

Therapeutic Potential of Metabotropic Glutamate Receptor Modulators

N. Hovelsø^{1,*}, F. Sotty¹, L.P. Montezinho¹, P.S. Pinheiro², K.F. Herrik¹ and A. Mørk¹

¹Department of Neurophysiology, H. Lundbeck A/S, Ottiliavej 9, 2500 Copenhagen-Valby, Denmark; ²Department of Neuroscience and Pharmacology, Faculty of Health Sciences, University of Copenhagen, 2200 Copenhagen N, Denmark

Abstract: Glutamate is the main excitatory neurotransmitter in the central nervous system (CNS) and is a major player in complex brain functions. Glutamatergic transmission is primarily mediated by ionotropic glutamate receptors, which include NMDA, AMPA and kainate receptors. However, glutamate exerts modulatory actions through a family of metabotropic G-protein-coupled glutamate receptors (mGluRs). Dysfunctions of glutamatergic neurotransmission have been implicated in the etiology of several diseases. Therefore, pharmacological modulation of ionotropic glutamate receptors has been widely investigated as a potential therapeutic strategy for the treatment of several disorders associated with glutamatergic dysfunction. However, blockade of ionotropic glutamate receptors might be accompanied by severe side effects due to their vital role in many important physiological functions. A different strategy aimed at pharmacologically interfering with mGluR function has recently gained interest. Many subtype selective agonists and antagonists have been identified and widely used in preclinical studies as an attempt to elucidate the role of specific mGluRs subtypes in glutamatergic transmission. These studies have allowed linkage between specific subtypes and various physiological functions and more importantly to pathological states. This article reviews the currently available knowledge regarding the therapeutic potential of targeting mGluRs in the treatment of several CNS disorders, including schizophrenia, addiction, major depressive disorder and anxiety, Fragile X Syndrome, Parkinson's disease, Alzheimer's disease and pain.

Keywords: Addiction, alzheimer's disease, anxiety, depression, epilepsy, fragile X syndrome, Huntington's disease, metabotropic glutamate receptors, pain, Parkinson's disease, schizophrenia.

INTRODUCTION

Glutamate is the main excitatory neurotransmitter in the central nervous system (CNS) and exerts its effects through the activation of several receptor subtypes. Glutamate mediates fast excitatory synaptic transmission between neurons through the ionotropic receptors α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA), N-methyl-D-aspartate (NMDA) and kainate receptors. Furthermore, glutamate exerts a prominent modulatory role of the fast excitatory tone set by the ionotropic receptors by activation of the metabotropic glutamate receptor (mGluR) family. Ionotropic and metabotropic glutamate receptors interact in the fine tuning of neuronal responses under different conditions of activity and their co-localisation indicates that glutamate modulates neurotransmission and neuronal excitability at the same synapses [1,2]. In addition, mGluRs are also present at synapses releasing neurotransmitters other than glutamate.

The actions of glutamate on ionotropic receptors are responsible for numerous physiological processes, including basic neuronal communication, axonal pathfinding, mood regulation, and memory formation. It has been suggested that dysregulation of the glutamatergic system is implicated in a variety of psychiatric and neurological disorders such as schizophrenia, major depression disorder, and Parkinson's

disease. In this review some basic features of mGluRs will briefly be revisited before a comprehensive overview of their therapeutic potential for the treatment of several psychiatric and neurological disorders will be discussed.

Classification, Structure, and Function

Metabotropic glutamate receptors belong to the superfamily of G-protein-coupled receptors. Structurally, mGluRs are formed by a large extracellular N-terminal domain containing the glutamate binding site and seven α -helical transmembrane segments [3]. The most conserved domains between the different mGluRs subtypes are the site involved in coupling to the G-protein and the glutamate binding site [2]. Functional mGluRs comprise homodimers stabilized by both an inter-subunit disulphide bond and hydrophobic interactions [2]. Eight different metabotropic receptors have been cloned (mGluRs 1-8). They are classified into three groups according to their sequence homology, pharmacological properties and intracellular signal transduction pathways Fig. (1). Group I consists of mGluR1 and mGluR5, and are positively coupled to phospholipase C through G-proteins of the G_q/G₁₁ type. Activation of group I receptors leads to stimulation of phospholipase C, production of inositol triphosphate, release of Ca²⁺ from intracellular stores and production of diacylglycerol, which in turn activates protein kinase C [4-6]. Group II, consisting of mGluR2 and mGluR3, and Group III, consisting of mGluR4, mGluR6, mGluR7 and mGluR8, are coupled to the inhibition of adenylyl cyclase activity through G-proteins of the G_i/G_o type. Approximately 70% sequence homology exists within each group of mGluRs,

*Address correspondence to this author at the Department of Neurophysiology, H. Lundbeck A/S, Ottiliavej 9, 2500 Copenhagen-Valby, Denmark; Tel: +4536434851; Fax: +4536438258; E-mail: nann@lundbeck.com

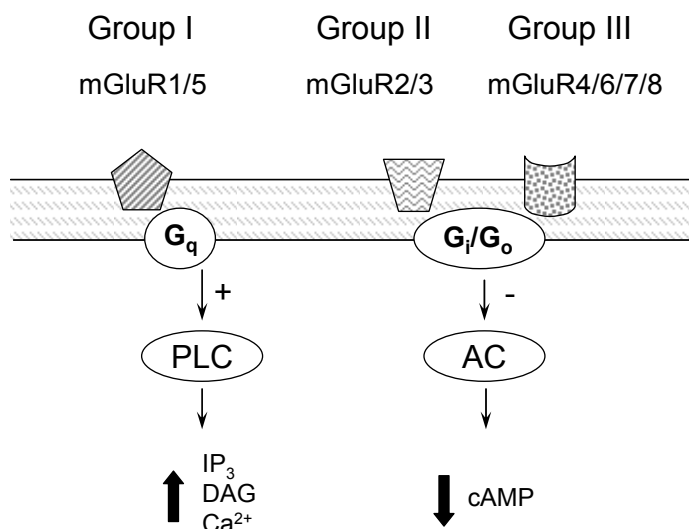


Fig. (1). Intracellular signalling pathways associated to the different mGluR subtypes. Group I mGluRs, including mGluR1 and mGluR5, are positively coupled to phospholipase C (PLC) through activation of a G-protein of the G_q type; in turn, production of inositol triphosphate (IP₃), release of Ca^{2+} from intracellular stores and production of diacylglycerol (DAG) activate protein kinase C. Group II and Group III mGluRs are negatively coupled to adenylyl cyclase (AC) through G-protein of the G_i/G_o type, leading to decreased formation of cyclic AMP (cAMP).

while approximately 45% sequence homology has been described between groups. Alternative splice variants have also been described for mGluR1, mGluR4, mGluR5, and mGluR7 [7].

Expression Pattern and Subcellular Localization

Group I mGluRs

Immunohistochemical studies have shown that high levels of mGluR1 are present in the hippocampus, globus pallidus, substantia nigra, thalamus, cerebellum and the olfactory bulb [7,8]. Lower levels of mGluR1 have also been found in the striatum, neocortex, and hypothalamus [7,9,10]. The mGluR5s are expressed in the cortex, striatum, caudate nucleus, nucleus accumbens and septum, as well as in the hippocampus and olfactory bulb, where they are highly co-expressed with mGluR1. Lower levels have been described in the cerebellum and thalamus. Both mGluR1 and mGluR5 are mainly concentrated in postsynaptic structures [11,12] and with a few exceptions almost undetectable in presynaptic structures [9,13]. At the subcellular level, mGluR5s are found both synaptically and extrasynaptically on postsynaptic spines of principal neurons [9,12-14] Fig. (2).

Group II mGluRs

Both mGluR2s and mGluR3s are highly expressed in the hippocampus, cortex, nucleus accumbens, striatum and amygdala. In addition, mGluR2s are found at high levels in the caudate nucleus, cerebellar cortex and olfactory bulb, while mGluR3s are enriched in the septum and substantia nigra [14]. In the hippocampus, mGluR2s are mainly found at presynaptic sites [15,16], whereas mGluR3s are mostly expressed postsynaptically [17]. Furthermore, presynaptically localized mGluR2s and mGluR3s are mostly found on axons and pre-terminal regions of both glutamatergic and GABAergic synapses [17]. In some brain structures, e.g.

hippocampus, cortex, and striatum, mGluR2s and mGluR3s have been found at both presynaptic and postsynaptic sites [17-22]. Postsynaptically, mGluR2s are mostly concentrated in cell bodies and dendritic shafts, whereas mGluR3s are found to be perisynaptic on spines [14,17] Fig. (2).

Group III mGluRs

The mGluR4s show a widespread brain distribution, but high distribution levels are only reported in the hippocampus and cerebellar cortex, while the expression of mGluR6s is limited to the retina [23]. The mGluR7s have a widespread distribution and can be found at high levels in several brain regions including hippocampus, cortex, globus pallidus, amygdala, colliculi and olfactory bulb. Lower expression levels are found in the striatum, substantia nigra, caudate nucleus and nucleus accumbens. The mGluR8s are found predominantly in the hippocampus, hypothalamus and olfactory bulb, but at low levels.

Group III mGluRs mainly serve as presynaptic autoreceptors involved in reducing glutamate release from presynaptic terminals [16,24-26], and in contrast to Group II mGluRs they are mostly located synaptically. Furthermore, mGluR7s are not only localized within the presynaptic active zone [25,26], but also on GABAergic terminals Fig. (2).

mGluRs in the Spinal Cord

Almost all mGluR subtypes have been described in the spinal cord [27-31]. In particular, both Group I and II mGluRs have been found in the soma and terminals of dorsal root ganglion neurons [32-34].

mGluRs in Non-Neuronal Cells

Several mGluR subtypes are expressed by non-neuronal brain cells, including microglia, oligodendrocytes and astrocytes. In microglial cells, activation of group III mGluRs

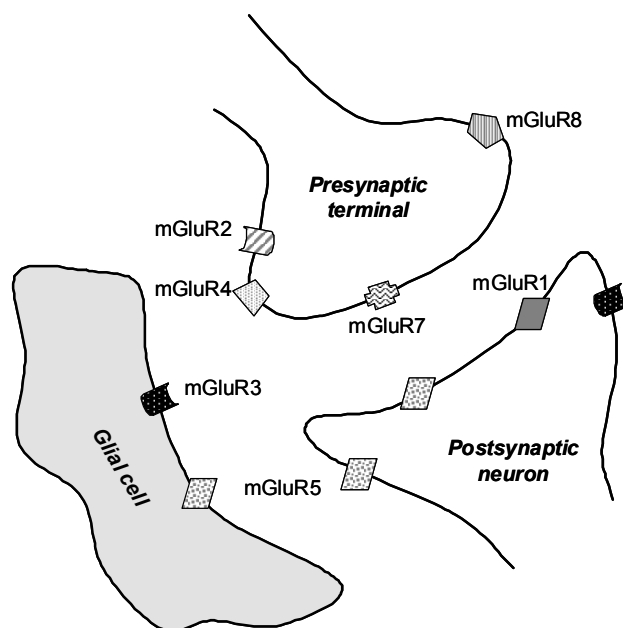


Fig. (2). Subcellular localization of the different mGluR subtypes in the glutamatergic synapse. Group I mGluRs, including mGluR1 and mGluR5, are almost exclusively located post-synaptically, and mGluR5 are present both synaptically and extrasynaptically. Group II mGluRs are predominantly located on presynaptic terminals in the extrasynaptic space, with the exception of mGluR3 which is highly expressed on glial cells as well as postsynaptically. Group III mGluRs are also principally expressed presynaptically; interestingly, mGluR7 is located in the synaptic cleft while mGluR4 and 8 are mostly extrasynaptic.

Note that mGluR3 and mGluR7 are also present on non-glutamatergic (e.g. GABAergic) terminals.

activates a neuroprotective pathway [35], while activation of group II mGluRs induces an apoptotic pathway [35,36]. In oligodendrocytes, mGluRs have been suggested to be involved in the control of oxidative stress and excitotoxicity [37]. In astrocytes, mGluRs have been suggested to play a role in the modulation of synaptic transmission within neighboring neurons. For instance, activation of astrocytic mGluR5 induces calcium oscillations, in turn leading to glutamate release that can affect neurotransmission [38]. The activation of astrocytic mGluRs has also been shown to alter the expression of glutamate transporters [39], and stimulate the release of inflammatory cytokines [40].

Molecular Mechanisms of Action

The mGluRs generally exert modulatory roles although some members of the family have been found to mediate synaptic transmission directly *via* activation of slow postsynaptic potentials [41]. Modulation is achieved either through activation of intracellular second messenger pathways, or through a direct action of the $\beta\gamma$ subunits of the heterotrimeric G-proteins in modulating, e.g. ion channels activity [42]. Despite their widespread distribution in the brain, there is currently limited knowledge on how mGluRs modulate the physiological release of glutamate [43]. However, the discovery of selective mGluR ligands has revealed that mGluRs localised presynaptically modulate neurotransmitter release in a number of ways which include the modulation of presynaptic Ca^{2+} and K^{+} channels and regulation of the release machinery.

Group I mGluRs trigger a signalling cascade involving protein kinase and ultimately leading to the inhibition of presynaptic K^{+} channels, thereby delaying repolarisation of

the nerve terminals [44,45]. They have also been shown to exert a positive effect by promoting the activity of presynaptic voltage dependent Ca^{2+} channels [46], although bidirectional actions have been reported [47]. A negative impact on transmitter release can also be exerted indirectly by postsynaptic Group I mGluRs through stimulation of endocannabinoid release and activation of presynaptic cannabinoid CB1 receptors [48].

Group II and III mGluRs generally act to reduce neurotransmitter release. For instance, their activation reduces glutamate release through depression of P/Q-type Ca^{2+} channels [49,50]. This might be achieved by a direct action of the dissociated G-protein $\beta\gamma$ subunits [51]. The mGluR7 mediated depression of Ca^{2+} channel function requires phospholipase C or protein kinase C activity [52,53]. Additionally, the actions of mGluR7s are independent of cAMP and protein kinase A, although group III mGluRs typically operate by reducing cyclic AMP at the synapses between mossy fibres and stratum lucidum interneurons [1,52]. Inhibitory action of presynaptic mGluRs may involve a direct regulation of the release/exocytosis machinery downstream of Ca^{2+} entry. Indeed, the $\beta\gamma$ subunits of G proteins can directly modulate release [54] through interactions with the SNARE complex [55,56] and also lead to a reduction in the number of active release sites [57].

Modulation of Synaptic Plasticity

The mGluRs have been implicated in a number of synaptic plasticity phenomena, including short-term modification of synaptic strength, long-term depression (LTD) and long-term potentiation (LTP).

Long-Term Depression

Activation of Group I mGluRs, either synaptically or pharmacologically, has been shown to induce long-term depression of synaptic transmission [58-64]. For instance, Group I mGluRs were found to play a critical role in the induction of long-term depression at the corticostriatal synapse, presumably by the regulation of intracellular calcium [65]. Some studies have suggested that the induced long-term depression was mediated indirectly through postsynaptic release of retrograde messengers [66].

A classical example of group II mGluR-mediated LTD is found at hippocampal mossy fiber synapses, where prolonged low-frequency stimulation activates Group II mGluRs localized preterminally, causing a persistent reduction in glutamate release [67,68] that is exclusively presynaptic in nature. At this particular synapse, both LTD and LTP share a common mechanism and the same protocols that induce LTD through presynaptic mGluRs can be used to reverse LTP, a phenomenon termed depotentiation [69]. Group II mGluR-dependent depotentiation is also observed in the amygdala [70].

Presynaptic Group III mGluRs were shown to be involved in the induction of LTD at synapses between hippocampal mossy fibers and stratum lucidum interneurons, as well as at excitatory inputs onto stratum radiatum interneurons [49,52,71]. This form of plasticity, however, requires a rise in postsynaptic Ca^{2+} through Ca^{2+} -permeable AMPA receptors.

The mechanisms of mGluR-dependent LTD are discussed in section "Fragile X Syndrome" [64,72].

Long-Term Potentiation

The activation of mGluRs has also been shown to be required for both NMDA receptor-dependent and independent forms of LTP [73,74]. Group I mGluRs have been involved in the induction of LTP at the cortico-striatal synapse, where co-activation of both mGluR1 and mGluR5 is required, thereby contributing to a strong increase in intracellular calcium levels [75]. In addition, Group I mGluRs are required for the induction of a new form of LTP specific of NMDA receptors at hippocampal mossy fiber synapses [76,77]. In NMDA receptor-dependent LTP, mGluR activation apparently serves as a priming factor or molecular switch and mGluR5 [78-80], but not mGluR1 [81] seems to be critically involved. Some forms NMDA receptor-independent LTP also require mGluR activation [82,83] and Group I mGluRs seems to be the major player [81,84,85]. A recent report showed a role for Group I mGluRs in the induction of long-term plasticity of non-synaptic, synchronized neuronal activity [86]. A role for group II mGluRs in LTP has also been evidenced [87,88], whereas Group III mGluRs presumably have an indirect effect on LTP induction at mossy fiber synapses onto stratum lucidum interneurons, a process that has been attributed to presynaptic mGluR7 functioning as a metaplastic switch [52].

Pharmacological Tools

In the present section, we will introduce compounds used in preclinical studies aimed at elucidating the role of specific

mGluR subtypes in the pathophysiology of several psychiatric and neurological disorders (Table 1).

SCHIZOPHRENIA

Schizophrenia is a psychiatric disorder affecting 1% of the population worldwide [89,90]. Symptoms associated with schizophrenia have traditionally been categorised into positive symptoms e.g. delusions and hallucinations, negative symptoms e.g. blunting of affect, social withdrawal, lack of motivation, cognitive deficits e.g. impairment in memory, executive function, working and long-term memory and affective symptoms [91]. The dopamine hypothesis of schizophrenia, implying a hyperactivity of the dopaminergic system, has been the main neurochemical hypothesis for many years [90,92]. Current antipsychotic drugs are all dopamine D_2 receptor antagonists with varying degrees of potency, and while these drugs to some extent are effective against the positive symptoms, the effects on negative and cognitive symptoms are very limited. Furthermore, the treatment is often accompanied by a range of side-effects such as extrapyramidal motor symptoms and the metabolic syndrome [90]. There is increasing evidence that a primary glutamatergic dysfunction is associated with schizophrenia, leading secondarily to a dopaminergic imbalance. More precisely, a current model for the pathophysiology of schizophrenia involves a mechanism by which NMDA receptor hypofunction would induce a dysregulation of GABA fast-spiking interneurons in cortical areas, leading in turn to a disinhibition of pyramidal glutamatergic output and contributing to a reduction in the synchronized neuronal activity of these neurons [93,94]. Further support of the involvement of an altered glutamatergic neurotransmission in the pathophysiology of schizophrenia stems from the observation that the NMDA antagonist, ketamine, induces schizophrenia-like symptoms in healthy volunteers, including transient psychosis, disrupted affect, and cognitive deficits, and exacerbates specific symptoms in schizophrenic patients [95,96]. This growing body of evidence involving disturbances in glutamatergic transmission in schizophrenia is at the basis of an increasing effort aimed at modulating metabotropic glutamate receptors as a possible therapeutic strategy (Table 2).

Selective Group I Modulation

Based on the purported disinhibition of pyramidal glutamatergic output as a core feature of schizophrenia, modulation of glutamate signalling at the postsynaptic levels *via* group I metabotropic receptors, and in particular mGluR5s, has gained interest as a potential promising therapeutic strategy.

mGluR1

There is only limited evidence that mGluR1 positive modulation may be of interest for the treatment of schizophrenia. On one hand, increased mGluR1 expression has been reported in the prefrontal cortex of schizophrenics [97]. In addition, mGluR1 knockout mice have impaired sensorimotor gating as assessed in a prepulse inhibition paradigm [98], a deficit also encountered in schizophrenic patients. However, the sensorimotor gating deficits observed in mGluR1 knockout mice were only reversed by lamotrigine, an anticonvulsant drug used in the treatment of bipolar

Table 1. Compounds with mGluR Affinity Cited in the Text with their Chemical Names, Main Target and Action

| Compound Abbreviation | Chemical Name | Target | Action |
|-----------------------|--|-------------------|---------------------|
| A794278 | 9-dimethylamino-3-cycloheptyl-3H-5-thia-1,3,6-triazafuoren-4-one | Group I, mGluR1 | Antagonist |
| A794282 | 9-dimethylamino-3-(4-ethylphenyl)-3H-5-thia-1,3,6-triazafuoren-4-one | Group I, mGluR1 | Antagonist |
| A841720 | 9-dimethylamino-3-(N-hexamethyleneiminy)-3H-5-thia-1,3,6-triazafuoren-4-one | Group I, mGluR1 | Antagonist |
| A850002 | 9-dimethylamino-3-(4-methylphenyl)-3H-5-thia-1,3,7-triazafuoren-4-one | Group I, mGluR1 | Antagonist |
| ACPT-I | (1S,3R,4S)-1-aminocyclopentane-1,3,4-tricarboxylic acid | Group III | Agonist |
| ADX47273 | (S)-(4-fluorophenyl)-(3-[3-(4-fluoro-phenyl)-[1,2,4]-oxadiazol-5-yl]piperidin-1-yl) methanone | Group I, mGluR5 | PAM |
| AIDA | 1-aminoindan-1,5-dicarboxylic acid | Group I, mGluR1 | Antagonist |
| AMN082 | N,N'-dibenzhydrylethane-1,2-diamine dihydrochloride | Group III, mGluR7 | Agonist |
| APDC | (2R,4R)-4-aminopyrrolidine-2,4-dicarboxylate | Group II | Agonist |
| (1S,3S)-APDC | aminopyrrolidine-2,4-dicarboxylate | Group II | Agonist |
| BAY367620 | [(3aS,6aS)-6a-naphtalen-2-yl-methyl-5-methyliden-hexahydro-cyclopental[c]furan-1-on] | Group I, mGluR1 | Antagonist |
| BINA | biphenyl-indanone A | Group II, mGluR2 | PAM |
| Compound 5 | 1-(2-Hydroxy-3-methyl-4-{4-[4-(2H-tetrazol-5-yl)phenoxy]butoxy}phenyl)ethanone | Group II, mGluR2 | PAM |
| Compound 8q | 3-cyano-N-(1,3-diphenyl-1H-pyrazol-5-yl)benzamide | Group I, mGluR5 | NAM |
| CDPPB | 3-cyano-N-(1,3-diphenyl-1H-pyrazol-5-yl)benzamide | Group I, mGluR5 | PAM |
| (S)-4C3HPG | 4-[(1S)-1-amino-2-hydroxy-2-oxoethyl]-2-hydroxybenzoic acid | Group I; Group II | Antagonist; Agonist |
| CPCCOEt | 7-(hydroxyimino)cyclopropa[b]chromen-1a-carboxylate ethyl ester | Group I, mGluR1 | Antagonist |
| (S)-4CPG | (S)-4-carboxyphenylglycine | Group I | Antagonist |
| DCG-IV | (2S,2'R,3'R)-2-(2',3'-dicarboxycyclopropyl)glycine | Group II | Agonist |
| (S)-3,4-DCPG | (S)-3,4-dicarboxyphenylglycine | Group III, mGluR8 | Agonist |
| DHPG | (S)-3,5-dihydroxyphenylglycine | Group I | Agonist |
| EMQMCM | 3-ethyl-2-methyl-quinolin-6-yl)-(4-methoxy-cyclohexyl)-methanone methanesulfonate | Group I, mGluR1 | Antagonist |
| Fenobam | 1-(3-chlorophenyl)-3-(3-methyl-5-oxo-4H-imidazol-2-yl) urea | Group I, mGluR5 | Antagonist |
| JNJ16259685 | 3,4-dihydro-2H-pyrano[2,3-b]quinolin-7-yl)-(cis-4-methoxycyclohexyl) methadone | Group I, mGluR1 | Antagonist |
| L-AP4 | L-(+)-2-amino-4-phosphonobutyric acid | Group III | Agonist |
| L-SOP | L-serine-O-phosphate | Group III | Agonist |
| LSP1-2111 | Undisclosed | Group III | Agonist |
| LY339764 | (R,S)-2-amino-2-(4-carboxycyclobutyl-3-(9-xanthen-9-yl)propanoic acid | Group I, mGluR5 | Antagonist |
| LY339840 | (S)-(+)-a-amino-4-carboxy-2-methylbenzeneacetic acid | Group I, mGluR1 | Antagonist |
| LY341495 | 1S,2S)-2-[(2S)-2-amino-3-(2,6-dioxo-3H-purin-9-yl)-1-hydroxy-1-oxopropan-2-yl]cyclopropane-1-carboxylic acid | Group II | Antagonist |
| LY354740 | (1S,2S,5R,6S)-2-aminobicyclo[3.1.0]hexane-2,6-dicarboxylic acid | Group II | Agonist |
| LY367335 | 2-amino-2-(3-cis and trans-carboxycyclobutyl)-3(9H-xanthen-9-yl)propionic acid | Group I | Antagonist |
| LY367366 | (R,S)-2-amino-2-(4-carboxyphenyl)-3-(9H-thioxanthen-9-yl) propanoic acid | Group I, mGluR5 | Antagonist |
| LY367385 | (+)-2-methyl-4-carboxyphenylglycine | Group I, mGluR1 | Antagonist |
| LY379268 | (2R,6R)-2-amino-4-oxabicyclo[3.1.0]hexane-2,6-dicarboxylic acid | Group II | Agonist |

Table 1. contd....

| Compound Abbreviation | Chemical Name | Target | Action |
|-----------------------|--|------------------|------------|
| LY389795 | 1R,4R,5S,6R)-4-amino-2-thiabicyclo[3.1.0]hexane-4,6-dicarboxylic acid | Group II | Agonist |
| LY393053 | 2-amino-2(S)-(3-cis-carboxycyclobutyl-3-(9-thioxanthen-9-yl)propionic acid) | Group I | Antagonist |
| LY404039 | (-)-(1R,4S,5S,6S)-4-amino-2-sulfonylbicyclo[3.1.0]hexane-4,6-dicarboxylic acid | Group II | Agonist |
| LY456066 | 2-[4-(In-dan-2-ylamino)-5,6,7,8-tetrahydro-quinazolin-2-ylsulfanyl]-ethanol | Group I, mGluR5 | Antagonist |
| LY456236 | 4-methoxy-phenyl)-(6-methoxy-quinazolin-4-yl)-amine | Group I, mGluR1 | Antagonist |
| LY487379 | 2,2,2-trifluoro-N-[4-(2-methoxyphenoxy)phenyl]-N-(3-pyr idinylmethyl)ethanesulfonamide | Group II, mGluR2 | PAM |
| MAP4 | α -methyl-aminophosphonobutyrate | Group III | Antagonist |
| (+)-MCPG | 4-[(2S)-2-amino-1-hydroxy-1-oxopropan-2-yl]benzoic acid | Group I and II | Antagonist |
| MGS00391 | R,2R,3R,5R,6R)-2-amino-3-(3,4-dichlorobenzyloxy)-6 fluorobicyclo [3.1.0]hexane-2,6-dicarboxylic acid | Group II | Antagonist |
| MPEP | 2-methyl-6-(2-phenylethynyl)pyridine | Group I, mGluR5 | Antagonist |
| MPPG | 2-amino-2-(4-phosphonophenyl)propanoic acid | Group II | Antagonist |
| MPTP | 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine | Group I, mGluR5 | Antagonist |
| MTEP | 3-((2-methyl-1,3-thiazol-4-yl)ethynyl)pyridine hydrochloride | Group I, mGluR5 | Antagonist |
| MTPG | 2-amino-2-[4-(tetrazol-1-yl)phenyl]propanoic acid | Group II | Antagonist |
| NAAG | N-acetylaspartylglutamate | Group II, mGluR3 | Agonist |
| PHCCC | N-phenyl-7-(hydroxyimino)cyclopropa[b]chromen-1ac-arbox amide | Group II, mGluR4 | PAM |
| (R,S)-PPG | (RS)-4-phosphonophenylglycine | Group III | Agonist |
| R214127 | 1-(3,4-dihydro-2H-pyran[2,3-b]quinolin-7-yl)-2-phenyl-1-ethanone | Group I, mGluR1 | NAM |
| SIB1757 | 6-methyl-2-(phenylazo)-3-pyridinol | Group I, mGluR1 | Antagonist |
| SIB1893 | (E)-2-methyl-6-styrylpyridine | Group I, mGluR5 | Antagonist |

Abbreviations and symbols: PAM, positive allosteric modulator; NAM, negative allosteric modulator.

disorder, but not by a classical antipsychotic. Pharmacological modulation of mGluR1s using the selective antagonist EMQMCM has no effect on either MK-801-induced locomotor hyperactivity or MK-801-induced deficit in prepulse inhibition [99]. These observations further weaken a potential link between mGluR1 and psychotic symptoms.

mGluR5

Extensive evidence exists in the literature supporting a therapeutic potential of mGluR5 positive modulation for the treatment of schizophrenia. The mGluR5s are expressed in both GABAergic interneurons and pyramidal neurons in cortical and hippocampal areas, and are functionally coupled to NMDA receptors. Activation of mGluR5 leads to potentiation of NMDA receptor-mediated currents in cortical areas [100]. Thus, based on the NMDA hypofunction hypothesis of schizophrenia, positive modulation of mGluR5 might provide a viable approach to restore NMDA receptor function. In line with this assumption, mGluR5 knockout mice display locomotor hyperactivity in response to a novel environment as well as in response to MK-801, an NMDA antagonist,

sensorimotor gating deficits in a prepulse inhibition paradigm, and short-term memory deficit in a Y-maze test [101]. Studies using pharmacological modulation of mGluR5s further strengthen the involvement of mGluR5 in the pathophysiology of schizophrenia. For instance, the selective mGluR5 antagonist MPEP was found to potentiate MK-801-induced locomotor hyperactivity and stereotypies [102], indicative of a pro-psychotic effect. On the other hand, amphetamine-disrupted PPI in rats was found to be reversed by a selective positive allosteric modulator of mGluR5, compound 8q [103]. Recently, another selective mGluR5 allosteric potentiator, ADX47273, was reported to reduce conditioned avoidance responding in rats, a standard screening model for antipsychotic efficacy, as well as prevent PCP- and amphetamine-induced locomotor hyperactivity, suggestive of an antipsychotic potential [104]. The antipsychotic potential of mGluR5 positive modulators may arise from a modulatory effect on the mesolimbic dopaminergic pathway, as suggested by the decrease in basal dopamine levels in the nucleus accumbens induced by ADX47273 [105]. Additionally, mGluR5 agonism has also been shown to modulate the

Table 2. Summary of Preclinical Studies Supporting a Role of mGluRs in Schizophrenia

| Receptor Subtype | Pharmacological Tool | Effect | References |
|------------------|------------------------|---|---------------------------------|
| Group I | | | |
| mGluR1 | EMQMCM (antagonist) | ↔ MK-801-induced locomotor hyperactivity in rats; ↔ MK-801-induced deficits in prepulse inhibition in rats | Pietraszek <i>et al.</i> 2005 |
| mGluR5 | MPEP (antagonist) | ↑ MK-801-induced locomotor hyperactivity and stereotypies in rats; ↑ MK-801-induced impairments of spatial working memory and instrumental learning in rats | Homayoun <i>et al.</i> 2004 |
| | | ↓ social interaction in rats | Koros <i>et al.</i> 2007 |
| | | ↓ burst firing activity in PFC neurons in awake rats; ↑ excitatory effect of MK-801 on PFC neurons | Homayoun and Moghaddam 2006 |
| | Compound 8q (PAM) | ↓ disruption of prepulse inhibition induced by amphetamine in rats | Lindsley <i>et al.</i> 2004 |
| | ADX47273 (PAM) | ↓ conditioned avoidance responding in rats ↓ PCP- and amphetamine-induced locomotor hyperactivity in rats ↓ dopamine levels in the nucleus accumbens ↑ recall after a 48h delay in a novel object recognition task | Liu <i>et al.</i> 2008 |
| | CDPPB (PAM) | ↓ set-shifting impairment induced by NMDA receptor blockade in rats | Darrah <i>et al.</i> 2008 |
| | | ↑ bursting in PFC neurons in awake rats; ↓ excitatory effect of MK-801 on PFC neurons | Lecourtier <i>et al.</i> 2007 |
| Group II | | | |
| mGluR2 | Compound 5 (PAM) | ↓ ketamine-induced hyperactivity in rats | Pinkerton <i>et al.</i> 2005 |
| | BINA (PAM) | ↓ PCP-, but not amphetamine-induced hyperactivity in mice; ↓ PCP-induced disruption in sensorimotor gating in mice | Galici <i>et al.</i> 2006 |
| | LY487379 (PAM) | ↓ neonatal PCP-induced deficits in social discrimination in rats | Harich <i>et al.</i> 2007 |
| mGluR2/3 | LY354740 (agonist) | ↓ PCP-induced glutamate efflux in the nucleus accumbens and prefrontal cortex; ↓ PCP-induced locomotor hyperactivity in rats; ↔ dopamine levels in nucleus accumbens | Moghaddam and Adams 1998 |
| | | ↓ PCP-induced deficits in delayed alternation task in rats; ↓ neonatal PCP-induced deficits in social discrimination in rats | Harich <i>et al.</i> 2007 |
| | | ↓ performances in a delayed alternation task in rats; ↔ PCP-induced deficits in delayed alternation task in rats | Schlumberger <i>et al.</i> 2009 |
| | | ↓ firing rate and ↑ burst firing of pyramidal cells in the prefrontal cortex in awake rats; reverses ↑ firing rate and ↓ burst firing induced by MK-801 | Homayoun <i>et al.</i> 2005 |
| | LY404039 (agonist) | ↓ PCP-induced and amphetamine-induced locomotor hyperactivity; ↓ conditioned avoidance responding in rats | Rorick-Kehn <i>et al.</i> 2007 |
| | | ↓ PCP-induced and amphetamine-induced locomotor hyperactivity abolished in mGluR2, but not mGluR3 knockout mice | Fell <i>et al.</i> 2008 |
| Group III | | | |
| mGluR7 | AMN082 (agonist) | Prevents ↓ EPSC frequency induced by L-AP4 in dopaminergic neurons | de Rover <i>et al.</i> 2008 |
| | | ↔ basal or cocaine-induced increase in dopamine levels in the nucleus accumbens; ↔ basal or cocaine-induced locomotor hyperactivity in rats; ↓ cocaine-induced decrease in GABA levels in the ventral pallidum | Li <i>et al.</i> 2009 |
| mGluR8 | (S)-3,4-DCPG (agonist) | ↓ amphetamine-induced locomotor hyperactivity in mice when administered i.c.v.; trend to ↓ spontaneous locomotor activity | Robbins <i>et al.</i> 2007 |

Abbreviations and symbols: ↔, unchanged; ↑, increased; ↓, decreased; MK-801, dizocilpine maleate; PAM, positive allosteric modulator; PFC, prefrontal cortex; PCP, phencyclidine; EPSC, excitatory postsynaptic currents; i.c.v., intracerebroventricular injection.

ventral striatopallidal GABAergic pathway [102], which is a common target for antipsychotic drugs.

With respect to modulation of cognitive function, blockade of mGluR5s by MPEP has been reported to potentiate MK-801-induced impairments of spatial working memory and instrumental learning in rats [102]. Conversely, allosteric potentiation of mGluR5s using ADX47273 was found to enhance recall after a 48h delay in a novel object recognition task [104]. A modulatory role of mGluR5s on cognitive function is also indicated by the finding that CDPPB, another selective mGluR5 positive allosteric modulator, reduces the characteristic set-shifting impairment induced by NMDA receptor blockade in rats [106]. The modulatory effect of mGluR5s on cognitive processing has been hypothesized to rely on changes in neuronal activity in the prefrontal cortex. In fact, inhibition of mGluR5s by MPEP was reported to decrease burst firing activity of cortical neurons in awake rats and potentiated the increase in firing rate induced by NMDA receptor blockade [107]. Conversely, the mGluR5 positive allosteric modulator CDPPB was shown to increase bursting in PFC neurons in awake rats, and to prevent the robust excitatory effect of NMDA receptor blockade by MK-801 [108]. Taken together, these studies indicate that mGluR5 positive modulation may be effective in ameliorating cognitive dysfunction induced by NMDA hypofunction by restoring the function of prefrontal cortical neurons, and thereby be of clinical usefulness for the treatment of cognitive symptoms associated with schizophrenia.

Besides an antipsychotic and procognitive potential of mGluR5 positive modulation, the selective mGluR5 antagonist, MPEP, has interestingly been reported to induce social interaction deficits in rats, suggesting a potential link between mGluR5 and social deficits characteristic of negative symptoms in schizophrenia [109].

Besides the above-mentioned preclinical evidence, an involvement of the GRM5 gene in schizophrenia has been suggested in a genetic study showing a significant difference in allele frequency distribution between schizophrenics and controls [110]. An increased neuronal mGluR5 expression has also been reported in the pyramidal cell layer in the prefrontal cortex of schizophrenics [111].

In summary, mGluR5 positive modulation may be a promising therapeutic strategy for the treatment of positive and cognitive symptoms associated with schizophrenia. Moreover, a potential benefit on negative symptoms, and particularly on social withdrawal, may also be achieved, although additional studies with positive modulators would be required to draw any conclusion in this regard.

Selective Group II Modulation

Stimulation of group II receptors inhibits synaptic release of glutamate presynaptically [112]. A considerable amount of evidence supports a therapeutic potential of group II receptor agonism for the treatment of schizophrenia. Recently, clinical proof of concept has been obtained in schizophrenic patients [113], thereby identifying mGluR2/3 modulation as one of the most attractive strategies amongst all mGluRs for the treatment of schizophrenia. The first preclinical evidence

was obtained using the highly selective agonist of group II mGluR, LY354740. In these early studies, activation of mGluR2/3 with LY354740 was found to block phencyclidine-induced glutamate efflux in the nucleus accumbens and prefrontal cortex as well as phencyclidine-induced locomotor hyperactivity in rats [114]. Interestingly, dopaminergic neurotransmission was not affected by LY354740. Another selective mGluR2/3 receptor agonist with improved bioavailability, LY404039, was also reported to produce similar antipsychotic effects in psychostimulant-induced locomotor hyperactivity as well as in a conditioned avoidance response paradigm without producing motor impairment [115]. Recently, selective positive allosteric modulators of mGluR2 have been identified, and have been shown to reduce NMDA blockade-induced hyperactivity in rats [116,117]. Another selective mGluR2 positive allosteric modulator, BINA, was found to block locomotor hyperactivity induced by phencyclidine, but not by amphetamine, as well as prevent phencyclidine-induced disruption in sensorimotor gating in mice [118]. In line with this assumption, the attenuation of phencyclidine- and amphetamine-induced locomotor hyperactivity produced by the mGluR2/3 agonists, LY379268 and LY404039, was completely abolished in mGluR2 knockout mice whereas it was not affected in mGluR3 knockout mice [119,120]. Taken together, these findings suggest that targeting mGluR2s, rather than mGluR3s, is a relevant mechanism for the treatment of schizophrenia.

Few preclinical studies have suggested a procognitive potential of mGluR2/3 agonism. For instance, LY354740 reversed cognitive deficits induced by phencyclidine in a delayed alternation task in rats [114]. Both mGluR2/3 agonism using LY354740 and selective allosteric potentiation of mGluR2 using LY487379 was reported to attenuate deficits in social discrimination induced by neonatal phencyclidine in rats, an effect attributed to a selective attention deficit, while no effect of either compound was found on the total time spent in social interaction [121]. However, others have reported that mGluR2/3 agonism produced no effect or impairment in cognitive function. For instance, LY354740 was found to impair performance in a delayed alternation task in rats, while it had no effect on phencyclidine-induced deficits in the same task [122]. The reason for these discrepancies has not been clearly established.

Neurons in the prefrontal cortex are assumed to play a critical role in the cognitive impairing and psychotomimetic effects of NMDA receptor antagonists. In this respect, it has been hypothesized that the putative antipsychotic and procognitive effects of mGluR2/3 agonism would attenuate the disruption of neuronal activity in the prefrontal cortex produced by NMDA hypofunction. In fact, stimulation of mGluR2/3 by LY354740 was reported to decrease the firing rate and increase burst firing of pyramidal cells in the prefrontal cortex in awake rats, while it was able to reverse the increased firing rate and decreased burst firing induced by the NMDA receptor antagonist MK-801 [123].

Ketamine is a non-competitive NMDA antagonist widely used in clinical studies as it induces schizophrenia-like symptoms, including hallucinations, delusions, and cognitive impairment [95]. For instance, it has been shown to disrupt

working memory in humans at subanesthetic doses, an effect attributed to NMDA receptor blockade in the prefrontal cortex. Interestingly, LY354740 was reported to attenuate working memory deficits induced by ketamine administration in healthy volunteers [124]. A study in post-mortem brain tissues from schizophrenic patients using an antibody directed against mGluR2 and 3 also revealed an increased mGluR2/3 expression in the prefrontal cortex [97]. It was shown recently that mGluR2 expression, rather than mGluR3, was increased in the prefrontal cortex of schizophrenics [125]. Interestingly, an oral prodrug of LY404039 (LY2140023) was evaluated in schizophrenic patients. Treated patients showed significant improvement in both positive and negative symptoms compared to placebo [113].

In summary, stimulation of mGluR2/3, and most likely selectively mGluR2, represents an attractive therapeutic strategy for the treatment of schizophrenia. More precisely, beneficial effects have been observed on both positive and negative symptoms in schizophrenic patients [113]. In addition, a beneficial effect on working memory has also been reported in humans [124], further suggesting that improvement of cognitive symptoms associated to schizophrenia may also be achieved. However, this clinical endpoint would need to be specially considered in future schizophrenia trials.

Selective Group III Modulation

mGluR7

Few genetic studies have suggested a role of mGluR7 in the pathophysiology of schizophrenia. GRM7 analysis in a population of Japanese schizophrenics identified a single nucleotide polymorphism (SNP) with lower promoter activity, suggesting that lower expression of mGluR7 may increase the risk of developing schizophrenia [126]. Another study from a Japanese sample has also identified highly significant association of schizophrenia with the two other SNPs in GRM7 [127]. A mutation in PICK-1 protein, which is crucial for clustering of mGluR7 at presynaptic release sites, has also been associated with schizophrenia [128].

In addition to these genetic findings, mGluR7 have been shown to modulate the mesolimbic dopaminergic system and the ventral striatopallidal feedback loop, suggesting a potential link to positive symptoms of schizophrenia. Interestingly, the selective mGluR7 antagonist, AMN082, was reported to decrease the frequency of mEPSCs in dopaminergic neurons of the ventral tegmental area *in vitro* [129]. Since glutamatergic innervation of the ventral tegmental area plays a critical role in burst firing of dopaminergic neurons, the ability of mGluR7 to modulate these excitatory inputs may be indicative of an antipsychotic potential of mGluR7 agonism. However, AMN082 did not affect basal or cocaine-induced increase in dopamine levels in the nucleus accumbens, while it decreased GABA and increased glutamate levels [130,131]. The effect of AMN082 on glutamate levels was further shown to be partly mediated through reduction of GABA levels. In line with the absence of modulation of dopamine levels by mGluR7 activation, AMN082 did not affect basal or cocaine-induced locomotor hyperactivity in rats [131]. However, AMN082 was found to block cocaine-induced decrease in GABA levels in the ventral pallidum. In

summary, mGluR7 agonism does not seem to affect dopaminergic neurotransmission in the nucleus accumbens, but is able to modulate the ventral striatopallidal pathway in condition of excessive dopaminergic tone in the nucleus accumbens, which may be relevant to the treatment of positive symptoms.

Several studies have also suggested that mGluR7 may regulate cognitive function. For instance, spatial and working memory has been investigated in mGluR7 knockout mice [132]. In a Morris water maze task, mGluR7 knockout mice show a significant delay in acquiring the location of the hidden platform, as well as in recall during the probe trial. In a working memory version of the Morris water maze, mGluR7 knockout mice were impaired and consistently slower to solve the matching-to-position task, possibly due to impairment in short-term memory. In the consecutive extinction trials, mGluR7 knockout mice were also delayed to adopt a new search strategy. Taken together, these data suggest that mGluR7 knockout mice have impaired reference memory acquisition and spatial working memory, and a dysfunctional glutamatergic signalling particularly in the hippocampus and prefrontal cortex where mGluR7 are expressed has been hypothesized to cause these deficits. Performances in complex working memory tasks such as 8-arm radial maze task were also impaired in mGluR7 knockout mice [133]. Interestingly, the working memory deficit was associated with an increased hippocampal theta power while performing the task, which was suggested to reflect a lack of modulation of local inhibition, in turn leading to decreased neuronal firing threshold and altered spike timing [134]. At the cellular level, mGluR7 knockout mice were reported to exhibit deficits in short-term, but not long-term potentiation in the hippocampus [135], findings in agreement with the hypothesis that short-term potentiation represents the cellular substrate for short-term memory and critical for working memory performances.

Taken together, these findings indicate that mGluR7 positive modulation may represent a new therapeutic strategy potentially beneficial for the treatment of positive as well as cognitive symptoms. In addition, since mGluR7s are also highly expressed in the amygdala and have been implicated in anxiety (see "Major Depression Disorder and Anxiety"), a potential effect on negative symptoms might also be achieved. However, since the lines of evidence rely on the use of knockout mice and a single pharmacological tool, additional studies using other selective positive or negative modulators of mGluR7 would be needed.

mGluR8

In a genetic study, one susceptibility locus for schizophrenia was identified within the GRM8 region in Japanese [136], suggesting that mGluR8 may have a therapeutic potential. In this respect, the antipsychotic potential of mGluR8 modulation was investigated in a preclinical study. While the selective mGluR8 agonist (S)-3,4-DCPG was devoid of any effect on PCP- or amphetamine-induced locomotor hyperactivity in rats when administered systemically, intracerebroventricular (S)-3,4-DCPG administration prevented amphetamine-induced locomotor hyperactivity in mice, although a trend to reduce spontaneous locomotor activity was also ob-

served [137]. In addition, mGluR8 knockout mice did not show any change in locomotor activity or deficit in sensorimotor gating as assessed in a prepulse inhibition of the startle response paradigm, further ruling out a possible antipsychotic effect of mGluR8 modulation. However, the same study reported that mGluR8 knockout mice exhibit an anxiogenic phenotype as assessed in an open field as well as an elevated plus maze, suggesting that mGluR8 modulation might be relevant for negative symptoms of schizophrenia.

ADDICTION

Drug addiction is a chronic illness arising as a consequence of frequent drug taking. It involves the progression from acute drug use to the development of drug-seeking behavior, the vulnerability to relapse, and the decreased, slowed ability to respond to naturally rewarding stimuli. Several lines of evidence suggest that glutamate neurotransmission plays a key role in the processes leading to drug addiction and relapse. In particular, hyperglutamatergic response in the corticostriatal pathway in response to drug-related cues and leading to drug-seeking behaviour is believed to be a core component underlying relapse.

Glutamate homeostasis refers to the regulation of extracellular glutamate levels in the synaptic and extrasynaptic spaces and is highly dependent on the balance between glial and synaptic glutamate release and elimination. Impairment in glutamate homeostasis leads in turn to altered synaptic activity and plasticity by affecting glutamate levels available for stimulation of ionotropic and metabotropic glutamate receptors. Preclinical studies have shown that chronic administration of several drug of abuse, including cocaine, heroin and nicotine, induced neuroadaptive changes at the glutamatergic synapse. These changes include reduced basal extrasynaptic glutamate levels, presumably as a consequence of reduced expression of the glial cystine-glutamate exchanger, in turn leading to down-regulation of presynaptic mGluR2/3 [138,139]. Since presynaptic mGlu2/3 regulates synaptic glutamate release, its down-regulation has been suggested to result in enhanced synaptic glutamate release during cue-, stress- or drug-induced reinstatement. This increased synaptic glutamate transmission results in postsynaptic adaptations, including up-regulation of AMPA Glu1 receptors (GRIA1), and a compensatory down-regulation of mGluR1/5, ultimately leading to neuroplasticity. These glutamate-induced neuroplastic changes have been suggested to be the common factor in relapse for many types of drugs. In addition, preclinical evidence supports the occurrence of similar glutamate dependent-neuroplastic alterations in other key brain areas such as the ventral tegmental area and the amygdala [140,141] (Table 3).

Selective Group I Modulation

mGluR1

Only a few preclinical studies have investigated the effect of selective mGluR1 negative allosteric modulators on drug-related behaviours. For instance, the mGluR1 antagonist, CPCCPOEt, was shown to reduce measures of ethanol reward in an operant self-administration paradigm, block the expression of ethanol-induced place conditioning, as well as

ethanol consumption under 24-h free-access conditions in alcohol-preferring mice [142]. At the neurochemical level, CPCCPOEt was found to inhibit dopamine and glutamate increases induced by ethanol administration in the nucleus accumbens, while it facilitated GABA-induced release, suggesting that mGluR1a blockade may regulate the rewarding properties of ethanol by facilitating its inhibitory effect in the nucleus accumbens. In a cocaine sensitization paradigm, EMQMCM, another mGluR1 negative allosteric modulator, was reported to reduce the expression of the sensitized ambulatory motor activity of cocaine-experienced rats acutely challenged with cocaine [143,144]. However, the same dose range of EMQMCM also reduced ambulatory horizontal activity in control animals, which may confound the apparent reduction observed in sensitized animals. Interestingly, another mGluR1 antagonist, JNJ 16259685, infused locally in the dorsal hippocampus was recently reported to block drug context-induced reinstatement of cocaine-seeking behaviour in rats, while it did not alter locomotor activity or food-reinforced instrumental behavior [145]. In another report, the role of mGluR1 in the reinstatement of nicotine-seeking behaviour in rats was investigated [146]. In this study, blockade of mGluR1 by EMQMCM (5 and 10 mg/kg) was found to prevent both cue-induced and nicotine priming-induced reinstatement of nicotine-seeking behaviour. However, the highest dose of EMQMCM (10 mg/kg) also inhibited cue-induced reinstatement of food-seeking behaviour. This observation is in contrast to that reported by Xie *et al.* [145] showing that mGluR1 blockade selectively in the hippocampus did not affect food-reinforced instrumental behaviour, but may indicate that non regional selective mGluR1 blockade may not only inhibit the motivational value of drugs, but also that of natural reinforcers. In another study, blockade of mGluR1 by EMQMCM (5, 10 and 20 mg/kg) was reported to prevent the expression of sensitization to the locomotor effect of morphine, but did not affect the expression of withdrawal syndrome in mice [147], further suggesting that mGluR1 blockade may be of potential benefit in the treatment of opiate seeking behaviour.

In conclusion, preclinical evidence suggests that mGluR1 blockade may represent a potential pharmacological mechanism relevant for the treatment of drug addiction, including alcohol, cocaine, nicotine, and opiates. However, one concern might be that such a mechanism would also affect the motivational value of natural reinforcers.

mGluR5

Numerous preclinical studies have investigated the role of mGluR5 in drug-related behaviours. For instance, the mGluR5 antagonist, MPEP, was shown to reduce measures of ethanol reward in an operant self-administration paradigm, block the expression of ethanol-induced place conditioning, as well as ethanol consumption under 24-h free-access conditions in alcohol-preferring mice [142]. Along the same line, mGluR5 null mutant mice show decreased alcohol consumption [148]. In addition, MPEP was reported to block cue-induced reinstatement of alcohol-seeking behavior in alcohol-preferring rats [149], further strengthening a potential benefit of mGluR5 blockade in alcohol addiction. With respect to other drugs of abuse, MPEP has been shown

Table 3. Summary of Preclinical Studies Supporting a Role of mGluRs in Addiction

| Receptor Subtype | Pharmacological Tool | Effect | References |
|------------------|--------------------------|---|--|
| Group I | | | |
| mGluR1 | CPCCPOEt (antagonist) | ↓ Ethanol reward, ethanol-induced CPP and ethanol consumption in mice | Lominac <i>et al.</i> 2006 |
| | EMQMCM (antagonist) | ↓ Expression of cocaine sensitization | Dravolina <i>et al.</i> 2006; Kotlinska and Bochenski 2009 |
| | | ↓ Cue-induced and nicotine priming-induced reinstatement of nicotine-seeking behaviour in rats | Dravolina <i>et al.</i> 2006 |
| | | ↓ Expression of sensitization to morphine | Kotlinska and Bochenski 2007 |
| | JNJ16259685 (antagonist) | ↓ Drug context-induced reinstatement of cocaine-seeking behaviour in rats when infused intra-hippocampus | Xie <i>et al.</i> 2010 |
| mGluR5 | MPEP (antagonist) | ↓ Ethanol reward, ethanol-induced CPP and ethanol consumption in mice | Lominac <i>et al.</i> 2006 |
| | | ↓ Cue-induced reinstatement of alcohol-seeking behaviour in alcohol-preferring rats | Schroeder <i>et al.</i> 2008 |
| | | ↓ Expression of sensitization to nicotine and nicotine-induced drug-seeking behaviour in rats | Tessari <i>et al.</i> 2004 |
| | | ↓ Cocaine self-administration cocaine-induced reinstatement of drug seeking in squirrel monkeys | Lee <i>et al.</i> 2005; Platt <i>et al.</i> 2008 |
| | | ↓ Expression of cocaine sensitization in rats | Tessari <i>et al.</i> 2004 |
| | MTEP (antagonist) | ↔ Expression of cocaine sensitization in rats | Dravolina <i>et al.</i> 2006 |
| | | ↓ Reinforcing effects of methamphetamine and cue- and drug-induced reinstatement of methamphetamine-seeking behaviour in rats | Gass <i>et al.</i> 2009 |
| | | ↓ Expression of morphine sensitization and naloxone-induced symptoms of morphine withdrawal in morphine-dependent mice | Kotlinska and Bochenski 2007; Palucha <i>et al.</i> 2004a |
| Group II | | | |
| mGluR2/3 | LY354740 (agonist) | ↓ Naloxone-induced symptoms of morphine withdrawal in morphine-dependent mice | Klodzinska <i>et al.</i> 1999) |
| | LY379268 (agonist) | ↓ Ethanol self-administration and cue-induced reinstatement of ethanol seeking in rats; ↓ locomotor activity at same doses | Bäckström and Hyytiä 2005 |
| | | ↓ Drug-induced reinstatement of cocaine seeking in rats | Peters and Kalivas 2006 |
| | | ↓ Nicotine self-administration and drug-induced reinstatement of nicotine seeking in rats | Liechti <i>et al.</i> 2007 |
| | | ↓ Stress- and drug context-induced reinstatement of ethanol seeking in rats | Zhao <i>et al.</i> 2006 |
| | | ↓ Cue-induced reinstatement of heroin seeking in rats after acute treatment; tolerance observed after 14 days of treatment | Bossert <i>et al.</i> 2006 |
| Group III | | | |
| mGluR7 | AMN082 (agonist) | ↓ Rewarding effects of cocaine | Li <i>et al.</i> 2009 |
| mGluR8 | (S)- 3,4-DCPG (agonist) | ↓ Ethanol self-administration and cue-induced reinstatement of ethanol seeking; ↓ Locomotor activity at same doses | Bäckström and Hyytiä 2005 |

Abbreviations and symbols: ↔, unchanged; ↑, increased; ↓, decreased; CPP, conditioned place preference.

to block the expression of sensitization to the locomotor effect of nicotine, as well as reduce nicotine-induced drug-seeking behaviour in a model of nicotine-triggered relapse to nicotine-seeking in rats [150]. Blockade of mGluR5 has also been reported to affect cocaine-induced behaviours. In particular, both acute and chronic blockade of mGluR5 by MPEP attenuate cocaine self-administration in squirrel monkeys [151,152]. Acute MPEP treatment has also been shown to prevent cocaine-induced reinstatement of drug seeking in squirrel monkeys [151], as well as block the expression of cocaine sensitization in rats [150]. In contrast to the latter findings, another mGluR5 antagonist, MTEP, was devoid of effect on the expression of cocaine sensitization in rats [143]. It is generally agreed that MTEP exhibits superior selectivity for mGluR5 compared to MPEP, with fewer off-targets identified compared to the former [153]. Of particular interest is the observation that MPEP is able to interact functionally with NMDA receptors in rat [154]. Therefore, the effects of MPEP should be interpreted cautiously, especially when considering behaviours expected to be affected by NMDA receptors. Another study investigating the role of mGluR5 in methamphetamine-induced behaviours revealed that MTEP dose-dependently reduced the reinforcing effects of methamphetamine and attenuated cue- and drug-induced reinstatement of methamphetamine-seeking behaviour in rats [155]. Blockade of mGluR5 with MTEP was further found to inhibit the expression of morphine sensitization [147] as well as naloxone-induced symptoms of morphine withdrawal in morphine-dependent mice [147,156]. Interestingly, blockade of mGluR5 has been shown to be devoid of effect on food-motivated instrumental behaviour and cue-induced reinstatement of food-seeking [152,155]. Taken together, these findings suggest that selective blockade of mGluR5 might represent an interesting mechanism for the treatment of addictive behaviours to various drug classes. With contrast to mGluR1 blockade, no effect on the motivational value of natural reinforcers would be expected based on the available preclinical data.

Selective Group II Modulation

Presynaptic mGluR2/3s act as autoreceptors, their stimulation attenuating synaptic glutamate release. Agonising mGluR2/3 has been suggested to be of potential interest for the treatment of drug addiction. The recent discovery of selective mGluR2/3 agonists has offered the opportunity to test this hypothesis in preclinical models. LY354740 has been shown to block naloxone-induced symptoms of morphine withdrawal in morphine-dependent mice [157]. Another mGluR2/3 agonist, LY379268, was reported to attenuate ethanol self-administration and cue-induced reinstatement of ethanol seeking, although at doses that also reduced spontaneous locomotor activity [158]. The same mGluR2/3 agonist was found to block drug-induced reinstatement of cocaine seeking in rats [159]. In the latter study, LY379268 was also found to attenuate reinstatement of food seeking, although at doses 3 times higher than those effective in reinstatement of cocaine seeking. These nonspecific inhibitory effects on responding for natural rewards at higher doses of LY379268 have been reported by others [160-162]. Whether these effects are a general consequence of stimulating mGluR2/3

or whether they are compound specific is currently unknown. Stimulation of mGluR2/3 by LY379268 was further found to prevent nicotine self-administration and drug-induced reinstatement of nicotine seeking in rats [162], as well as stress- and drug context-induced reinstatement of ethanol seeking in rats [163]. It was also reported to be effective in preventing cue-induced reinstatement of heroin seeking [161]. However, a tolerance to the effect of LY379268 on nicotine self-administration was observed after daily administration for 14 days [162].

Studies using intracerebral administration of mGluR2/3 agonists have also identified critical brain areas involved in the modulation of drug seeking and relapse-like behaviours by mGluR2/3. For instance, LY379268 administered into the ventral tegmental area or the nucleus accumbens blocked reinstatement of heroin and cocaine seeking [159,161,164] and nicotine self-administration [162], further supporting the importance of mGluR2/3 located at excitatory synapses in the nucleus accumbens as well as on mesolimbic dopaminergic neurons in the ventral tegmental area.

In summary, mGluR2/3 agonism might represent a novel approach for the treatment of addiction to various drugs, including ethanol, cocaine, opiates and nicotine. A potential concern may arise from the observation that mGluR2/3 agonism induced an inhibitory effect on the motivational value of natural rewards in animals and may need further clarification.

Selective Group III Modulation

Similarly to metabotropic receptors of group II, subtypes belonging to group III are also located presynaptically, where they act as autoreceptors, regulating synaptic glutamate release. One major difference however between the two groups is their subcellular localization. While group II receptors are mostly perisynaptic, group III receptors are mainly expressed within the synapse. Evidence linking mGluRs belonging to group III to drug addiction is only limited, including a potential role of mGluR7 and mGluR8 subtypes, but may still be of possible value.

Glutamate receptor metabotropic 7 (Grm7) has been identified as a candidate risk gene linked to alcohol addiction, as suggested by the observation that Grm7 mRNA expression levels in different mouse strains were inversely correlated to their alcohol consumption in a preference drinking behaviour paradigm [165]. In a recent report, the selective mGluR7 agonist, AMN082, was found to inhibit the rewarding effects of cocaine, while the rewarding effects of a natural reinforcer were not affected [131]. AMN082 injection in the nucleus accumbens or the ventral pallidum mimicked the effects observed after systemic administration. In addition, activation of mGluR7 by AMN082 was found to prevent cocaine-induced inhibition of GABA release in the ventral pallidum. Taken together, these data suggest that mGluR7 modulate cocaine reward by regulating GABAergic transmission in the ventral striatopallidal pathway. The recent availability of selective ligands for mGluR7 will hopefully help to further evaluate the therapeutic potential of mGluR7 modulation in the treatment of addictive behaviours.

A genome-wide association approach has identified *Grm8*, the gene encoding mGluR8, as having a strong association with heroin addiction [166]. Only few preclinical studies have investigated the role of mGluR8 in drug-induced behaviours. Amongst these, the reported selective mGluR8 agonist, (S)-3,4-DCPG, was demonstrated to attenuate ethanol self-administration and cue-induced reinstatement of ethanol seeking, although at doses that also reduced spontaneous locomotor activity [158]. In conclusion, there is only a very weak biological rationale supporting a potential interest of mGluR8 agonism for the treatment of drug addiction.

MAJOR DEPRESSION DISORDER AND ANXIETY

Major depressive disorder (MDD) and anxiety disorders are severe, disabling diseases that are highly prevalent and associated with negative impact on medical health, life quality and productivity [167-170]. Preclinical and clinical evidence suggest that MDD arises due to a decreased availability of serotonin and norepinephrine, since tricyclic antidepressants such as imipramine block the transporters for serotonin and norepinephrine leading to increased levels of serotonin and norepinephrine in the cerebrospinal fluid [171,172]. Furthermore, anxiety is believed to result mainly from a hyperactive state of the serotonergic system, where especially dysfunction of the 5-HT_{1A} receptors is of significance [173]. The introduction of selective serotonin reuptake inhibitors (SSRIs) and combined serotonin and norepinephrine reuptake inhibitors (SNRIs) into clinical practice led to an improvement in the treatment of MDD and anxiety disorders by producing therapeutic benefit without the serious side-effects associated with the older tricyclic antidepressants [174]. Although SSRIs and SNRIs are effective, a meaningful therapeutic improvement is only apparent after several weeks of treatment [175]. Furthermore, many depressed patients respond only partially and a substantial proportion of patients fail to respond at all to first-line treatment [168]. Moreover, in those patients that do respond, side-effects such as sexual dysfunction, sleep disturbances and gastrointestinal disturbances have been reported [168]. During the past years various data support the idea that compounds working through multitarget mechanisms will have a better effect on both cardinal and comorbid symptoms of depression than selective compounds [176]. The above studies emphasize the need for improved treatment against MDD working through new mechanisms of action, either as monotherapy or add-on therapy.

Interestingly, the NMDA receptor antagonist, ketamine, is effective acutely in treatment-resistant depression [177], suggesting that improved treatment for MDD may be possible by targeting the glutamatergic neurotransmission (Table 4).

Selective Group I Modulation

Group I mGluR activation in rat hippocampus by DHPG was decreased after subchronic treatment with the antidepressant treatments, electroconvulsant stimulation and imipramine [178,179]. The immunoreactivity of both hippocampal mGluR1 and mGluR5 was upregulated following subchronic electroconvulsant stimulation [180]. The observed receptor upregulations may reflect a compensatory mechanism

caused by the receptor subsensitivity provoked the antidepressant treatment.

Although effects of mGluR1 antagonists have not been investigated as frequently as mGluR5 antagonists for anxiolytic and antidepressant effects, recent studies reported that the selective mGluR1 antagonists, JNJ16259685 and AIDA, exerted anxiolytic-like effect in the rat lick suppression test [181] and blocked anxiogenic behaviour in the rat light-dark test and open-field test [182], respectively.

mGluR5

The potential use of mGluR5 negative modulators for the treatment of anxiety and depression has been broadly investigated. The mGluR5 antagonist, MPEP was shown to exert anxiolytic-like effect in several anxiety-like behaviour tests, including elevated-plus maze, social exploration, fear-potentiated startle, Vogel-conflict and light-dark box test [182-187]. In addition, MTEP exerted anxiolytic-like effects in contextual fear conditioning following acute or subchronic treatment, indicating that tolerance does not develop to the anxiolytic effect of MTEP [188]. Furthermore, it has been suggested that mGluR5 antagonism exerts its anxiolytic effect in the conditioned emotional response paradigm by a mechanism different from that of diazepam [189].

In the chronic mild stress model, a validated model to screen for antidepressant activity, increased expression of hippocampal mGluR5 has been reported [190]. It has been demonstrated that the mGluR5 antagonists, MPEP and MTEP, shortened the immobility time in the tail-suspension test and forced-swim test in mice [186,191-193], indicative of an antidepressant-like effect. Both MPEP and MTEP displayed antidepressant-like effects in an animal model of depression, the olfactory bulbectomy model [191,194]. Interestingly, it has been reported that mGluR5 knockout mice display an antidepressant-like behavioural phenotype [193]. In these mice imipramine, but not MPEP, exerted an antidepressant effect [193]. In the same study a synergy of MPEP and imipramine was observed in the mouse forced-swim test. The above studies indicate that mGluR1 and mGluR5 antagonists may have therapeutic potential in the treatment of MDD and anxiety disorders. Moreover, the antidepressant efficacy of tricyclic antidepressants and SSRIs might be enhanced by concomitant treatment with mGluR5 negative modulators.

Selective Group II Modulation

A human postmortem study showed increased levels of mGluR2/3 in samples from the prefrontal cortex [195]. In a Japanese population an association between *GRM3* (group II mGluR gene) and MDD has been reported [196]. Variations in *GRM3* have been found to affect prefrontal glutamatergic neurotransmission and cognitive functions [197]. Preliminary evidence suggests that hippocampal levels of mGluR2/3 are decreased in the olfactory bulbectomy model [198] and in depressed flinders sensitive line rats [199]. However, the functional consequences of these alterations are unknown.

An association between classical antidepressants and mGluR2/3 compounds might exist. Chronic imipramine treatment was observed to reduce mGluR2/3 agonist-

Table 4. Summary of Preclinical Studies Supporting a Role of mGluRs in Major Depression Disorder and Anxiety

| Receptor Subtype | Pharmacological Tool | Effect | References |
|------------------|----------------------------------|--|--|
| Group I | | | |
| mGluR1 | JNJ16259685 (antagonist) | ↓ Anxiety-like behaviour in the rat lick suppression test | Steckler <i>et al.</i> 2005 |
| | AIDA (antagonist) | ↓ Anxiogenic behaviour in the rat light-dark test and open-field test | Mikulecka and Mares 2009 |
| mGluR5 | MPEP (antagonist) | ↓ Anxiogenic behaviour in elevated-plus maze, social exploration, fear-potentiated startle, Vogel-conflict and light-dark box test | Ballard <i>et al.</i> 2005; Spooren <i>et al.</i> 2000; Spooren and Gasparini 2004; Tatarczynska <i>et al.</i> 2001; Mikulecka and Mares 2009; Spanka <i>et al.</i> 2010 |
| | MTEP (antagonist) | ↓ Anxiogenic behaviour contextual fear conditioning | Gravius <i>et al.</i> 2008 |
| | MPEP & MTEP (antagonists) | ↓ Immobility time in the tail-suspension test and forced-swim test | Tatarczynska <i>et al.</i> 2001; Palucha <i>et al.</i> 2005; Belozertseva <i>et al.</i> 2007; Li <i>et al.</i> 2006 |
| | | Antidepressant-like effects in the olfactory bulbectomy model | Palucha <i>et al.</i> 2005; Pilc <i>et al.</i> 2002 |
| Group II | | | |
| mGluR2/3 | LY341495 (antagonist) | ↑ Firing rate of serotonergic dorsal raphe neurones | Kawashima <i>et al.</i> 2005 |
| | MGS0039 (antagonist) | ↑ Firing rate of serotonergic dorsal raphe neurones | Kawashima <i>et al.</i> 2005 |
| | | ↑ Extracellular levels of serotonin in the rat prefrontal cortex | Karasawa <i>et al.</i> 2005 |
| | LY341495 & MGS0039 (antagonists) | ↓ Immobility and in the tail-suspension test and forced-swim test | Witkin <i>et al.</i> 2007; Chaki <i>et al.</i> 2004 |
| | LY341495 (antagonist) | Effective in the marble burying test ↔ Anxiogenic behaviour in elevated plus maze and stress-induced hyperthermia tests | Bespalov <i>et al.</i> 2008 |
| | MGS0039 (antagonist) | ↓ Anxiogenic behaviour in the conditioned fear model | Yoshimizu <i>et al.</i> 2006 |
| Group III | | | |
| | ACPT-I (agonist) | ↓ Immobility time in the forced-swim test | Tatarczynska <i>et al.</i> 2002; Palucha <i>et al.</i> 2004c; Klak <i>et al.</i> 2007 |
| | | ↓ Anxiogenic behaviour in the stress-induced hyperthermia, elevated plus-maze tests and in the Vogel test | Stachowicz <i>et al.</i> 2009 |
| mGluR4 | PHCCC (PAM) | ↓ Immobility time in the forced-swim test | Klak <i>et al.</i> 2007 |
| mGluR7 | AMN082 (agonist) | ↓ Immobility time in the forced swim test and tail suspension test | Palucha <i>et al.</i> 2007 |
| mGluR8 | RSPPG (agonist) | ↓ Immobility time in the forced-swim test | Palucha <i>et al.</i> 2004b |

Abbreviations and symbols: ↔, unchanged; ↑, increased; ↓, decreased; PAM, positive allosteric modulator.

mediated inhibition of forskolin-stimulated cAMP production in rat hippocampus [200]. Moreover, administration of the selective mGluR2/3 antagonists, LY341495 and MGS0039, increased the firing rate of serotonergic dorsal raphe neurones [201], and administration of MGS0039 increased extracellular levels of serotonin in the rat prefrontal cortex [202]. Chronic treatment with MGS0039, furthermore, increased hippocampal neurogenesis [203], a mechanism demonstrated for some of the current antidepressants [204].

Both LY341495 and MGS0039 reduced immobility and in the tail-suspension test and forced-swim test [205,206]. In

the forced-swim test, the two compounds increased swimming behaviour without affecting climbing behaviour [206]. Those antidepressants that increase serotonergic neurotransmission predominantly increase swimming behaviour whereas those that increase catecholaminergic neurotransmission increase climbing behaviour [207]. Thus, mGluR2/3 compounds may indirectly interact with the serotonergic neurotransmission. mGluR2/3 knockout mice displayed antidepressant-like behaviour, i.e. increased mobility, in the forced-swim test but not in the tail suspension test [208]. It has been reported that following administration of imipramine in combination with LY-341495 neuroadaptations

to imipramine occurred faster than with imipramine alone [209], indicating that mGluR2/3 antagonism may shorten the lag time required to obtain full therapeutic improvement of current antidepressant, which is only apparent after several weeks of treatment [174].

LY341495 was effective in the marble burying test in mice [210]. However, in the same study LY341495 had no effects in the elevated plus maze and stress-induced hyperthermia tests in mice or on punished drinking. Thus, the behavioural profile of an mGluR2/3 antagonist seems to be different from that of conventional anxiolytic and antidepressant drugs. However, MGS0039 was reported to show anxiolytic-like effects in the conditioned fear model [211], indicating that mGluR2/3 antagonists may be beneficial in the treatment of some anxiety disorders.

Despite the fact that mGluR2/3 antagonists affect serotonergic activity the above studies suggest that mGluR2/3 antagonists work through different mechanism of actions compared to those of SSRIs and SNRIs and may provide beneficial effects in the treatment of MDD and anxiety disorders.

Selective Group III Modulation

Chronic treatment with citalopram, but not imipramine, decreased immunoreactivity of mGluR7 but not mGluR4 in the rat hippocampus and cerebral cortex [212]. However, the same study reported that chronic citalopram or imipramine did not change the effect of ACPT-1, a group III mGluR agonist, on forskolin-stimulated cAMP production. The lack of effect of chronic imipramine on mGluR4 expression in the rat brain has also been observed by another group [200]. Further studies are needed to clarify the effect of antidepressant treatments on group III mGluRs.

Pharmacological studies of group III mGluRs have been limited due to lack of subtype-selective compounds. However, antidepressant-like effect of the group III mGluR agonist, ACPT-I, has been described in the forced-swim test [156,213,214]. Moreover, the mGluR4 positive allosteric modulator, PHCCC [214] and the selective mGluR8 agonist, RSPPG [156] induced an antidepressant-like in the forced-swim test. Moreover, mGluR7 knockout mice displayed an antidepressant-like phenotype in the forced-swim test and the tail suspension test [215]. In line with the latter study, the selective mGluR7 agonist AMN082 induced a dose-dependent decrease in the immobility time in the forced swim test and tail suspension test, supporting the hypothesis of antidepressant-like potency of mGluR7 agonists [216]. In the same study, AMN082 did not change the behaviour of mGluR7 knockout mice in the tail suspension test, whilst imipramine significantly reduced their immobility, indicating an mGluR7-dependent mechanism of the antidepressant-like activity of AMN082. ACPT-I, a group III mGluR agonist, produced anxiolytic-like effect after central administration [217] as assessed by the stress-induced hyperthermia and elevated plus-maze tests in mice, and the Vogel test in rats. The potential anxiolytic effect of ACPT-I in the stress-induced hyperthermia test was inhibited by the benzodiazepine receptor antagonist flumazenil and by the 5-HT_{1A} receptor antagonist, WAY100635. At the same time, ritan-

serin, a 5-HT_{2A/C} receptor antagonist, did not change the anxiolytic-like effects of ACPT-I. The results of these studies indicate that both GABAergic and serotonergic systems are involved in the potential anxiolytic action of ACPT-I [217].

Currently, little is known about the dysfunction of the mGluR8. A recent study evaluated the behaviour of mGluR8 knockout mice in different behavioural tasks commonly used in neuropsychiatric research [218]. These mice expressed no anxiogenic phenotype in unconditioned anxiety models, including elevated plus maze, elevated zero maze and light-dark box test. However, a contextual fear deficit was observed in the mGluR8 knockout mice, indicating that these receptors may be implicated in some anxiety disorders, such as generalised anxiety [219].

Despite the limited number of studies on selective group III mGluR ligands the present studies indicate that compounds belonging to this group may possess antidepressant and anxiolytic properties.

PARKINSON'S DISEASE

Parkinson's disease (PD) is the second most common neurodegenerative disease affecting up to 3% of the elderly population worldwide [220,221]. It is characterized by motor symptoms such as rigidity, tremor, bradykinesia, postural instability and gait disturbances. PD is a progressive neurodegenerative disease affecting dopaminergic neurons in the substantia nigra pars compacta selectively. The resulting loss of dopaminergic innervation in the striatum is believed to be the primary event underlying the motor symptoms of PD. This loss of dopaminergic tone within the striatum leads to secondary disturbances in the two efferent striatal systems arising from the medium spiny neurons, termed the direct and indirect pathways [222]. Both pathways converge in substantia nigra pars reticulata/globus pallidus interna. However, the indirect pathway comprises projections to the globus pallidus externa, then to the subthalamic nucleus, and finally to the substantia nigra pars reticulata/globus pallidus interna. Accordingly, stimulation of these two pathways oppositely regulates the main basal ganglia output pathway, the nigrothalamic pathway. In this respect, a loss of dopamine tone in the striatum is assumed to induce an imbalance between the two pathways, namely a disinhibition of striatopallidal neurons and an inhibition of striatonigral neurons, ultimately resulting in an increased activity of the GABAergic nigrothalamic pathway [223]. It is generally believed that restoration of a normal level of activity within the indirect pathway would provide symptomatic effect in PD patients. Within this pathway, the subthalamic nucleus seems to play a critical role since it displays a continuous abnormal bursting mode of activity in PD patients [224,225]. Interestingly, similar findings have also been reported in rodent models based on the use of a neurotoxin, e.g. 6-hydroxydopamine (6-OHDA) [226]. In the clinic, long-term use of L-DOPA, the most prescribed anti-Parkinsonian drug, often results in a loss of efficacy and the apparition of dyskinesias [227]. It is thus essential to develop novel pharmacotherapies exhibiting a sustained symptomatic effect in the advanced stage of the disease, but also the potential to delay or stop the progression of the disease in its early stage. Since several subtypes of

mGluRs are expressed at relevant synapses along the indirect pathway, they may represent a promising strategy for the treatment of PD and have been suggested to provide symptomatic as well as neuroprotective potential (Table 5).

Selective Group I Modulation

mGluR1 and mGluR5 are located postsynaptically to glutamatergic terminals in almost all striatal medium spiny

neurons, as well as in the globus pallidus, subthalamic nucleus and substantia nigra reticulata [228], and are therefore in key positions to modulate neuronal activity within the indirect pathway. Activation of group I mGluRs has been shown to enhance NMDA currents in striatal neurons [229], as well as induce overactivity of the striatopallidal pathway as measured by proenkephalin mRNA expression [230]. Based on these and other evidence, it was hypothesized that blockade of group I mGluRs might induce antiparkinsonian-

Table 5. Summary of Preclinical Studies Supporting a Role of mGluRs in Parkinson's Disease

| Receptor Subtype | Pharmacological Tool | Effect | References |
|------------------|-----------------------|---|---|
| Group I | | | |
| | DHPG (agonist) | ↓ Amphetamine-induced rotations in 6-OHDA rats | Agari <i>et al.</i> 2008 |
| mGluR1 | EMQMCM (antagonist) | ↓ Haloperidol-induced catalepsy; ↔ haloperidol-induced locomotor Hypoactivity in rats; ↔ L-DOPA-induced dyskinesias | Dekundy <i>et al.</i> 2006 |
| | AIDA (antagonist) | ↔ L-DOPA-induced dyskinesias | Dekundy <i>et al.</i> 2006 |
| | LY367385 (antagonist) | Neuroprotection and rescue against 6-OHDA toxicity in rats | Vernon <i>et al.</i> 2005; 2007 |
| mGluR5 | MPEP (antagonist) | ↓ Reaction time in partial bilaterally 6-OHDA-lesioned rats after chronic treatment | Breyse <i>et al.</i> , 2002; 2003 |
| | | Neuroprotection and rescue against 6-OHDA toxicity in rats | Vernon <i>et al.</i> 2005; 2007 |
| | | ↓ Motor asymmetry in unilateral 6-OHDA-lesioned rats after STN administration | Phillips <i>et al.</i> 2006 |
| | MTEP (antagonist) | ↓ Priming and expression of L-DOPA-induced dyskinesia in rats | Rylander <i>et al.</i> 2009; Dekundy <i>et al.</i> 2006 |
| | | ↓ Haloperidol induced catalepsy; ; ↔ haloperidol-induced locomotor hypoactivity in rats | Dekundy <i>et al.</i> 2006 |
| | SIB-1893 (antagonist) | ↓ L-DOPA-induced dyskinesias in MPTP primates; ↔ symptomatic effect of L-DOPA | Hill <i>et al.</i> 2001 |
| Group II | | | |
| mGluR2/3 | DCG-IV (agonist) | ↓ Reserpine-induced akinesia in rats | Dawson <i>et al.</i> 2000 |
| | LY354740 (agonist) | ↓ Haloperidol-induced catalepsy in rats | Bradley <i>et al.</i> 2000 |
| | LY379268 (agonist) | ↔ Reserpine-induced akinesia in rats; ↔ rotations in 6-OHDA rats ↓ Nigrostriatal degeneration after 6-OHDA in rats | Murray <i>et al.</i> 2002 |
| | | ↓ Nigrostriatal degeneration after MPTP in mice | Battaglia <i>et al.</i> 2003 |
| Group III | | | |
| | L-AP4 (agonist) | ↓ Haloperidol-induced catalepsy; ↓ reserpine-induced akinesia in rats; ↓ forelimb asymmetry in 6-OHDA-lesioned rats | Valenti <i>et al.</i> 2003 |
| | | ↓ Reaction time in partial bilaterally 6-OHDA-lesioned rats | Lopez <i>et al.</i> 2007 |
| | | Neuroprotection and rescue following 6-OHDA lesion in rats | Vernon <i>et al.</i> 2005; 2007 |
| | | Neuroprotection against rotenone toxicity | Jiang <i>et al.</i> 2006 |
| mGluR4 | PHCCC | ↓ Reserpine-induced akinesia in rats | Marino <i>et al.</i> 2003 |
| | LSP1-2111 (agonist) | ↓ Haloperidol induced catalepsy in rats; ↓ reaction time in partial bilaterally 6-OHDA-lesioned rats; ↓ glutamate transmission at the striatopallidal synapse | Beurrier <i>et al.</i> 2009 |
| mGluR7 | AMN082 (agonist) | ↓ Haloperidol induced catalepsy; ↓ apomorphine-induced rotations in unilateral 6-OHDA rats; ↓ reaction time deficits in bilateral 6-OHDA rats | Greco <i>et al.</i> 2009 |

Abbreviations and symbols: ↑, increased; ↓, decreased ; ↔, unchanged; 6-OHDA, 6- hydroxydopamine; ACh, Acetylcholine; MPTP, L-3,4-dihydroxyphenylalanine; PAM, positive allosteric modulator; STN, subthalamic nucleus.

like effects *in vivo*. In line with this assumption, it was reported that intra-striatal blockade of mGluR1/5 by AIDA, reversed haloperidol-induced catalepsy in rats, an effect attributed to normalization of the activity of striatopallidal neurons [231]. Several studies have used subtype-selective antagonists of group I mGluRs to further dissect out the contribution of mGluR1 and mGluR5 to a potential antiparkinsonian-like effect in relevant animal models. For instance, intrapallidal injections of the selective mGluR1 agonist DHPG have been shown to reduce amphetamine-induced rotational behavior in 6-OHDA-lesioned rats [232]. In addition, the selective mGluR1 antagonist EMQMCM, was shown to slightly inhibit haloperidol-induced catalepsy, while it was devoid of effect on hypoactivity induced by a lower dose of haloperidol [233]. Similar findings were reported with the selective mGluR5 antagonist MTEP. In the same study, both EMQMCM and MTEP were ineffective to induce rotations in unilateral DA-depleted rats, further weakening a potential symptomatic benefit of group I antagonism. Interestingly, an antiparkinsonian-like profile has been reported for another non-competitive mGluR5 antagonist, MPEP [234]. However, it was shown that chronic, but not acute treatment with MPEP, reversed akinesia in a reaction time test in bilateral 6-OHDA-lesioned rats. Furthermore, acute treatment with MPEP did not induce rotations in unilateral 6-OHDA-lesioned rats, finding in agreement with the study by Dekundy *et al.* [233]. In contrast, chronic administration of MPEP induced a significant increase in the number of rotations in unilateral 6-OHDA-lesioned rats. In a more recent study, it was shown that the increased metabolic activity in the subthalamic nucleus of dopamine-depleted rats, as detected by cytochrome oxidase staining, was reversed in animals chronically treated with MPEP [235]. Taken together, these data indicate that mGluR5 blockade would normalize the hyperactive state of the subthalamic nucleus, most likely indirectly through modulation of striatopallidal neurons [236], thereby leading to antiparkinsonian-like effects. In line with this assumption, local injections of MPEP into the subthalamic nucleus attenuated motor asymmetries in unilateral 6-OHDA-lesioned rats [237]. In summary, mGluR5 antagonism may offer symptomatic improvement based on the reported antiparkinsonian-like effects in animal models, whereas mGluR1 antagonism holds less promise.

Besides a potential symptomatic effect, group I mGluR antagonism has been suggested to be of potential interest with respect to alleviation of L-DOPA-induced dyskinesias. Whereas selective mGluR1 antagonists such as EMQMCM or AIDA were ineffective in a rodent model of L-DOPA-induced dyskinesias [233], several preclinical studies have suggested such a potential for mGluR5 blockade. Dyskinesias induced by an acute challenge of L-DOPA after three weeks of priming with L-DOPA were prevented by acute administration, as well as by chronic co-administration of MTEP in 6-OHDA-lesioned rats [233,238]. These data suggest that mGluR5 antagonism can prevent priming of dyskinesias, as well as reverse the expression of dyskinesias in primed animals, an effect attributed to the normalization of an excessive GABA overflow in the substantia nigra reticulata [239]. Furthermore, it was reported in an early study that

another mGluR5 antagonist, SIB-1893, alleviated dyskinesia induced by L-DOPA without changing its therapeutic effect in a primate model of PD [240]. Further supporting a role of mGluR5 in L-DOPA-induced dyskinesias, it was recently reported that mGluR5 binding was increased in the putamen and pallidum of dyskinetic MPTP-treated primates, while mGluR5 binding was normalized when dyskinesias were prevented by NMDA receptor blockade [241]. Taken together, these findings strongly suggest a potential benefit of mGluR5 antagonism in the alleviation of L-DOPA-induced dyskinesias typically observed after long-term treatment in patients. Furthermore, the preclinical findings also indicate that the symptomatic effect of L-DOPA would not be affected when considering mGluR5 antagonism as an adjunct therapy to L-DOPA.

In addition to putative symptomatic and antidyskinetic effects of mGluR5 blockade relevant in the late stage of Parkinson's disease, a neuroprotective potential relevant in the early stage of the disease has also been suggested. Preclinical studies have shown that repeated intranigral injection of either LY367385, a mGluR1 antagonist or MPEP, a mGluR5 antagonist, produced robust neuroprotection of nigral dopaminergic neurons in 6-OHDA-lesioned rats [242]. Interestingly, subchronic intranigral injections with LY367385 or MPEP were also reported to slow down dopamine cell loss in rats already undergoing nigrostriatal degeneration, a more clinically relevant model [243], indicative of a potential rescuing effect of group I mGluR antagonism. One of the caveats in the latter studies is related to the fact that MPEP also acts as a NMDA receptor antagonist depending on the concentration used, which may account for the observed neuroprotective effect. Glutamate excitotoxicity has thus been hypothesized to contribute to the progression of dopamine neurodegeneration in Parkinson's disease, and would occur as a consequence of the hyperactive state of glutamatergic neurons in the subthalamic nucleus [244,245]. However, it was reported that MPEP lost its neuroprotective properties against MPTP toxicity in mGluR5 knockout mice, ruling out a possible contribution of NMDA receptors [246]. Taken together, preclinical findings indicate that mGluR5, and possibly mGluR1 antagonism, may offer neuroprotective benefit slowing down the progression of the disease in its early stages.

Selective Group II Modulation

The mGluR2/3 are expressed presynaptically on glutamatergic and GABAergic terminals in several nuclei of the basal ganglia, including striatum, globus pallidus and substantia nigra [16,16], where their activation acts to reduce synaptic neurotransmitter release. Interestingly, an adaptive downregulation of mGluR2/3 in Parkinsonian patients has been reported and suggested to compensate for increased glutamatergic transmission [247]. In preclinical studies, several group II mGluR agonists have been shown to reduce motor abnormalities observed in animal models of Parkinson's disease. For instance, intraventricular administration of DCG-IV reversed reserpine-induced akinesia in rats [248]. In addition, another group II mGluR agonist, LY354740, was reported to reverse haloperidol-induced catalepsy in rats, an effect attributed at least partly to mGluR2/3-mediated reduc-

tion in glutamate release from subthalamic terminals [249]. However, contrasting results were obtained with LY379268, a selective mGluR2/3 agonist, which failed to reverse reserpine-induced akinesia and failed to affect rotational behaviour in 6-OHDA-lesioned rats [250]. In neuroprotective studies, LY379268 provided some protection against 6-OHDA neurotoxicity in rats, an effect correlated to both functional improvement and correction of dopamine turnover [250], as well as against MPTP neurotoxicity in mice as assessed by reduction of the extent of nigrostriatal degeneration [251]. In agreement with the assumption that glutamate excitotoxicity contributes to dopamine cell loss, mGluR2/3 may exert neuroprotective effects *via* regulation of glutamatergic transmission at the subthalamonigral synapse onto dopaminergic neurons [252].

In summary, activation of mGluR2/3 may offer symptomatic as well as neuroprotective benefit in the treatment of Parkinson's disease. However, more studies would be needed especially with respect to motor improvement as contradictory data have been reported in the literature.

Selective Group III Modulation

Group III mGluRs are presynaptically localized on GABAergic and glutamatergic terminals in several basal ganglia nuclei, including globus pallidus and substantia nigra, and therefore represent potential targets for reduction of abnormal activity associated with PD [253]. Several pre-clinical studies support a possible therapeutic benefit of mGluR4 agonism, while only few studies have investigated the role of other subtypes, e.g. mGluR7 and mGluR8.

mGluR4

Positive modulation of mGluR4 has recently gained interest as a potential strategy aimed at improving motor symptoms in both early and late stages of Parkinson's disease, as well as offering neuroprotection in early stages. Early pre-clinical studies have shown that the selective group III agonist L-AP4 produced symptomatic improvement in both acute and chronic rodent models of Parkinson's disease, including haloperidol-induced catalepsy, reserpine-induced akinesia, and forelimb asymmetry in unilateral 6-OHDA-lesioned rats [254]. It was further reported in the same study that L-AP4 modulated glutamatergic transmission at the striato-pallidal synapse through activation of mGluR4, as indicated by the loss of effect in brain slices from mGluR4 knockout mice. This mechanism was also suggested to contribute to the observed behavioral improvements. In line with this hypothesis, intrapallidal administration of L-AP4 was later reported to alleviate 6-OHDA-induced akinesia assessed in a reaction time task in rats [255]. A selective allosteric potentiator of mGluR4, PHCCC, was also reported to reverse reserpine-induced akinesia in rats, an effect again attributed to the modulation of striatopallidal glutamate transmission [256]. Recently, a novel mGluR4-preferring agonist, LSP1-2111, was shown to reverse akinesia assessed by a reaction time task in 6-OHDA-lesioned rats after intrapallidal injection, and block haloperidol-induced catalepsy after systemic administration [257]. Furthermore, LSP1-2111 reduced glutamate transmission at the striatopallidal synapse by a presynaptic mechanism, as shown for L-AP4.

Taken together, a symptomatic potential of mGluR4 agonism is strongly supported by preclinical studies in animal models of Parkinson's disease.

In neuroprotection studies, both acute and subchronic intranigral injections of L-AP4 are found to prevent dopaminergic cell loss induced by 6-OHDA in rats [242,243]. Subchronic intranigral injections of L-AP4 were further shown to slow down dopaminergic neurodegeneration in 6-OHDA-lesioned rats already undergoing degeneration [243]. In addition, L-AP4 was also reported to protect cultured dopaminergic neurons against rotenone toxicity [258]. Based on the glutamate excitotoxicity hypothesis of dopaminergic degeneration, mGluR4 activation on glutamatergic terminals of subthalamonigral neurons have been suggested to contribute to such a neuroprotective effect. In fact, L-AP4 was shown to inhibit excitatory transmission in the substantia nigra pars compacta, and the effect was potentiated by PHCCC, further involving a selective effect of mGluR4 [259]. These studies suggest that a decrease in excitatory glutamatergic transmission from the subthalamic nucleus may contribute to a possible neuroprotective benefit of mGluR4 positive modulation.

In summary, preclinical evidence support a putative symptomatic and neuroprotective benefit of mGluR4 positive modulation in the treatment of Parkinson's disease. However, more selective and brain penetrant compounds would be needed to strengthen this assumption.

mGluR7

Recently, the selective allosteric mGluR7 agonist, AMN082, was shown to reverse haloperidol-induced catalepsy in rats after intrastriatal or systemic administration [260]. The same anticataleptic effect of AMN082 was also found in wild-type, but not in mGluR7 knockout mice, indicative of a selective mGluR7-mediated effect. In addition, AMN082 was found to reverse apomorphine-induced rotations in unilateral 6-OHDA-lesioned rats as well as reverse akinesia in a reaction time task in bilateral 6-OHDA-lesioned rats [260]. This unique study suggests that mGluR7 positive modulation may represent an interesting strategy with respect to alleviation of motor symptoms in Parkinson's disease. Additional studies are needed to address whether a neuroprotective potential can also be achieved.

FRAGILE X SYNDROME

Fragile X syndrome (FXS) is the most common form of inherited mental retardation and is characterized by intellectual disabilities, autistic features, hyperactivity, audiogenic seizures, and certain physical features, e.g. elongation of the face, enlargement of the ears and postpubertal macroorchidism [261,262]. It is usually caused by a mutation of the fragile X mental retardation-1 gene (FMR1), leading to either decreased levels or complete loss of the FMR1 gene product fragile X mental retardation protein (FMRP) [263,264]. FMRP is an RNA binding protein that controls for instance synthesis of certain components of the postsynaptic density (PSD) in both the neocortex and hippocampus, as well as translational efficiency of dendritic mRNAs in response to stimulation of mGluRs [265-271].

Fmr1 knockout mice display physical and behavioural phenotypes comparable to those of the human counterpart, such as macroorchidism, increased locomotor activity, audiogenic seizures, and impaired fear-conditioned memory [272-274]. Similarly to post-mortem observations in FXS patients [275-277], Fmr1 knockout mice showed altered morphology of cortical and hippocampal dendritic spines, e.g. more spines are prolonged and immature, [276,278]. These observations suggest that FMRP plays a role in spine development and stabilisation [279], and a loss of FMRP function may result in impaired synaptic activity underlying mental retardation [280].

Selective Group I Modulation

Bear *et al.* (2004) proposed that group I mGluRs might be involved in FXS. More precisely, FXS would involve an increased occurrence of a non-NMDA receptor-dependent form of long-term depression (LTD) induced by activation of the group I mGluRs [267,281]. This mGluR-LTD is an activity-dependent synaptic weakening, at least partly mediated by group I mGluRs [280,282]. In line with this hypothesis, it has been demonstrated that Fmr1 knockout mice display an increased occurrence of mGluR-LTD [281], while NMDA receptor-LTD has been found to be normal in the Fmr1 knockout mice in the hippocampus and cerebellum [61,281]. Furthermore, activation of group I mGluRs by DHPG has been shown to induce mGluR-LTD at excitatory synapses onto CA1 pyramidal cells [281,283,284]. It is believed that activation of group I mGluRs initiates postsynaptic protein synthesis, while FMRP suppresses the translation of the mRNA encoding proteins involved in mGluR-LTD, thus leading to functionally opposite roles of group I mGluRs and FMRP. It has therefore been hypothesized that FXS symptoms would result from an aberrant group I mGluR-mediated protein synthesis, in turn leading to mGluR-LTD involving internalization of AMPA receptors, GluR1 and GluR2, at dendritic synapses [264]. Since both LTD and long-term potentiation (LTP) are believed to contribute to learning and memory [281], an increased mGluR-LTD may interfere with the establishment and maintenance of strong synapses required for memory formation.

Recently, heterozygotes double Fmr1/Grm5 (encoding mGluR5) knockout mice and have been generated [285]. Interestingly, these mice showed no difference from wild type mice with regard to spine density, thus suggesting that a 50% decrease of group I mGluR signalling is sufficient to rescue the increased spine density phenotype in the Fmr1 knockout. In this and other examples, it was demonstrated that heterozygote Fmr1/Grm5 knockout mice were devoid of the phenotypes, except macroorchidism, characterizing the Fmr1 knockout mice. These data suggest that decreased group I mGluR signalling may be a promising target for FXS.

In recent years, pharmacological studies using mGluR5 antagonists have been performed in animal models of FXS. It was demonstrated that the non-competitive antagonist MPEP increased inhibitory phosphorylation of glycogen synthase kinase-3 (GSK3) in Fmr1 knockout mice, but not in wild type mice [286]. Interestingly, it was reported that Fmr1 knockout mice display impaired inhibitory serine-

phosphorylation of GSK3, and inhibition of GSK-3 by lithium ameliorated behavioural deficits in models of FXS. Taken together, these studies indicate that increased mGluR signalling in Fmr1 knockout mice may contribute to the deficit in inhibitory control of GSK3. In line with this hypothesis, studies in primary cortical neurons have shown that FMRP acted as a repressor of the translation of Shank1 mRNAs, which controls dendritic spine morphology, and that DHPG-mediated mGluR stimulation enhanced the translation of Shank1 [263].

Furthermore, McBride *et al.* (2005) showed that MPEP restored mushroom bodies, memory deficits and courtship behaviour activity in a *Drosophila* FXS model [287]. Courtship behaviour activity was recovered in larvae and adults treated with MPEP, suggesting that the observed effects did not result from the prevention of developmental defects. Even though the doses of MPEP used have been demonstrated to antagonize mammalian NMDA receptor activity, other competitive mGluR5 antagonists such as MPPG, MTPG, and LY341495, have shown similar rescue of the mutant flies, suggesting that the observed effects result from mGluR5 blockade. In a follow up study, it was shown that learning deficits, which appeared at an older age than the deficits in training and memory, were rescued by the same mGluR antagonists as used previously [288]. Interestingly, the learning deficits were rescued with treatment during development alone. However, the loss of mushroom bodies could not be restored in older flies. In a zebrafish model of FXS, MPEP has been shown to either completely or partially rescue the phenotypes, including craniofacial and neurite branching abnormalities [289]. In Fmr1 knockout mice, MPEP has been demonstrated to rescue heightened audiogenic seizures susceptibility, abnormal center-field behaviour [267], and impaired pre-pulse inhibition [290]. In addition, both MPEP and fenobam rescued the protrusion morphology observed in hippocampal neurons of Fmr1 knockout mice [267].

The mGluR5 antagonist, fenobam, has been demonstrated to reduce hyperactivity and anxiety in patients suffering from FXS in a small clinical trial [291]. Furthermore, 50% of the treated patients showed improvement in prepulse inhibition. However, this effect was not always well-correlated with subjective clinical improvement nor with the pharmacokinetics of fenobam, which showed great inter-individual differences. In preclinical studies, fenobam and other mGluR5 antagonists, including MTEP, have been reported to produce impairments in various cognitive tests, including water maze and passive avoidance tests [292,293], while other studies have not found any effect of fenobam on working memory or spatial learning at therapeutic relevant doses [183]. In line with a potential effect on cognitive function, MPEP has been demonstrated to suppress theta and gamma oscillations and impair LTP in the dentate gyrus, whereas enhanced LTP was observed in the CA1 region of rats [294]. Currently, no preclinical or clinical evidence supporting a role of selective group II or II modulators in the treatment of FXS has been published.

Since the Fmr1 gene has been identified as an autism-related gene and the most common cause of autism [295], modulation of mGluRs for the treatment of FXS might also

be beneficial for autism, which is also supported by the observation that MPEP blocked repetitive features in a mice model of autism [296].

In conclusion, reduction in mGluR5 signalling may represent a promising target for treating many of the aspects of FXS. However, possible adverse effects of mGluR5 antagonism on cognitive function remain to be further addressed.

HUNTINGTON'S DISEASE

Huntington's Disease (HD) is an autosomal dominant neurodegenerative disorder characterized by involuntary body movements, cognitive deficits, and changes in personality [297-299]. Symptoms generally start appearing between mid thirties and middle age, and patients usually die 15 to 20 years after the symptomatic onset [297,299]. The neurodegeneration occurs preferentially in the striatum, extends at later stages to other brain regions including the deep layers of cortex, globus pallidus, thalamus, subthalamic nuclei, substantia nigra and gliosis formation appears [297]. In the striatum, the neuronal loss selectively affects GABAergic medium spiny neurons, whereas large aspiny cholinergic neurons are spared [297,299].

As for fragile X syndrome, HD is caused by a mutation in the gene encoding the protein huntingtin (Htt) [297,299]. Even though Htt is expressed throughout the nervous system, the neurodegeneration remains limited to specific brain areas even at late stages of HD. Some studies have suggested that Htt is required for cell survival, and loss of its function may therefore be involved in neurodegeneration [298]. Glutamate excitotoxicity has been proposed to play a central role in the pathogenesis of HD [300], and the involvement of the NMDA receptors in glutamate-mediated excitotoxicity has been especially investigated [301]. However, there is emerging evidence that mGluRs may also play a role in glutamate-mediated excitotoxicity [302]. More precisely, the identification of proteins interacting with both Htt and mGluR signalling indicates that Htt might indeed be involved in the regulation of mGluR signaling [303,304].

Pharmacological studies using a combined group I mGluR antagonist and group II mGluR agonist, (S)-4C3HPG, have shown a reduction in the lesion volume induced by intra-striatal quinolinolate *in vivo* [305]. In contrast, no effect on lesion volume was observed with (+)-MCPG, a less potent group I/II mGluR antagonist. These data suggest that a reduction in mGluR signalling may have neuroprotective effects. Further characterization of the precise receptor subtypes involved is unfortunately lacking.

R6/2 HD transgenic mice, which express part of the mutated human HD gene, have been reported to develop a progressive neurological phenotype with reduction in brain weight, formation of neuronal intranuclear inclusions (NII) and have limited survival [306]. They also display decreased expression of several neurotransmitter receptors including striatal mGluR1, mGluR2 and mGluR3. The decrease in the mGluR2 has been suggested to contribute to glutamate-mediated excitotoxicity by increasing glutamate release from corticostriatal terminals [307]. Decreased levels of glial glutamate transporters, which are necessary for the clearance of glutamate from the synaptic cleft, have also been reported in

R6/2 HD mice, and may also contribute to excitotoxicity [308]. Schiefer *et al.* (2004) investigated the effect of the non-competitive mGluR5 antagonist, MPEP, and the mGluR2/3 agonist, LY379268, on the disease course in R6/2 HD transgenic mice. Chronic administration of both MPEP and LY379268 from a presymptomatic stage mildly, but significantly, increased survival time and reduced early hyperactivity, effects attributed to the inhibition of glutamate neurotransmission in the basal ganglia circuitry [309]. In the same study, MPEP was also found to attenuate the progressive loss of motor coordination, a robust measure of disease progression in R6/2 HD transgenic mice, while only a trend towards attenuation was observed with LY379268. These data suggest that decreasing glutamatergic transmission through modulation of mGluRs may provide symptomatic relief as well as slow down the progression of the disease. In line with a putative glutamatergic dysfunction in HD, the non-selective glutamate receptor antagonist, riluzole, which is used in the treatment of amyotrophic lateral sclerosis, has been demonstrated to increase survival time in R6/2 HD transgenic mice [310]. However, riluzole has since been demonstrated to lack efficacy in HD patients [311], which may be due to non-specific effects on other transmitter systems.

In conclusion, mGluRs modulation may provide therapeutic benefits in HD. However, more studies aimed at elucidating the precise molecular mechanisms underlying the disease would be needed to further support a role of specific mGluR subtypes.

ALZHEIMER'S DISEASE

Alzheimer's disease (AD) is characterized by the deposition of β -amyloid ($A\beta$) into senile plaques, the formation of neurofibrillary tangles, and neuronal death [312]. Glutamatergic pathways have been implicated in the pathophysiology of AD [313]. It has been suggested that $A\beta$ triggers neurodegeneration by a complex interaction of processes, including increased levels of extracellular glutamate and intracellular calcium, leading to apoptosis and neuronal death [314].

A down-regulated and desensitized group I mGluR/phospholipase C signalling has been demonstrated in the frontal cortex of AD patients, which was found to correlate with disease progression in the cerebral cortex [315]. Furthermore, a reduction of mGluR1 has been found in the frontal cortex of patients with AD, and this reduction was correlated with the progression of the disease [315]. These findings suggest that group I mGluR dysfunction may be involved in AD. Activation of group I mGluRs has been shown to accelerate processing of amyloid precursor protein by α -secretase into non-amylogenic products, leading to protection against $A\beta$ deposit *in vitro* [316,317]. Thus, decreased levels and activity of group I mGluR in cerebral cortex may affect the deposition of $A\beta$ in AD patients.

Pharmacological activation of Group II and III mGluR has been shown to reduce neuronal death *in vitro*, an effect attributed to reduced glutamate release [318,319]. Moreover, the group II mGluR agonist LY379268, was shown to protect neurons from hippocampus against excitotoxicity *in vivo* [320]. mGluR2 has been suggested to play a role in the

pathogenesis of neuronal cell death and survival, as indicated by an upregulation of its expression in the hippocampus of AD patients, and a close association with hyperphosphorylated tau deposition [318,319,321]. Thus, mGluR2 might play a key role in the pathogenesis of AD [321]. Interestingly, agonists of mGluRs induced Tau phosphorylation, and several lines of evidence suggest that the differential expression of glutamate receptors in specific populations of neurons may account for specific neuronal vulnerability [314].

In conclusion, there is limited information about the role of mGluRs in the pathogenesis of AD, but it is likely that mGluRs may play a significant role in the pathophysiology of this disease. However, there is a lack of published reports, showing efficacy with mGluR ligands in AD models.

PAIN

Inflammatory and neuropathic pain is a major health problem affecting up to 5% of the population worldwide. The mechanisms underlying pain remain unclarified. However, it is thought that continuous activation of peripheral afferent fibers by noxious stimulation results in sensitization of dorsal horn neurons, which would subsequently produce aberrant activity in primary afferent fibers [322]. As a result, peripheral and central mechanisms contribute to a cycle of persistent nociception. Persistent activation of peripheral afferents may result in central changes in neurotransmitter release or receptor states, resulting in chronic nociceptive activation. Glutamate is released in the spinal dorsal horn, in which it acts *via* activation of ionotropic glutamate receptors as well as mGluRs [1,322]. Increasing evidence supports a specific role of mGluRs in nociceptive transmission, given the wide expression of these receptors along the nociceptive neuroaxis, such as the dorsal root ganglia, midbrain periaqueductal grey region, spinal cord, thalamus and amygdala [323]. For instance, Group I mGluRs are expressed both at the spinal and supra-spinal levels, including the thalamus, a brain area critically involved in the signaling of nociceptive information [324]. Moreover, Group I and II mGluRs are expressed by peripheral terminals and in the soma of dorsal root ganglia neurons [323]. Almost all mGluR subtypes are expressed in the spinal cord [10,12,16,17,323,325]. The mGluR3, 5, 7 and 8 have been reported to be present in the midbrain periaqueductal grey region [323], an important center for the processing of nociceptive information, and some reports suggest that group I and group III mGluRs are expressed in the amygdala [326,327], a region involved in emotional pain (Table 6). Besides being expressed in neurons, mGluRs are also found in glial cells, which may play a role in neuropathic pain [328], but this topic will not be reviewed here.

Selective Group I Modulation

mGluR1

Early studies have reported that intrathecal injection of the non-selective mGluR1/5 agonist DHPG induced hyperalgesia and spontaneous pain in rats [329-332]. Other studies inactivating the mGluR1 by a selective antibody have shown that intrathecal blockade of mGluR1 reduced nociceptive behaviors in several models of inflammatory and neuropathic

pain, including complete Freund's adjuvant (CFA)-induced chronic inflammatory pain, formalin-induced persistent nociception, nerve injury-induced neuropathic pain and DHPG-induced spontaneous nociceptive behaviours in rodents [333-337]. In line with an involvement of mGluR1 in pain mechanisms, up-regulation of mGluR1 was also demonstrated in spinal dorsal horn in response to persistent inflammatory hyperalgesia [338]. In addition to a role of spinal mGluR1 in nociception, a role of thalamic mGluR1 is also suggested by electrophysiological studies showing that the selective mGluR1 antagonist LY367385, was able to reduce the response of somatosensory neurons of the rat thalamus to noxious stimuli [339]. Several mGluR1 antagonists, e.g. CPCCOEt, AIDA, LY456236 and LY367385, have shown antinociceptive effects in various pain models such as thermal noxious, formalin-induced pain, CFA, mechanical allodynia [32,340-346]. In addition, systemic administration of the non-competitive mGluR1 antagonist, A841720, has been reported to reverse inflammatory and neuropathic pain in rodents, further supporting a therapeutic potential of mGluR1 antagonism in the treatment of chronic pain states [347,348]. Several other non-competitive mGluR1 antagonists, such as A841720, A794282, A794278 and A850002, have also been reported to attenuate spontaneous nociception in a pre-clinical model of postoperative pain [349]. Beside these pharmacological evidences, antisense knockdown of mGluR1 receptors has been reported to decrease spinal nociceptive neurotransmission and neuropathic hyperalgesia [334,336,337], further strengthening the antinociceptive potential of mGluR1 blockade.

mGluR5

A large body of literature supports the assumption that mGluR5 modulates pain and that mGluR5 antagonism may be used in the treatment of chronic pain conditions. As described for mGluR1, mGluR5 are also expressed both at the spinal and supra-spinal levels where they control nociceptive transmission. At the supra-spinal level, pharmacological blockade of mGluR1 by MPEP prevented the nociceptive response of sensory neurons in the rat thalamus [350]. At the behavioural level, the selective mGluR5 antagonist, SIB1757, fully reversed hyperalgesia in a neuropathic pain model in rats [351]. In addition, two other mGluR5 antagonists, MPEP and MTEP, have been shown to produce antinociceptive effects in a wide range of rat nociceptive assays, including CFA-induced chronic inflammatory pain, hyperalgesia induced by formalin and mechanical allodynia following spinal nerve ligation [32,34,343,346,352]. MPEP was also reported to prevent the increased nociceptive reaction induced by the cannabinoid receptor agonist WIN 55,212-2 in rats [353]. Moreover, MPEP abolished acetic acid-induced writhing activity in mice, and was shown to reduce mechanical allodynia and thermal hyperalgesia in a model of postoperative hypersensitivity [352]. Taken together, preclinical studies support the concept of mGluR5 antagonism for the treatment of chronic pain.

Selective Group II Modulation

N-acetylaspartylglutamate (NAAG) is an endogenous peptide agonist that activates mGluR2/3, with preference for mGluR3 [354]. Systemic administration of NAAG or of

Table 6. Summary of Preclinical Studies Supporting a Role of mGluRs in Pain

| Receptor Subtype | Pharmacological Tool | Effect | References |
|------------------|--|--|---|
| Group I | | | |
| mGluR1 | A841720 (antagonist) | ↓ CFA-induced pain in rats ↓ Mechanical allodynia in rats | El-Kouhen <i>et al.</i> 2006;Zheng <i>et al.</i> 2005 |
| | A841720 A794282 A794278 A850002 (antagonists) | ↓ Post-operative pain in rats | Zhu <i>et al.</i> 2008 |
| | CPCCOEt (antagonist) | ↓ Formalin- induced pain in mice ↓ Thermal hyperalgesia in mice ↓ Noxious stimulation- induced pain in rats | Bhave <i>et al.</i> 2001;Han and Neugebauer 2005 |
| | AIDA LY456236 LY36738 (antagonists) | ↓ Formalin- induced pain in mice ↓ Thermal hyperalgesia in mice and rats ↓ Mechanical allodynia in rats | Lee <i>et al.</i> 2007;Varty <i>et al.</i> 2005;Zhang <i>et al.</i> 2002 |
| mGluR5 | SIB1757 (antagonist) | ↓ Thermal hyperalgesia in rats | Dogrul <i>et al.</i> 2000 |
| | MPEP & MTEP (antagonists) | ↓ Formalin- induced pain in mice ↓ Post-operative pain in rats ↓ Thermal hyperalgesia in mice ↓ Noxious stimulation- induced pain in rats ↓ Carrageenan- induced pain in mice ↓ CFA- induced pain in rats ↓ Touch-evoked allodynia in mice ↓ Mechanical allodynia in mice | Bhave <i>et al.</i> 2001;Varty <i>et al.</i> 2005;Zhu <i>et al.</i> 2004;Lee <i>et al.</i> 2007;Walker <i>et al.</i> 2001 |
| Group II | | | |
| mGluR2/3 | LY354740 LY379268 LY389795 (agonists) | ↓ Carrageenan- induced pain in rats ↓ CFA-induced pain in rats ↓ Formalin-induced pain in rats ↓ Mechanical allodynia in rats ↓ Thermal hyperalgesia in rats | Simmons <i>et al.</i> 2002;Jones <i>et al.</i> 2005 |
| | APDC (agonist) | ↓ Thermal-induced pain in rats ↓ Formalin-induced pain in rats ↓ Carrageenan- induced pain in mice ↓ CFA- induced pain in rats ↓ Thermal hyperalgesia in rats | Neugebauer and Carlton 2002;Yang and Gereau 2003 |
| Group III | | | |
| mGluR4/7/8 | L-AP4 (agonist) | ↓ Mechanical allodynia in rats | Chen and Pan 2005 |
| | ACPT-I (agonist) | ↓ Carrageenan-induced pain in rats ↓ Vincristine-induced pain in rats | Goudet <i>et al.</i> 2008 |
| mGluR4 | PHCCC (PAM) | ↓ Carrageenan-induced pain in rats ↓ Vincristine-induced pain in rats | Goudet <i>et al.</i> 2008 |
| mGluR7 | AMN082 (agonist) | ↓ Carrageenan-induced pain in rats | Palazzo <i>et al.</i> 2008 |
| | (S)-3,4-DCPG (agonist) | ↓ Carrageenan-induced pain in rats ↓ Vincristine-induced pain in rats | Palazzo <i>et al.</i> 2008 |

Abbreviations and symbols: ↓, decreased; CFA, complete Freund's adjuvant; NAM, negative allosteric modulator; PAM, positive allosteric modulator.

NAAG peptidase inhibitors, ZJ-43 and 2-PMPA, have been proven to be effective in reducing perception of inflammatory and neuropathic pain in rat models [355,356]. These findings are consistent with the hypothesis that NAAG may play a physiologic role in moderation of pain perception, and that pharmacologically increasing the level of NAAG activation of mGluR2/3 may be promising in pain management. Furthermore, several studies have suggested that activation of mGluR2/3 have analgesic properties when applied centrally, or peripherally at the inflammatory site [357-363]. For instance, selective activation of peripheral mGluR2/3 with APDC was shown to reduce inflammation-induced thermal and mechanical hypersensitivity as well as formalin-induced hyperalgesia, and to contribute to the recovery from mechanical hypersensitivity following carrageenan-induced inflammation [361]. Three other mGluR2/3 agonists LY354740, LY379268 and LY389795 were also found to attenuate formalin-induced paw licking behavior [363,364] as well as carrageenan-induced thermal hyperalgesia, whereas mechanical allodynia was not affected [365].

Selective Group III Modulation

The role of group III mGluRs in nociceptive processing has not been thoroughly investigated, but some evidence may indicate a therapeutic potential of certain subtypes. It was indeed reported that activation of group III mGluRs with L-AP4 attenuated allodynia in spinal nerve-ligated rats, but did not affect pain threshold in normal rats [366]. In addition, intrathecal injection of another group III mGluR agonist, ACPT-I, inhibited the nociceptive responses to formalin as well as the mechanical hyperalgesia associated with inflammatory pain in carrageenan-treated and monoarthritic rats or neuropathic pain in mononeuropathic and vincristine-treated rats, while it did not affect pain threshold to mechanical or thermal stimuli in normal rats [367]. Similar antinociceptive effects were also observed following intrathecal injection of PHCCC, a mGluR4 positive allosteric modulator [367]. At the cellular level, activation of the group III mGluRs by L-AP4 reduced pain-related synaptic plasticity in the amygdala under normal conditions as well as in a model of arthritis pain [368]. Recently, Palazzo *et al.* (2008) investigated the role of mGluR7 and mGluR8 in the amygdala in pain related behaviours [369]. In their study, activation of mGluR7 by the selective agonist AMN082 was found to have pro-nociceptive effects under normal conditions, but not in the arthritic pain model, while activation of mGluR8 by the selective receptor agonist S-3,4-DCPG exhibited antinociceptive effects in arthritic, but not in normal rats [369]. Taken together, evidence supports a role of group III mGluRs in chronic pain. However, more studies using subtype selective compounds would be needed in order to identify which of group III mGluRs would be most promising.

In conclusion, mGluRs modulation may represent a promising strategy for the treatment of different types of pain, including inflammatory and neuropathic pain. At present, there are very few effective and well-tolerated therapies for neuropathic pain. Current medications include opioids, antidepressants and anticonvulsants. Modulation of mGluRs may offer better efficacy and more importantly better side-effect profile than current therapies, since the use of opioids

in neuropathic pain remains controversial, and both antidepressants and anticonvulsants are associated with side-effects.

EPILEPSY

Glutamate is widely implicated in the mechanisms underlying epilepsy [370,371]. Therefore, targets that potentially control glutamatergic transmission are of special interest to investigate as candidates to prevent epileptogenesis. In rodent models of epilepsy changes in glutamate receptor or glutamate transporter expression were shown to affect epileptic seizures [372]. Independent of the primary cause, synaptically released glutamate appears to play a major role in the initiation and spread of seizure activity. Antagonists of ionotropic glutamate receptors reduced seizures in several animal models of epilepsy [373-375]. However, these drugs failed early in clinical trials due to multiple side effects, including motor and cognitive impairment [376,377]. With the more recent discovery of mGluRs, there is a renewed interest in targeting glutamate receptors for the treatment of epilepsy. Several preclinical studies have suggested a role for mGluRs in epileptogenesis and neuronal injury [378,379] and support pharmacological modulation of specific subtypes as a potential therapeutic strategy for the treatment of epilepsy (Table 7).

Selective Group I Modulation

Activation of group I mGluRs enhances neuronal excitability leading to a potentiation of NMDA and AMPA/KA receptor functions. Long-lasting functional enhancement of group I mGluR activity has been reported in amygdala-kindled rats, and up-regulation of mGluR5 immunoreactivity has been described in temporal lobe epilepsy and in focal cortical dysplasia patients [380]. Group I mGluR agonists, such as (1*S*,3*R*)-ACPD and 3,5-DHPG, were reported to induce limbic seizures and neuronal injury in rats [381], while other non-selective mGluR1/5 antagonists such as LY393053, LY339764, LY367335, LY367366, and LY339840 have demonstrated potent anticonvulsant activity in models of DHPG-induced limbic seizures [341]. In addition, the selective mGluR1 agonist (S)-4CPG reduced audiogenic seizures in DBA/2 mice [382], and attenuated pentylenetetrazol (PTZ)- and DMCM-induced seizures in rats [383]. Further supporting mGluR1 involvement in epilepsy, the mGluR1-preferring antagonists LY367385 and AIDA, blocked spike-wave discharge (SWD) in lethargic mice (*lh/lh*) [384], a genetic model of absence epilepsy [385], and reduced sound-induced clonic seizures in DBA/2 mice [384]. Moreover, these two compounds were also shown to reduce seizures in genetically epilepsy prone rats (GEPR) [341,385] and attenuate PTZ- [386,387] and DHPG- [341,388] induced seizures in rodents. AIDA further attenuated KA-induced seizures in immature rats [389] as well as kindling-related learning deficits [386]. A non-competitive mGluR1 antagonist, BAY36-7620, was reported to reduce sound-induced clonic seizures in DBA/2 mice and to attenuate PTZ- induced seizures in animals [3], further supporting an anticonvulsant effect of mGluR1 antagonism.

Two non-competitive mGluR5 antagonists, MPEP and SIB1893, were reported to suppress seizures induced by the

Table 7. Summary of Preclinical Studies Supporting a Role of mGluRs in Epilepsy

| Receptor Subtype | Pharmacological Tool | Effect | References |
|------------------|--|--|--|
| Group I | | | |
| | LY393053 LY339764 LY367335 LY367366 LY339840 (antagonists) | ↓ Seizures induced by DHPG in mice | Kingston <i>et al.</i> 2002 |
| mGluR1 | (S)-4CPG (antagonist) | ↓ Seizures in DBA/2 mice ↓ Seizures induced by PTZ and DMCM in mice | Dalby and Thomsen 1996; Thomsen <i>et al.</i> 1994 |
| | LY367385 (antagonist) | ↓ Seizures in DBA/2 mice ↓ SWD in <i>lh/lh</i> mouse ↓ Seizures in GEPR ↓ Seizures induced by DHPG ↓ Seizures induced by kindling ↔ KA-induced and pilocarpine-induced seizures in rats | Burgess <i>et al.</i> 1997; Chapman <i>et al.</i> 1999; Kingston <i>et al.</i> 2002; Nagaraja <i>et al.</i> 2004; Renaud <i>et al.</i> 2002; Smolders <i>et al.</i> 2004; Thomsen and Dalby 1998 |
| | AIDA (antagonist) | ↓ Seizures in DBA/2 mice ↓ SWD in <i>lh/lh</i> mouse ↓ Seizures in GEPR ↓ Seizures induced by DHPG in mice ↓ Seizures induced by kindling and KA in rats ↔ Pilocarpine-induced seizures in rats | Burgess <i>et al.</i> 1997; Chapman <i>et al.</i> 1999; Kingston <i>et al.</i> 2002; Nagaraja <i>et al.</i> 2004; Renaud <i>et al.</i> 2002; Smolders <i>et al.</i> 2004; Thomsen and Dalby 1998 |
| | BAY367620 (antagonist) | ↓ Seizures in DBA/2 mice ↓ Seizures induced by PTZ in mice | De Vry <i>et al.</i> 2001 |
| mGluR5 | MPEP (antagonist) | ↓ Seizures in DBA/2 mice ↓ Seizures induced by DHPG and CHPG in mice ↓ SWD in <i>lh/lh</i> mouse ↔ KA-induced and pilocarpine-induced seizures in rats | Chapman <i>et al.</i> 2000 |
| | SIB1893 (antagonist) | ↓ Seizures induced by DHPG and CHPG in mice ↓ Seizures in DBA/2 mice ↓ SWD in <i>lh/lh</i> mouse ↔ Kindling-induced and pilocarpine-induced seizures in rats | Chapman <i>et al.</i> 2000 |
| Group II | | | |
| mGluR2/3 | (1S,3S)-APDC (agonist) | ↓ Seizures in DBA/2 mice ↓ Seizures induced by DMCM ↔ PTZ-induced seizures in mice ↔ ES in mice | Attwell <i>et al.</i> 1998; Dalby and Thomsen 1996 |
| mGluR2/3 | DCG-IV (agonist) | ↓ Seizures induced by kindling in rats ↔ KA-induced seizures in rats | Attwell <i>et al.</i> 1998; Miyamoto <i>et al.</i> 1997 |
| Group III | | | |
| mGluR4/7/8 | ACPT-1 (agonist) | ↓ Seizures in DBA/2 mice ↓ Seizures induced by DHPG in mice ↓ GEPR | Chapman <i>et al.</i> 2001 |
| mGluR4/7/8 | (R,S)-PPG (agonist) | ↓ Seizures in DBA/2 mice ↓ ES in mice ↓ Seizures in GEPR | Chapman <i>et al.</i> 1999; Gasparini <i>et al.</i> 1999 |
| mGluR8 | (S)-3,4-DCPG (agonist) | ↓ Seizures induced by DL-HCA in rats | Folbergrova <i>et al.</i> 2008 |

Abbreviations and symbols: ↔, unchanged; ↓, decreased; DHPG, 3,5-dihydroxyphenylglycine; PTZ, pentylenetetrazol; DMCM, Methyl-6,7-dimethoxy-4-ethyl-beta-carboline-2-carboxylate; GEPR, Genetically epilepsy prone rats; KA, kainic acid (KA); SWD, spike-wave discharge; C3HPG, (S)-4-carboxy-3-hydroxyphenylglycine; ES, Electroshock seizure; DL-HCA, DL-homocysteic acid.

selective mGluR5 agonist, CHPG [390]. These compounds also reduced sound-induced clonic seizures and DHPG-induced seizures in DBA/2 mice [390]. At the cellular level, MPEP suppressed DHPG-induced neuronal firing in rats, and decreased the incidence of SWD in lethargic mice (*lh/lh*) [390].

Selective Group II modulation

Supporting a role of mGluR2/3 in epilepsy, altered expression and function of these receptors has been reported in pilocarpine-treated rats [391,392]. Following evidence that (*S*)-4-C3HPG exhibited anticonvulsant activity, compounds with preferential action on group II mGluRs have been tested in several models of epilepsy. The mGluR2 agonist, (*1S,3S*)-APDC, reduced sound-induced seizures in GEPR and DBA/2 mice, as well as DMCM-induced seizures in rats, and enhanced the generalised seizure threshold in kindled rats [383,393,394]. Activation of mGluR2 by (*1S,3S*)-APDC or DCG-IV was also found to enhance the generalised seizure threshold in kindled rats [393]. In contrast, DCG-IV was ineffective against KA-induced seizures [395]. Further supporting an anticonvulsant effect of group II mGluRs agonism, LY379268 and LY389795 reduced SWD activity in *lh/lh* mice [396].

Selective Group III Modulation

The lack of optimal subtype selective compounds has hampered the investigation of the role of individual group III mGluRs in epileptogenesis. However, knockout animals have implicated an important role for mGluR7 in seizure activity [397,398], and a down-regulation of mGluR8 has been described in pilocarpine-epileptic rats [399]. In addition, the use of various mouse strains with differential susceptibility to pilocarpine-induced epilepsy has indicated that mGluR4 expression levels in the dentate gyrus of the hippocampus were inversely correlated with seizure susceptibility [400]. Pharmacological studies using activation of group III mGluR have yielded mixed results in animal models of epilepsy. Early studies using two group III mGluR agonists, L-AP4 and L-SOP, indicated a pro-convulsant effect of these compounds [401]. However, subsequent studies have described anticonvulsant effects of group III mGluR agonists. For instance, the group III mGluR agonist ACPT-1, with affinity for mGluR4, mGluR6 and mGluR8, reduced sound-induced seizures in GEPR and DBA/2 mice, as well as DHPG-induced seizures in rats. These data suggest that non-specific effects may be responsible for the convulsant action of L-SOP and L-AP4 [402]. Further supporting an anticonvulsant effect of group III mGluR agonism, (*R,S*)-PPG, an mGluR8-preferring agonist, was found to reduce sound-induced seizures in DBA/2 mice and GEPR, as well as electroshock-induced seizures in mice, with little evidence of any excitatory or pro-convulsant actions [402-404]. Moreover, the selective agonist for subtype 8 of group III mGluR, (*S*)-3,4-DCPG, was reported to reduce DL-homocysteic acid-induced seizures in immature rats and suppress generalized clonic-tonic seizures. Cortical energy metabolite changes associated with clonic-tonic seizures were also either normalized measured as a decrease of glucose and glycogen or

markedly reduced measured as an accumulation of lactate [405].

In summary, mGluRs modulation has shown antiepileptic activity in various animal models of generalized seizures, including sound-induced clonic seizures in DBA/2 mice, a model sensitive to all drugs clinically effective in primary generalised seizures. Suppression of SWD has been shown with some mGluR modulators in lethargic mice, a model predictive of clinical efficacy in absence seizures [406]. Spontaneous seizures following status epilepticus induced by KA or pilocarpine in rats are regarded as models of human temporal lobe epilepsy. In these models, the effects of mGluR modulators have not been thoroughly investigated and mixed results have been observed. In conclusion, the existing preclinical data positively support the therapeutic potential of mGluR ligands in epilepsy. However, studies using subtype specific agents in different models are still required.

CONCLUSION AND PERSPECTIVES

Earlier attempts to target ionotropic glutamate receptors for the treatment of central nervous system disorders have failed due to severe side effects that included, among others, cognitive and motor impairments. As outlined in this review, targeting glutamatergic neurotransmission through modulation of the mGluR family of receptors holds great promise for the treatment of a number of central nervous system disorders, with the advantage of potential fewer side effects. From the collection of evidence presented here, one can extract that the therapeutic effects of drugs targeting mGluRs involve, in the majority of cases, a reduction in the excitatory drive either through antagonism of Group I mGluRs or activation of Group II and III mGluRs. One clear exception is the use of mGluR5 positive modulators for the treatment of cognitive deficits associated with schizophrenia. Obviously, the final result will depend on whether the targeted glutamatergic pathways are directly or indirectly involved in the pathological condition of interest, and the clinical efficacy may reside in an indirect potentiation of GABAergic, dopaminergic or other neurotransmitter systems. In addition, targeting mGluRs may also have therapeutic actions independent of glutamatergic signalling altogether, since non-glutamatergic terminals also express mGluRs.

In recent years, an intense effort has been concentrated in the synthesis and characterization of novel, more selective drugs acting on selective subtypes of mGluRs. However, whereas the advantages of some compounds resides in their selectivity, in some cases a mixed Group I antagonism/Group II or III agonism may be the key for their effectiveness, and novel therapeutic approaches may benefit from the combined selective targeting of multiple mGluRs. Many of the drugs discussed here have shown promising performance in preclinical studies. However, there is still limited clinical evidence supporting the putative therapeutic benefit of mGluR modulation in the treatment of psychiatric and neurological disorders, with the exception of a positive clinical trial of the mGluR2/3 agonist, LY2140023, for the treatment of schizophrenia [113]. Several clinical trials of subtype-

selective mGluR modulators are ongoing for the treatment of schizophrenia, anxiety and depression among others. The outcome of these clinical studies will be met with great interest, since they will reveal whether targeting subtypes of mGluRs is a viable strategy for the treatment of various central nervous system disorders both in terms of therapeutic benefits as well as side-effect liability.

CONFLICT OF INTEREST

Declared none.

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