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Review Article

Alterations of the *TP53* Gene in Gastric and Esophageal Carcinogenesis

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TP53 genes is one of more important tumor suppressor gene, which acts as a potent transcription factor with fundamental role in the maintenance of genetic stability. The development of esophageal and gastric cancers is a multistep process resulting in successive accumulation of genetic alterations that culminates in the malignant transformation. Thus, this study highlights the participation of the main genetic alterations of the TP53 gene in esophageal and gastric carcinogenesis. Among these changes, high frequency of TP53 mutations, loss of heterozygosity (LOH), overexpression of the p53 protein, and consequently loss of p53 function, which would be early events in esophageal and gastric cancers, as well as an important biomarker of the prognosis and treatment response. Furthermore, Single Nucleotide Polymorphisms (SNPs) of TP53 have been implicated in the development and prognosis of several cancers, mainly TP53 codon 72 polymorphism whose role has been extensively studied in relation to susceptibility for esophageal and gastric cancer development.

1. Introduction

Gastric and esophageal cancers together are responsible by high rates of incidence and mortality worldwidely [1]. These neoplasms are histologically and genetically heterogeneous, but in the meantime sharing common aspects, such as a multifactorial origin involving risk factors such as smoking consumption and alcohol intake, progression through precancerous lesions, and the occurrence of an inflammatory process [2, 3].

Esophageal cancer is classified in two major histological types as esophageal squamous cell carcinoma (ESCC) and esophageal adenocarcinoma (EA). The ESCC may occur as consequence of a premalignant lesion knows as megaesophagus (esophagus dilatation) due achalasia leading to food retention or esophageal stasis. In consequence, chronic esophagitis may occur due increased bacterial proliferation in the liquid stasis [4], acanthosis, parakeratose and

leukoplakia [5]. The megaesophagus increases the risk of the 3% to 8% of developing ESCC [6]. The EA is related with Barrett's esophagus (BE), an acquired metaplastic abnormality in which the normal stratified squamous epithelium of the esophagus is replaced by an intestinal-like columnar epithelium containing goblet cells (intestinal metaplasia). Such condition is widespread and provides a 100-fold increased risk for the development of EA [7].

The gastric adenocarcinoma accounts for approximately 95% of cases of gastric malignancies. It is classified by histopathological characteristics in diffuse and intestinal subtypes [8] and occurs as distinct clinical and epidemiological entities. The gastric cancer (GC) can progress through of multistep process from a chronic gastritis frequently resulting from *Helicobacter pylori* infection to gastric atrophy, intestinal metaplasia, dysplasia, and finally to carcinoma [9]. This bacterium, due the inflammatory process in gastric mucosa is considered the major risk factor of GC. It is present

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in 77% of noncardia gastric cancers [10] and in 90% of all chronic gastritis patients, so has been associated with increased risk of cancer up to nine times [11, 12].

Although different genetic and epigenetic alteration involving oncogenes activation, tumor suppressor genes mutations, DNA repair genes, microsatellite instability, loss of heterozygosity (LOH) have been reported in both esophageal and gastric cancers [2, 3, 13, 14], genetic alterations in *TP53* tumor suppressor gene are fundamental events related in both early stage and advanced tumor.

In this study, we summarize the main molecular alterations of the *TP53* gene in esophageal and gastric carcinogenesis reported in literature and also our contribution to studies of this gene in precancerous and malignant lesions of the esophagus and stomach, such as frequency and types of *TP53* mutations, LOH, overexpression of the mutant p53 protein, and consequently loss of p53 function, which may act as important biomarker of the prognosis and treatment response. In addition, we also focused on the role of *TP53* codon 72 polymorphism, which has been extensively studied in relation to risk for esophageal and gastric cancer development.

2. TP53 Gene

TP53 gene mapped on 17p13.1 [15] is one of more important tumor suppressor gene composed by 11 exons (\sim 20 KB), which genomic integrity of exons 5–8 is particularly important for its activity [16, 17]. TP53 gene encodes a nuclear p53 protein of 393 amino acids, which acts as a potent transcription factor with key role in the maintenance of genetic stability [18]. This protein regulates the expression of hundreds of genes and noncoding RNAs, as well as the RNA processing complexes activity. When activated, in response to cellular stress (Figure 1), p53 triggers adequate cellular response, including cell-cycle arrest, DNA repair and programmed cell death (apoptosis) [19], and preventing the multiplication of damaged cells [20], being named "the guardian of the genome" [21]. The p53 protein has also others biological functions: senescence, DNA metabolism, angiogenesis, cellular differentiation, and the immune response [22].

The function of *TP53* gene is usually altered through LOH, mutations, and rarely by DNA methylation. Over 50% of human cancers present inactivated *TP53*, due loss of function mutations [23], among 95% of them occurred within the genomic region encoding the sequence-specific DNA-binding domain of *TP53*. These mutations disrupted the proper conformation of that sequence so mutant forms of *TP53* lacked the sequence-specific transactivation ability. Thus, impaired *TP53* activity promotes the accumulation of DNA damage in cells, which leads to a cancer phenotype.

In general, *TP53* exons 5–9 are investigated because they contain the zinc-finger domain and the transactivating domain, which are mutational hotspots; by the way, more than 80% of *TP53* mutations are clustered there [24]. The *TP53* mutations consist of primarily missense substitutions

(75%) nonrandomly distributed along the molecule, particularly the central DNA-binding-domain [25]. These single aminoacid changes affect *TP53* transcriptional activity to various degrees. The *TP53* mutational spectrum is characterized by the presence of mutations at six discrete hotspot codons within the DNA binding domain of the molecule: codons 175, 245, 248, 249, 273, and 282 [26]. Furthermore, other alterations include frameshift insertions and deletions (9%), nonsense mutations (7%), silent mutations (5%), and some infrequent alterations [27]. More than 27,000 somatic mutation data of *TP53* appear in the International Agency for Research on Cancer (IARC) *TP53* database version R15 [25, 28].

For the p53 protein expression, the wild type has short-life and the mutant forms have a longer half-life [29, 30], and show the dominant-negative behavior toward wild type [31, 32], so overexpression and accumulation of mutant p53 protein by immunohistochemistry assay has been widely used as marker for detection of p53 abnormalities in neoplasms.

More than two decades after the *TP53* gene discovery and knowledge about its function in cell cycle control and normal cells homeostasis, mutations of this gene remain the prevalent genetic alteration involved in cancer etiology.

3. Mutations of TP53 Gene in Esophageal and Gastric Carcinogenesis

3.1. Esophageal Adenocarcinoma (EA) and Esophageal Squamous Cell Carcinoma (ESCC). The EA is a multistep process, in which the metaplastic epithelium characteristic of Barrett's esophagus (BE) is thought to sequentially develop low-grade dysplasia, high-grade dysplasia, early EA, and, eventually, invasive carcinoma [33, 34].

The genetic and epigenetic alteration more common in BE is the inactivation of CDKN2A on chromosome 9p [35, 36]. However, the loss of TP53 is an important event in BE progression [37, 38], because patients with LOH in TP53 are 16 times more likely to progress to EA [39], since both mutations and LOH in TP53 appear to provide a competitive advantage to the mutant clone [40]. Mutations in TP53 can be a predictor of significantly reduced postoperative survival following surgical resection of EA and would appear to be a clinically useful molecular prognostic biomarker. In a study that assessed the prognostic value of TP53 mutations in EA was observed that 47% of the tumors analyzed had TP53 mutations, predominantly G:C to A:T transitions at CpG dinucleotides. Such mutations have been associated with overexpression of p53 protein, tumor differentiation, and significantly reduced postoperative survival following surgical resection of EA [41].

Many studies has also focused on genetic alterations in ESCC at different loci of the chromosomes, because some of the microsatellite markers may be useful for the early detection of this type of cancer. The LOH was found in at least one of the eight markers, including *TP53*. However, the most of the 38 microsatellite markers analysis did not display microsatellite instability, suggesting these regions are

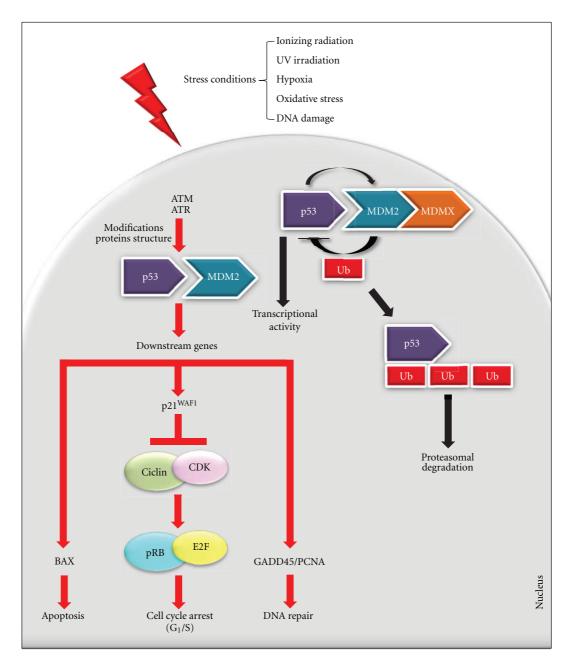


FIGURE 1: The p53 signaling pathway: *In normal conditions* (*black arrows*), p53 is maintained at very low levels. p53 is downregulated by MDM2 (murine double minute 2) and MDMX (Mdm4 p53 binding protein homolog mouse). MDM2 is an E3 ubiquitin ligase, which controls p53 proteasomal degradation. MDMX lacks the E3 ligase function and suppresses the transcriptional activity of p53, which is independent of MDM2. It also forms a heterodimeric complex with MDM2 and stimulates MDM2-mediated p53 degradation. The expression of MDM2 is controlled by p53 itself through a negative feedback loop. *In stress conditions* (*red arrows*) p53 responds to a range of environmental and intracellular stresses, including agents that cause DNA damage, ultraviolet radiation, and oxidative stress. In damage response are activated several kinases (ataxia telangiectasia mutated (ATM) and ataxia telangiectasia and Rad3 related (ATR)), which cause conformational changes in p53, MDMX, and MDM2 blocking their interactions and resulting in p53 stabilization. Activated p53 protein subsequently transactivates expression of several target genes, such as the cyclin-dependent kinase inhibitor protein p21^{WAF1}, which induce G₁/S arrest, proapoptotic genes particularly those involved in the mitochondrial pathway of apoptosis, such as BAX, and genes involved in DNA repair, such as GADD45/PCNA.

possible targets of genomic instability in early-stage ESCC carcinogenesis [42].

The esophagus is most frequently exposed to carcinogens as the stomach or colon, such carcinogens present in food or

dietary factors act as inducers of *TP53* mutations in ESCC in some areas considered high risk, such as China, Southern Brazil, and Taiwan [43]. A high frequency of *TP53* mutations and p53 protein expression in the ESCC has been reported,

and loss of p53 function would be the early events in ESCC development [44]. The study performed in ESCC patients in Japanese population reported mutations in exons 5–9 of the *TP53* gene in 48% of them, whereas transversions were the most prevalent, followed by transitions. Transversion G:C to T:A occurred preferentially at codons 157, 248, and 273, considered known sites of adducts formation on DNA. Among the sources of transversion, oxidative DNA damage, and metabolites of benzo(a)pyrene are associated with esophageal carcinogenesis, since smoking is the major risk factor for the development of this neoplasm and the fact that this substance is an important component of cigarette smoke [43].

One of the highest incidences of ESCC in the world is found in northeastern Iran, Golestan Province, with rates over 50 per 100,000 person-years in both genders [45]. In this high-risk geographic area was found a total of 120 TP53 mutations in 107/119 cases (89.9%), including 11 patients with double or triple mutations, which mutation pattern was heterogeneous with infrequent mutations at common TP53 "hotspots," but with frequent transversions attributable to environmental carcinogens forming bulky DNA adducts, including 40% at bases known as site of mutagenesis by polycyclic aromatic hydrocarbons (PAHs). The authors no observed relation of the mutation pattern with ethnicity, tobacco or opium use, and alcoholic beverage consumption or urban versus rural residence. Thus, the multiple environmental carcinogens seem to be the cause of this heterogeneous mutation pattern [45].

Our research group, in a small sample of ESCC patients of southeastern Brazil described two novel mutations in the TP53 exons 5 (codon 147) and 6 (codon 197) in 2/10 cases of ESCC, but no mutation was found in the 30 cases of chronic esophagitis assessed. While one of them was a silent mutation (codon 147) the other was a missense mutation (codon 197) resulting in a change from valine to alanine that could affect the structure and function of the p53 protein [46]. In addition, Egashira et al. [24] identified several mutations in exons 2, 3, 10, and 11 of TP53 gene in ESCC and some of these mutations might be deleterious because they are expected to lead to a truncated protein. A significant correlation between the presence of TP53 gene mutation and LOH was found, whereas there was no significant correlation between LOH and protein expression.

Recently, 10 esophageal cancer cell lines and 91 surgically resected specimens were examined for LOH at the *TP53* using microsatellite analysis, CGH (comparative genomic hybridization), FISH (fluorescence in situ hybridization), and SNP-CGH (single nucleotide polymorphism-CGH) [47]. It was verified that LOH without copy number change at the *TP53* locus was observed in *TP53* mutant ESCC, suggesting that copy-neutral LOH occurring as a result of chromosomal instability might be the major mechanism for inactivation of the intact allele in esophageal squamous cell carcinogenesis associated with *TP53* mutation. These results emphasize the pivotal role of genetic alterations in *TP53* in the esophageal carcinogenesis, with serious consequences for the deregulation of the cell cycle.

3.2. Gastric Carcinogenesis. Molecular studies have supplied important information on the genetic events in GC involving a number of genetic and epigenetic alterations including oncogenes as amplification of c-MYC, c-ERBB2, c-MET, E-cadherin (CDH1), tumor suppressor genes with mutations of APC, TP53, and cell cycle regulators, cell adhesion molecules and DNA repair genes [13, 48, 49]. Other genetic factors, such as DNA polymorphisms and genetic instability, may also be implicated in the two distinct major genetic pathways of gastric carcinogenesis [50]. However, LOH at chromosome 17p and TP53 mutations are implicated in the development of both intestinal and diffuse type gastric cancer [50].

TP53 mutation is one of the most prevalent genetic alterations in GC. More than one mutation may be present in a single tumor resulting in heterogeneity of the TP53 mutational status. There are conflicting results with respect to the prevalence of TP53 mutations and their relationship to histological type or tumor stage of GC. Some studies showed that mutations tend to affect mainly intestinal-type tumors, while others found that the incidence of mutation is similar in both intestinal and diffuse-type tumors, ranging between 25% and 40% of the cases studied. According tumor stage, TP53 abnormalities appear to occur early in intestinaltype cancers, but some studies showed that the frequency of TP53 mutation in both early and advanced intestinal-type is consistent at around 40% each, similar to that observed in advanced diffuse-type, while in early diffuse-type TP53 mutations are uncommon [43, 51–53].

In Japanese patients with GC, Oki et al. [43] found TP53 mutations in 16% (18/112) of the cases, more often in intestinal-type. The TP53 mutational spectrum was wide, including in a decreasing order of frequency, codons 175, 248, 273, 282, 245, and 213, all of which are CpG sites. Transitions of the G:C to A:T are the most common type of mutation, regardless of the histological type of the tumor, followed by transversions. Interestingly, it appears to be a difference in the frequency of G:C to A:T and A:T to G:C transitions in European compared to Asian populations [53]. The observed pattern of mutations are consistent with that for dietary mutagens associated with the metabolism of nitrogenous compounds involved in gastric carcinogenesis, thus resulting in the deamination of nucleic acids. C to T mutations are also induced by nitric oxide, a substance produced during infection by *H. pylori bacterium* [54].

Similarly, in Chinese population, Chen et al. [55] reported *TP53* mutations in GC occurring in four exons affecting codons 131, 132, 133, 135, 149, 151, 162, 167, 173, 174, and 175 of exon 5, codons 193, 197, 213, and 215 of exon 6, codons 245, 246, 248, 249, and 270 of exon 7, and codons 271, 272, 273, and 282 of exon 8. Among the mutations, G:C to A:T transitions was the highest (41.7%), followed by A:T to G:C (25%), G:C to C:G (11.1%), G:C to T:A (8.3%), A:T to T:A (2.8%), and frameshift mutation. The authors also reported an association between *TP53* mutation and patients with high/high-middle differentiated type-GC, indicating that these mutations are responsible for the initiation stages of gastric carcinoma, rather than the slowing of differentiation.

The tumor suppressor functions of p53 protein are largely demonstrated by its apoptosis-inducing ability that may be dependent or independent of *de novo* gene transcription. As a transcription factor, p53 targets multiple elements involved in the apoptotic pathway [56].

Apart from transcriptionally targeting elements, p53 is also able to mediate transcription-independent apoptosis. Under cellular stress, p53 accumulates in the cytosol or mitochondria and leads to the direct activation of proapoptotic Bcl-2 family members, such as Bax and/or Bak [57, 58], so it selectively activates p53-mediated apoptosis. This selectivity may help to avoid the unwanted side effects associated with conventional p53 treatment based on transcription [59, 60].

Several studies have assessed the relationship between apoptosis and *TP53* alterations. In gastric epithelium a balance between cell proliferation rate and programmed cell death or apoptosis maintain the homeostasis. An imbalance of these two processes leading to increased proliferation of the gastric epithelial cells may enhance the effect of carcinogens on DNA, increasing the risk of mutational changes, and the development of gastric cancer [61, 62].

In focus, we investigated the association of apoptosis with infection by H. pylori in benign gastric lesions and GC [63]. Although not observed significant differences in apoptotic index (AI) between the different groups of benign gastric lesions, whether by the TUNEL technique or by the CPP32 (caspase-3 activated) antibody, the CAG (chronic atrophic gastritis) group showed a statistically increased AI, compared to normal mucosa (NM), as well as a higher number of TUNEL-positive cases. Furthermore, the CG (chronic gastritis) group had a statistically higher AI than did the NM, as well as a higher number of CPP32positive cases. However, the GC group displayed a low AI, and no significant differences were found regarding the histological subtypes, intestinal or diffuse. Also in this study, was found statistically higher AI in individuals infected by H. pylori in GU (gastric ulcer) and IM (intestinal metaplasia) groups compared to NM from patients without infection. In general, this study showed high AI in both groups of CG and CAG regardless of infection by H. pylori, aneuploidy, and overexpression of the protein p53. However, the precise involvement of H. pylori infection in the balance between apoptosis and proliferation has yet to be elucidated.

Regarding the histological subtypes of GC, Triantafyllou et al. [64] investigated both apoptotic and proliferation indices and found higher AI in advanced intestinal type tumors, as well as p53 protein expression significantly higher in advanced cancers and in the nondysplastic tissue adjacent. According to the authors these data indicate similar cell turnover during tumorigenesis between both tumors types.

Considering the variation of the prognosis among patients with the same tumor stage, Liu et al. [65] assessed the relationship between some apoptotic markers, such as p53, bcl-2, bax, and c-myc expression to clinicopathological characteristics and their prognostic significance in GC. The authors observed a strong correlation between the expression of p53, bax, and c-myc, as well as with histological grade, but a reverse correlation between histological type and p53

expression, so demonstrating that deregulation of p53 might result in uncontrolled proliferation in gastric cancer.

So far, many efforts have been applied for understanding the mechanisms involving *TP53* mutations in carcinogenesis and development of the gastrointestinal tract; it is clear participation of *TP53* gene alterations in early stages and progression of these tumor types.

4. p53 Protein Overexpression in Esophageal and Gastric Carcinogenesis

The expression analysis on p53 protein level by immunohistochemical staining has been performed on routine paraffin embedded material and the overexpression and an accumulation of protein is used as an indicator of mutant form of *TP53* gene, which has been shown to be a powerful marker of malignancy.

One of the most common abnormality detected in EA is overexpression of p53 protein, that is restricted to more advanced areas of dysplasia and malignancy [66, 67]. Among 137 primarily resected EA samples, after immunohistochemical staining, showed accumulation of p53 in 52% cases [68]. Increased p53 expression, as measurement of *TP53* mutations, was observed also in BE with high-grade dysplasia (HGD) and in BE-associated to EA suggesting the involvement of the *TP53* in the pathogenesis of this entity [69]. Moreover, p53 expression confirmed multifocal dysplasia in BE esophageal mucosectomies and the patients displayed increased aneusomy for chromosome 17 along the sequence of cancer progression [70].

Other esophageal precursor lesions have also evidenced alterations in p53 expression. For example, study performed by researchers of our laboratory [71] observed that the proportion of p53-positive cases increased progressively according to the severity of the esophageal pathology. Positive immunostaining for p53 protein was observed in a few cells in the normal mucosa, which was interpreted as expression of wild-type p53 protein. However, a progressive increase of p53 protein expression was observed as follows: chagasic megaesophagus (26.1%), chronic esophagitis (52.2%), and ESCC (100%). A strong and diffuse nuclear staining in the ESCC probably arose from the high expression of mutant p53 protein, whereas in chronic esophagitis and chagasic megaesophagus, it was not possible to indicate p53 as mutated protein. It may also have been due to the expression of wild-type p53 that accumulates in the cells as a consequence of the physiological and inflammatory processes in the esophageal epithelium [71].

To characterize p53 alterations in multiple esophageal carcinomas and to study their roles in carcinogenesis, p53 immunohistochemical and mutation analyses using laser capture microdissection on surgically resected were performed in esophageal carcinomas. p53 protein accumulation was observed in 72.7% of tumors. In the 9 cases of multiple esophageal carcinomas, *TP53* mutations were detected in the whole tumor in 1 (11.1%) case, in the microdissected tumor samples of main lesions in 3 (33.3%) cases, and in the microdissected tumor samples of concomitant lesions in 3

(33.3%) cases. For the microdissected tumor samples, there was a 54.5% (12/22) concordance rate between the results of immunostaining and molecular analysis. The finding of different *TP53* gene mutations among multiple esophageal carcinomas suggests further evidence of multicentric or field carcinogenesis of the esophagus [72].

In ESSC, the aberrant expression of p53 protein has been observed during the tumoral progression and appears to be associated with lymph node metastasis [73, 74]. When p53 protein expression was examined in 148 ESCC cases using immunohistochemistry combined with tissue microarray, Lin et al. [75] showed p53 protein accumulation in 86% high-grade dysplasia/carcinoma in situ (HGD/CIS), 81% of low-grade dysplasia (LGD), and in none of reactive atypical epithelium (RAE) and normal epithelium (NE). Of HGD/CIS and LGD with p53 protein accumulation, similar percentages had mutations (83% and 82%, resp.). p53 expression has also been reported in 65.5% lymph node metastasis, whereas p53 was in 50% of cases of ESCC, with the specificity of 90.9% and sensitivity rate of 65.5% in detected lymph node metastasis and positive predictive value was 95%. Expression of p53 was significantly correlated with stage and lymph node metastasis, suggesting that when preoperative staging has been insufficient in ESCC, immunohistochemical analysis of p53 in tissues could be an aid to clinicians regarding lymph node metastasis [73].

The p53 status, both gene mutation and immunohistochemical staining, was assessed as potential predictive markers of chemotherapy response and prognosis for ESCC [76]. The results of retrospective study showed mutant *TP53* and p53-positive protein in 46.8% and 55.8% of patients, respectively, which was not associated with clinicopathological findings of patient including initial tumor stage. Response to chemotherapy was observed in only 16.7% of patients with mutation of TP53 gene, which showed significantly poorer prognosis. However, there was no correlation of p53 protein status with response to chemotherapy, curative resection rate, or prognosis. These parameters were also investigated in a group of patients in the prospective study. Similarly to the retrospective study, TP53 mutation was associated with poorer response to chemotherapy and prognosis. Thus, these findings showed that TP53 genotype is a potentially useful predictor of poorer response to chemotherapy and prognosis in ESCC patients.

In gastric benign lesions and gastric cancer, our research group assessed the p53 overexpression and occurrence of aneuploidy for chromosome 17 and *TP53* gene deletion [77, 78]. In intestinal metaplasia (IM) from cancer-free patients, immunohistochemistry revealed p53 overexpression in 12% of the analyzed cases, as well as *TP53* gene deletion in 60% of the cases. All GC cases presented higher frequencies of trisomy or tetrasomy of chromosome 17 and *TP53* deletion, and immunohistochemistry detected overexpression of p53 protein in 80% of the assessed cases. These results suggest that IM and GC may share the same genetic alterations [77]. Similarly, Khayat et al. [79] evidenced positive immunoreactivity of p53 in IM samples and, in these cases, the frequency of cells with two chromosome 17 and two *TP53* signals was higher than in p53-negative cases. In other benign lesions

as chronic gastritis (CG) and gastric ulcer (GU) has also been reported p53-positive immunoreactivity, that is, in 45% and 12% of cases, respectively [78]. In this study, trisomy of 17 was increasingly found from CG to GU, but no *TP53* deletion was found in these gastric lesions. The occurrence of aneuploidies in benign lesions evidences chromosomal instability in early stages of gastric carcinogenesis.

Overexpression of p53 protein and aneuploidy of chromosome 17 has been observed in GC and nontumoral tissues in others studies. Lu et al. [80] showed that in tumor tissues aneuploidy of chromosome 17 occurred in 58.6% of cases and 45.5% of GC samples overexpressed p53 protein, significantly higher than those in nontumor strict mucosa (0 and 12.1%, resp.). The expression of p53 in nontumor gastric mucosa with dysplasia was significantly higher than that in the mucosa without dysplasia. Overexpression of p53 protein was associated with the size of tumors that may help in diagnosis and prognostic prediction of GC [80].

In addition, association of p53 expression with the tumor biological behavior and prognosis of GC patients was also reported. However, the prognostic impact of p53 abnormalities in this neoplasm remains controversial. It was described that the degree of p53 expression correlates with the proliferative rate of the gastric cancer. Furthermore, a significant association between p53 overexpression and the metastatic spread to lymph nodes or shortened survival has been described by some studies on GC, but not by others [81, 82].

Although *H. pylori* eradication has some inhibitory effects on the subsequent development of GC, there are sporadic cases of malignant progression even after successful eradication. The pathogenesis of GC emerging after *H. pylori* eradication remains to be clarified. Iijima et al. [83] assessed the relationship of the acid secretion pattern to the occurrence site of GC emerging after bacterial eradication in order to estimate the individual cancer risk. The p53 protein frequently was accumulated in non-acid-secreting areas, suggesting that genetic alteration such as *TP53* mutation seems to be already present in the residual non-acid-secreting areas that could be the origin site of gastric carcinogenesis even after eradication.

Esophageal and gastric carcinomas show multiple and distinct molecular alterations, which indicate that progression of cancer is a multistep complex process with many different pathways and accumulation of various alterations. Presumably, it is not only one molecular factor that can predict the biological behavior of these cancers, but patterns of *TP53* mutations and protein overexpression would appear to be an useful biomarker of tumor progression, prognosis, and prediction of response to treatment of gastroesophageal cancer patients.

5. TP53 Polymorphisms and Risk of Esophageal and Gastric Cancer

The tumor suppressor *TP53* pathway plays a crucial role in preventing carcinogenesis, thus single nucleotide polymorphisms (SNPs) of *TP53* gene naturally occurring in human

populations are expected to cause measurable perturbation on p53 function [84]. It is known that functional polymorphisms can impact tumor biology and have been implicated in the development and prognosis of several cancers, being highlighted as a potential candidate of the susceptibility for cancer development [85–89]. These genetics variants in *TP53* may modulate cancer risk because they are also supposed to influence cell cycle progression, apoptosis, and DNA repair [90].

At least 85 SNPs are described on *TP53* [25]. However, the most investigated polymorphism in this gene is a nonsynonymous single base pair change in a proline-rich domain located in exon 4 codon 72 (*TP53* Arg72Pro, rs 1042522), which consists in a substitution of cytosine (C) for guanine (G) and results in the substitution of arginine (Arg72—CGC) by proline (Pro72—CCC) [91].

The Arg72Pro polymorphism shows differences in its biochemical or biological functions [92, 93]. This change in amino acid sequence may alter the ability of p53 to bind to response elements in target genes and thus induce gene transcription, its interaction with p73 and its targeting of the proteasome. In addition, alter recognition motifs for posttranslational modifications or p53 stability, and still the susceptibility to degradation by human papillomavirus E6 protein [94, 95]. It may also modulate the apoptosis at differing rates [94] and modify sensitivity to chemotherapeutic agents [96]. The Pro72 variant exhibits decreased apoptotic potential than the Arg72 variant [87, 91, 93, 97], indicating that the two polymorphic variants of *TP53* are functionally distinct, which may influence the cancer risk or treatment [88, 97].

Some studies have reported the identification of Arg72Pro polymorphism and its role in many kinds of cancers such as cervical [98], lung [99–102], breast [103–108], colorectal [109, 110], esophageal [111–113], and gastric [114].

Even if *TP53* gene is highly polymorphic, the *TP53* codon 72 polymorphism is the only whose role has been extensively studied in relation to esophageal and gastric cancer and the results have been inconsistent (Table 1). Some studies in esophageal and gastric cancer of different populations have evidenced association of the Pro72 variant with cancer risk, while others with the Arg72 variant.

5.1. Association with Esophageal Cancer. In the last decades, several studies had been focused in the association between Arg72Pro polymorphism and esophageal cancer (EC) susceptibility, but the results are still conflicting and heterogeneous [88, 111–113, 115–119, 121, 132].

Some studies showed that in EC, the Pro allele was associated to protection [93] or that the Arg allele was associated with increased risk of EC [112], but others found the opposite, that Arg allele was associated to protection [88], or that the Pro allele was associated with increased risk of this disease [46, 87, 111, 113, 117, 120, 121]. Moreover, maintaining this functional change had been associated not only with increased risk, but also with earlier age of onset, reduced response to chemotherapy, and early recurrence in

a variety of cancers [85, 87]. However, other studies did not find any association between Arg72Pro polymorphism and susceptibility to ESCC [118, 132], either to EA, age of onset and stage of disease at the time of the diagnosis detection [74].

Piao et al. [120], in South Korea population, observed the Arg72Pro polymorphism was associated with an increased risk of EC and also found that smoking status changed the association between the Arg72Pro polymorphism and the risk of this cancer, so that the Odds Ratio of the Arg/Pro genotype was higher in ever-smokers than in never-smokers. Another study had indicated significant association between this polymorphism and smoking with risk of development of ESCC being the highest risk in smokers carrying Pro/Pro genotypes [111]. In addition, a study in a Chinese mainland population found that the Pro/Pro genotype was significantly associated with an increased risk of ESCC and the association was especially noteworthy in women and in younger patients [113].

Besides, Cescon et al. [87] showed that, among all EC patients treated with standard therapy, the Pro/Pro genotype was associated with shorter overall survival and progression, independent of standard clinical prognostic features, thus the authors suggest that TP53 could help the prediction of prognosis in EC, identifying high-risk patient's subgroups that might benefit themselves from new therapeutic strategies. Some recent meta-analyses have focused on Arg72Pro polymorphism on EC risk. For example, Jiang et al. [88] verified a significantly reduced risk of EC associated with TP53 genotypes (Arg/Arg + Arg/Pro versus Pro/Pro) and their analysis was restricted to welldesigned studies. Moreover, the Arg allele was significantly associated with decreased EC risk. Other meta-analysis showed that the Arg72Pro was associated with an increased risk of EC (Pro/Arg + Pro/Pro versus Arg/Arg) and the authors have observed no heterogeneity between the studies [133]. However, when the authors performed a stratified analysis by ethnicity, the increased risk of EC associated with Arg72Pro polymorphism (Pro/Arg + Pro/Pro versus Arg/Arg) was more evident in Asian group, thus their results suggest that Arg72Pro polymorphism may contribute to EC development, especially in Asians.

Despite of various studies assessing the functional *TP53* Arg72Pro polymorphism in relation to EC susceptibility, the results remain conflicting probably due to methodological errors such as selection bias, inappropriate specimens used for genotyping, or limited statistical power [88] and also ethnicity. Therefore, additional well-designed large studies still are required for the validation of this association.

5.2. Association with Gastric Cancer. Similarly to studies of association with EC, the relationship between the Arg72Pro polymorphism, and GC susceptibility is also controversial. Studies performed in southern China [122] and in Venezuela [134] suggest that Arg allele-carriers could be associated with the development of GC. In contrast, studies in Korea reported that Pro/Pro genotype was associated with increased risk of this neoplasm [95, 126]. While in a Chinese

Tumor site	Country (Ethnicity)	Case/Control (n)	Genotype frequency case/Control (%)			D. C
			Arg/Arg	Arg/Pro	Pro/Pro	Reference
Esophagus	China (Asians)	758/1420	26.2/29.9	44.9/51.5	28.9/18.6	[111]
	China (Asians)	435/550	85.7/49.6	4.4/35.8	9.9/14.6	[112]
	China (Asians)	673/694	24.2/28.1	45.5/52.7	30.3/19.2	[113]
	China (Asians)	62/50	43.0/20.0	34.0/52.0	23.0/28.0	[115]
	China (Asians)	120/232	24.0/29.0	50.0/52.0	27.0/18.0	[116]
	China (Asians)	218/415	20.1/30.1	43.6/45.8	36.3/24.2	[117]
	Japan (Asians)	102/241	36.3/37.7	50.0/44.4	13.7/18.0	[118]
	South Africa (Africans)	73/115	36.0/32.0	56.0/54.0	7.0/14.0	[119]
	South Korea (Asians)	340/1700	39.4/43.2	45.6/45.6	15.0/11.2	[120]
	Taiwan (Asians)	90/254	22.2/37.0	51.1/45.7	26.7/17.3	[121]
	United States (Caucasians)	312/454	53.0/57.0	39.0/35.0	8.0/8.0	[89]
Stomach	China (Asians)	324/317	29.6/29.6	55.6/50.5	14.8/19.9	[122]
	China (Asians)	140/125	15.7/25.6	60.0/54.4	24.3/20.0	[123]
	China (Asians)	500/1000	24.6/31.6	49.0/48.6	26.4/19.8	[124]
	Japan (Asians)	144/239	35.4/37.7	48.6/44.4	16.0/18.0	[118]
	Japan (Asians)	117/116	41.9/43.1	44.4/44.8	13.7/12.1	[125]
	Korea (Asians)	2213/1700	42.4/43.2	44.1/45.6	13.4/11.2	[95]
	Korea (Asians)	292/216	34.6/41.2	43.1/47.7	22.3/11.1	[126]
	Korea (Asians)	84/43	35.7/39.5	50.0/41.9	14.3/18.6	[127]
	Taiwan (Asians)	123/126	28.5/34.1	58.5/42.1	13.0/23.8	[128]
	Italy (Caucasians)	114/295	62.3/57.3	29.8/34.6	7.9/8.1	[129]
	United States (Caucasians)	155/134	33.1/36.3	46.8/45.5	20.1/18.2	[130]

120/277

47.6/45.1

Table 1: Frequency distribution of *TP53* codon 72 polymorphism genotype and association with risk of the gastric and esophageal cancers in the worldwide.

population, Ke-Xiang et al. [123] showed that Arg/Pro + Pro/Pro genotypes increased risk of GC. Corroborating these data, a recent meta-analysis suggests that the Pro/Pro genotype was associated with several types of cancer, including GC [93].

United Kingdom (Caucasians)

In contrast, other recent meta-analysis with 21 case-control studies did not associate the Arg72Pro polymorphism with the risk of GC. However, when subgroups were assessed according to anatomical site, it was found that Pro/Pro genotype was significantly associated with increased risk of cardia GC [89]. Others case-control studies performed in Asian [122, 124, 127, 132, 135] and Mexico [136] showed a higher frequency of Pro allele in cardia GC [124, 135] and of Arg allele in noncardia GC [122, 127, 136]. On the other hand, studies performed in United Kingdom revealed an increase in the frequency of the Arg allele in patients with cardia GC [131, 137], suggesting an effect of ethnical group beyond of location region in the stomach.

Corroborating these findings, recent meta-analyses about the distributions of the two polymorphic variants of *TP53* codon 72 and their effect on the risk of GC indicated that the anatomical site of tumor and ethnicity may contribute to the differences in the risk of gastric tumorigenesis [89]. The meta-analyses study by Zhou et al. [114] performed among eight studies in Asians and four in Caucasians showed that cardia GC had a significantly higher frequency of Pro/Pro genotype among Asians.

Another variable that should also be considered in the studies on association of the Arg72Pro polymorphism is the histological type of the GC. Whereas some authors found no relationship [126, 127, 129, 138], studies performed in China [139] and Korea [95] observed that the Pro allele carriers had a higher risk of developing the intestinal-type GC and others studies performed in Japan [125] and United States [130] found that this polymorphic allele was associated with an increased risk of developing the diffuse-type GC. The exact biological mechanism underlying the association between Arg72Pro polymorphism with the histological type is still unclear [95]. However, it has been suggested that the intestinal-type predominates in high-risk geographic areas such as East Asia, and it is related to the prevalence of H. pylori infection among the elderly, whereas the diffusetype is found more uniformly worldwide and is apparently unrelated to *H. pylori* prevalence [140].

7.3/8.3

[131]

45.1/46.6

Although the potential effects of Arg72Pro polymorphism and their interactions with location, histology, ethnicity, and environmental exposures on GC risk have been assessed in several studies in different population worldwide, their exact effect are still unknown. The genetic susceptibility to gastric carcinogenesis related to *TP53* polymorphisms may be attributed to several factors, including the accumulation and interaction of SNPs [124, 129], gene-environmental interactions [93], age [128, 131], *H. pylori* infection [123], bile or acid reflux, and smoking [137].

In general, these data suggest that the *TP53* polymorphism alone is insufficient to explain its effect on risk of cancer, but together with others genetic polymorphisms and environmental factors may modulate the individual risk of developing cancer.

6. Conclusions

In this study we focused on the participation of genetic alterations of *TP53* gene, such as mutational inactivation, LOH, SNPs, and expression of mutant form of p53 protein in the esophageal and gastric carcinogenesis. The studies emphasize the fundamental role of molecular alterations of "the guardian of the genome" in these neoplasms, with serious consequences for the deregulation of the cell cycle, loss of proapoptotic function and reduced sensitivity for anticancer drugs. Considering the involvement of *TP53* alterations both in early stages as in tumor progression, it is an important biomarker for the diagnosis, tumor progression, and poor prognosis associated with lymph nodes metastasis.

Recent studies have demonstrated another pathway of participation of the *TP53* gene in carcinogenesis, through the regulation of miRNAs. *TP53* regulates the transcription expression and the maturation of a group of miRNAs. On the other hand, miRNAs can regulate the activity and function of *TP53* through its direct repression or its regulators in cells. Thus these findings have demonstrated that miRNAs are important components in the *TP53* network [141].

Despite major advances and growing number of publications on the role of *TP53* tumor suppressor gene in carcinogenesis, still several other functions has emerged as cell metabolism, stem cells renewal and the occurrence of p53 isoform variants that may alter the function of wild-type p53. Therefore, p53 and other members of its family as p63 and p73 act in an intricate regulatory network controlling the expression of hundreds of target genes that regulate the cell cycle for the maintenance of genetic stability and preventing cancer formation.

Authors' Contribution

The authors M. Succi and M. A. Proença had the same contribution in the realization of the manuscript.

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References

- [1] D. M. Parkin, F. Bray, J. Ferlay, and P. Pisani, "Global cancer statistics, 2002," *Ca-A Cancer Journal for Clinicians*, vol. 55, no. 2, pp. 74–108, 2005.
- [2] E. Tahara, "Genetic pathways of two types of gastric cancer," *IARC Scientific Publications*, no. 157, pp. 327–349, 2004.
- [3] S. Bandla, A. Pennathur, J. D. Luketich et al., "Comparative genomics of esophageal adenocarcinoma and squamous cell

- carcinoma," Annals of Thoracic Surgery, vol. 93, no. 4, pp. 1101–1106, 2012.
- [4] A. V. Safatle-Ribeiro, U. Ribeiro Jr., P. Sakai et al., "Integrated p53 histopathologic/genetic analysis of premalignant lesions of the esophagus," *Cancer Detection and Prevention*, vol. 24, no. 1, pp. 13–23, 2000.
- [5] R. E. Kraichely and G. Farrugia, "Achalasia: physiology and etiopathogenesis," *Diseases of the Esophagus*, vol. 19, no. 4, pp. 213–223, 2006.
- [6] B. L. D. M. Brücher, H. J. Stein, H. Bartels, H. Feussner, and J. R. Siewert, "Achalasia and esophageal cancer: incidence, prevalence, and prognosis," *World Journal of Surgery*, vol. 25, no. 6, pp. 745–749, 2001.
- [7] A. M. Macedo and S. D. J. Pena, "Genetic variability of Trypanosoma cruzi: implications for the pathogenesis of Chagas disease," Parasitology Today, vol. 14, no. 3, pp. 119– 124, 1998.
- [8] P. Lauren, "The two histological main types of gastric carcinoma: diffuse and so-called intestinal-type carcinoma—an attempt at a histo-clinical classification," *Acta Pathologica et Microbiologica Scandinavica*, vol. 64, pp. 31–49, 1965.
- [9] P. Correa, "A human model of gastric carcinogenesis," *Cancer Research*, vol. 48, no. 13, pp. 3554–3560, 1988.
- [10] L. T. Nguyen, T. Uchida, K. Murakami, T. Fujioka, and M. Moriyama, "Helicobacter pylori virulence and the diversity of gastric cancer in Asia," Journal of Medical Microbiology, vol. 57, no. 12, pp. 1445–1453, 2008.
- [11] E. J. Kuipers, "Review article: exploring the link between *Helicobacter pylori* and gastric cancer," *Alimentary Pharmacology and Therapeutics, Supplement*, vol. 13, no. 1, pp. 3–11, 1999.
- [12] G. H. A. Scholte, L. J. Van Doorn, A. Cats et al., "Genotyping of *Helicobacter pylori* in paraffin-embedded gastric biopsy specimens: relation to histological parameters and effects on therapy," *The American Journal of Gastroenterology*, vol. 97, no. 7, pp. 1687–1695, 2002.
- [13] M. G. Smith, G. L. Hold, E. Tahara, and E. M. El-Omar, "Cellular and molecular aspects of gastric cancer," World Journal of Gastroenterology, vol. 12, no. 18, pp. 2979–2990, 2006.
- [14] C. A. J. Ong, P. Lao-Sirieix, and R. C. Fitzgerald, "Biomarkers in Barrett's esophagus and esophageal adenocarcinoma: predictors of progression and prognosis," World Journal of Gastroenterology, vol. 16, no. 45, pp. 5669–5681, 2010.
- [15] S. Benchimol, P. Lamb, and L. V. Crawford, "Transformation associated p53 protein is encoded by a gene on human chromosome 17," *Somatic Cell and Molecular Genetics*, vol. 11, no. 5, pp. 505–509, 1985.
- [16] W. S. El-Deiry, S. E. Kern, J. A. Pietenpol, K. W. Kinzler, and B. Vogelstein, "Definition of a consensus binding site for p53," *Nature Genetics*, vol. 1, no. 1, pp. 45–49, 1992.
- [17] J. A. Pietenpol, T. Tokino, S. Thiagalingam, W. S. El-Deiry, K. W. Kinzler, and B. Vogelstein, "Sequence-specific transcriptional activation is essential for growth suppression by p53," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 91, no. 6, pp. 1998–2002, 1994.
- [18] V. A. Belyi, P. Ak, E. Markert et al., "The origins and evolution of the p53 family of genes," *Cold Spring Harbor Perspectives in Biology*, vol. 2, no. 6, Article ID a001198, 2010.
- [19] K. H. Vousden and D. P. Lane, "p53 in health and disease," Nature Reviews Molecular Cell Biology, vol. 8, no. 4, pp. 275– 283, 2007.

- [20] Y. Aylon and M. Oren, "New plays in the p53 theater," Current Opinion in Genetics and Development, vol. 21, no. 1, pp. 86– 92, 2011.
- [21] D. P. Lane, "p53, guardian of the genome," *Nature*, vol. 358, no. 6381, pp. 15–16, 1992.
- [22] K. Suzuki and H. Matsubara, "Recent advances in p53 research and cancer treatment," *Journal of Biomedicine and Biotechnology*, vol. 2011, Article ID 978312, 7 pages, 2011.
- [23] A. Nakagawara and T. Ozaki, "P53: the attractive tumor suppressor in the cancer research field," *Journal of Biomedicine* and *Biotechnology*, vol. 2011, Article ID 603925, 13 pages, 2011.
- [24] A. Egashira, M. Morita, R. Yoshida et al., "Loss of p53 in esophageal squamous cell carcinoma and the correlation with survival: analyses of gene mutations, protein expression, and loss of heterozygosity in Japanese patients," *Journal of Surgical Oncology*, vol. 104, no. 2, pp. 169–175, 2011.
- [25] International Agency for Research on Cancer (IARC), "Polymorphisms in TP53 gene sequence," 2012, http://www-p53.iarc.fr.
- [26] A. I. Robles and C. C. Harris, "Clinical outcomes and correlates of TP53 mutations and cancer," Cold Spring Harbor Perspectives in Biology, vol. 2, no. 3, Article ID a001016, 2010.
- [27] M. Olivier, R. Eeles, M. Hollstein, M. A. Khan, C. C. Harris, and P. Hainaut, "The IARC *TP53* database: new online mutation analysis and recommendations to users," *Human Mutation*, vol. 19, no. 6, pp. 607–614, 2002.
- [28] A. Petitjean, M. I. W. Achatz, A. L. Borresen-Dale, P. Hainaut, and M. Olivier, "TP53 mutations in human cancers: functional selection and impact on cancer prognosis and outcomes," Oncogene, vol. 26, no. 15, pp. 2157–2165, 2007.
- [29] L. V. Crawford, D. C. Pim, and P. Lamb, "The cellular protein p53 in human tumours," *Molecular Biology and Medicine*, vol. 2, no. 4, pp. 261–272, 1984.
- [30] G. Cattoretti, F. Rilke, S. Andreola, L. D'Amato, and D. Delia, "P53 expression in breast cancer," *International Journal of Cancer*, vol. 41, no. 2, pp. 178–183, 1988.
- [31] S. E. Kern, J. A. Pietenpol, S. Thiagalingam, A. Seymour, K. W. Kinzler, and B. Vogelstein, "Oncogenic forms of p53 inhibit p53-regulated gene expression," *Science*, vol. 256, no. 5058, pp. 827–830, 1992.
- [32] M. Hachiya, A. Chumakov, C. W. Miller, M. Akashi, J. Said, and H. P. Koeffler, "Mutant p53 proteins behave in a dominant, negative fashion in vivo," *Anticancer Research*, vol. 14, no. 5, pp. 1853–1859, 1994.
- [33] G. W. Falk, "Barrett's esophagus," *Gastroenterology*, vol. 122, no. 6, pp. 1569–1591, 2002.
- [34] C. P. Wild and L. J. Hardie, "Reflux, Barrett's oesophagus and adenocarcinoma: burning questions," *Nature Reviews Cancer*, vol. 3, no. 9, pp. 676–684, 2003.
- [35] K. Schulmann, A. Sterian, A. Berki et al., "Inactivation of p16, RUNX3, and HPP1 occurs early in Barrett's-associated neoplastic progression and predicts progression risk," *Oncogene*, vol. 24, no. 25, pp. 4138–4148, 2005.
- [36] G. Clément, R. Braunschweig, N. Pasquier, F. T. Bosman, and J. Benhattar, "Methylation of APC, TIMP3, and TERT: a new predictive marker to distinguish Barrett's oesophagus patients at risk for malignant transformation," *Journal of Pathology*, vol. 208, no. 1, pp. 100–107, 2006.
- [37] P. M. Schneider, A. G. Casson, B. Levin et al., "Mutations of p53 in Barrett's esophagus and Barrett's cancer: a prospective study of ninety-eight cases," *The Journal of Thoracic and Cardiovascular Surgery*, vol. 111, no. 2, pp. 323–333, 1996.

- [38] C. M. Gleeson, J. M. Sloan, D. T. McManus et al., "Comparison of p53 and DNA content abnormalities in adenocarcinoma of the oesophagus and gastric cardia," *British Journal of Cancer*, vol. 77, no. 2, pp. 277–286, 1998.
- [39] B. J. Reid, L. J. Prevo, P. C. Galipeau et al., "Predictors of progression in Barrett's esophagus II: baseline 17p (p53) loss of heterozygosity identifies a patient subset at increased risk for neoplastic progression," *The American Journal of Gastroenterology*, vol. 96, no. 10, pp. 2839–2848, 2001.
- [40] C. C. Maley, P. C. Galipeau, X. Li, C. A. Sanchez, T. G. Paulson, and B. J. Reid, "Selectively advantageous mutations and hitchhikers in neoplasms: p16 lesions are selected in Barrett's esophagus," *Cancer Research*, vol. 64, no. 10, pp. 3414–3427, 2004.
- [41] K. Madani, R. Zhao, H. J. Lim, and A. G. Casson, "Prognostic value of p53 mutations in oesophageal adenocarcinoma: final results of a 15-year prospective study," *European Journal of Cardio-Thoracic Surgery*, vol. 37, no. 6, pp. 1427–1432, 2010.
- [42] Y. C. Cai, C. K. So, A. Y. Nie et al., "Characterization of genetic alteration patterns in human esophageal squamous cell carcinoma using selected microsatellite markers spanning multiple loci," *International Journal of Oncology*, vol. 30, no. 5, pp. 1059–1067, 2007.
- [43] E. Oki, Y. Zhao, R. Yoshida et al., "The difference in p53 mutations between cancers of the upper and lower gastrointestinal tract," *Digestion*, vol. 79, no. 1, pp. 33–39, 2009.
- [44] S. G. Kim, S. J. Hong, K. W. Kwon et al., "The expression of p53, p16, cyclin D1 in esophageal squamous cell carcinoma and esophageal dysplasia," *The Korean Journal of Gastroenterology*, vol. 48, no. 4, pp. 269–276, 2006.
- [45] B. Abedi-Ardekani, F. Kamangar, M. Sotoudeh et al., "Extremely high *TP53* mutation load in esophageal squamous cell carcinoma in Golestan Province, Iran," *PLoS ONE*, vol. 6, no. 12, Article ID e29488, 2011.
- [46] A. P. F. Silveira, F. D. S. Manoel-Caetano, S. Aoki, L. H. T. Yamasaki, P. Rahal, and A. E. Silva, "Gene mutations and polymorphisms of *TP53* and FHIT in chronic esophagitis and esophageal carcinoma," *Anticancer Research*, vol. 31, no. 5, pp. 1685–1690, 2011.
- [47] H. Saeki, H. Kitao, K. Yoshinaga et al., "Copy-neutral loss of heterozygosity at the p53 locus in carcinogenesis of esophageal squamous cell carcinomas associated with p53 mutations," *Clinical Cancer Research*, vol. 17, no. 7, pp. 1731– 1740, 2011.
- [48] G. Tamura, "Alterations of tumor suppressor and tumorrelated genes in the development and progression of gastric cancer," *World Journal of Gastroenterology*, vol. 12, no. 2, pp. 192–198, 2006.
- [49] P. Vogiatzi, C. Vindigni, F. Roviello, A. Renieri, and A. Giordano, "Deciphering the underlying genetic and epigenetic events leading to gastric carcinogenesis," *Journal of Cellular Physiology*, vol. 211, no. 2, pp. 287–295, 2007.
- [50] S. Nobili, L. Bruno, I. Landini et al., "Genomic and genetic alterations infuence the progression of gastric cancer," World Journal of Gastroenterology, vol. 17, no. 3, pp. 290–299, 2011.
- [51] H. Iwamatsu, K. Nishikura, H. Watanabe et al., "Heterogeneity of p53 mutational status in the superficial spreading type of early gastric carcinoma," *Gastric Cancer*, vol. 4, no. 1, pp. 20–26, 2001.
- [52] X. P. Liu, K. Tsushimi, M. Tsushimi et al., "Expression of p53 protein as a prognostic indicator of reduced survival time in diffuse-type gastric carcinoma," *Pathology International*, vol. 51, no. 6, pp. 440–444, 2001.

- [53] C. M. Fenoglio-Preiser, J. Wang, G. N. Stemmermann, and A. Noffsinger, "*TP53* and gastric carcinoma: a review," *Human Mutation*, vol. 21, no. 3, pp. 258–270, 2003.
- [54] T. Nguyen, D. Brunson, C. L. Crespi, B. W. Penman, J. S. Wishnok, and S. R. Tannenbaum, "DNA damage and mutation in human cells exposed to nitric oxide in vitro," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 89, no. 7, pp. 3030–3034, 1992.
- [55] H. C. Chen, H. J. Chen, M. A. Khan et al., "Genetic mutations of p53 and k-ras in gastric carcinoma patients from Hunan, China," *Tumor Biology*, vol. 32, no. 2, pp. 367–373, 2011.
- [56] J. Zhao, Y. Lu, and H.-M. Shen, "Targeting p53 as a therapeutic strategy in sensitizing TRAIL-induced apoptosis in cancer cells," *Cancer Letters*, vol. 314, no. 1, pp. 8–23, 2012.
- [57] J. E. Chipuk, T. Kuwana, L. Bouchier-Hayes et al., "Direct activation of Bax by p53 mediates mitochondrial membrane permeabilization and apoptosis," *Science*, vol. 303, no. 5660, pp. 1010–1014, 2004.
- [58] J. I. J. Leu, P. Dumont, M. Hafey, M. E. Murphy, and D. L. George, "Mitochondrial p53 activates Bak and causes disruption of a Bak-Mcl1 complex," *Nature Cell Biology*, vol. 6, no. 5, pp. 443–450, 2004.
- [59] J. J. Fuster, S. M. Sanz-González, U. M. Moll, and V. Andrés, "Classic and novel roles of p53: prospects for anticancer therapy," *Trends in Molecular Medicine*, vol. 13, no. 5, pp. 192–199, 2007.
- [60] D. Speidel, "Transcription-independent p53 apoptosis: an alternative route to death," *Trends in Cell Biology*, vol. 20, no. 1, pp. 14–24, 2010.
- [61] Y. Yang, C. S. Deng, J. Z. Peng, B. C. Y. Wong, S. K. Lam, and H. H. X. Xia, "Effect of *Helicobacter pylori* on apoptosis and apoptosis related genes in gastric cancer cells," *Molecular Pathology*, vol. 56, no. 1, pp. 19–24, 2003.
- [62] M. Naumann and J. E. Crabtree, "Helicobacter pylori-induced epithelial cell signalling in gastric carcinogenesis," Trends in Microbiology, vol. 12, no. 1, pp. 29–36, 2004.
- [63] A. C. Targa, A. C. G. César, P. M. Cury, and A. E. Silva, "Apoptosis in different gastric lesions and gastric cancer: relationship with *Helicobacter pylori*, overexpression of p53 and aneuploidy," *Genetics and Molecular Research*, vol. 6, no. 3, pp. 554–565, 2007.
- [64] K. Triantafyllou, P. Kitsanta, D. G. Karamanolis, C. Kittas, and S. D. Ladas, "Epithelial cell turnover, p53 and bcl-2 protein expression during oncogenesis of early and advanced gastric cancer in a Western population," *Digestive and Liver Disease*, vol. 40, no. 1, pp. 39–45, 2008.
- [65] X. Liu, H. Cai, H. Huang, Z. Long, Y. Shi, and Y. Wang, "The prognostic significance of apoptosis-related biological markers in chinese gastric cancer patients," *PLoS ONE*, vol. 6, no. 12, Article ID e29670, 2011.
- [66] D. L. Persons, W. S. Croughan, K. A. Borelli, and R. Cherian, "Interphase cytogenetics of esophageal adenocarcinoma and precursor lesions," *Cancer Genetics and Cytogenetics*, vol. 106, no. 1, pp. 11–17, 1998.
- [67] J. Mueller, M. Werner, and J. R. Siewert, "Malignant progression in Barrett's esophagus: pathology and molecular biology," *Recent Results in Cancer Research*, vol. 155, pp. 29–41, 2000.
- [68] R. Langer, B. H. A. Von Rahden, J. Nahrig et al., "Prognostic significance of expression patterns of c-erbB-2, p53, p16 INK4A, p27KIP1, cyclin D1 and epidermal growth factor receptor in oesophageal adenocarcinoma: a tissue microarray study," *Journal of Clinical Pathology*, vol. 59, no. 6, pp. 631–634, 2006.

- [69] Y. Yang, J. Fruehauf, S. Xiang, and C. J. Li, "Genomic instability in precancerous lesions before inactivation of tumor suppressors p53 and APC in patients," *Cell Cycle*, vol. 5, no. 13, pp. 1443–1447, 2006.
- [70] R. Cestari, V. Villanacci, E. Rossi et al., "Fluorescence in situ hybridization to evaluate dysplasia in Barrett's esophagus: a pilot study," *Cancer Letters*, vol. 251, no. 2, pp. 278–287, 2007.
- [71] M. F. Bellini, K. R. M. Leite, P. M. Cury, and A. E. Silva, "p53, p16 and Fhit proteins expressions in chronic esophagitis and Chagas disease," *Anticancer Research*, vol. 28, no. 6 A, pp. 3793–3799, 2008.
- [72] S. Ito, T. Ohga, H. Saeki et al., "p53 mutation profiling of multiple esophageal carcinoma using laser capture microdissection to demonstrate field carcinogenesis," *International Journal of Cancer*, vol. 113, no. 1, pp. 22–28, 2005.
- [73] U. Han, O. I. Can, S. Han, B. Kayhan, and B. U. Onal, "Expressions of p53, VEGF C, p21: could they be used in preoperative evaluation of lymph node metastasis of esophageal squamous cell carcinoma?" *Diseases of the Esophagus*, vol. 20, no. 5, pp. 379–385, 2007.
- [74] G. Liu, D. W. Cescon, R. Zhai et al., "p53 Arg72Pro, MDM2 T309G and CCND1 G870A polymorphisms are not associated with susceptibility to esophageal adenocarcinoma," Diseases of the Esophagus, vol. 23, no. 1, pp. 36–39, 2010.
- [75] D. C. Lin, Z. Z. Shi, L. Y. Xue et al., "Expression of cell cycle related proteins cyclin D1, p53 and p21 WAFI/CIP1 in esophageal squamous cell carcinoma," *Yi Chuan*, vol. 32, no. 5, pp. 455–460, 2010.
- [76] M. Yamasaki, H. Miyata, Y. Fujiwara et al., "P53 genotype predicts response to chemotherapy in patients with squamous cell carcinoma of the esophagus," *Annals of Surgical Oncology*, vol. 17, no. 2, pp. 634–642, 2010.
- [77] A. C. G. César, A. A. Borim, A. Caetano, P. M. Cury, and A. E. Silva, "Aneuploidies, deletion, and overexpression of TP53 gene in intestinal metaplasia of patients without gastric cancer," Cancer Genetics and Cytogenetics, vol. 153, no. 2, pp. 127–132, 2004.
- [78] A. C. Gobbo César, M. de Freitas Calmon, P. M. Cury, A. Caetano, A. A. Borim, and A. E. Silva, "Genetic alterations in benign lesions: chronic gastritis and gastric ulcer," World Journal of Gastroenterology, vol. 12, no. 4, pp. 625–629, 2006.
- [79] A. S. Khayat, A. C. Guimarães, D. Q. Calcagno et al., "Interrelationship between *TP53* gene deletion, protein expression and chromosome 17 aneusomy in gastric adenocarcinoma," *BMC Gastroenterology*, vol. 9, article 55, 2009.
- [80] H. Z. Lu, Y. P. Wu, W. Luo et al., "Correlation between aneuploidy of chromosome 17, over-expression of *TP53* and TOP-IIalpha, and the clinicopathological features and diagnosis of gastric adenocarcinoma," *Zhonghua Zhong Liu Za Zhi*, vol. 31, no. 10, pp. 754–758, 2009.
- [81] X. Li, F. Luo, Q. Li et al., "Identification of new aberrantly expressed miRNAs in intestinal-type gastric cancer and its clinical significance," *Oncology Reports*, vol. 26, no. 6, pp. 1431–1439, 2011.
- [82] B.-G. Jang and W. H. Kim, "Molecular pathology of gastric carcinoma," *Pathobiology*, vol. 78, no. 6, pp. 302–310, 2011.
- [83] K. Iijima, Y. Abe, T. Koike et al., "Gastric cancers emerging after *H. pylori* eradication arise exclusively from non-acid-secreting areas," *The Tohoku Journal of Experimental Medicine*, vol. 226, no. 1, pp. 45–53, 2012.
- [84] C. Whibley, P. D. P. Pharoah, and M. Hollstein, "p53 polymorphisms: cancer implications," *Nature Reviews Cancer*, vol. 9, no. 2, pp. 95–107, 2009.

- [85] E. C. Pietsch, O. Humbey, and M. E. Murphy, "Polymorphisms in the p53 pathway," *Oncogene*, vol. 25, no. 11, pp. 1602–1611, 2006.
- [86] G. L. Bond and A. J. Levine, "A single nucleotide polymorphism in the p53 pathway interacts with gender, environmental stresses and tumor genetics to influence cancer in humans," *Oncogene*, vol. 26, no. 9, pp. 1317–1323, 2007.
- [87] D. W. Cescon, P. A. Bradbury, K. Asomaning et al., "p53 Arg72Pro and MDM2 T309G polymorphisms, histology, and esophageal cancer prognosis," Clinical Cancer Research, vol. 15, no. 9, pp. 3103–3109, 2009.
- [88] D. K. Jiang, L. Yao, W. Z. Wang et al., "TP53 Arg72Pro polymorphism is associated with esophageal cancer risk: a meta-analysis," World Journal of Gastroenterology, vol. 17, no. 9, pp. 1227–1233, 2011.
- [89] L. Liu, K. Wang, Z. M. Zhu, and J. H. Shao, "Associations between *p53 Arg72Pro* and development of digestive tract cancers: a meta-analysis," *Archives of Medical Research*, vol. 42, no. 1, pp. 60–69, 2011.
- [90] M. Umar, R. Upadhyay, R. Khurana, S. Kumar, U. C. Ghoshal, and B. Mittal, "Role of *p53* and *p73* genes polymorphisms in susceptibility to esophageal cancer: a case control study in a northern Indian population," *Molecular Biology Reports*, vol. 39, no. 2, pp. 1153–1162, 2011.
- [91] G. J. Matlashewski, S. Tuck, and D. Pim, "Primary structure polymorphism at amino acid residue 72 of human p53," *Molecular and Cellular Biology*, vol. 7, no. 2, pp. 961–963, 1987.
- [92] M. Thomas, A. Kalita, S. Labrecque, D. Pim, L. Banks, and G. Matlashewski, "Two polymorphic variants of wild-type p53 differ biochemically and biologically," *Molecular and Cellular Biology*, vol. 19, no. 2, pp. 1092–1100, 1999.
- [93] G. Francisco, P. R. Menezes, J. Eluf-Neto, and R. Chammas, "Arg72Pro TP53 polymorphism and cancer susceptibility: a comprehensive meta-analysis of 302 case-control studies," *International Journal of Cancer*, vol. 129, no. 4, pp. 920–930, 2011.
- [94] I. J. Dahabreh, H. Linardou, P. Bouzika, V. Varvarigou, and S. Murray, "TP53 Arg72Pro polymorphism and colorectal cancer risk: a systematic review and meta-analysis," Cancer Epidemiology Biomarkers and Prevention, vol. 19, no. 7, pp. 1840–1847, 2010.
- [95] H. R. Song, S. S. Kweon, H. N. Kim et al., "Erratum to: p53 codon 72 polymorphism in patients with gastric and colorectal cancer in a Korean population," *Gastric Cancer*, vol. 14, no. 3, article 248, 2011.
- [96] F. Gianfagna, E. De Feo, C. M. van Duijn, G. Ricciardi, and S. Boccia, "A systematic review of meta-analyses on gene polymorphisms and gastric cancer risk," *Current Genomics*, vol. 9, no. 6, pp. 361–374, 2008.
- [97] P. Dumont, J. I. J. Leu, A. C. Della Pietra, D. L. George, and M. Murphy, "The codon 72 polymorphic variants of p53 have markedly different apoptotic potential," *Nature Genetics*, vol. 33, no. 3, pp. 357–365, 2003.
- [98] S. J. Klug, M. Ressing, J. Koenig et al., "TP53 codon 72 polymorphism and cervical cancer: a pooled analysis of individual data from 49 studies," The Lancet Oncology, vol. 10, no. 8, pp. 772–784, 2009.
- [99] O. Popanda, L. Edler, P. Waas et al., "Elevated risk of squamous-cell carcinoma of the lung in heavy smokers carrying the variant alleles of the *TP53* Arg72Pro and p21 Ser31Arg polymorphisms," *Lung Cancer*, vol. 55, no. 1, pp. 25–34, 2007.

- [100] S. A. Nadji, M. Mahmoodi, A. A. Ziaee et al., "An increased lung cancer risk associated with codon 72 polymorphism in the *TP53* gene and human papillomavirus infection in Mazandaran province, Iran," *Lung Cancer*, vol. 56, no. 2, pp. 145–151, 2007.
- [101] S. Dai, C. Mao, L. Jiang, G. Wang, and H. Cheng, "P53 poly-morphism and lung cancer susceptibility: a pooled analysis of 32 case-control studies," *Human Genetics*, vol. 125, no. 5-6, pp. 633–638, 2009.
- [102] Y. Lili, Z. Deqiang, C. Chengwen et al., "TP53 Arg72Pro polymorphism and lung cancer risk: a meta-analysis," International Journal of Cancer, vol. 125, no. 12, pp. 2903–2911, 2009.
- [103] A. P. S. Damin, A. P. G. Frazzon, D. C. Damin et al., "Evidence for an association of *TP53* codon 72 polymorphism with breast cancer risk," *Cancer Detection and Prevention*, vol. 30, no. 6, pp. 523–529, 2006.
- [104] S. Gochhait, S. I. Bukhari, N. Bairwa et al., "Implication of BRCA2 -26G>A 5' untranslated region polymorphism in susceptibility to sporadic breast cancer and its modulation by p53 codon 72 Arg>Pro polymorphism," *Breast Cancer Research*, vol. 9, no. 5, article R71, 2007.
- [105] C. Baynes, C. S. Healey, K. A. Pooley et al., "Common variants in the ATM, BRCA1, BRCA2, CHEK2 and TP53 cancer susceptibility genes are unlikely to increase breast cancer risk," Breast Cancer Research, vol. 9, no. 2, article R27, 2007.
- [106] M. M. Gaudet, M. D. Gammon, J. T. Bensen et al., "Genetic variation of *TP53*, polycyclic aromatic hydrocarbon-related exposures, and breast cancer risk among women on Long Island, New York," *Breast Cancer Research and Treatment*, vol. 108, no. 1, pp. 93–99, 2008.
- [107] S. Costa, D. Pinto, D. Pereira et al., "Importance of *TP53* codon 72 and intron 3 duplication 16bp polymorphisms in prediction of susceptibility on breast cancer," *BMC Cancer*, vol. 8, article 32, 2008.
- [108] Z. Zhang, M. Wang, D. Wu et al., "P53 codon 72 polymorphism contributes to breast cancer risk: a meta-analysis based on 39 case-control studies," *Breast Cancer Research and Treatment*, vol. 120, no. 2, pp. 509–517, 2010.
- [109] X. L. Tan, A. Nieters, M. Hoffmeister, L. Beckmann, H. Brenner, and J. Chang-Claude, "Genetic polymorphisms in TP53, nonsteroidal anti-inflammatory drugs and the risk of colorectal cancer: evidence for gene-environment interaction?" Pharmacogenetics and Genomics, vol. 17, no. 8, pp. 639–645, 2007.
- [110] A. Dakouras, N. Nikiteas, E. Papadakis et al., "p53Arg72 homozygosity and its increased incidence in left-sided sporadic colorectal adenocarcinomas, in a Greek-Caucasian population," *Anticancer Research*, vol. 28, no. 2, pp. 1039– 1043, 2008.
- [111] Y. Hong, X. Miao, X. Zhang et al., "The role of P53 and MDM2 polymorphisms in the risk of esophageal squamous cell carcinoma," *Cancer Research*, vol. 65, no. 20, pp. 9582–9587, 2005.
- [112] W. Yang, Y. Zhang, X. Tian, T. Ning, and Y. Ke, "p53 codon 72 polymorphism and the risk of esophageal squamous cell carcinoma," *Molecular Carcinogenesis*, vol. 47, no. 2, pp. 100–104, 2008.
- [113] Y. Shao, W. Tan, and S. Zhang, "P53 gene codon 72 polymorphism and risk of esophageal squamous cell carcinoma: a case/control study in a Chinese population," *Diseases of the Esophagus*, vol. 21, no. 2, pp. 139–143, 2008.
- [114] Y. Zhou, N. Li, W. Zhuang et al., "P53 codon 72 polymorphism and gastric cancer: a meta-analysis of the literature,"

- International Journal of Cancer, vol. 121, no. 7, pp. 1481–1486, 2007.
- [115] T. Li, Z. M. Lu, M. Guo et al., "p53 Codon 72 polymorphism (C/G) and the risk of human papillomavirus-associated carcinomas in China," *Cancer*, vol. 95, no. 12, pp. 2571–2576, 2002.
- [116] N. Hu, W. J. Li, H. Su et al., "Common genetic variants of TP53 and BRCA2 in esophageal cancer patients and healthy individuals from low and high risk areas of northern China," Cancer Detection and Prevention, vol. 27, no. 2, pp. 132–138, 2003.
- [117] L. Cai, L. N. Mu, H. Lu et al., "Dietary selenium intake and genetic polymorphisms of the GSTP1 and p53 genes on the risk of esophageal squamous cell carcinoma," *Cancer Epidemiology Biomarkers and Prevention*, vol. 15, no. 2, pp. 294–300, 2006.
- [118] N. Hamajima, K. Matsuo, T. Suzuki et al., "No associations of p73 G4C14-to-A4T14 at exon 2 and *p53 Arg72Pro* polymorphisms with the risk of digestive tract cancers in Japanese," *Cancer Letters*, vol. 181, no. 1, pp. 81–85, 2002.
- [119] M. Vos, C. H. Adams, T. C. Victor, and P. D. Van Helden, "Polymorphisms and mutations found in the regions flanking exons 5 to 8 of the *TP53* gene in a population at high risk for esophageal cancer in South Africa," *Cancer Genetics and Cytogenetics*, vol. 140, no. 1, pp. 23–30, 2003.
- [120] J. M. Piao, H. N. Kim, H. R. Song et al., "p53 codon 72 polymorphism and the risk of esophageal cancer: a Korean case-control study," *Diseases of the Esophagus*, vol. 24, no. 8, pp. 596–600, 2011.
- [121] J.-M. Lee, Y.-C. Lee, S.-Y. Yang et al., "Genetic polymorphisms of p53 and GSTP1, but not NAT2, are associated with susceptibility to squamous-cell carcinoma of the esophagus," *International Journal of Cancer*, vol. 89, no. 5, pp. 458–464, 2000.
- [122] H. Shen, A. Solari, X. Wang et al., "P53 codon 72 polymorphism and risk of gastric cancer in a Chinese population," *Oncology Reports*, vol. 11, no. 5, pp. 1115–1120, 2004.
- [123] Z. Ke-Xiang, L. Yu-Min, L. Xun, Z. Wen-Ce, S. Yong, and L. Tao, "Study on the association of p53 codon 72 polymorphisms with risk of gastric cancer in high incidence Hexi area of Gansu Province in China," *Molecular Biology Reports*, vol. 39, no. 1, pp. 723–728, 2011.
- [124] M. Yang, Y. Guo, X. Zhang et al., "Interaction of *p53 Arg72Pro* and *MDM2 T309G* polymorphisms and their associations with risk of gastric cardia cancer," *Carcinogenesis*, vol. 28, no. 9, pp. 1996–2001, 2007.
- [125] T. Hiyama, S. Tanaka, Y. Kitadai et al., "p53 codon 72 polymorphism in gastric cancer susceptibility in patients with Helicobacter pylori-associated chronic gastritis," *International Journal of Cancer*, vol. 100, no. 3, pp. 304–308, 2002.
- [126] S. Y. Yi and W. J. Lee, "A p53 genetic polymorphism of gastric cancer: difference between early gastric cancer and advanced gastric cancer," World Journal of Gastroenterology, vol. 12, no. 40, pp. 6536–6539, 2006.
- [127] W. C. Chung, K. M. Lee, B. I. Lee et al., "p53 genetic polymorphism of gastric cancer in Korea," *Korean Journal of Internal Medicine*, vol. 21, no. 1, pp. 28–32, 2006.
- [128] K. C. Lai, W. C. Chen, L. B. Jeng, S. Y. Li, M. C. Chou, and F. J. Tsai, "Association of genetic polymorphisms of MK, IL-4, p16, p21, p53 genes and human gastric cancer in Taiwan," *European Journal of Surgical Oncology*, vol. 31, no. 10, pp. 1135–1140, 2005.
- [129] E. De Feo, R. Persiani, A. La Greca et al., "A case-control study on the effect of p53 and p73 gene polymorphisms on gastric

- cancer risk and progression," *Mutation Research*, vol. 675, no. 1-2, pp. 60–65, 2009.
- [130] J. Sul, G. P. Yu, Q. Y. Lu et al., "P53 Codon 72 polymorphisms: a case-control study of gastric cancer and potential interactions," *Cancer Letters*, vol. 238, no. 2, pp. 210–223, 2006.
- [131] Z. W. Zhang, P. Newcomb, A. Hollowood et al., "Age-associated increase of codon 72 arginine p53 frequency in gastric cardia and non-cardia adenocarcinoma," *Clinical Cancer Research*, vol. 9, no. 6, pp. 2151–2156, 2003.
- [132] D. Peixoto Guimaraes, S. Hsin Lu, P. Snijders et al., "Absence of association between HPV DNA, TP53 codon 72 polymorphism, and risk of oesophageal cancer in a high-risk area of China," Cancer Letters, vol. 162, no. 2, pp. 231–235, 2001.
- [133] Y. Zhao, F. Wang, S. Shan et al., "Genetic polymorphism of p53, but not GSTP1, is association with susceptibility to esophageal cancer risk-a meta-analysis," *International Journal of Medical Sciences*, vol. 7, no. 5, pp. 300–308, 2010.
- [134] M. Cañas, Y. Morán, M. E. Camargo et al., "*TP53* codon 72 polymorphism and gastric cancer risk: a case-control study in individuals from the central-western region of Venezuela," *Investigacion Clinica*, vol. 50, no. 2, pp. 153–161, 2009.
- [135] Y. Zhou, N. Li, W. Zhuang, and X. Wu, "P53 codon 72 polymorphism and gastric cancer risk in a Chinese Han population," *Genetic Testing and Molecular Biomarkers*, vol. 14, no. 6, pp. 829–833, 2010.
- [136] G. I. Pérez-Pérez, F. J. Bosques-Padilla, M. L. Crosatti, R. Tijerina-Menchaca, and E. Garza-González, "Role of p53 codon 72 polymorphism in the risk of development of distal gastric cancer," *Scandinavian Journal of Gastroenterology*, vol. 40, no. 1, pp. 56–60, 2005.
- [137] Z. W. Zhang, P. Newcomb, A. Hollowood et al., "A comparison study of gastric cancer risk in patients with duodenal and gastric ulcer: roles of gastric mucosal histology and p53 codon 72 polymorphism," *Digestive Diseases and Sciences*, vol. 49, no. 2, pp. 254–259, 2004.
- [138] A. S. Khayat, L. Lobo Gatti, E. Moura Lima et al., "Polymorphisms of the TP53 codon 72 and WRN codon 1367 in individuals from Northern Brazil with gastric adenocarcinoma," Clinical and Experimental Medicine, vol. 5, no. 4, pp. 161–168, 2005.
- [139] Y. G. Xi, K. Y. Ding, X. L. Su et al., "p53 polymorphism and p21^{WAF1/CIP1} haplotype in the intestinal gastric cancer and the precancerous lesions," *Carcinogenesis*, vol. 25, no. 11, pp. 2201–2206, 2004.
- [140] K. D. Crew and A. I. Neugut, "Epidemiology of gastric cancer," World Journal of Gastroenterology, vol. 12, no. 3, pp. 354– 362, 2006.
- [141] Z. Feng, C. Zhang, R. Wu, and W. Hu, "Tumor suppressor p53 meets microRNAs," *Journal of Molecular Cell Biology*, vol. 3, no. 1, pp. 44–50, 2011.