2012; 18(15): 4173–4182.
- Patnaik A, Appleman LJ, Tolcher AW, et al. First-in-human phase I study of copanlisib (BAY 80-6946), an intravenous pan-class I phosphatidylinositol 3-kinase inhibitor, in patients with advanced solid tumors and non-Hodgkin's lymphomas. *Ann Oncol* 2016; 27(10): 1928–1940.
- 88. Sarker D, Ang JE, Baird R, et al. First-in-human phase I study of pictilisib (GDC-0941), a potent pan-class I phosphatidylinositol-3-kinase (PI3K) inhibitor, in patients with advanced solid tumors. *Clin Cancer Res* 2015; 21(1): 77–86.
- Shapiro GI, LoRusso P, Kwak E, et al. Phase Ib study of the MEK inhibitor cobimetinib (GDC-0973) in combination with the PI3K inhibitor pictilisib (GDC-0941) in patients with advanced solid tumors. *Invest New Drugs* 2020; 38(2): 419–432.
- 90. Xie P, Tan SY, Li HF, et al. Transcriptome data-based status of PI3K/AKT/mTOR pathway indicates heterogeneity and immune modulation in patients with pancreatic ductal adenocarcinoma. *J Gene Med* 2024; 26(1): e3570.
- 91. Marsh RW, Rocha Lima CM, Levy DE, et al. A phase II trial of perifosine in locally advanced, unresectable, or metastatic pancreatic adenocarcinoma. *Am J Clin Oncol* 2007; 30(1): 26–31.
- 92. Hedley D, Moore MJ, Hirte H, et al. A phase II trial of perifosine as second line therapy for advanced pancreatic cancer. A study of the Princess Margaret Hospital [PMH] Phase II Consortium. *J Clin Oncol* 2005; 23(16 Suppl): 4166.
- 93. Azaro A, Rodon J, Calles A, et al. A first-inhuman phase I trial of LY2780301, a dual p70 S6 kinase and Akt Inhibitor, in patients with advanced or metastatic cancer. *Invest New Drugs* 2015; 33(3): 710–719.

- 94. Tolcher AW, Patnaik A, Papadopoulos KP, et al. Phase I study of the MEK inhibitor trametinib in combination with the AKT inhibitor afuresertib in patients with solid tumors and multiple myeloma. *Cancer Chemother Pharmacol* 2015; 75(1): 183–189.
- 95. Tolcher AW, Kurzrock R, Valero V, et al. Phase I dose-escalation trial of the oral AKT inhibitor uprosertib in combination with the oral MEK1/ MEK2 inhibitor trametinib in patients with solid tumors. *Cancer Chemother Pharmacol* 2020; 85(4): 673–683.
- 96. Tolcher AW, Khan K, Ong M, et al. Anti-tumour activity in RAS-driven tumours by blocking AKT and MEK. *Clin Cancer Res* 2015; 21(4): 739–748.
- 97. Murphy AG, Zahurak M, Shah M, et al. A phase I study of dinaciclib in combination with MK-2206 in patients with advanced pancreatic cancer. *Clin Transl Sci* 2020; 13(6): 1178–1188.
- 98. Chung V, McDonough S, Philip PA, et al. Effect of selumetinib and MK-2206 vs oxaliplatin and fluorouracil in patients with metastatic pancreatic cancer after prior therapy: SWOG S1115 study randomized clinical trial. *JAMA Oncol* 2017; 3(4): 516–522.
- 99. Pan Y, Rhea P, Tan L, et al. PBI-05204, a supercritical CO extract of Nerium oleander, inhibits growth of human pancreatic cancer via targeting the PI3K/mTOR pathway. *Invest New Drugs* 2015; 33(2): 271–279.
- 100. Hong DS, Henary H, Falchook GS, et al. First-in-human study of pbi-05204, an oleanderderived inhibitor of akt, fgf-2, nf-κB and p70s6k, in patients with advanced solid tumors. *Invest New Drugs* 2014; 32(6): 1204–1212.
- 101. Roth MT, Cardin DB, Borazanci EH, et al. A phase II, single-arm, open-label, Bayesian adaptive efficacy and safety study of PBI-05204 in patients with stage IV metastatic pancreatic adenocarcinoma. *Oncologist* 2020; 25(10): e1446–e1450.
- 102. Ichikawa K, Ito S, Kato E, et al. TAS0612, a novel RSK, AKT, and S6K inhibitor, exhibits antitumor effects in preclinical tumor models. *Mol Cancer Ther* 2024; 23(2): 174–186.
- 103. Serra V, Eichhorn PJA, García-García C, et al. RSK3/4 mediate resistance to PI3K pathway inhibitors in breast cancer. J Clin Invest 2013; 123(6): 2551–2563.
- 104. ClinicalTrials.gov. A study of TAS0612 in participants with advanced or metastatic solid tumor cancer—full text view, https://classic.

clinicaltrials.gov/ct2/show/NCT04586270 (2024, accessed 8 April 2024).

- 105. Garrido-Laguna I, Tan AC, Uson M, et al. Integrated preclinical and clinical development of mTOR inhibitors in pancreatic cancer. Br J Cancer 2010; 103(5): 649–655.
- 106. Gangadhar TC, Cohen EEW, Wu K, et al. Two drug interaction studies of sirolimus in combination with sorafenib or sunitinib in patients with advanced malignancies. *Clin Cancer Res* 2011; 17(7): 1956–1963.
- 107. Zhang JW, Zhao F and Sun Q. Metformin synergizes with rapamycin to inhibit the growth of pancreatic cancer in vitro and in vivo. *Oncol Lett* 2018; 15(2): 1811–1816.
- 108. Bever KM, Borazanci EH, Thompson EA, et al. An exploratory study of metformin with or without rapamycin as maintenance therapy after induction chemotherapy in patients with metastatic pancreatic adenocarcinoma. *Oncotarget* 2020; 11(21): 1929–1941.
- 109. Hou S, Schmid AN and Desai NP: ABI-009 (nab-Sirolimus) improves tumor accumulation and antitumor activity over oral mTOR inhibitors. *Cancer Res* 2019; 79(13 Suppl): 348.
- 110. Aadi Bioscience, Inc. A phase 1, open-label, dose-escalation study to assess safety and pharmacokinetics of nab-sirolimus in patients with locally advanced or metastatic solid tumors and moderate liver impairment, clinicaltrials. gov, Report No. NCT05661461, https:// clinicaltrials.gov/study/NCT05661461 (2023, accessed 1 January 2024).
- 111. Aadi Bioscience, Inc. A phase 2 multi-center open-label basket trial of nab-sirolimus for adult and adolescent patients with malignant solid tumors harboring pathogenic inactivating alterations in TSC1 or TSC2 genes, clinicaltrials.gov, Report No. NCT05103358, https://clinicaltrials.gov/study/NCT05103358 (2024, accessed 1 January 2024).
- 112. Mirati Therapeutics Inc. A phase 1/2 trial of adagrasib in combination with nab-sirolimus in patients with advanced solid tumors and non-small cell lung cancer with a KRAS G12C mutation, clinicaltrials.gov, Report No. NCT05840510, https://clinicaltrials.gov/study/ NCT05840510 (2024, accessed 1 January 2024).
- 113. Wolpin BM, Hezel AF, Abrams T, et al. Oral mTOR inhibitor everolimus in patients with gemcitabine-refractory metastatic pancreatic cancer. *J Clin Oncol* 2009; 27(2): 193–198.
- 114. Kordes S, Klümpen HJ, Weterman MJ, et al. Phase II study of capecitabine and the oral

mTOR inhibitor everolimus in patients with advanced pancreatic cancer. *Cancer Chemother Pharmacol* 2015; 75(6): 1135–1141.

- 115. Kordes S, Richel DJ, Klümpen HJ, et al. A phase I/II, non-randomized, feasibility/safety and efficacy study of the combination of everolimus, cetuximab and capecitabine in patients with advanced pancreatic cancer. *Invest New Drugs* 2013; 31(1): 85–91.
- 116. Second Affiliated Hospital of Guangzhou Medical University. Trametinib combined with everolimus and lenvatinib in the treatment of recurrent/refractory advanced solid tumors: a phase II clinical trial, clinicaltrials.gov, Report No. NCT04803318, https://clinicaltrials.gov/ study/NCT04803318 (2023, accessed 1 January 2024).
- 117. Royal Marsden NHS Foundation Trust. A phase I trial of VS-6766 (RO5126766) (a dual RAF/MEK inhibitor) exploring intermittent, oral dosing regimens in patients with solid tumours or multiple myeloma, with an expansion to explore intermittent dosing in combination with everolimus, clinicaltrials.gov, Report No. NCT02407509, https://clinicaltrials.gov/study/ NCT02407509 (2024, accessed 1 January 2024).
- 118. Javle MM, Shroff RT, Xiong H, et al. Inhibition of the mammalian target of rapamycin (mTOR) in advanced pancreatic cancer: results of two phase II studies. *BMC Cancer* 2010; 10: 368.
- 119. Karavasilis V, Samantas E, Koliou GA, et al. Gemcitabine combined with the mTOR inhibitor temsirolimus in patients with locally advanced or metastatic pancreatic cancer. A Hellenic Cooperative Oncology Group Phase I/ II Study. *Target Oncol* 2018; 13(6): 715–724.
- 120. Nemunaitis J, Hochster HS, Lustgarten S, et al. A phase I trial of oral ridaforolimus (AP23573; MK-8669) in combination with bevacizumab for patients with advanced cancers. *Clin Oncol (R Coll Radiol)* 2013; 25(6): 336–342.
- 121. Basu B, Dean E, Puglisi M, et al. First-inhuman pharmacokinetic and pharmacodynamic study of the dual m-TORC 1/2 inhibitor AZD2014. *Clin Cancer Res* 2015; 21(15): 3412–3419.
- 122. Soares HP, Ming M, Mellon M, et al. Dual PI3K/mTOR inhibitors induce rapid overactivation of the MEK/ERK pathway in human pancreatic cancer cells through suppression of mTORC2. *Mol Cancer Ther* 2015; 14(4): 1014–1023.
- 123. Hassan Z, Schneeweis C, Wirth M, et al. MTOR inhibitor-based combination therapies

for pancreatic cancer. *Br J Cancer* 2018; 118(3): 366–377.

- 124. Papadopoulos KP, Tabernero J, Markman B, et al. Phase I safety, pharmacokinetic, and pharmacodynamic study of SAR245409 (XL765), a novel, orally administered PI3K/mTOR inhibitor in patients with advanced solid tumors. *Clin Cancer Res* 2014; 20(9): 2445–2456.
- 125. Schram AM, Gandhi L, Mita MM, et al. A phase Ib dose-escalation and expansion study of the oral MEK inhibitor pimasertib and PI3K/MTOR inhibitor voxtalisib in patients with advanced solid tumours. *Br J Cancer* 2018; 119(12): 1471–1476.
- 126. Knight SD, Adams ND, Burgess JL, et al. Discovery of GSK2126458, a highly potent inhibitor of PI3K and the mammalian target of rapamycin. ACS Med Chem Lett 2010; 1(1): 39–43.
- 127. Munster P, Aggarwal R, Hong D, et al. Firstin-human phase I study of GSK2126458, an oral pan-class I phosphatidylinositol-3-kinase inhibitor, in patients with advanced solid tumor malignancies. *Clin Cancer Res* 2016; 22(8): 1932–1939.
- 128. Wainberg ZA, Alsina M, Soares HP, et al. A multi-arm phase I study of the PI3K/mTOR inhibitors PF-04691502 and gedatolisib (PF-05212384) plus irinotecan or the MEK inhibitor PD-0325901 in advanced cancer. *Target Oncol* 2017; 12(6): 775–785.
- 129. ClinicalTrials.gov. Study of the CDK4/6 inhibitor palbociclib (PD-0332991) in combination with

the PI3K/mTOR inhibitor gedatolisib (PF-05212384) for patients with advanced squamous cell lung, pancreatic, head & neck and other solid tumors—full text view, https://classic.clinicaltrials.gov/ct2/show/NCT03065062 (2024, accessed 4 April 2024).

- 130. Wright SCE, Vasilevski N, Serra V, et al. Mechanisms of resistance to PI3K inhibitors in cancer: adaptive responses, drug tolerance and cellular plasticity. *Cancers* 2021; 13(7): 1538.
- 131. Tan J and Yu Q. Molecular mechanisms of tumor resistance to PI3K-mTOR-targeted therapy. *Chin J Cancer* 2013; 32(7): 376–379.
- Porta C, Paglino C and Mosca A. Targeting PI3K/Akt/mTOR signaling in cancer. Front Oncol 2014; 4: 64.
- 133. Hoxhaj G and Manning BD. The PI3K-AKT network at the interface of oncogenic signalling and cancer metabolism. *Nat Rev Cancer* 2020; 20(2): 74–88.
- 134. Ando Y, Inada-Inoue M, Mitsuma A, et al. Phase I dose-escalation study of buparlisib (BKM120), an oral pan-class I PI3K inhibitor, in Japanese patients with advanced solid tumors. *Cancer Sci* 2014; 105(3): 347–353.
- 135. Bardia A, Gounder M, Rodon J, et al. Phase Ib study of combination therapy with MEK inhibitor binimetinib and phosphatidylinositol 3-kinase inhibitor buparlisib in patients with advanced solid tumors with RAS/RAF alterations. *Oncologist* 2020; 25(1): e160–e169.

Visit Sage journals online journals.sagepub.com/ home/tam

Sage journals