Research Article

Synergistic Effect of Network-Based Multicomponent Drugs: An Investigation on the Treatment of Non-Small-Cell Lung Cancer with Compound Liuju Formula

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Lung cancer is the most common cause of cancer death with high morbidity and mortality, which non-small-cell lung cancer (NSCLC) accounting for the majority. Traditional Chinese Medicine (TCM) is effective in the treatment of complex diseases, especially cancer. However, TCM is still in the conceptual stage. The interaction between different components remains unknown due to its multicomponent and multitarget characteristics. In this study, compound Liuju formula was taken as an example to isolate compounds with synergistic biological activity through systems pharmacology strategy. Through pharmacokinetic evaluation, 37 potentially active compounds were screened out. Meanwhile, 116 targets of these compounds were obtained by combing with the target prediction model. Through network analysis, we found that multicomponent drugs can present a synergistic effect through regulating inflammatory signaling pathway, invasion pathway, proliferation, and apoptosis pathway. Finally, it was confirmed that the bioactive compounds of compound Liuju formula have not only a killing effect on NSCLC tumor cells but also a synergistic effect on inhibiting the secretion of correlative inflammatory mediators, including TNF- α and IL-1 β . The systems pharmacology method was applied in this study, which provides a new direction for analyzing the mechanism of TCM.

1. Introduction

With increasing incidence and mortality, cancer has become the leading cause of death and has caused serious public health problems worldwide. Among them, lung cancer is the most common type of cancer [1, 2]. Nonsmall-cell lung cancer (NSCLC) accounts for 80–85% of all lung cancers [3]. At present, the main effective methods in reducing the mortality of non-small-cell lung cancer include chemotherapy, radiotherapy, targeted therapy, and surgery [4]. Nonetheless, the overall 5-year survival rate of NSCLC remains low and is only 18% [5], while the drug resistance and side effects are getting more serious [2]. Therefore, there is an urgent need for novel methods for treating NSCLC that are effective and safe.

In recent years, Traditional Chinese Medicine (TCM) is widely used all over the world. TCM has been effective to relieve complex diseases for over 4000 years due to multicomponent, multitarget, and multilevel characteristics [6]. For instance, compound Liuju formula is widely used in the treatment of lung diseases in China. Clinical medicines such as clinical compound Liuju tablets and compound Liuju granules contain leaves of Hanliuye (Salix matsudana Koidz., SMK), Yejuhua (Chrysanthemum indicum L., CIL), and Baihuasheshecao (Hedyotis diffusa Willd, HDW). The SMK was used in the Chinese dictionary [7] as a traditional anti-inflammatory and analgesic [7, 8] medicine. Research has shown that CIL has been used to cure inflammationrelated diseases and malignant tumors [9, 10]. It is reported that EEHDW has an effective inhibitory activity on human lung cancer cells by inhibiting cell proliferation and reducing cell activity [11]. TCM had fewer side effects, wide availability and better effect, and can availably improve the life quality of NSCLC patients. However, it is very hard to explain the interaction among the collaborative compositions that we confront a huge complex system for TCM formula.

With the development of analytical tools such as biology network [12], network pharmacology [13], and systems biology [14], the complex and comprehensive mechanism of TCM syndrome differentiation is expected to be quickly and efficiently clarified. In previous research, we have successfully constructed a new systems pharmacology method in order to explore the potential mechanism of Traditional Chinese Medicine. This method combines pharmacokinetics (absorption, distribution, metabolism, excretion, and toxicity (ADME/T) characteristics of drugs), molecular evaluation, target prediction, and pathway analysis to explore the effects of drugs [15, 16], which provides a platform for identifying multiple mechanisms of action of drugs. This platform has successfully developed four herbs, including Radix Astragali Mongolici, Radix Puerariae Lobatae, Radix Ophiopogonis Japonici, and Radix Salviae miltiorrhiza [17], that were applied in the comprehensive treatment of cardiovascular disease. systems pharmacology has become a widely used new tool to reveal the mechanism of drug action and drug development.

In our current work, the systems pharmacology method was utilized to study synergic bioactive compounds isolated from the compound Liuju formula treatment of NSCLC. We screened out bioactive ingredients from the constructed compound Liuju formula via ADME by calculating pharmacokinetic properties and evaluating oral bioavailability (OB) and drug-likeness (DL). Then, a comprehensive target prediction method combining biological model and mathematical model was used to predict the homologous target of the selected bioactive ingredients. Next, the obtained target was verified by functional enrichment analysis and targetdisease interaction analysis. Finally, the system revealed potential collaborative interactions between bioactive ingredients, active targets, and pathways through systematic pharmacology theory and NSCLC-related signaling pathway evaluation. And, in vitro experiments were conducted to further verify the inhibitory effect of potential bioactive ingredients on tumor cells. These results not only provide new treatments for NSCLC but also promote the elucidation and development of the molecular mechanism of TCM.

2. Materials and Methods

2.1. Construction of Molecular Database and ADME Screening. All compounds of SMK, CIL, and HDW were obtained from our formerly established database named Traditional Chinese Medicine Systems Pharmacology Database (TCMSP, http://lsp.nwu.edu.cn/) [18] and were detected artificially. Because glycosyl molecules are usually hydrolyzed into free glycosides *in vivo*, which can be easily absorbed by intestinal mucosa, we wrote the molecule without glycosyl viscose as _qt [19]. To gain the bioactive molecules, we employed a comprehensive model comprising predict OB (oral bioavailability) and DL (drug-likeness) to appraise pharmacokinetic and pharmaceutical properties.

2.1.1. Oral Bioavailability. Oral bioavailability (OB) refers to the speed and degree of drug absorption into human circulation, which reflects the proportion of drugs in human circulation and plays a critical role in drug screening. In this work, OB value was calculated by an in-house model OBioavail1.1 [18]. And, the threshold of OB value was positioned as 25% by the following conditions: firstly, get as much information as possible from the herbs studied with the fewest compounds. Secondly, the acquired model is correctly interpreted by the reported pharmacological data [20]. In this work, for further analysis, we limited the OB threshold as 25%.

2.1.2. Drug-Likeness. Drug-likeness (DL) refers to the similarity between compounds and known drugs, which is an important factor in determining the success of final clinical trials of drugs. In this work, we utilized the previously developed internal model (Tanimoto coefficient) [20] to predict drug-like properties of expected molecules. The DL appraisal formula is as follows:

$$T(A, B) = \frac{A \cdot B}{|A|^2 + |B|^2 - A \cdot B}.$$
 (1)

Among them, A represents the molecular descriptor of herbal compounds, and B represents the average molecular properties of all compounds in the database (http://www. drugbank.ca/) [21]. For further study, we defined $DL \ge 0.18$ (average of drug library) as the criterion for screening candidate compounds.

In order to obtain the potential bioactive ingredients, the screening standard was defined as $OB \ge 25\%$, $DL \ge 0.18$.

2.2. Drug Targeting. In order to structure a direct link between potential bioactive ingredients and targets, we utilized the in-house developed system drug targeting tool (SysDT) [22] and weighted integration similarity (WES) [23] algorithm to predict the target of the compound and to improve the comprehensiveness and accuracy of the target data bank.

Firstly, the weighted ensemble similarity (WES) and systematic drug targeting tool (SysDT) were applied to explore the target information of active compound. As for SysDT, which includes two mathematical tools, Random Evidence-Based Complementary and Alternative Medicine

Forest (RF) and Support Vector Machine (SVM) can determine the interaction between composite targets more completely [22]. There was another computing model, WES, which combines CDK parameters, Dragon parameters, and CDK-Dragon mixing parameters, to predict the direct target of the actual bioactive ingredients [23]. Secondly, the collected protein targets were mapped to the UniProt database (http://www.uniprot.org) for standardization [24]. Finally, the normalized compound targets were mapped to Therapeutic Target Database (TTD, http:// database.idrb.cqu.edu.cn/TTD/) [25], Comparative Toxicogenomics Database (CTD, http://ctdbase.org/) [26], Pharmacogenomics Knowledgebase (PharmGKB, https:// www.pharmgkb.org/) [27], and Kyoto Encyclopedia of Genes and Genomes (KEGG, http://www.kegg.jp/) [28] to obtain their corresponding diseases and to screen out a relationship between target and disease network.

2.3. Gene Ontology (GO) Analysis and Pathway Enrichment. In order to further analyze the specific biological processes and approaches of the potential targets we have obtained, Gene Ontology (GO) enrichment analysis was performed by linking the targets to the KEGG. KEGG is a collection of databases for systematic analysis of gene functions, biological pathways, diseases, drugs, and chemicals [29]. Finally, the pathway and process enrichment analyses were carried out by using Metascape (Metascape, http:// metascape.org) [30, 31] software.

2.4. Network Construction. In order to visualize the action mechanism of active compounds treating NSCLC and further clarify the relationship between active targets and compounds, we constructed two relational networks: Compound-Target network (C-T network) and Target-Pathway network (T-P network). In these networks, compounds, targets, and pathways were represented by nodes, while the relationship between them was represented by line segments. The degree represents the number of edges associated with a node, and the larger the number, the more node relationships it represents. The topological properties of these networks were analyzed using Cytoscape 3.6.0 [32], which is fashionable bioinformatics software.

2.5. Pathway Construction. In terms of pathway, in order to explore the integrative mechanism of action of compound Liuju formula on NSCLC, an integrated pathway related to NSCLC was established based on existing pathological knowledge of NSCLC. Firstly, in order to obtain the basic information of the pathway, we mapped the screened human target proteins to the KEGG database. Secondly, the integrated KEGG pathways of targets with false discovery rated (FDR) less than 0.05 by Fisher's Exact test in the Database for Annotation, Visualization, and Integrated Discovery (DA-VID, http://david.abcc.ncifcrf.gov) (evaluated to Fisher's exact test, FDR < 0.05) were inspected [33]. Finally, we manually assembled a relatively complete NSCLC-related pathway to further analyze the molecular action mechanism.

2.6. Experimental Detection

2.6.1. Sample Treatment. Chemicals apigenin (B20981, HPLC \geq 98%), kaempferol (B21126, HPLC \geq 98%), and ursolic acid (B21403, HPLC \geq 98%) were purchased from Shanghai Yuanye BioTechnology Co., Ltd., (Shanghai, China), and the concentration of the original solution prepared with dimethyl sulfoxide (DMSO) (American, Sigma) was 100 mmol/L. In order to ensure that the survival of cells was not affected, the final concentration of DMSO should not exceed 0.1%.

2.6.2. Cell Cultures. The murine macrophage line RAW264.7 cell and human NSCLC cell lines H1975 cells were obtained from Cell Resource Center, Shanghai Institutes for Biological Sciences, and CAS. RAW264.7 and H1975 cells were cultured in DMEM and RPMI 1640 (Gibco, USA) medium, respectively. Supplemented with 10% heat inactivated foetal bovine serum (FBS) and antibiotics (100 units/mL penicillin and 100 μ g/mL streptomycin). Cells were survived in the incubator of 5% CO₂ at 37°C. Culture medium was changed every other day.

2.6.3. Establishment of Inflammation Model. RAW264.7 cells were used to construct inflammatory models. Firstly, the cells were cultured in 150 mm Petri dish for 24 hours and treated with drugs for 2 hours. Then, 0.1 μ g/mL lipopoly-saccharide (LPS) was added to culture for 18 hours. Finally, the cells were collected, and the expression level of inflammatory mediators was detected.

2.6.4. Cell Cytotoxicity Analysis. Determination of cell cytotoxicity was conducted by Cell Counting Kit-8 (CCK-8) assay (Best Bio, Shanghai, China). In brief, H1975 cells were cultured in 96-well plates at a density of 1×10^5 cells/well. After 24-hour training, cells were exposed to different concentrations of apigenin, kaempferol, and ursolic acid. After treatment for 48 h, $10 \,\mu$ L of CCK-8 assay was added to each well, and the cells were hatched for 1–4 h at 37°C and 5% CO2. The absorbance value at 550 nm was surveyed using a microplate reader (Molecular Devices, USA). The cell viability was calculated as: OD of treatment/OD of control × 100%.

2.6.5. Expression Levels of TNF- α and IL-1 β . Enzyme-linked immunosorbent assay (ELISA) kit (R&D Systems, USA) was used to measure the expression of TNF- α and IL-1 β . The cell supernatant after drug treatment was collected according to the protocol of the specification, and 50 μ L of the sample was used for detection. The sample concentration was calculated according to the standards provided in the kit.

2.7. Statistical Analysis. Variables were analyzed by Student's *t*-test and one-way ANOVA and post hoc analysis of variance (GraphPad Prism version 7). Results were reported as mean values S.E. *p < 0.05; **p < 0.01, and ***p < 0.001.

3. Results

3.1. Active Compound Screening. To screen out the potential bioactive ingredients of SMK, CIL, and HDW, we appraised the ingredients' ADME properties including OB and DL. As shown in Supporting Information Table 1, the results showed that among 156 compounds, 37 compounds reached the standard of $OB \ge 25\%$, $DL \ge 0.18$. It was worth noting that 6 shared compounds met the screening conditions of SMK, CIL, or HDW, such as β -sitosterol, apigenin, luteolin, kaempferol, ursolic acid, and sitogluside-qt, indicating that these active compounds may exhibit effective pharmacological effects on NSCLC. Further, in order to verify whether virtual screening results were consistent with NSCLC, we conducted a literature review of the potential components. Many of the 37 active components had been reported of having significant antitumor and anti-inflammatory effects. For example, β -sitosterol (MOL004, OB = 36.91%, DL = 0.75) induced G0/G1 cell cycle arrest and inhibited cell proliferation in A549 cells [34]. Studies have shown that ursolic acid (MOL074, OB = 37.73%, DL = O.75) induces apoptosis via activation of caspases and phosphorylation of glycogen synthase kinase 3 beta in ovarian cancer cells [35]. Apigenin (MOL009, OB = 45.09, DL = 0.21) was affirmed to inhibit the migration/invasion of NSCLC cells harboring different EGFR statuses via suppressing the Snail/Slug-mediated EMT [36]. These potential compounds may be key components in the treatment of NSCLC.

3.2. Medicine Targeting and Analysis. To explore the target of compounds for NSCLC, we enriched the targets of the compounds. Therefore, we identified 116 targets of these active compounds by means of the WES and SysDT algorithms (as shown in Table 1). The results indicated that most compounds act on more than one target and exhibit multiple pharmacological effects of biologically active molecules. For example, target peroxisome proliferatoractivated receptor gamma (PPARG) corresponds to 23 compounds accounting for 62% of the total active compound. Studies have shown that activation of PPARG, γ subtype, could cause proliferation inhibition or differentiation of tumor cells [37]. In addition, elevated coexpression of PTGS2 and NOS2 (51% and 48% of the total compounds, respectively) proteins is a strong predictor of poor survival among cancer patients [38]. Hence, these targets involved in the biological processes of NSCLC will be further researched.

3.3. Pathway and Process Enrichment Analyses. Next, we used Metacape software to enrich and analyze the gene ontology (GO) of proteins targeting potential bioactive components to verify whether these proteins are related to NSCLC and to set threshold of *P* value ≤ 0.05 . As shown in Figure 1, we discovered that they all participate in the biological processes such as "cellular response to organic cyclic compound," "inflammatory response," "response to

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inorganic substance," and "cellular response to nitrogen compound". Thus, the targets of active molecules we filter from the compound Liuju formula could be regarded as the NSCLC therapeutic targets.

3.4. Compound-Target Network Analysis. In this section, we used Cytoscape 3.6.0 to generate the C-T relation network diagram (Figure 2) which contains 950 interactions between 37 molecules and 116 targets to reveal the relationship between the target and the compound more directly. Subsequently, the C-T network topology analysis showed that the average degree of the compound was 31 and the average target degree was 8, respectively. This may mean that each active compound is associated with multiple targets and all play key roles in disease mechanisms.

Notably, apigenin (MOL009, Degree = 51) effectively suppressed lung cancer progression by targeting the CD26-Akt-Snail/Slug signaling pathway [36]. And, the research indicated that kaempferol (MOL023, degree = 48) increased tumor cell killing effect through inhibition of the AKT/PI3K and ERK pathways [39]. Also, ursolic acid (MOL047, Degree = 14) was one of the action components that was present in extracts of CIL and HDW. In recent years, anticancer, anti-inflammation, and regulating immune cell effects of ursolic acid have been discovered [40-42]. Therefore, these three ingredients with a high degree play crucial roles in NSCLC treatment. Significantly, prostaglandin-endoperoxide synthase 2 (PTGS2, Degree = 19) has been found to be highly expressed in many cancer types, and it contributes to tumorigenesis via the inhibition of apoptosis, increased angiogenesis, and invasiveness [43]. All these suggest that compounds probably treat NSCLC by inhibition of tumor cell cycle, anti-inflammation, and inhibiting tumor angiogenesis.

3.5. Target-Pathway Network Analysis. The results are shown in Table 2; the 24 dramatically enriched pathways (p value \leq 0.05, multiple targets \geq 8) may be the main pathway of action and play a key role in NSCLC disease. As shown in Figure 3, the T-P network includes 88 nodes (64 targets and 24 pathways) and 232 edges. Meanwhile, numerous pathways are regulated by multiple target proteins, which might be the main factor contributing to the anti-NSCLC effect of the herbal formula for NSCLC. pathways in cancer (p value = $7.7 * 10^{(-12)}$, hsa05200, degree = 28), PI3K-Akt signaling pathway (p value = $1.0 * 10^{(-5)}$, hsa04151, degree = 18), MicroRNAs in cancer (p value = $1.0 \times 10^{(-3)}$, hsa05206, degree = 13), Proteoglycans in cancer (p value = $7.2 \times 10^{(-4)}$, hsa05205, degree = 11), and TNF signaling pathway (pvalue = $1.7 * 10^{(-4)}$, hsa04668, degree = 9) may be crucial pathways. For instance, the PI3K-AKT pathway may be a key pathway regulating the proliferation and apoptosis of NSCLC cells [44]. Meanwhile, the activation of TNF signaling by inflammatory signaling plays an important role in the development of tumors [45]. Also, heparin sulfate proteoglycans (HSPGs) in the proteoglycans in cancer pathway are key components of the extracellular matrix that mediate cell

UniProt-ID	Protein names	Gene names	Degree
P03372	Estrogen receptor	ESR1	25
P04150	Glucocorticoid receptor	NR3C1	13
P05067	Amyloid-beta A4 protein	APP	15
P06401	Progesterone receptor	PGR	4
P06746	DNA polymerase beta	POLB	4
P10275	Androgen receptor	AR	27
P10636	Microtubule-associated protein tau	MAPT	13
P11413	Glucose-6-phosphate 1-dehydrogenase	G6PD	13
P11473	Vitamin D3 receptor	VDR	13
P37231	Peroxisome proliferator-activated receptor gamma	PPARG	23
Q08828	Adenylate cyclase type 1	ADCY1	12
Q13887	Krueppel-like factor 5	KLF5	1
Q9NYA1	Sphingosine kinase 1	SPHK1	3
Q9UBM7	7-dehydrocholesterol reductase	DHCR7	14
P00915	Carbonic anhydrase 1	CA1	6
P00918	Carbonic anhydrase 2	CA2	17
P04798	Cytochrome P450 1A1	CYP19A1	11
007072	1,25-dihydroxyvitamin D(3) 24-hydroxylase,	CVD24A1	F
Q0/9/3	mitochondrial	CIP24AI	5
Q92731	Estrogen receptor beta	ESR2	18
O15439	Multidrug resistance-associated protein 4	ABCC4	4
O15118	NPC intracellular cholesterol transporter 1	NPC1	4
Q96RI1	Bile acid receptor	NR1H4	4
Q12908	Ileal sodium/bile acid cotransporter	SLC10A2	6
P51449	Nuclear receptor ROR-gamma	RORC	4
O75751	Solute carrier family 22 member 3	SLC22A3	5
P04278	Sex hormone-binding globulin	SHBG	5
Q12772	Sterol regulatory element-binding protein 2	SREBF2	4
P02774	Vitamin D-binding protein	GC	2
O95622	Adenylate cyclase type 5	ADCY5	13
P04798	Cytochrome P450 1A1	CYP1A1	12
P05091	Aldehyde dehydrogenase, mitochondrial	ALDH2	11
P05177	Cytochrome P450 1A2	CYP1A2	10
P07900	Heat shock protein HSP 90-alpha	HSP90AA1	16
P09917	Arachidonate 5-lipoxygenase	ALOX5	13
P11309	Serine/threonine-protein kinase pim-1	PIM1	15
P18031	Tyrosine-protein phosphatase non-receptor type 1	PTPN1	18
P19438	Tumor necrosis factor receptor superfamily member	TNFRSF1A	9
P23219	Prostaglandin G/H synthase 1	PTGS1	14
P24941	Cvclin-dependent kinase 2	CDK2	14
P33527	Multidrug resistance-associated protein 1	ABCC1	14
P35228	Nitric oxide synthase, inducible	NOS2	18
P35354	Prostaglandin G/H synthase 2	PTGS2	19
P36888	Receptor-type tyrosine-protein kinase FLT3	FLT3	7
P47989	Xanthine dehydrogenase/oxidase (includes xanthine dehydrogenase)	XDH	12
P48736	Phosphatidylinositol 4,5-bisphosphate 3-kinase	PIK3CG	9
P49841	Glycogen synthase kinase-3 beta	GSK3B	15
P68400	Casein kinase II subunit alpha	CSNK2A1	4
000534	Cyclin-dependent kinase 6	CDK6	12
Q12791	Calcium-activated potassium channel subunit alpha-	KCNMA1	4
012882	Dihydronyrimidine dehydrogenase [NADD(+)]	מעפת	4
Q12002 Q16539	Mitogen_activated protein kinase 14		4 15
016678	Cytochrome D450 1R1	CVD1R1	13
O9RVA1	Tubulin beta 2R chain	TURROR	14
O9UNO0	ATP-hinding cassette subfamily G member 2	ABCG2	13
O9Y263	Phospholipase A-2-activating protein	PLA A	13
	r noophonpaoe 11 2 activating protein		14

TABLE 1: Continued.

UniProt-ID	Protein names	Gene names	Degree
P00519	Tyrosine-protein kinase ABL1	ABL1	7
P35869	Aryl hydrocarbon receptor	AHR	8
O43570	Carbonic anhydrase 12	CA12	18
P51679	C-C chemokine receptor type 4	CCR4	10
P13569	Cystic fibrosis transmembrane conductance regulator	CFTR	2
P16220	Cyclic AMP-responsive element-binding protein 1	CREB1	12
P53355	Death-associated protein kinase 1	DAPK1	11
P60568	Interleukin-2	IL2	18
P51812	Ribosomal protein S6 kinase alpha-3	RPS6KA3	11
P43405	Tyrosine-protein kinase SYK	SYK	4
Q04760	Lactoylglutathione lyase	GLOI	15
P08183	Multidrug resistance protein 1	ABCBI	12
P141/4	Macrophage migration inhibitory factor	MIF	3
P15559	NAD(P)H dehydrogenase [quinone] I	NQOI	6
014/46	Telomerase reverse transcriptase	TERI	11
P02/66	Iranstnyretin		12
P15692	Vascular endotnelial growth factor A	VEGFA	1/
P00/34	Protnrombin	F2	9
P08514	Integrin alpha-IID	IIGA2B STS	2
PU8842	Steryi-sullatase	515	3
P15090	Fatty acid-binding protein, adipocyte		2
P28223	5-nydroxytryptamine receptor 2A	CA9	ð 0
Q10/90	Eibroblest growth factor 1	CA9 ECE1	8 7
P03230 D00038	Fibroblast growth factor 2	FGF1 ECE2	7
P10145	Interleukin 8	CYCL8	1
D00382	Galectin_1	LCALS1	1
P17931	Galectin-3	LGAIS3	2
P01112	GTPase HRas	HRAS	1
P14679	Tyrosinase	TYR	9
P08253	72 kDa type IV collagenase	MMP2	4
P08254	Stromelysin-1	MMP3	4
P09874	Poly [ADP-ribose] polymerase 1	PARP1	2
P14780	Matrix metalloproteinase-9	MMP9	4
P24864	G1/S-specific cvclin-E1	CCNE1	4
P39900	Macrophage metalloelastase	MMP12	4
O60285	NUAK family SNF1-like kinase 1	NUAK1	6
P37840	Alpha-synuclein	SNCA	5
P11712	Cytochrome P450 2C9	CYP2C9	3
P51684	C-C chemokine receptor type 6	CCR6	2
P27695	DNA-(apurinic or apyrimidinic site) lyase	APEX1	4
P11388	DNA topoisomerase 2-alpha	TOP2A	5
O60218	Aldo-keto reductase family 1 member B10	AKR1B10	9
P25116	Proteinase-activated receptor 1	F2R	5
P43681	Neuronal acetylcholine receptor subunit alpha-4	CHRNA4	4
D62151	Serine/threonine-protein phosphatase 2A 55 kDa	DDD2D2A	2
P03131	regulatory subunit B alpha isoform	FFF2R2R	5
Q04206	Transcription factor p65	RELA	3
P14867	Gamma-aminobutyric acid receptor subunit alpha-1	GABRA1	3
P06881	Calcitonin gene-related peptide 1	CALCA	1
P13501	C-C motif chemokine 5	CCL5	1
P32245	Melanocortin receptor 4	MC4R	1
P68104	Elongation factor 1-alpha 1	EEF1A1	3
Q13822	Ectonucleotide pyrophosphatase/phosphodiesterase family member 2	ENPP2	3
Q9Y251	Heparanase	HPSE	3
P53985	Monocarboxylate transporter 1	SLC16A1	2
P11509	Cytochrome P450 2A6	CYP2A6	3
Q15788	Nuclear receptor coactivator 1	NCOA1	1
Q99814	Endothelial PAS domain-containing protein 1	EPAS1	3
P00533	Epidermal growth factor receptor	EGFR	4
Q16665	Hypoxia-inducible factor 1-alpha	HIF1A	2
P40926	Malate dehydrogenase, mitochondrial	MDH2	2



FIGURE 1: Pathway and process enrichment analyses of the potential targets. (a) Heatmap of enriched terms across input gene lists, colored by p values. (b) Network of enriched terms: colored by cluster ID, where nodes that share the same cluster ID are typically close to each other.



FIGURE 2: Compound-target network. A compound node and a protein node are linked if the protein is targeted by the corresponding compound. Node size is proportional to its degree.

proliferation, invasion, and cell signaling [46, 47]. Therefore, tumor invasion is an important process of tumor growth and metastasis. So, we infer that the compound activates multiple signaling pathways to inhibit inflammation, enhance immune response, and delay invasion of NSCLC. It is noteworthy that the multitarget enrichment of these pathways provides further theoretical support for the treatment of NSCLC. 3.6. NSCLC Disease Pathway Analysis. To explain the therapeutic mechanism of the active ingredients at the pathway level, based on the target pathway information obtained in the KEGG database, the key pathways obtained from the T-P network analysis were integrated to construct a complete "NSCLC pathway." As shown in Figure 4, the "NSCLC pathway" includes three signaling pathways,

TABLE 2: The information of disease-related pathways.

Term	Pathways	Degree
hsa05200	Pathways in cancer	28
hsa04151	PI3K-AKT signaling pathway	18
hsa05206	MicroRNAs in cancer	13
hsa04915	Estrogen signaling pathway	11
hsa05205	Proteoglycans in cancer	11
hsa04015	Rap1 signaling pathway	11
hsa05166	HTLV-I infection	11
hsa05215	Prostate cancer	10
hsa05142	Chagas disease (American trypanosomiasis)	10
hsa04062	Chemokine signaling pathway	10
hsa05203	Viral carcinogenesis	10
hsa04976	Bile secretion	9
hsa04668	TNF signaling pathway	9
hsa05160	Hepatitis C	9
hsa05161	Hepatitis B	9
hsa04010	MAPK signaling pathway	9
hsa05222	Small-cell lung cancer	8
hsa04914	Progesterone-mediated oocyte maturation	8
hsa04726	Serotonergic synapse	8
hsa04071	Sphingolipid signaling pathway	8
hsa04611	Platelet activation	8
hsa05162	Measles	8
hsa05152	Tuberculosis	8
hsa04024	cAMP signaling pathway	8
hsa04810	Regulation of actin cytoskeleton	8

hsa04151: PI3K-Akt signaling pathway, hsa05205: polysaccharide in cancer, and hsa04668: TNF signaling pathway. The integrated pathway reflects multiple modules such as cell proliferation, apoptosis, angiogenesis, and migration.

3.6.1. Cell Cycle Progression Module. PI3K-AKT pathway and TNF signaling pathway are involved in regulating the cell cycle progress. As we mentioned in Figure 4, apigenin (MOL009) was observed to affect cyclin-dependent kinase 2 (CDK2). It plays a key role in cell cycle progression which could accelerate the transition from G1 to S phase, and the dysregulation of CDK2 is closely related to many cancers [48]. Besides, luteolin (MOL022) was predicted to impact the heat shock protein 90 (HSP90), which regulates DNA methyltransferase transcription and silences tumor suppressor and DNA repair gene methylation in tumor development, growth, and therapeutic response plays an important role [49]. Above results indicate that the compound can effectively regulate the expression of PI3K, CDK/ cyclin, GSK3, and other genes, thereby activating the pathway and regulating the cell cycle progression.

3.6.2. Inflammation Effects Module. Inflammatory mediators and inflammatory cells are important components of tumor microenvironment and play an important role in the occurrence, development, and metastasis of tumor [50]. Hence, anti-inflammatory is an integral strategy for the compound treatment. As an important indicator of inflammation, the transcription factor p65 (NF- κ B) reduces the expression of the downstream protein prostaglandin G/H synthase 2 (COX-2) and nitric oxide synthase (iNOS) by activated luteolin (MOL022) [51]. Studies have shown that COX2 may be affected by systemic inflammation, and the prognostic impact of COX2 expression depends on tumor characteristics [52]. Also, iNOS is the main mediator of inflammation, and iNOS can enhance inflammation and plays an important role in apoptosis [53]. Therefore, the analysis showed that the compound alleviated the symptoms of inflammatory disorders in patients with NSCLC by regulating the anti-inflammatory activities of NF- κ B, cox-2, and iNOS.

3.6.3. Invasion Module. Invasion and metastasis of tumor cells are critical to the development of tumors and exacerbate the progression of tumors. In the "NSCLC pathway" shown in Figure 4, the polysaccharide in cancer pathway was involved in regulating the invasion and metastasis progress. Research have shown that tumor cells migration was associated with an increase of $\alpha\gamma\beta3$ (MOL022, luteolin) integrin proteasome degradation [54, 55]. Meanwhile, FGF2, also known as basic fibroblast growth factor (bFGF) and FGF- β , is a growth factor and signaling protein encoded by the FGF2 gene. It was involved in a variety of biological processes, including cell growth, tissue repair, angiogenesis, tumor growth, and invasion [56, 57]. In addition, VEGFA was modulated by apigenin (MOL009) and luteolin (MOL022). It has been reported in literature that VEGFA has many functions such as increasing vascular permeability, inducing angiogenesis, angiogenesis and endothelial cell growth, promoting cell migration, and inhibiting apoptosis [58-60]. Thus, these above analyses show that compound may treat NSCLC by regulating the angiogenesis, migration, and invasion.

3.7. In Vitro Experimental Detection

3.7.1. Cell Cytotoxicity Assay. In this section, we used H1975 cell line to verify the compound efficacy. We selected three highly active multitarget components apigenin (MOL009), kaempferol (MOL023) and ursolic acid (MOL047) common to three plants described above for further verification. We used different concentrations of medicine concentration to detect the cytotoxicity of apigenin, kaempferol, and ursolic acid on H1975 and RAW264.7 cell and explored its inhibition rate. The results of cellular cytotoxicity assay (Figures 5(a) and 5(b)) showed that different concentrations of H1975 and RAW264.7 cells. These results demonstrated that apigenin, kaempferol, and ursolic acid had obvious proliferation inhibition activity on cancer cells.

3.7.2. Expression Levels of TNF- α and IL-1 β . To further confirm the anti-inflammatory activity of apigenin, kaempferol, and ursolic acid, we used RAW264.7 macrophages which were treated by LPS, with or without apigenin, kaempferol, and ursolic acid. Based on previous toxicity results, we chose a dose of 20 μ M for further test. Therefore, we examined TNF- α and IL-1 β by ELISA, whose results



FIGURE 3: Target-pathway network. The T-P network is built by a target and a pathway if the pathway is lighted at the target. Node size is proportional to its degree.

demonstrated that the apigenin, kaempferol, and ursolic acid can downregulate the levels of TNF- α . The combination of drugs is more effective, especially in the three coadministered groups (Figures 5(c) and 5(d)). It is worth noting that the expression of IL-1 β was not as obvious as TNF- α , and the apigenin group was not different from the model group. However, compared with the model group, the expression level of the combined group was significantly reduced. In summary, the results of two groups showed that the combined group was better than the single drug group, and the combination of three drugs was the most obvious one with significant differences.

4. Discussion

With increasing incidence and mortality, lung cancer has become the most common cancer and the leading cause of cancer death [61]. In addition, high metastatic rate of lung cancer and its poor prognosis leading to the search for antilung cancer drugs has become an important issue to be solved.

Hence, in this study, we chose compound Liuju formula, a clinically used Chinese medicine compound, as an example

to interpret the combination effect of Traditional Chinese Medicine treatment. In order to further reveal the potential action mechanism of active compounds in Traditional Chinese Medicine formulas, we proposed a systematic pharmacological approach to gain a deeper understanding of the synergistic pharmacological mechanisms of compound Liuju formula. Firstly, based on the evaluation method, 37 active ingredients were obtained, and 116 potential diseaserelated targets were predicted. The results showed that the compound Liuju formula has the characteristics of multicomponent and multitarget synergistic treatment. Then, active compounds and C-T analysis showed that several active compounds in the compound Liuju formula are essential for the treatment of NSCLC, including apigenin, kaempferol, and ursolic acid. Moreover, some targets such as CDK2, COX2, iNOS, and VEGF have anti-inflammatory, antimigration, and antiproliferation effects on NSCLC. In addition, the result of pathway and process enrichment analyses, T-P analyses, and the integrated "NSCLC pathway" suggest that compound Liuju formula mainly treats NSCLC by regulating cellular process, inflammatory response, migration, and invasion. Finally, we have shown that apigenin, kaempferol, ursolic acid has obvious anti-proliferative effect



FIGURE 4: The integrated NSCLC pathway for compound Liuju formula.







FIGURE 5: Effect of compound Liuju formula active component at the cellular level. (a, b) Cellular cytotoxicity assay. The *x*-axis shows the drugs concentrations. The *y*-axis shows the inhibition rate of drugs on cells. (c, d) Effect of apigenin, kaempferol, and ursolic acid on LPS-induced production of inflammatory mediators. Expression levels TNF- α and IL-1 β in RAW264.7 cells: experience group cells treated with different dose medicines and analyzed by ELISA. A + K: apigenin and kaempferol; A + U: apigenin and ursolic acid; K + U: kaempferol and ursolic acid; A + K + U: apigenin, kaempferol, and ursolic acid. Data are presented as the mean ± SD (*n* = 3). **p* < 0.05, ***p* < 0.01, and ****p* < 0.001.

on lung cancer cells by *in vitro* cytotoxicity test results. Furthermore, we determined by ELISA kit that apigenin, kaempferol, and ursolic acid have significant anti-inflammatory effects, especially in the combination treatment group, confirming the synergistic effect between apigenin, kaempferol, and ursolic acid. The formula has a promoting effect on the regulation of tumor inflammatory microenvironment and has a potential research value in the treatment of NSCLC.

In summary, the systems pharmacology method reveals the characteristics of compound Liuju formula with multicomponent Chinese medicine treatment and multitarget effective treatment. And, this strategy provides a potential method for the rational discovery of new medicines.

However, the current methods of systems pharmacology are still in the early stage of development, and the content of the platform needs to be further enriched. Some models, such as ADME screening, need to be optimized, and some database bases should be expanded and updated. And, developing new algorithms, adding more drug-like properties, and improving screening accuracy could enrich the content of the platform. Moreover, the inhibitory effect on the lung cancer of compound Liuju formula was investigated by us in the study, and then we can carry out systems pharmacology prediction on other diseases and other malignant cancers in in-depth exploration. A more comprehensive therapeutic effect of compound Liuju formula would be developed and may contribute new strategies to cancer therapy.

Data Availability

The data such as active constituent ADME parameters, disease-related targets, and disease-related pathways to support the findings of this study are included within the article. The data such as compound structure, pathway and process enrichment analyses, compound-target network, Target-Pathway network, and the integrated NSCLC pathway used to support the findings of this study are included within the supplementary information file. The cellular level effect data used to support the findings of this study are available from the corresponding author upon request because part of the data for this result is included in the mentor's fund project, and it is temporarily not suitable for disclosure.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Y. L. conceived and designed the study. X. S., M. J., and X. T. C. carried out target prediction and analysis. Y. P. L. and J. L. Z. performed network building and analysis. X. S. and M. J. carried out experiments and acquisition of data. X. S., C. L. Z., and J. Z. analyzed and interpreted the data. W. X. provided financial support. Y. L. and Y. H. W. approved the final version of the manuscript.

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Supplementary Materials

Supplementary Figure 1: Graphical Abstract. Table 1: Active constituents and their corresponding ADME parameters. (*Supplementary Materials*)

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References

- M.-l. Han, Y.-f. Zhao, C.-h. Tan et al., "Cathepsin L upregulation-induced EMT phenotype is associated with the acquisition of cisplatin or paclitaxel resistance in A549 cells," *Acta Pharmacologica Sinica*, vol. 37, no. 12, pp. 1606–1622, 2016.
- [2] S.-F. Zhou, S.-T. Pan, Z.-W. Zhou et al., "Proteomic response to 5,6-dimethylxanthenone 4-acetic acid (DMXAA, vadimezan) in human non-small cell lung cancer A549 cells determined by the stable-isotope labeling by amino acids in cell culture (SILAC) approach," *Drug Design, Development* and Therapy, vol. 2015, pp. 937–968, 2015.
- [3] L. Liu and S. Wei, "Research progress of KRAS mutation in non-small cell lung cancer," *Chinese Journal of Lung Cancer*, vol. 21, no. 5, pp. 419–424, 2018.
- [4] R. L. Keith and Y. E. Miller, "Lung cancer chemoprevention: current status and future prospects," *Nature Reviews Clinical Oncology*, vol. 10, no. 6, pp. 334–343, 2013.
- [5] R. L. Siegel, K. D. Miller, and A. Jemal, "Cancer statistics, 2015," *CA: A Cancer Journal for Clinicians*, vol. 65, no. 1, pp. 5–29, 2015.
- [6] S. L. Chen and J.-G. Jiang, "Application of gene differential expression technology in the mechanism studies of nature product-derived drugs," *Expert Opinion on Biological Therapy*, vol. 12, no. 7, pp. 823–839, 2012.
- [7] N. Morita, M. Shimizu, M. Arisawa, and S. Kitanaka, "Studies on medicinal resources. XXXV. The components of salix plants (salicaceae) in Japan. (2) the components of the leaves of salix matsudana KOIDZ. f.tortuosa REHD., and S.gilgiana SEEMEN," Yakugaku Zasshi, vol. 94, no. 7, pp. 875–877, 1974.
- [8] J. Zhang, J. N. Zhang, and L. K. Han, "Studies on chemical constituents of leaves of Salix matsudana Koidz and their influence on lipolysis," *Zhongguo Zhong Yao Za Zhi = Zhongguo Zhongyao Zazhi = China Journal of Chinese Materia Medica*, vol. 25, no. 9, pp. 538–541, 2000.
- [9] C. Kim, M.-C. Kim, S.-M. Kim et al., "Chrysanthemum indicum L. extract induces apoptosis through suppression of constitutive STAT3 activation in human prostate cancer DU145 cells," *Phytotherapy Research*, vol. 27, no. 1, pp. 30–38, 2013.
- [10] H. J. Lee, H. S. Seo, J. Ryu, Y. P. Yoon, S. H. Park, and C. J. Lee, "Luteolin inhibited the gene expression, production and secretion of MUC5AC mucin via regulation of nuclear factor kappa B signaling pathway in human airway epithelial cells," *Pulmonary Pharmacology & Therapeutics*, vol. 31, pp. 117– 122, 2015.
- [11] Y.-J. Kuo, J.-S. Yang, C.-C. Lu, S.-y. Chiang, J.-G. Lin, and J.-G. Chung, "Ethanol extract of hedyotis diffusawilld upregulates G0/G1 phase arrest and induces apoptosis in human leukemia cells by modulating caspase cascade signaling and altering associated genes expression was assayed by cDNA microarray," *Environmental Toxicology*, vol. 30, no. 10, pp. 1162–1177, 2015.
- [12] T. Ideker and N. J. Krogan, "Differential network biology," *Molecular Systems Biology*, vol. 8, no. 1, p. 565, 2014.
- [13] A. L. Hopkins, "Network pharmacology," Nature Biotechnology, vol. 25, no. 10, pp. 1110-1111, 2007.
- [14] J. K. Nicholson and J. C. Lindon, "Metabonomics," *Nature*, vol. 455, no. 7216, pp. 1054–1056, 2008.
- [15] C. Huang, C. Zheng, Y. Li, Y. Wang, A. Lu, and L. Yang, "Systems pharmacology in drug discovery and therapeutic insight for herbal medicines," *Briefings in Bioinformatics*, vol. 15, no. 5, pp. 710–733, 2013.

- [16] C. Zheng, T. Pei, C. Huang et al., "A novel systems pharmacology platform to dissect action mechanisms of traditional Chinese medicines for bovine viral diarrhea disease," *European Journal of Pharmaceutical Sciences*, vol. 94, pp. 33-45, 2016.
- [17] X. Wang, X. Xu, W. Tao, Y. Li, Y. Wang, and L. Yang, "A systems biology approach to uncovering pharmacological synergy in herbal medicines with applications to cardiovascular disease," *Evidence-Based Complementray and Alternative Medicine*, vol. 2012, Article ID 519031, 15 pages, 2012.
- [18] J. Ru, L. Peng, J. Wang et al., "TCMSP: a database of systems pharmacology for drug discovery from herbal medicines," *Journal of Cheminformatics*, vol. 6, no. 1, p. 13, 2014.
- [19] Y. Yang, C. Huang, X. Su et al., "Deciphering the multicomponent synergy mechanism from a systems pharmacology perspective: application to Gualou Xiebai Decoction for coronary heart disease," *Journal of Functional Foods*, vol. 47, pp. 143–155, 2018.
- [20] X. Wang, X. Xu, Y. Li et al., "Systems pharmacology uncovers Janus functions of botanical drugs: activation of host defense system and inhibition of influenza virus replication," *Integrative Biology*, vol. 5, no. 2, pp. 351–371, 2013.
- [21] D. S. Wishart, Y. D. Feunang, A. C. Guo et al., "DrugBank 5.0: a major update to the DrugBank database for 2018," *Nucleic Acids Research*, vol. 46, no. 1, pp. D1074–D1082, 2017.
- [22] Y. Hua, J. Chen, X. Xue et al., "A systematic prediction of multiple drug-target interactions from chemical, genomic, and pharmacological data," *PLoS One*, vol. 7, no. 5, Article ID e37608, 2012.
- [23] C. Zheng, Z. Guo, C. Huang et al., "Large-scale direct targeting for drug repositioning and discovery," *Scientific Reports*, vol. 5, no. 1, p. 11970, 2015.
- [24] C. H. Wu, A. Rolf, B. Amos et al., "The universal protein resource (UniProt): an expanding universe of protein information," *Nucleic Acids Research*, vol. 34, pp. 187–191, 2006.
- [25] Z. Feng, S. Zhe, Q. Chu et al., "Therapeutic target database update 2012: a resource for facilitating target-oriented drug discovery," *Nucleic Acids Research*, vol. 40, no. 1, pp. 1128–1136, 2012.
- [26] A. P. Davis, C. J. Grondin, R. J. Johnson et al., "The comparative toxicogenomics database: update 2017," *Nucleic Acids Research*, vol. 45, no. 1, pp. D972–D978, 2019.
- [27] C. F. Thorn, T. E. Klein, and R. B. Altman, "PharmGKB: the Pharmacogenomics knowledge base," in *Methods in Molecular Biology*, vol. 1015, pp. 311–320, Humana Press, Totowa, NJ, USA, 2013.
- [28] M. Kanehisa, M. Furumichi, M. Tanabe, Y. Sato, and K. Morishima, "KEGG: new perspectives on genomes, pathways, diseases and drugs," *Nucleic Acids Research*, vol. 45, no. D1, pp. D353–D361, 2017.
- [29] S. Lobert, M. E. Graichen, R. D. Hamilton et al., "Prognostic biomarkers for HNSCC using quantitative real-time PCR and microarray analysis: β-tubulin isotypes and the p53 interactome," *Cytoskeleton*, vol. 71, no. 11, pp. 628–637, 2014.
- [30] G. Bindea, B. Mlecnik, H. Hackl et al., "ClueGO: a cytoscape plug-in to decipher functionally grouped gene ontology and pathway annotation networks," *Bioinformatics*, vol. 25, no. 8, pp. 1091–1093, 2009.
- [31] S. Liu and G. Wang, "Bioinformatic analysis reveals CYP2C9 as a potential prognostic marker for HCC and liver cancer cell lines suitable for its mechanism study," *Cellular and Molecular Biology*, vol. 64, no. 7, pp. 70–74, 2018.
- [32] J. Liu, T. Pei, J. Mu et al., "Systems pharmacology uncovers the multiple mechanisms of xijiao dihuang decoction for the

treatment of viral hemorrhagic fever," *Evidence-Based Complementary and Alternative Medicine*, vol. 2016, Article ID 9025036, 17 pages, 2016.

- [33] D. W. Huang, B. T. Sherman, and R. A. Lempicki, "Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources," *Nature Protocols*, vol. 4, no. 1, pp. 44–57, 2009.
- [34] X. Wang, M. Li, M. Hu, P. Wei, and W. Zhu, "BAMBI overexpression together with β-sitosterol ameliorates NSCLC via inhibiting autophagy and inactivating TGF-β/Smad2/3 pathway," Oncology Reports, vol. 37, no. 5, pp. 3046–3054, 2017.
- [35] Y.-H. Song, S.-J. Jeong, H.-Y. Kwon, B. Kim, S.-H. Kim, and D.-Y. Yoo, "Ursolic acid from oldenlandia diffusa induces apoptosis via activation of caspases and phosphorylation of glycogen synthase kinase 3 beta in SK-OV-3 ovarian cancer cells," *Biological & Pharmaceutical Bulletin*, vol. 35, no. 7, pp. 1022–1028, 2012.
- [36] J.-H. Chang, C.-W. Cheng, Y.-C. Yang et al., "Downregulating CD26/DPPIV by apigenin modulates the interplay between Akt and Snail/Slug signaling to restrain metastasis of lung cancer with multiple EGFR statuses," *Journal of Experimental* & Clinical Cancer Research, vol. 37, no. 1, p. 199, 2018.
- [37] Y. Langle, C. Lodillinsky, D. Belgorosky, E. O. Sandes, and A. M. Eiján, "Role of peroxisome proliferator activated receptor-gamma in bacillus Calmette-Guérin bladder cancer therapy," *The Journal of Urology*, vol. 188, no. 6, pp. 2384–2390, 2012.
- [38] D. Basudhar, S. A. Glynn, M. Greer et al., "Coexpression of NOS2 and COX2 accelerates tumor growth and reduces survival in estrogen receptor-negative breast cancer," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 114, no. 49, pp. 13030–13035, 2017.
- [39] W.-T. Kuo, Y.-C. Tsai, H.-C. Wu et al., "Radiosensitization of non-small cell lung cancer by kaempferol," *Oncology Reports*, vol. 34, no. 5, pp. 2351–2356, 2015.
- [40] S. C. d. A. Lopes, M. V. M. Novais, C. S. Teixeira et al., "Preparation, physicochemical characterization, and cell viability evaluation of long-circulating and pH-sensitive liposomes containing ursolic acid," *BioMed Research International*, vol. 2013, Article ID 467147, 7 pages, 2013.
- [41] J. S. Ruan, H. Zhou, L. Yang et al., "Ursolic acid attenuates TGF-β1 induced epithelial-ymesenchymal transition in NSCLC by targeting integrin avβ5/MMPs signaling," Oncology Research Featuring Preclinical and Clinical Cancer Therapeutics, vol. 27, no. 5, pp. 593–600, 2019.
- [42] S.-E. Jang, J.-J. Jeong, S. R. Hyam, M. J. Han, and D.-H. Kim, "Ursolic acid isolated from the seed of cornus officinalis ameliorates colitis in mice by inhibiting the binding of lipopolysaccharide to toll-like receptor 4 on macrophages," *Journal of Agricultural and Food Chemistry*, vol. 62, no. 40, pp. 9711–9721, 2014.
- [43] J. Bertrand, B. Liagre, L. Ghezali, J.-L. Beneytout, and D. Y. Leger, "Cyclooxygenase-2 positively regulates Akt signalling and enhances survival of erythroleukemia cells exposed to anticancer agents," *Apoptosis: An International Journal on Programmed Cell Death*, vol. 18, no. 7, pp. 836–850, 2013.
- [44] B. Zhou, D. Wang, G. Sun, F. Mei, Y. Cui, and H. Xu, "Effect of miR-21 on apoptosis in lung cancer cell through inhibiting the PI3K/akt/NF-κB signaling pathway in vitro and in vivo," *Cellular Physiology and Biochemistry*, vol. 46, no. 3, pp. 999–1008, 2018.

- [45] M. A. T. Hildebrandt, J. A. Roth, A. A. Vaporciyan et al., "Genetic variation in the TNF/TRAF2/ASK1/p38 kinase signaling pathway as markers for postoperative pulmonary complications in lung cancer patients," *Scientific Reports*, vol. 5, no. 1, 2015.
- [46] E. Hull, M. Montgomery, and K. Leyva, "Epigenetic regulation of the biosynthesis & enzymatic modification of heparan sulfate proteoglycans: implications for tumorigenesis and cancer biomarkers," *International Journal of Molecular Sciences*, vol. 18, no. 7, p. 1361, 2017.
- [47] R. K. Okolicsanyi, A. J. van Wijnen, S. M. Cool, G. S. Stein, L. R. Griffiths, and L. M. Haupt, "Heparan sulfate proteoglycans and human breast cancer epithelial cell tumorigenicity," *Journal of Cellular Biochemistry*, vol. 115, no. 5, pp. 967–976, 2014.
- [48] Y. Wang, Y. Chen, X. Cheng et al., "Design, synthesis and biological evaluation of pyrimidine derivatives as novel CDK2 inhibitors that induce apoptosis and cell cycle arrest in breast cancer cells," *Bioorganic & Medicinal Chemistry*, vol. 26, no. 12, pp. 3491–3501, 2018.
- [49] G. P. Nagaraju, C. Wu, N. Merchant, Z. Chen, G. B. Lesinski, and B. F. El-Rayes, "Epigenetic effects of inhibition of heat shock protein 90 (HSP90) in human pancreatic and colon cancer," *Cancer Letters*, vol. 402, pp. 110–116, 2017.
- [50] L. Guo, Y. Zhang, L. Zhang, F. Huang, J. Li, and S. Wang, "MicroRNAs, TGF-β signaling, and the inflammatory microenvironment in cancer," *Tumour Biology*, vol. 37, no. 1, pp. 115–125, 2016.
- [51] K. Natarajan, P. Abraham, R. Kota, and B. Isaac, "NF-κBiNOS-COX2-TNF α inflammatory signaling pathway plays an important role in methotrexate induced small intestinal injury in rats," *Food and Chemical Toxicology*, vol. 118, pp. 766–783, 2018.
- [52] Y. Sano, Y. Kogashiwa, R. Araki et al., "Correlation of inflammatory markers, survival, and COX2 expression in oral cancer and implications for prognosis," *Otolaryngology— Head and Neck Surgery*, vol. 158, no. 4, pp. 667–676, 2018.
- [53] H. Nakazawa, K. Chang, S. Shinozaki et al., "iNOS as a driver of inflammation and apoptosis in mouse skeletal muscle after burn injury: possible involvement of sirt1 S-nitrosylationmediated acetylation of p65 NF-κB and p53," *PLoS One*, vol. 12, no. 1, Article ID e0170391, 2017.
- [54] Y.-L. Hsu, L.-Y. Wu, M.-F. Hou et al., "Glabridin, an isoflavan from licorice root, inhibits migration, invasion and angiogenesis of MDA-MB-231 human breast adenocarcinoma cells by inhibiting focal adhesion kinase/Rho signaling pathway," *Molecular Nutrition & Food Research*, vol. 55, no. 2, pp. 318–327, 2011.
- [55] Z. Jing, Z. Qiong, W. Yonggang, and L. Yanping, "Rat bone marrow mesenchymal stem cells improve regeneration of thin endometrium in rat," *Fertility and Sterility*, vol. 101, no. 2, pp. 587–594, 2014.
- [56] C. A. Dionne, G. Crumley, F. Bellot et al., "Cloning and expression of two distinct high-affinity receptors crossreacting with acidic and basic fibroblast growth factors," *The EMBO Journal*, vol. 9, no. 9, pp. 2685–2692, 1990.
- [57] M. Lafage-Pochitaloff, F. Galland, J. Simonetti, H. Prats, M. G. Mattei, and D. Birnbaum, "The human basic fibroblast growth factor gene is located on the long arm of chromosome 4 at bands q26-q27," *Oncogene Research*, vol. 5, no. 3, pp. 241–244, 1990.
- [58] H. Jiang, J. F. Toscano, M. Schiraldi et al., "Differential expression of vascular endothelial growth factor-A isoforms between intracranial atherosclerosis and moyamoya disease,"

Journal of Stroke and Cerebrovascular Diseases, vol. 28, no. 2, pp. 360–368, 2019.

- [59] S. Karaman, V.-M. Leppänen, and K. Alitalo, "Vascular endothelial growth factor signaling in development and disease," *Development*, vol. 145, no. 14, 2018.
- [60] Y. Song, J. Hu, Q. Chen et al., "Association between vascular endothelial growth factor rs699947 polymorphism and the risk of three major urologic neoplasms (bladder cancer, prostate cancer, and renal cell carcinoma): a meta-analysis involving 11,204 subjects," *Gene*, vol. 679, pp. 241–252, 2018.
- [61] R. L. Siegel, K. D. Miller, and A. Jemal, "Cancer statistics, 2017," CA: A Cancer Journal for Clinicians, vol. 67, no. 1, pp. 7–30, 2017.