

A new species of the genus *Xenophrys* (Anura: Megophryidae) from northern Thailand

DEAR EDITOR,

Species of *Xenophrys* are conserved morphologically and live primarily in forests. In Thailand, the genus harbors many cryptic species. Herein we report the collection of specimens from Doi Inthanon, Chiang Mai Province, northern Thailand, which were identified previously as *X. minor*. Molecular and morphological analyses find that these specimens differ significantly from other known congeners, and therefore we describe a new species. Further, our phylogenetic analyses indicate that *X. latidactyla* is a junior synonym of *X. palpebralespinosa*.

The Indo-Burma and Sundaland biodiversity hotspots span Thailand and host a high diversity of amphibian species (Myers et al., 2000). In Thailand, at least 32 of 193 species of amphibians appear to be endemic (Frost, 2019). Some recent studies of *Megophrys sensu lato*, *Leptobranchella* and *Fejervarya* have shown that the diversity of amphibians in Thailand is underestimated (Chen et al., 2017, 2018; Suwannapoom et al., 2016). Further, 10 new species of amphibians have been discovered in the past three years alone (Frost, 2019; Pawangkhanant et al., 2018; Poyarkov et al., 2018; Suwannapoom et al., 2018). Thus, the extensively rich amphibian diversity in Thailand appears to remain underestimated. Like many other regions, high rates of deforestation owe to increased development (Royal Forest Department, 2006) and this drives a high risk of extinction even before the discovery of new species.

Genus *Xenophrys* Günther, 1864 (family Megophryidae) occurs in the southern and eastern Himalayas, Indochina, and northward to the Qinling and Huangshan mountains of mainland China (Chen et al., 2017). Of the 66 species, Thailand has seven species only: *X. aceras*, *X. lekaguli*, *X. longipes*, *X. major*, *X. minor*, *X. parva* and *X. takensis* (Chan-

ard, 2003; Mahony, 2011; Nutphund, 2001; Stuart et al., 2006). *Xenophrys minor* Stejneger was described originally from Kwanghsien (now Guan Xian, Dujiangyan City), 55 kilometers northwest of Chengtu (Chengdu), Szechwan (Sichuan), China. Several researchers recorded it for montane areas of northern Thailand (Chan-ard, 2003; Chuaynkern & Chuaynkern, 2012; Nabhitabhata et al., 2000; Nabhitabhata and Chan-ard, 2005). Several recent publications revised the *X. minor* complex (Li et al., 2014; Mahony et al., 2013; Wang et al., 2012). Further, the type locality occurs in Sichuan, China and a distance of over 1 500 km separates them from the known localities in northern Thailand. Thus, the taxonomic status of Thai populations previously referred to as *X. minor* requires further investigations.

Our recent fieldwork in Thailand resulted in the collection of specimens of *Xenophrys* cf. "*minor*" from Chiang Mai Province (Doi Inthanon). Further phylogenetic analyses of mtDNA sequences and morphological examinations showed that this species is distantly related to *X. minor* from China and can be distinguished from all known congeners both by molecular and morphological characters. Based on an integrative taxonomic approach, we describe this population as a new

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species of the genus *Xenophrys*.

Six individuals were collected from Doi Inthanon, Chiang Mai Province, Thailand in August 2017 (Figure 1). After specimens were euthanized using benzocaine, liver tissue was taken and preserved in 95% alcohol. The specimens were fixed with 10% formalin for 24 hours and subsequently transferred to 70% ethanol. All specimens were deposited in the herpetological collection of the Museum of the Kunming Institute of Zoology (KIZ), Chinese Academy of Sciences (CAS) and University of Phayao (AUP). Six preserved adult specimens were examined and measured to the nearest 0.1 mm using digital calipers. Morphological terminology followed Fei et al. (2009) and Poyarkov et al. (2017) (Supplementary Methods).

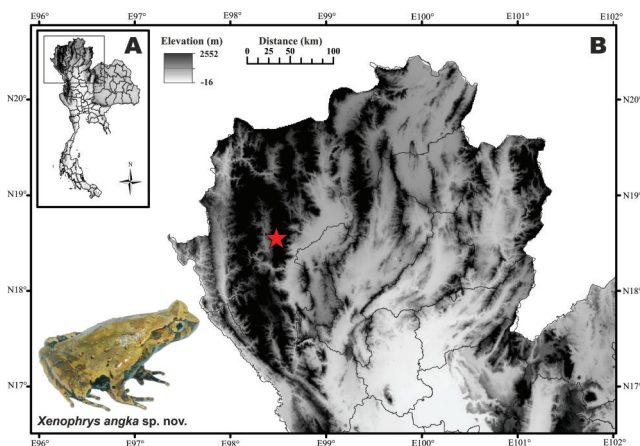


Figure 1 Known distribution of *Xenophrys angka* sp. nov. from Doi Inthanon, Chiang Mai Province, Thailand

A: Map of Thailand; B: Inset map of northern Thailand. Star shows the type locality of the new species. Photo by N.A. Poyarkov.

Whole genomic DNA was extracted, and a partial fragment of the mitochondrial 16S rRNA were amplified and sequencing. DNA extraction, primers and PCR cycle protocols are in Supplementary Methods. Matrilineal genealogies were reconstructed to study the phylogenetic relationships among *Xenophrys* based on the partial mitochondrial 16S rRNA gene. Homologous sequences of related species of *Xenophrys*, and those of the outgroups *Leptobranchella oshanensis*, *L. ventripunctata*, *Leptobranchium boringii*, *Megophrys nasuta*, and *M. baluensis*, were downloaded from GenBank (Supplementary Table S1). Trees were reconstructed using maximum likelihood (ML) and Bayesian inference (BI) (Supplementary Methods). Apart from tree-based methods, we also calculated row pairwise sequence divergence using uncorrected p-distances implemented in MEGA v6.0.6 (Tamura et al., 2013).

The results of the ML and BI analyses yielded essentially identical topologies with relatively high nodal support values for most terminal nodes (Figure 2). The tree resolved monophyly of *Xenophrys* with two major lineages, corresponding to the subgenera *Xenophrys sensu stricto* and *Panophrys* (Figure 2). *Xenophrys* cf. "*minor*" from Chiang Mai

Province assigned to *Panophrys* and phylogenetic relationships within which remained essentially unresolved. The Chiang Mai population of *Xenophrys* cf. "*minor*" was a strongly supported lineage (BPP=1, BS=100; Figure 2), which different notably from *X. minor sensu stricto* from China (Figure 2). Genetic distance between the Chiang Mai population and other species of *Xenophrys* ranged from 4.5% (*X. rubrimera*, subgenus *Panophrys*) to 14.4% (*X. mangshanensis*, subgenus *Xenophrys*) (Supplementary Table S2). The tree nested *X. latidactyla* within the radiation of *X. palpebralespinosa* with high support (BPP=1.0, BS=94) (Figure 2). The genetic divergence between *X. latidactyla* and *X. palpebralespinosa* was 0.9%.

The Chiang Mai population of *Xenophrys* cf. "*minor*" differs in a number of taxonomically important diagnostic characters from other congeners, including *X. minor* from China. Thus, both mtDNA and morphological analyses clearly indicate that this population represents a separately evolving lineage and an undescribed species, which we describe below.

Taxonomic account

Xenophrys angka sp. nov.

Figures 3–4; Table 1.

Chresonymy: *Xenophrys minor* (partim) — Chan-ard, 2003; Nabhitabhata and Chan-ard, 2005; Nabhitabhata et al., 2000; Chuaynkern and Chuaynkern, 2012.

Holotype: KIZ040591, an adult female collected from Kiew Mae Pan nature trail in Doi Inthanon, Chiang Mai Province, Thailand (N18.556187°, E98.482229°; elevation 2 190 m a.s.l.), collected by Chatmongkon Suwannapoom, Parinya Pawangkhanant and Nikolay A. Poyarkov on 29, August, 2017.

Paratypes: Two males KIZ040595 and AUP-00077, three females KIZ040592, AUP-00076 and AUP-00055; collected at the same locality and same collection information as the holotype.

Etymology: The specific epithet "*angka*" is given as a noun in apposition and refers to the name of the highest mountain of Thailand, Doi Angka, located in the Doi Inthanon, Chiang Mai Province, Thailand, where the new species occurs.

Diagnosis: *Xenophrys angka* sp. nov. is a member of genus *Xenophrys* based on the following combination of morphological characters: head large, somewhat narrow and comparatively non-compressed, angular; tympanum distinct; vertical pupil; transverse skin fold at head basis absent; no large horny spines on dorsum; and two narrow glandular mid-dorsal ridges present, forming X, H or Y-shaped figure (Chen et al., 2017). The following combination of characters diagnoses the new species: (1) small body size, adult snout-vent length (SVL) 31.2–32.1 mm in males ($n=2$), 37.5–39.2 mm in females ($n=4$); (2) tympanum distinct and circular; (3) vomerine ridges indistinct and vomerine teeth absent; (4) maxillary teeth present; (5) tongue heart-shaped, not

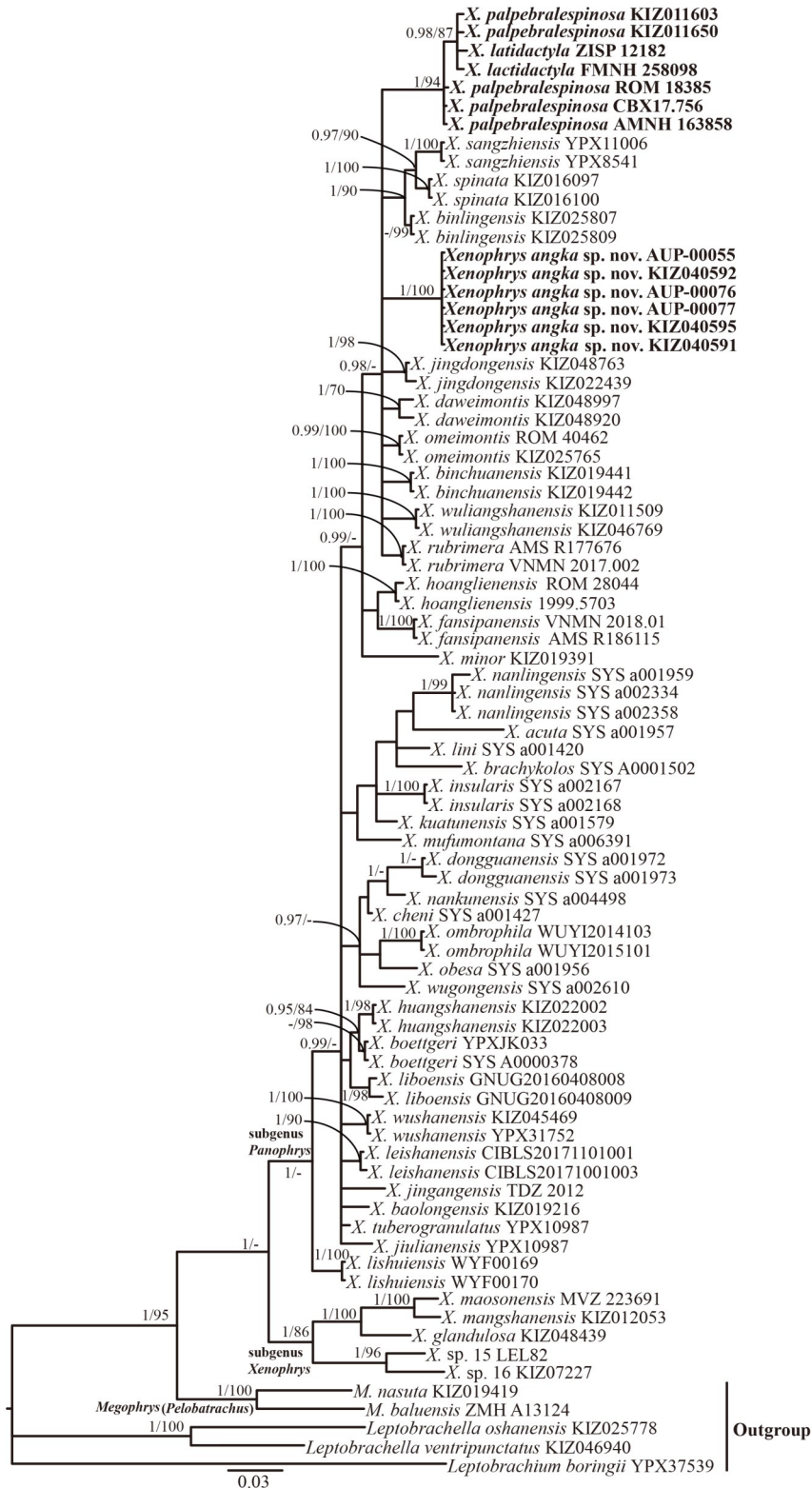


Figure 2 BI tree resulting from 541 bp length fragment of mitochondrial 16S rRNA gene for *Xenophrys* species and outgroups Bayesian posterior probabilities (BPP)>95%/ML inferences (ML-BS)>70% are shown for each node. Hyphen (“-”) denotes a Bayesian posterior probabilities<95% and bootstrap support<70%.

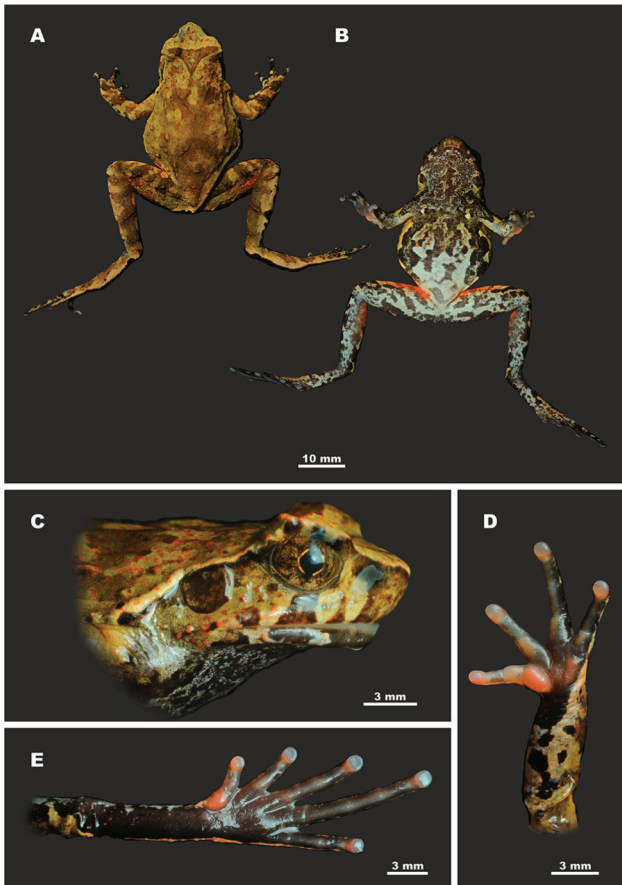


Figure 3 Holotype *Xenophrys angka* sp. nov. (KIZ040591, field number AUP-00074, female) in life

A: Dorsal view; B: Ventral view; C: Head, lateral view; D: Volar view of the left hand; E: Plantar view of the left foot. Photos by N.A. Poyarkov.

notched posteriorly; (6) supratympanic fold distinct, extending from the posterior corner of eye to shoulder; (7) webbing between toes rudimentary; (8) lateral fringes on toes absent; (9) tibio-tarsal articulation reaching the area between eye and snout tip; (10) nuptial pads present on finger I; (11) subarticular tubercles present on the base of fingers I–II, but absent on fingers III–IV; (12) subarticular tubercle present at base of toes I, absent on toes II–IV; (13) heels meeting or overlapping when tibiae positioned at right angle to body axis; (14) inner metatarsal tubercle big, outer metatarsal tubercle absent; (15) protruding projection posterior to cloaca of males present; (16) dorsal surface with a complete dark brown interorbital triangle with light blotch in the middle and two distinct thin opposing "V" -shaped reddish glandular ridges with ridge on dorsum; (17) orange coloration of groin contrasting with surrounding regions on males; and (18) inner metacarpal tubercle and outer metatarsal tubercles distinct reddish color.

Description of holotype (measurements in Table 1): KIZ040591, sexually mature female, body habitus stocky (Figure 3), body size small (SVL=38.2 mm); head large (head

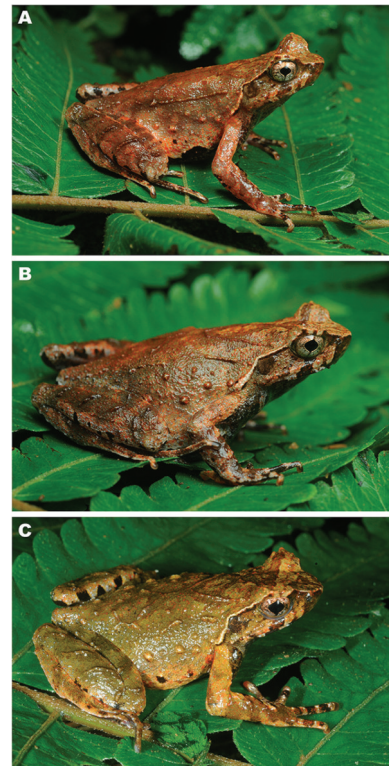


Figure 4 Type series of *Xenophrys angka* sp. nov. in life

A: Holotype KIZ040591, female; B: Paratype AUP-00055, female; C: Paratype AUP-00077, male. Photos by N. A. Poyarkov and C. Suwannapoom.

length (HDL)/SVL 33.8%, maximum head width (HDW)/SVL 32.2%), slightly longer than wide (HDW/HDL 95.3%); triangular in dorsal view; top of head flat; snout short (snout length (SNT)/HDL 32.6%) and wide (the distance between anterior orbital borders (IFE)/HDW 50.4%), snout obtusely pointed in dorsal view (Figure 3A), sharply protruding in profile, without rostral appendage, notably projecting beyond lower jaw (Figure 3C); loreal region vertical and concave; canthus rostralis distinct, sharp; dorsal region of snout flattened; eyes large (eye diameter (ED)/HDL 31.8%); slightly protuberant in dorsal view and notably protruding in profile (Figure 3), eye less than twice as long as maximum tympanum diameter (tympanum diameter (TD)/ED 56.1%) and subequal to snout length (ED/SNT 97.6%); eye-tympanum distance less than maximum tympanum diameter (tympanum–eye distance (TED)/TD 87.0%); tympanum distinct, circular in shape, relatively small (TD/HDL 17.8%), eye diameter notably larger than tympanum diameter (ED/TD 178.3%); nostril rounded, laterally orientated, nostril closer to anterior corner of eye than to tip of snout (distance from nostril to eye (DNE)/snout–nostril distance (SN) 85.7%); internarial distance equal to width of upper eyelid (internarial distance (IND)/width of upper eyelid (UEW) 100.0%), slightly larger than interorbital distance (IND/interorbital distance (IOD) 103.0%); pineal ocellus not visible externally (Figure 3A); tongue heart-

Table 1 Measurements (in mm) of type series of *Xenophrys angka* sp. nov.

	KIZ 040591*	KIZ 040592	AUP- 00076	AUP- 00055	KIZ 040595	AUP- 00077
Sex	F	F	F	F	M	M
SVL	38.2	37.5	38.2	39.2	32.1	31.2
HDL	12.9	12.8	12.7	13.0	11.3	11.5
HDW	12.3	12.6	12.4	12.6	10.5	11.0
SNT	4.2	4.4	4.0	4.1	3.5	3.7
DNE	1.8	1.7	1.7	1.8	1.4	1.5
IND	3.4	3.2	3.7	3.7	3.1	3.3
IOD	3.3	3.2	3.2	3.5	3.0	2.7
UEW	3.4	3.1	3.2	3.2	2.5	2.9
ED	4.1	4.0	4.2	4.1	3.9	3.7
TD	2.3	1.8	1.8	2.3	1.7	1.8
TED	2.0	1.4	1.4	2.1	1.5	1.3
SN	2.1	2.5	2.4	2.2	2.0	2.1
IFE	6.2	6.4	6.6	6.6	5.3	5.3
IBE	10.4	10.1	10.0	9.6	9.0	9.2
FAL	7.8	8.0	7.4	8.6	6.4	6.4
HL	9.4	9.7	9.7	9.3	8.7	8.6
THL	17.6	18.5	18.1	17.6	14.8	14.0
TL	18.7	18.8	18.7	18.6	16.0	15.5
FL	15.7	16.8	17.2	16.5	15.6	14.3
IMTL	1.6	2.4	1.9	1.7	1.4	1.7
FLI	3.6	3.5	3.6	3.8	3.5	3.9
FLII	4.1	4.1	4.1	4.2	3.8	4.1
FLIII	6.2	6.3	6.4	6.5	6.0	6.5
FLIV	4.4	4.2	4.2	4.4	4.1	4.3
TLI	2.1	2.3	2.2	2.2	2.4	2.3
TLII	4.2	4.4	5.0	4.6	4.2	3.9
TLIII	7.0	6.1	6.9	7.0	6.0	5.6
TLIV	9.8	10.1	10.4	10.4	8.7	8.2
TLV	5.3	5.5	5.6	5.1	4.7	4.0

For abbreviations, see text and Supplementary Methods. Asterisk (*) indicates holotype. F: Female; M: Male.

shaped, not notched posteriorly; vomerine ridges indistinct and vomerine teeth absent; maxillary teeth present; pupil diamond-shaped (Figure 3C), vertical.

Forelimbs moderately long and robust; forearm not enlarged, length shorter than hand length (forearm length

(FAL)/hand length (HL) 83.0%); fingers long and narrow, not flattened dorsoventrally, lateral fringes on fingers absent, relative finger lengths: I<II<IV<III; tips of all fingers rounded, slightly dilated relative to digit widths, with circular pads, terminal grooves absent; no webbing between fingers; a large subarticular tubercle present at base of fingers I–II, absent on fingers III – IV; more distal subarticular tubercles absent, replaced by low callous dermal ridges; supernumerary tubercles absent; inner metacarpal tubercle big, oval, outer metacarpal tubercle small, flattened (Figure 3D).

Hindlimbs long and robust, thigh length shorter than tibia length (thigh length (THL)/tibia length (TL) 94.1%), but greater than foot length (THL/foot length (FL) 112.1%); tibio-tarsal articulation of straightened limb reaching eye level; heels slightly overlapping when tibias positioned at right angles to body axis; toes long and slightly dorsoventrally flattened, relative toes lengths: I<II<V<III<IV; tips of all toes rounded, slightly dilated, terminal grooves absent; notably expanded relative to digit widths forming circular pads; terminal grooves absent; lateral dermal fringes on absent; rudimentary webbing present between all toes; tarsal fold absent; subarticular tubercle present at base of toes I, absent on toes II – IV, replaced by indistinct callous dermal ridges; inner metatarsal tubercle large, ca. 1.5x longer than wide, oval-shaped; and outer metatarsal tubercle absent (Figure 3E).

Dorsal surface of body and both dorsal and lateral surfaces of head weakly granular; several large distinct warts scattered on flanks; horn-like tubercle and several smaller tubercles at edge of eyelids present; and supratympanic fold distinct, glandular, starting immediately at posterior corner of upper eyelid (palpebrum) and running posteriorly towards dorsal edge of tympanum, where it sharply curves ventrally becoming more prominent and swollen and gently continues towards axilla. Dorsolateral folds well-developed, glandular, almost straight, running from scapular region posteriorly towards sacral region becoming less distinct and interrupted posteriorly; two opposing "V"-shaped glandular skin folds present on dorsum joined by a ca. 10 mm long dorsomedial fold in a hourglass-shape (Figure 3A); dorsal surfaces of limbs with small tubercles forming distinct transverse skin folds on hindlimbs and irregular reticulate folds on forelimbs; ventral surfaces of limbs, chest, abdomen and throat smooth; pectoral glands prominent, rounded, located closely to axilla (Figure 3D); femoral glands small, oval-shaped, positioned on posterior surface of thighs closer to groin than to the knee.

Color of the holotype in life: Coloration of holotype in life shown in Figure 3 and Figure 4A. In life, dorsal surface light brown with olive green tint, with a complete dark brown inverted triangle with light-ochre central blotch present between eyes; all small tubercles and glandular ridges on dorsal surfaces of head and body reddish-brown, hourglass-shaped dorsal glandular skin ridges also reddish brown surrounded with darker grey-brown "X"-shaped marking with indistinct borders; flanks with irregular dark brown spots and reddish mottling getting denser ventrally (Figure 4A); lateral surfaces of head light brownish-gray to beige, upper lips with

3 to 4 dark reddish-brown vertical bars, the one below eye largest and most distinct; tympanum entirely dark brown; white supratympanic fold with black to dark-brown lower margin; loreal region dark-brown; the edge of upper eyelid (palpebrum) with 3 distinct reddish palpebral tubercles, the medial one largest forming horn-like projection; dorsally palpebral tubercles edged with dark brown; dorsal parts of limbs reddish-brown with dark brown crossbars and reddish mottling; dorsal surface of the fingers with dark brown crossbars; ventral surfaces of body and limbs primarily off-white to grey, mottled with dense black and white blotches and flecks on belly; black and white marbling getting denser anteriorly; gular and chest dark grey and mottled with white; posterior part of belly off-white with large grey blotches; ventral surfaces of hindlimbs with contrasting black and white marbling; groin area, anterior surfaces of thighs and posterior surfaces of heels bright reddish-orange in life; pectoral and femoral glands small, cream-white in color; iris golden-bronze with dense black mottling; pupil edged with thin golden line; thenar and palmar surface of limbs dark grey; and inner metacarpal tubercle, outer metacarpal tubercle, and inner metatarsal tubercle bright pink.

Color of the holotype in preservative: For coloration see Supplementary Figure S1. After 2 years in preservative, dorsum faded to dark brown; a complete dark brown inverted triangle between eyes and crossbars on dorsal limbs and fingers still clear; two opposing "V"-shaped skin folds present on dorsum becoming less distinct; throat and chest turns dark brown; abdomen and ventral surface of limbs dark brown mottled with whitish-grey marbling; inner metacarpal tubercle and inner metatarsal tubercle become off-white, outer metacarpal tubercle brown; dark brown vertical bars present upper lips still clear; supratympanic folds dark brown; pectoral glands and femoral glands still distinct, cream; ventral side of the hands and toes brown and digit tips brown or cream.

Variation: Morphometric variation of types reported in Table 1. All 5 paratypes are very similar in morphology the holotype, but also show variation in color in life. Dorsal surface of holotype and paratype AUP-00055 light brown, but paratype AUP-00077 orange (Figure 4).

Male secondary sexual characteristics: The new species shows slight differences in body size between the sexes: females have slightly larger SVL (37.5–39.2 mm, mean 38.3 mm; $n=4$) than males (31.2–32.1 mm, mean 31.7 mm; $n=2$). All adult males have nuptial pads pinkish-red in life covering the dorsal surface of the base of FI. Male has external, single subgular vocal sac with slit-like openings at posterior corners of jaws and a prominent protruding projection posterior to cloaca (Figure 4).

Distribution and ecology: *Xenophrys angka* sp. nov. is currently known only from Doi Inthanon, Chiang Mai province, Thailand (Figure 1) at elevations from 1800 to 2200 m a.s.l. along forest cascade streams and waterfalls. Most males were

located by calls made while sitting on vegetation 1–2 m from the stream; females were recorded in forest litter along the streams. *Xenophrys angka* sp. nov. appears to be a strict forest specialist, restricted to patches of undisturbed montane evergreen forests and is likely endemic to the Doi Inthanon – Thanon Thong Chai Range. It inhabits forest floor, leaf litter and the nearby undergrowth rocky mountainous surrounded by moist evergreen broadleaved forests. The new species was found in sympatry with *Leptobrachium huashen* Fei et Ye and *Limnonectes taylori* Matsui, Panha, Khonsue et Kuraishi.

Comparisons: We compared *Xenophrys angka* sp. nov. with their 66 known congeners on the basis of morphology. Comparisons with each subgenus are discussed separately below.

Subgenus *Panophrys*: *Xenophrys angka* sp. nov. differs from the following large-sized species by having a smaller adult male size, SVL 31.2–32.1 mm (vs. adult-male 42.0–45.0 mm, $n=5$, in *X. baolongensis*, Ye et al., 2007; 45.0–51.0 mm, $n=3$, in *X. binlingensis*, Fei et al., 2009; 81.3 mm, $n=1$, in *X. caudoprocta*, Fei et al., 2009; 53.0–56.5 mm, $n=2$, in *X. jingdongensis*, Fei et al., 2009; 56.0–59.5 mm, $n=10$, in *X. omeimontis*, Fei et al., 2009; 54.7 mm, $n=1$, in *X. sangzhiensis*, Jiang et al., 2008; 99.8–115.6 mm, $n=6$, in *X. shuichengensis*, Tian et al., 2000; and 47.2–54.4, $n=18$, in *X. spinata*, Fei et al., 2009); from *X. liboensis* by having a smaller adult female body size, SVL 37.5–39.2 mm (vs. 60.8–70.6 mm, $n=8$, in *X. liboensis*, Zhang et al., 2017); further from *X. acuta* by meeting or overlapping when tibias positioned at right angle to body axis (vs. not meeting in *X. acuta*, Li et al., 2014), and head slightly longer than wide (vs. head length slightly shorter than head width in *X. acuta*, Li et al., 2014); from *X. binchuanensis* by lateral dermal fringes on toes absent (vs. lateral dermal fringes on toes wide in *X. binchuanensis*, Fei et al., 2009), and horn-like tubercle above eyelids present (vs. absent in *X. binchuanensis*, Fei et al., 2009); from *X. boettgeri* by tongue not notched behind (vs. tongue notched behind in *X. boettgeri*, Fei et al., 2009), lateral dermal fringes on absent (vs. present in *X. boettgeri*, Fei et al., 2009), and male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. boettgeri*, Fei et al., 2009); from *X. brachykolos* by head length longer than wide (vs. head width larger than head length in *X. brachykolos*, Fei et al., 2009), male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. brachykolos*, Fei et al., 2009), and heels meeting or overlapping (vs. not meeting in *X. brachykolos*, Fei et al., 2009); from *X. cheni* by lateral fringes on fingers and toes absent (vs. lateral fringes on fingers and toes wide in *X. cheni*, Wang et al., 2014), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. subarticular tubercles indistinct in *X. cheni*, Wang et al., 2014), golden-bronze (vs. iris dark brown in *X. cheni*, Wang et al., 2014), and tongue not notched behind (vs. margin of tongue notched behind in *X. cheni*, Wang et al., 2014); from *X. daweimontis* by vomerine ridges indistinct and vomerine teeth absent (vs. present in *X. daweimontis*, Rao & Yang, 1997),

and male with external single subgular vocal sac (vs. male with internal vocal sac in *X. daweimontis*, Rao & Yang, 1997); from *X. dongguanensis* and *X. nankunensis* by vomerine ridges indistinct and vomerine teeth absent (vs. strong vomerine ridge bearing vomerine teeth in *X. dongguanensis* and *X. nankunensis*, Wang et al., 2019), subarticular tubercle present at base of toes I, absent on toe II–IV (vs. subarticular tubercles only present at base of each toe in *X. dongguanensis* and *X. nankunensis*, Wang et al., 2019), and heels meeting or overlapping (vs. heels not meeting in *X. dongguanensis* and *X. nankunensis*, Wang et al., 2019); from *X. fansipanensis* and *X. hoanglienensis* by vomerine ridges indistinct and vomerine teeth absent (vs. present in *X. fansipanensis* and *X. hoanglienensis*, Tapley et al., 2018), inner and outer metacarpal tubercles present (vs. absent in *X. fansipanensis* and *X. hoanglienensis*, Tapley et al., 2018), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. absent in *X. fansipanensis* and *X. hoanglienensis*, Tapley et al., 2018), inner metatarsal tubercle large (vs. inner metatarsal tubercle very weakly in *X. fansipanensis* and *X. hoanglienensis*, Tapley et al., 2018), and protruding projection posterior to cloaca of males present (vs. absent in *X. fansipanensis* and *X. hoanglienensis*, Tapley et al., 2018); from *X. huangshanensis* by tympanum distinct (vs. indistinct in *X. huangshanensis*, Fei et al., 2009), male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. huangshanensis*, Fei et al., 2009), subarticular tubercle present at base of toes I, absent on toe II–IV (vs. present in *X. huangshanensis*, Fei et al., 2009), and heels meeting or overlapping (vs. heels not meeting in *X. huangshanensis*, Fei et al., 2009); from *X. insularis* by vomerine ridges indistinct and vomerine teeth absent (vs. vomerine ridge strong with vomerine teeth in *X. insularis*, Wang et al., 2017), subarticular tubercle present at base of toes I, absent on toe II–IV (vs. subarticular tubercle only present at base of each toe in *X. insularis*, Wang et al., 2017), heels meeting or overlapping (vs. not meeting in *X. insularis*, Wang et al., 2017), and tibio-tarsal articulation reaching area between eye and snout tip (vs. reaching forward to posterior edge of tympanum in *X. insularis*, Wang et al., 2017); from *X. jinggangensis* by subarticular tubercle present at base of toes I, absent on toe II–IV (vs. a large subarticular tubercle at base of each toe in *X. jinggangensis*, Wang et al., 2012), relative finger lengths: I<II<IV<III (vs. II<I<IV<III in *X. jinggangensis*, Wang et al., 2012), and lateral fringes on fingers absent (vs. present in *X. jinggangensis*, Wang et al., 2012); from *X. julianensis* by tongue not notched posteriorly (vs. tongue weakly notched posteriorly in *X. julianensis*, Wang et al., 2019), subarticular tubercle present at base of toes I, absent on toe II–IV (vs. subarticular tubercles only present at base of toe I and II in *X. julianensis*, Wang et al., 2019); from *X. kuatunensis* by male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. kuatunensis*, Fei et al., 2009), and heels meeting or overlapping (vs. not meeting in *X. kuatunensis*, Fei et al., 2009); from *X. leishanensis* by male with external single subgular vocal sac (vs. male with internal

single subgular vocal sac in *X. leishanensis*, Li et al., 2018), relative finger lengths: I<II<IV<III (vs. II<I<V<III in *X. leishanensis*, Li et al., 2018), and head slightly longer than wide (vs. head width slightly larger than head length in *X. leishanensis*, Li et al., 2018); from *X. lini* by subarticular tubercle present at base of toes I, absent on toe II–IV (vs. subarticular tubercle distinct at base of each toe in *X. lini*, Wang et al., 2014), and lateral dermal fringes on fingers and toes absent (vs. wide in *X. lini*, Wang et al., 2014); from *X. lishuiensis* by male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. lishuiensis*, Wang et al., 2017); from *X. minor* by a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. absent in *X. minor*, Fei et al., 2009), subarticular tubercle present at base of toes I, absent on toe II–IV (vs. absent in *X. minor*, Fei et al., 2009), lateral dermal fringes on toes absent (vs. absent in *X. minor*, Fei et al., 2009), male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. minor*, Fei et al., 2009), and horn-like tubercle above eyelids present (vs. absent in *X. minor*, Fei et al., 2009); from *X. mufumontana* by relative finger lengths: I<II<IV<III (vs. II=IV<I<III in *X. mufumontana*, Wang et al., 2019), subarticular tubercle present at base of toes I, absent on toe II–IV (vs. subarticular tubercles only present at base of each toe in *X. mufumontana*, Wang et al., 2019), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. presence of a subarticular tubercle at base of each finger in *X. mufumontana*, Wang et al., 2019), and outer metacarpal tubercle small, flattened (vs. indistinct in *X. mufumontana*, Wang et al., 2019); from *X. nanlingensis* by vomerine ridges indistinct and vomerine teeth absent (vs. vomerine ridge and vomerine teeth present in *X. nanlingensis*, Wang et al., 2019), tongue not notched posteriorly (vs. tongue notched posteriorly in *X. nanlingensis*, Wang et al., 2019), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. presence of a subarticular tubercle at base of each finger in *X. nanlingensis*, Wang et al., 2019), subarticular tubercle present at base of toes I, absent on toe II–IV (vs. presence of a subarticular tubercle at base of each toe in *X. nanlingensis*, Wang et al., 2019), and adult males have nuptial pads pinkish-red in life covering dorsal surface of base of FI (vs. nuptial pads and nuptial spines invisible in males during breeding season in *X. nanlingensis*, Wang et al., 2019); from *X. obesa* by vomerine ridges indistinct (vs. two vomerine ridges moderately developed in *X. obesa*, Li et al., 2014), heels meeting or overlapping (vs. not meeting in *X. obesa*, Li et al., 2014), and head slightly longer than wide (vs. head width slightly larger than head length in *X. obesa*, Li et al., 2014); from *X. ombrophila* by heels meeting or overlapping (vs. not meeting in *X. ombrophila*, Messenger et al., 2019), and tibio-tarsal articulation reaching area between eye and snout tip (vs. reaching posterior corner of eye in *X. ombrophila*, Messenger et al., 2019); from *X. palpebralespinosa* by vomerine teeth absent (vs. present in *X. palpebralespinosa*, Fei et al., 2009), lateral dermal fringes on fingers and toes absent (vs. slight lateral fringes on fingers, lateral fringes wide on toes in *X. palpebralespinosa*, Fei et al.,

2009), and male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. palpebralespinosa*, Fei et al., 2009); from *X. rubrimera* by vomerine teeth absent (vs. present in *X. rubrimera*, Tapley et al., 2017), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. absence of subarticular tubercles on fingers in *X. rubrimera*, Tapley et al., 2017), and head slightly longer than wide (vs. head width slightly larger than head length in *X. rubrimera*, Tapley et al., 2017); from *X. tuberogranulata* by male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. tuberogranulata*, Mo et al., 2010), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. indistinct in *X. tuberogranulata*, Mo et al., 2010), and relative finger lengths: I<II<IV<III (vs. II<IV=I<III in *X. tuberogranulata*, Mo et al., 2010); from *X. wugongensis* by heels meeting or overlapping (vs. not meeting in *X. wugongensis*, Wang et al., 2019), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. presence of a subarticular tubercle at base of each finger in *X. wugongensis*, Wang et al., 2019), subarticular tubercle present at base of toes I, absent on toe II–IV (vs. presence of a subarticular tubercle at base of each toe in *X. wugongensis*, Wang et al., 2019); from *X. wuliangshanensis* by a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. indistinct in *X. wuliangshanensis*, Fei et al., 2009), male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. wuliangshanensis*, Fei et al., 2009), horn-like tubercle at edge of eyelids present (vs. absent in *X. wuliangshanensis*, Fei et al., 2009), and relative finger lengths: I<II<IV<III (vs. IV<III<II=I in *wuliangshanensis*, Fei et al., 2009); from *X. wushanensis* by horn-like tubercle and some smaller tubercles at edge of eyelids present (vs. absent in *X. wushanensis*, Fei et al., 2009), male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. wushanensis*, Fei et al., 2009), and lateral dermal fringes on toes absent (vs. male with wide lateral fringes in *X. wushanensis*, Fei et al., 2009).

Subgenus *Xenophrys*: *Xenophrys angka* sp. nov. differs from the following large-sized species by having a smaller adult male size, SVL 31.2–32.1 mm (vs. adult-male 55.8–62.4 mm, $n=6$, in *X. aceras*, Wang et al., 2017; 39.1–45.0 mm, $n=8$, in *X. ancræ*, Mahony et al., 2013; 76.7 mm, $n=20$, in *X. auralensis*, Ohler., 2002; 57.1 mm, $n=1$, in *X. damrei*, Mahony, 2011; 56.9–68.4 mm, $n=4$, in *X. flavipunctata*, Mahony et al., 2018; 76.3–81.0 mm, $n=10$, in *X. glandulosa*, Fei et al., 2009; 68.0–73.5 mm, $n=7$, in *X. himalayana*, Mahony et al., 2018; 55.6–66.6 mm, $n=8$, in *X. lekaguli*, Stuart et al., 2006; 47.0 mm, $n=1$, in *X. longipes*, Wang et al., 2017; 65.5 mm, $n=1$, in *X. major*, Fei et al., 2009; 62.5 mm, $n=1$, in *X. mangshanensis*, Fei et al., 2009; 58.0–76.0 mm, $n=6$, in *X. maosonensis*, Yang et al., 2018; 57.2–68.0 mm, $n=16$, in *X. medogensis*, Fei et al., 2009; 71.3–93.8 mm, $n=12$, in *X. periosa*, Mahony et al., 2018; 73.5–83.1 mm, $n=6$, in *X. robusta*, Mahony et al., 2018; 45.9–53.4 mm, $n=7$, in *X. megacephala*, Mahony et al., 2011; 47.4–53.0 mm, $n=33$, in *X. takensis*, Mahony, 2011); from *X.*

oreocrypta by having a smaller adult female body size, SVL 37.5–39.2 mm (vs. 94.9 mm, $n=1$, in *X. oreocrypta*, Mahony et al., 2018); further from *X. monticola* by vomerine ridges indistinct (vs. distinct in *X. monticola*, Mahony et al., 2018), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. absent in *X. monticola*, Mahony et al., 2018), inner and outer metacarpal tubercles distinct (vs. absent in *X. monticola*, Mahony et al., 2018), and inner metatarsal tubercle large (vs. indistinct in *X. monticola*, Mahony et al., 2018); from *X. oropedion* by vomerine ridges indistinct and vomerine teeth absent (vs. present in *X. oropedion*, Mahony et al., 2013), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. absent in *X. oropedion*, Mahony et al., 2013), protruding projection posterior to cloaca of males present (vs. absent in *X. oropedion*, Mahony et al., 2013), and inner and outer metacarpal tubercles distinct (vs. absent in *X. oropedion*, Mahony et al., 2013); from *X. pachyproctus* and *X. parva* by vomerine ridges indistinct and vomerine teeth absent (vs. present in *X. pachyproctus* and *X. parva*, Fei et al., 2009), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. absent in *X. pachyproctus* and *X. parva*, Fei et al., 2009), and male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. pachyproctus* and *X. parva*, Fei et al., 2009); from *X. serchhipii* by toes with rudimentary webbing (vs. at least one-fourth webbing in *X. serchhipii*, Mathew & Sen, 2007), relative finger lengths: I<II<IV<III (vs. 2nd and 4th fingers subequal in length in *X. serchhipii*, Mathew & Sen, 2007), and vomerine teeth absent (vs. present in *X. serchhipii*, Mathew & Sen, 2007); from *X. vegrandis* by a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. absent in *X. serchhipii*, Mathew & Sen, 2007), protruding projection posterior to cloaca of males present (vs. absent in *X. serchhipii*, Mathew & Sen, 2007), and pupil vertical, iris golden-bronze (vs. pupil horizontally orientated, iris metallic yellowish-orange in *X. serchhipii*, Mathew & Sen, 2007); from *X. zhangji* by vomerine ridges indistinct and vomerine teeth absent (vs. present in *X. zhangji*, Fei et al., 2009), a large subarticular tubercle present at base of fingers I–II, absent on fingers III–IV (vs. absent in *X. zhangji*, Fei et al., 2009), lateral dermal fringes on toes absent (vs. present in *X. zhangji*, Fei et al., 2009), and male with external single subgular vocal sac (vs. male with internal single subgular vocal sac in *X. zhangji*, Fei et al., 2009); from *X. zunhebotoensis* by horn-like tubercle and a number of smaller tubercles at edge of eyelids present (vs. absent in *X. zunhebotoensis*, Mathew & Sen, 2007), tongue not notched behind (vs. tongue notched behind in *X. zunhebotoensis*, Mathew & Sen, 2007), and vomerine teeth absent (vs. present in *X. zunhebotoensis*, Mathew & Sen, 2007).

From species not yet assigned to a subgenus: *Xenophrys angka* sp. nov. differs from *X. feii* by lateral dermal fringes on toes absent (vs. lateral fringes on toes moderate to wide in *X. feii*, Yang et al., 2018), lateral fringes on fingers absent (vs. moderate lateral fringes present on outer most three fingers in *X. feii*, Yang et al., 2018), groin coloration in life contrasting

with surrounding regions on males (vs. groin coloration not contrasting with surrounding regions on males in *X. feii*, Yang et al., 2018), and nuptial pads present on finger I (vs. absent in *X. feii*, Yang et al., 2018).

The diversity of *Xenophrys* is underestimated greatly (Chen et al., 2017; Liu et al., 2018). In the last three years, 19 new species were described mostly from China, Vietnam and India (Li et al., 2018; Mahony et al., 2018; Messenger et al., 2019; Tapley et al., 2017, 2018; Wang et al., 2017; Wang et al., 2017; Wang et al., 2019; Yang et al., 2018; Zhang et al., 2017). Compared to other countries in the Indochinese region, the diversity of *Xenophrys* in Thailand attracted less attention and no new species were described for the country within the last seven years. Herein, we describe a new species of *Xenophrys* from northern Thailand based on morphological and molecular analyses. Our discovery increases the number of amphibian species recorded in Thailand to 194, and the species number of *Xenophrys* to 27. The diversity of amphibians known from Thailand has increased remarkably from 125 (Khonsue and Thirakhuat, 2001) to 193 (Frost, 2019) and the diversity remains underestimated (Chan et al., 2018; Chen et al., 2017, 2018; Grismer et al., 2016; Laopichienpong et al., 2016; Matsui et al., 2018; Pawangkhanant et al., 2018; Poyarkov et al., 2018; Sheridan & Stuart, 2018; Suwannapoom et al., 2017; Suwannapoom et al., 2018). Further field surveys and taxonomic studies on the Thai herpetofauna will likely result in further discoveries of yet unknown lineages and species of amphibians.

Our study also elucidates taxonomic status of *Xenophrys latidactyla* (Orlov et al., 2015), which a recently described from northern Vietnam. It was described as a member of *Megophrys sensu lato* based on a single male specimen collected from Pu Mat National Park in south-western Nghe An Province. Orlov et al. (2015) indicated that this species is most closely resembles *X. palpebralespinosa*, inhabiting montane areas of northern Vietnam (recorded from provinces Lao Cai, He Giang, Cao Bang, Vinh Phuc, Son La, Thanh Hoa, Nghe An) and northern Laos, eastern part of Yunnan and the westernmost part of Guangxi, China (Bourret, 1937, 1942; Fei et al., 2009, 2010; Orlov et al., 2015; Tapley et al., 2017). The main morphological differences of these two species were the presence of very wide dermal fringes along all length of the toes in *X. latidactyla* (vs. broad dermal fringes absent in *X. palpebralespinosa*), presence of prominent subarticular tubercles in *X. latidactyla* (vs. absent), and differences in tympanum size. Our mtDNA genealogy unambiguously places two specimens of *X. latidactyla*, including the type specimen ZISP 12182, within radiation of *X. palpebralespinosa* from northern Vietnam and Laos. The genetic divergence between *X. latidactyla* and *X. palpebralespinosa* is small, ranging from 0.3%–1.4%, suggesting that these two taxa might be conspecific. Morphological differences that were used by Orlov et al. (2015) to diagnose these species might be subject to significant variation, which was not assessed due to small sample size ($n=1$): feet webbing and dermal fringes on toes in *X. palpebralespinosa* are normally significantly reduced out of the breeding season and get enlarged during the reproduction

(personal observation); the reported distinct subarticular tubercles in *X. latidactyla* might be the result of partial dehydration of limbs due to preservation in ethanol. Thus, no unambiguous morphological or molecular characters distinguishing these two species. Therefore, we transfer *X. latidactyla* to the synonymy of *X. palpebralespinosa* as a subjective junior synonym. Further, this result indicates that molecular analyses are essential in assessing species diversity of *Xenophrys*, and megophryids in general.

NOMENCLATURE ACTS REGISTRATION

The electronic version of this article in portable document format represents a published work according to the International Commission on Zoological Nomenclature (ICZN), and hence the new names contained in the electronic version are effectively published under that Code from the electronic edition alone (see Articles 8.5 – 8.6 of the Code). This published work and the nomenclature acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information can be viewed through any standard web browser by appending the LSID to the prefix <http://zoobank.org/>.

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SCIENTIFIC FIELD SURVEY PERMISSION INFORMATION

Specimen collection protocols were approved by the Institutional Ethical Committee of Animal Experimentation of the University of Phayao (certificate number UP-AE59-01-04-0022 issued to Chatmongkon Suwannapoom) and the Institute of Animal for Scientific Purposes Development Thailand (No. U1-01205-2558).

SUPPLEMENTARY DATA

Supplementary data to this article can be found online.

COMPETING INTERESTS

The authors declare that they have no competing interests.

AUTHORS' CONTRIBUTIONS

J.C., C.S., and Y.H.W. designed the study. C.S., P.P., and N.A.P. collected specimens in the field. J.Q.J. performed molecular experiments. Y.H.W. and K.X. measured the specimens. Y.H.W. performed data analyses. Y.H.W., C.S., N.A.P., R.W.M., and J.C. wrote the manuscript. All authors read and approved the final version of the manuscript.

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REFERENCES

- Bourret R. 1937. Notes herpétologiques sur l'Indochine française. XIV. Les batraciens de la collection du Laboratoire des Sciences Naturelles de l'Université. Descriptions de quinze espèces ou variétés nouvelles. Hanoi: Annexe au Bulletin Général de l'Instruction Publique, 5–56. (in French)
- Bourret R. 1942. Les Batraciens de l'Indochine. Hanoi: Institut Océanographique de l'Indochine, 1–547. (in French)
- Chan-ard T. 2003. A Photographic Guide to Amphibians in Thailand. Bangkok: Darnsutha Press, 175. (In Thai)
- Chan KO, Grismer LL, Brown RM. 2018. Comprehensive multi-locus phylogeny of old world tree frogs (Anura: Rhacophoridae) reveals taxonomic uncertainties and potential cases of over- and underestimation of species diversity. *Molecular Phylogenetics and Evolution*, **127**: 1010–1019.
- Chen JM, Poyarkov Jr NA, Suwannapoom C, Lathrop A, Wu YH, Zhou WW, Yuan ZY, Jin JQ, Chen HM, Liu HQ, Nguyen TQ, Nguyen SN, Duong TV, Eto K, Matsui M, Orlov NL, Stuart BL, Brown RF, Rowley JLL, Murphy RW, Wang YY, Che J. 2018. Large-scale phylogenetic analyses provide insights into unrecognized diversity and historical biogeography of Asian leaf-litter frogs, genus *Leptolalax* (Anura: Megophryidae). *Molecular Phylogenetics and Evolution*, **124**: 162–171.
- Chen JM, Zhou WW, Poyarkov Jr AN, Stuart LB, Brown MR, Lathrop A, Wang YY, Yuan ZY, Jiang K, Hou M, Chen HM, Suwannapoom C, Nguyen NS, Duong VT, Papenfuss JT, Murphy WR, Zhang YP, Che J. 2017. A novel multilocus phylogenetic estimation reveals unrecognized diversity in Asian horned toads, genus *Megophrys sensu lato* (Anura: Megophryidae). *Molecular Phylogenetics and Evolution*, **106**: 28–43.
- Chuaynkern Y, Chuaynkern C. 2012. A checklist of amphibians in Thailand. *Journal of Wildlife in Thailand*, **19**(1): 163–211.
- Fei L, Hu SQ, Ye CY, Huang YZ. 2009. Fauna Sinica, Amphibia. Vol. 2. Beijing: Science Press, 1–957. (in Chinese)
- Fei L, Ye CY, Jiang JP. 2010. Colored Atlas of Chinese Amphibians. Chengdu: Sichuan Publishing House of Science and Technology. (in Chinese)
- Frost DR. 2019. Amphibian species of the World 6.0, an online reference. New York, USA: American Museum of Natural History, <http://research.amnh.org/herpetology/amphibia/index.html>.
- Grismer LL, Wood Jr PL, Aowphol A, Cota M, Grismer MS, Murdoch ML, Aguilar C, Grismer JL. 2016. Out of borneo, again and again: biogeography of the stream toad genus *Ansonia* Stoliczka (Anura: Bufonidae) and the discovery of the first limestone cave-dwelling species. *Biological Journal of the Linnean Society*, **120**(2): 371–395.
- Jiang JP, Ye CY, Fei L. 2008. A new horn toad *Megophrys sangzhiensis* from Hunan, China (Amphibia, Anura). *Zoological Research*, **29**(2): 219–222. (in Chinese)
- Khonsue W, Thirakhupt K. 2001. A checklist of the amphibians in Thailand. *The Tropical Natural History*, **1**(1): 69–82.
- Laopichienpong N, Muangmai N, Supikamolseini A, Twilprawat P, Chanhom L, Suntrarachun S, Peyachoknagul S, Srikulnath K. 2016. Assessment of snake DNA barcodes based on mitochondrial COI and Cytb genes revealed multiple putative cryptic species in Thailand. *Gene*, **594**(2): 238–247.
- Li S, Xu N, Liu J, Jiang J, Wei G, Wang B. 2018. A new species of the Asian toad genus *Megophrys sensu lato* (Amphibia: Anura: Megophryidae) from Guizhou Province, China. *Asian Herpetological Research*, **9**(4): 224–239.
- Li YL, Jin MJ, Zhao J, Liu ZY, Wang YY, Pang H. 2014. Description of two new species of the genus *Megophrys* (Amphibia: Anura: Megophryidae) from Heishiding Nature Reserve, Fengkai, Guangdong, China, based on molecular and morphological data. *Zootaxa*, **3795**(4): 449–471.
- Liu Z, Chen G, Zhu T, Zeng Z, Lyu Z, Wang J, Messenger K, Greenberg AJ, Guo Z, Yang Z, Shi S, Wang Y. 2018. Prevalence of cryptic species in morphologically uniform taxa—fast speciation and evolutionary radiation in Asian frogs. *Molecular Phylogenetics and Evolution*, **127**: 723–731.
- Mahony S. 2011. Two new species of *Megophrys kuhl* & van hasselt (Amphibia: Megophryidae), from western Thailand and southern Cambodia. *Zootaxa*, **2734**(1): 23–39.
- Mahony S, Kamei RG, Teeling EC, Biju SD. 2018. Cryptic diversity within the *Megophrys major* species group (Amphibia: Megophryidae) of the Asian horned frogs: phylogenetic perspectives and a taxonomic revision of South Asian taxa, with descriptions of four new species. *Zootaxa*, **4523**(1): 1–96.
- Mahony S, Sengupta S, Kamei RG, Biju SD. 2011. A new low altitude species of *Megophrys kuhl* and van hasselt (Amphibia: Megophryidae), from Assam, Northeast India. *Zootaxa*, **3059**(1): 36–46.
- Mahony S, Teeling EC, Biju SD. 2013. Three new species of horned frogs, *Megophrys* (Amphibia: Megophryidae), from northeast India, with a resolution to the identity of *Megophrys boettgeri* populations reported from the region. *Zootaxa*, **3722**(2): 143–169.
- Mathew R, Sen N. 2007. Description of two new species of *Xenophrys* (Amphibia: Anura: Megophryidae) from north-east India. *Cobra*, **1**(2): 18–28.
- Matsui M, Khonsue W, Panha S. 2018. Two new species of *Ansonia* from Thailand (Anura: Bufonidae). *Zoological Science*, **35**(1): 39–49.

- Messenger KR, Dahn HA, Liang Y, Xie P, Wang Y, Lu C. 2019. A new species of the genus *Megophrys* gunther, 1864 (Amphibia: Anura: Megophryidae) from Mount Wuyi, China. *Zootaxa*, **4554**(2): 561–583.
- Mo XY, Shen YH, Li HH, Wu XS. 2010. A new species of *Megophrys* (Amphibia: Anura: Megophryidae) from the northwestern Hunan Province, China. *Current Zoology*, **56**(4): 432–436.
- Myers N, Mittermeier RA, Mittermeier CG, Da Fonseca GA, Kent J. 2000. Biodiversity hotspots for conservation priorities. *Nature*, **403**(6772): 853–858.
- Nabhitabhata J, Chan-ard T. 2005. Thai Red Data: Mammals, Reptiles and Amphibians. *Office of Natural Resources and Environmental Policy and Planning*, 234.
- Nabhitabhata J, Chan-ard T, Chuaynkern Y. 2000 “2004”. Checklist of Amphibians and Reptiles in Thailand. *Office of Environmental Policy and Planning*, 152.
- Nutphund W. 2001. Amphibians of Thailand. Bangkok: Amarin Printing and Publishing Public Co.
- Ohler A, Swan SR, Daltry JC. 2002. A recent survey of the amphibian fauna of the Cardomom Mountains, Southwest Cambodia with descriptions of three new species. *The Raffles Bulletin of Zoology*, **50**(2): 465–482.
- Orlov NL, Poyarkov Jr NA, Nguyen TT. 2015. Taxonomic notes on *Megophrys* frogs (Megophryidae: Anura) of Vietnam, with description of a new species. *Russian Journal of Herpetology*, **22**(3): 206–218.
- Pawangkhanant P, Poyarkov NA, Duong TV, Naiduangchan M, Suwannapoom CA. 2018. New species of *Leptobrachium* (Anura, Megophryidae) from western Thailand. *PeerJ*, **6**: e5584.
- Poyarkov NA Jr, Duong TV, Orlov NL, Gogoleva SS, Vassilieva AB, Nguyen LT, Nguyen VHD, Nguyen SN, Che J, Mahony S. 2017. Molecular, morphological and acoustic assessment of the genus *Ophryophryne* (Anura, Megophryidae) from Langbian Plateau, southern Vietnam, with description of a new species. *ZooKeys*, (672): 49–120.
- Poyarkov NA Jr, Suwannapoom C, Pawangkhanant P, Aksornneam A, Duong TV, Korost DV, Che J. 2018. A new genus and three new species of miniaturized microhylid frogs from Indochina (Amphibia: Anura: Microhylidae: Asterophryinae). *Zoological Research*, **39**(3): 130–157.
- Rao DQ, Yang DT. 1997. The karyotypes of Megophryinae (Pelobatidae) with a discussion on their classification and phylogenetic relationships. *Asiatic Herpetological Research*, **7**: 93–102.
- Royal Forest Department. 2006. Forestry in Thailand. Available at: <http://www.fao.org/forestry/10809-09f8870885bd8d85106e0a87cd906b784.pdf>.
- Sheridan JA, Stuart BL. 2018. Hidden species diversity in *Sylvirana nigrovittata* (Amphibia: Ranidae) highlights the importance of taxonomic revisions in biodiversity conservation. *PLoS One*, **13**(3): e0192766.
- Stuart BL, Chuaynkern Y, Chan-ard T, Inger RF. 2006. Three new species of frogs and a new tadpole from eastern Thailand. *Fieldiana Zoology*, **111**: 1–19.
- Suwannapoom C, Sumontha M, Tunprasert J, Ruangsuan W, Pawangkhanant P, Korost DV, Poyarkov NA Jr. 2018. A striking new genus and species of cave-dwelling frog (Amphibia: Anura: Microhylidae: Asterophryinae) from Thailand. *PeerJ*, **6**: e4422.
- Suwannapoom C, Yuan ZY, Jiang K, Yan F, Gao W, Che J. 2017. A new species of rain-pool frog (Dicroglossidae: *Fejervarya*) from western Thailand. *Zoological Research*, **38**(5): 243–250.
- Suwannapoom C, Yuan ZY, Poyarkov NA Jr, Yan F, Kamtaeja S, Murphy RW, Che J. 2016. A new species of genus *Fejervarya* (Anura: Dicroglossidae) from northern Thailand. *Zoological Research*, **37**(6): 327–337.
- Tamura K, Stecher G, Peterson D, Filipksi A, Kumar S. 2013. MEGA6: molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, **30**(12): 2725–2729.
- Tapley B, Cutajar T, Mahony S, Nguyen CT, Dau VQ, Luong AM, Le DT, Nguyen TT, Nguyen TQ, Portway C, Luong HV, Rowley JLL. 2018. Two new and potentially highly threatened *Megophrys* horned frogs (Amphibia: Megophryidae) from Indochina’s highest mountains. *Zootaxa*, **4508**(3): 301–333.
- Tapley B, Cutajar T, Mahony S, Nguyen CT, Dau VQ, Nguyen TT, Luong HV, Rowley JJ. 2017. The Vietnamese population of *Megophrys kuatunensis* (Amphibia: Megophryidae) represents a new species of Asian horned frog from Vietnam and southern China. *Zootaxa*, **4344**(3): 465–492.
- Tian Y, Gu X, Sun A. 2000. A new species of *Megophrys* in China (Amphibia: Pelobatidae). *Acta Zootaxonomica Sinica*, **25**(4): 462–466. (in Chinese)
- Wang J, Liu ZY, Lyu ZT, Zeng ZC, Wang YY. 2017. A new species of the genus *Xenophrys* (Amphibia: Anura: Megophryidae) from an offshore island in Guangdong Province, southeastern China. *Zootaxa*, **4324**(3): 541–556.
- Wang J, Lyu ZT, Liu ZY, Liao CK, Zeng ZC, Zhao J, Li YL, Wang YY. 2019. Description of six new species of the subgenus *Panophrys* within the genus *Megophrys* (Anura, Megophryidae) from southeastern China based on molecular and morphological data. *ZooKeys*, **851**: 113–164.
- Wang Y, Zhao J, Yang J, Zhou Z, Chen G, Liu Y. 2014. Morphology, molecular genetics, and bioacoustics support two new sympatric *Xenophrys* toads (Amphibia: Anura: Megophryidae) in Southeast China. *PLoS One*, **9**(4): e93075.
- Wang YY, Zhang TD, Zhao J, Sung YH, Yang JH, Pang H, Zhang Z. 2012. Description of a new species of the genus *Xenophrys* Günther, 1864 (Amphibia: Anura: Megophryidae) from Mount Jinggang, China, based on molecular and morphological data. *Zootaxa*, **3546**(1): 53–67.
- Yang JH, Wang J, Wang YY. 2018. A new species of the genus *Megophrys* (Anura: Megophryidae) from Yunnan Province, China. *Zootaxa*, **4413**(2): 325–338.
- Ye CY, Fei L, Xie F. 2007. A new species of Megophryidae – *Megophrys baolongensis* from China (Amphibia, Anura). *Herpetologica Sinica*, **11**: 38–41. (in Chinese)
- Zhang Y, Li G, Xiao N, Li J, Pan T, Wang H, Zhang BW, Zhou J. 2017. A new species of the genus *Xenophrys* (Amphibia: Anura: Megophryidae) from Libo County, Guizhou, China. *Asian Herpetological Research*, **8**(2): 75–85.