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# Letter to the Editor

# An autopsied FTDP-17 case with MAPT IVS 10 + 14C > T mutation presenting with frontotemporal dementia

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# Dear Editor,

Frontotemporal dementia and parkinsonism linked to chromosome 17 (FTDP-17) due to *MAPT* mutation is a heterogeneous genetic neurodegenerative disorder associated with familial frontotemporal

dementia (FTD) and/or parkinsonism [1–3]. Many mutational loci have been defined in the *MAPT* gene on chromosome 17, which encodes tau protein [3], and mutations of *MAPT* can cause brain pathology resembling frontotemporal lobar degeneration with tau inclusions (FTLD-tau)



**Fig. 1.** Horizontal T1-weighted brain MR images at age 53 (A-D) and age 58 (*E*-H), 6 and 11 years after onset, respectively. The images show progression of the marked atrophy of the bilateral frontal lobes (A, B, E, F), and the temporal lobes including hippocampus (C, D, G, H). The involvement of white matter is mild and brainstem regions, such as the pons and midbrain, are relatively preserved.

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**Fig. 2.** Neuropathological, biochemical, and genetic findings. A: Severe macroscopic atrophy of the left frontal lobe and temporal pole. B: Klüver-Barrera staining of the temporal lobe shows severe atrophy in the lower parts (T3-4). C: Hematoxylin-eosin staining of the inferior temporal cortex shows marked neuronal loss and gliosis. The inserted image shows increased astrocytes. D–F: Gallyas-Braak staining shows diffuse dystrophic neurites and coiled bodies in amygdala (D) and hippocampal CA1 (E with a magnified image) and threads and coiled bodies in the white matter adjacent to the middle temporal cortex (F). G–M: Neuronal and glial inclusions were positive for phosphorylated tau. G: PS422 staining of the middle temporal cortex showing a pretangle and diffuse dot-like dystrophic neurites. H: PHF1 staining shows some pretangles and diffuse dystrophic neurites in the dentate gyrus. I: A tuft-shaped astrocyte in the inferior temporal cortex detected by AT8 antibody. *J*, K: Globular inclusions in the proximal processes of astrocyte in the inferior temporal cortex detected by PS422 (*J*) and AT180 (K) antibodies. L, M: Tau deposits were detected by anti-4R (L) but not by RD3 (M) antibody in the white matter of the middle temporal lobe. N: T46 antibody immunoblot of the brain extract showing a strong 37 kDa band similar to CBD. A PSP brain sample was also analyzed for comparison. O: Immunoelectron microscopy of the brain extract shows ribbon-like twisted tau filaments labelled with AT8 antibody (indicated with arrowheads in the enlarged image). P: Sanger sequence showing the *MAPT* IVS 10 + 14C > T mutation. Scale bar in each image represents: 100  $\mu$ m (C, E), 50  $\mu$ m (D, H, L, M), 20  $\mu$ m (insets of C and E, F, G, I–K), or 100 nm (O).

[4]. However, there are few reports on the neuropathology of some mutations, and the pathogenesis has not been fully elucidated. Here, we describe the pathological and biochemical features of an FTDP-17 case harboring the very rare *MAPT* intervening sequence (IVS) 10 + 14C > T mutation.

# 1. Case presentation

A Japanese male in his late 40s, with no obvious neuropsychiatric family history, presented stereotyped behavior, disinhibition, and loss of sympathy, which gradually became more apparent. Hyperorality, dietary changes, repetitive speech, hoarding, and apathy followed within a few years. Donepezil was not effective, and at age 51, he was admitted to our hospital. Neurological examination and blood test findings were normal. The Cognistat test showed normal orientation but impaired memory, language, and reasoning function, and the Frontal Assessment Battery (FAB) score was 14/18, indicating slight frontal lobe dysfunction. Brain MR imaging revealed marked atrophy of the bilateral hippocampus, amygdala, and frontal and anterior temporal lobes (Fig. 1). The patient was diagnosed as probable behavioral variant FTD (bvFTD) [5]. The behavioral symptoms improved slightly on treatment with carbamazepine and valproic acid, but the cognitive dysfunction progressed. The patient's spontaneous activity gradually decreased, followed by muscle wasting, although other motor symptoms including parkinsonism were not apparent in the advanced stages. The patient eventually became bedridden and died of aspiration pneumonia at age 58.

Postmortem examination showed macroscopically severe brain atrophy in the bilateral cerebral frontotemporal lobes (Fig. 2A). The temporal lobe showed severer atrophy in the lower parts (T3-4) than in the upper parts (T1-2) (Fig. 2B), and atrophy of the caudate nuclei, amygdala, and hippocampus was also apparent. The brainstem was preserved, and the substantia nigra and locus coeruleus showed mildly reduced pigmentation. Histologically, neuronal loss and gliosis were the most pronounced in the middle and inferior temporal cortices (Fig. 2C), entorhinal cortex, hippocampus, and striatum. In the locus coeruleus and substantia nigra, some free melanin was found, but neuronal loss was not apparent. Degeneration of precentral gyrus and pyramidal tract was not evident. Gallyas-Braak (GB) staining revealed diffuse neuronal and glial staining in the cerebral neocortex, limbic regions, subcortical nuclei, and white matter (Fig. 2D-F). Immunostaining revealed phosphorylated 4-repeat tau inclusions (Fig. 2G-M), as summarized in Table 1. The major neuronal tau inclusions were GB-positive dots and thread-like dystrophic neurites. Pretangles were also seen but were not very frequent (Fig. 2E, G, H). Glial deposits were mostly GB-positive oligodendroglial inclusions and coiled bodies (Fig. 2F). Astrocytic inclusions, observed as tuft-shaped (Fig. 2I) or globular inclusions (Fig. 2J, K), were considerably fewer and were GB-negative. The accumulation of tau deposits was less in the brainstem and cerebellum. The pathological hallmarks of CBD, such as prominent ballooned neurons and astrocytic plaques, were absent. Argyrophilic grains were moderately seen in amygdala and related limbic cortexes. Anti-phosphorylated synuclein immunostaining detected only a few Lewy neurites in the hippocampus and temporal cortex, and methenamine-silver staining and anti-

#### Table 1

Neurodegeneration and tau-positive deposits in each brain region are classified as severe (++), moderate (+), mild (+/-), or not apparent (-).

	NEURONAL loss/ gliosis	Neuronal tau inclusions	Glial tau inclusions
Cortical areas			
Frontal	+	+	+
Motor	+	+/-	+
Parietal	+	+/-	+
Temporal	++	++	++
Entorhinal	++	++	++
Subiculum	+	++	++
Hippocampal CA	++	++	++
Cingulate	+	+	+
Occipital	+/-	NA	NA
Subcortical areas			
White matter	++	_	++
Nucleus	++	NA	NA
Accumbens			
Amygdala	+	+	+
Striatum	++	++	++
Globus pallidus	+	++	++
Thalamus	-	+	+
Subthalamic	++	++	+
nucleus			
Midbrain			
Red nucleus	_	+	+
Substantia nigra	+/-	+	+
Davas			
POIDS			
Locus ceruieus	+	+	+
Ponume nuclei	+/-	+	+
Medulla oblongata			
Hypoglossal	-	-	-
nucleus			
Inferior olivary	+/-	-	-
nucleus			
Cerebellum			
Purkinje cell	+	+	-
Granule cell	-	+	-
Dentate nucleus	+	+	+

phosphorylated TDP-43 immunostaining were consistently negative. Immunoblotting of the sarkosyl-insoluble fraction from the temporal lobes revealed a 4-repeat tau banding pattern with predominant 37 kDa fragments (Fig. 2N). Electron micrographs of sarkosyl-insoluble fraction from the temporal lobes with AT8 antibody labeling revealed ribbon-like filamentous structures with a regular twist at approximately every 200 nm (Fig. 2O). The DNA sequence analysis revealed the presence of *MAPT* IVS 10 + 14C > T heterozygous mutation in the frontal cortex (Fig. 2P).

# 2. Discussion

FTDP-17 with *MAPT* mutation is extremely rare. There have been only two reports of neuropathologically examined IVS 10 + 14C > T mutation, with seven autopsied patients in total (Table 2), and detailed immunohistochemical and biochemical analyses were performed only in one case [1,6].

Cases with this mutation clinically presented both psychiatric and neurological symptoms in various proportions. Our case showed moderate involvement of the prefrontal and anterior cingulate cortices, which may account for the behavioral symptoms [7]. Atrophy of the medial prefrontal cortex along the dilated longitudinal fissure resembled that in a previously reported bvFTD case harboring *MAPT* IVS 10 + 3 mutation [3]. Our case also showed severe pathology in the limbic regions, which, in general, are associated with behavioral responses. A recent study indicated that FTDP-17 may exhibit greater volume loss of the amygdala than the other FTLD subtypes [8]. Limbic-prefrontal dysfunction might have accounted for our case's behavioral symptoms. We found that neurological symptoms, such as parkinsonism, were not

#### Table 2

Summary of cases of FTDP-17 with *MAPT* IVS 10 + 14C > T mutation.

	Lynch T (1994) [1]	Omoto M (2012) [6]	Present case
Age of death	58 on average	54	58
Gender	Males and females	Female	Male
Number of	6	1	1
autopsied cases			
Family	13 members in an	2 Japanese siblings	NA
linkage	Irish-American	0	
0	family		
Age at onset	45 on average	44	Late 40s
Disease	13 years on	10 years	11 years
duration	average		
Psychiatric	Changes in	Apathy followed by	Changes in
symptoms	personality,	memory loss and	personality,
	amnesia, poor	disorientation	disinhibition,
	construction with		apathy, dietary
	preservation of		changes, and
	orientation,		stereotypy followed
	speech, and		by memory loss and
	calculation		disorientation
Neurological	Rigidity,	Mask-like face,	Dysphagia
findings	bradykinesia,	hypophonic voice,	
	postural	bradykinesia,right-	
	formulation	bunoreative doop	
	lasculation	tendon reflex	
Clinical	disinhibition-	FTDP-17	behavioral variant
diagnosis	dementia-	1101 17	FTD
ulugiloolo	parkinsonism-		112
	amyotrophy		
	complex		
Major affected	Frontal and	Globus pallidus,	Middle and inferior
brain	temporal cortex,	cerebellum,	temporal cortex,
regions	amygdalae, and	subthalamic	entorhinal cortex,
	substantia nigra	nucleus, substantia	hippocampus, and
		nigra, and locus	striatum.
		coeruleus	
Tau isoform	NA	4-repeat	4-repeat
Biochemical	NA	PSP pattern	CBD pattern
analysis		(predominant 33	(predominant 3/
Tou flow out	NTA	KDa Dands)	KDa Dands)
subtype	INA	INA	filament
Mutational	MAPT IVS $10 \pm 14C$	<ul> <li>т</li> </ul>	mailCit
site	140 IV 7 I40	/ 1	

prominent and the substantia nigra was relatively preserved in comparison with previous cases [6]. Such variability in pathology might account for the heterogeneity of clinical phenotypes.

All reported IVS10 mutations [2,3,9] were associated with 4-repeat tauopathies such as corticobasal degeneration (CBD), progressive supranuclear palsy (PSP), and recently proposed globular glial tauopathy (GGT) [10]. In the present case, the biochemical findings correspond well with the CBD pattern, but the pathology was atypical for CBD. Immunohistochemistry showed common CBD pathological features, such as abundant pretangles, dystrophic neurites, and glial inclusions, but the pathological hallmarks of CBD such as prominent ballooned neurons and astrocytic plaques were absent, and the main pathology was focused on the cerebral cortex and hippocampus, with only mild neurodegeneration in the substantia nigra. Moreover, a small number of pathological structures exhibited PSP-like features. These results collectively suggest a heterogeneous and multiple tau pathology, but not typical CBD or PSP, and the IVS mutation might be related to this. In addition, we identified electron-microscopic tau filaments presumably caused by the IVS 10 + 14 mutation. The appearance of those ribbon-like filamentous structures was similar to that seen in adjacent IVS mutations [2]. Similarities of filamentous tau structure due to different IVS mutations might cause specific clinical and pathological phenotypes [3]. Further evidence is needed to clarify the geneticpathological-clinical correlations in detail.

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# Ethics approval and consent

The patient's next to kin gave written consent for autopsy and postmortem analysis for research purposes. This study was approved by the ethics committee in the Tokyo Metropolitan Institute of Medical Science and was performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments.

#### Declaration of interest

None.

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#### References

- T. Lynch, M. Sano, K.S. Marder, K.L. Bell, N.L. Foster, et al., Clinical characteristics of a family with chromosome 17-linked disinhibition-dementia-parkinsonismamyotrophy complex, Neurology 44 (10) (1994 Oct) 1878–1884.
- [2] M. Goedert, M.G. Spillantini, Tau mutations in frontotemporal dementia FTDP-17 and their relevance for Alzheimer's disease, Biochim. Biophys. Acta 1502 (1) (2000 Jul) 110–121.
- [3] B. Ghetti, A.L. Oblak, B.F. Boeve, K.A. Johnson, B.C. Dickerson, et al., Invited review: Frontotemporal dementia caused by microtubule-associated protein tau gene (MAPT) mutations: a chameleon for neuropathology and neuroimaging, Neuropathol. Appl. Neurobiol. 41 (1) (2015 Feb) 24–46.
- [4] S.L. Forrest, J.J. Kril, C.H. Stevens, J.B. Kwok, M. Hallupp, et al., Retiring the term FTDP-17 as MAPT mutations are genetic forms of sporadic frontotemporal tauopathies, Brain 141 (2) (2018 Feb) 521–534.

- [5] K. Rascovsky, J.R. Hodges, D. Knopman, M.F. Mendez, J.H. Kramer, et al., Sensitivity of revised diagnostic criteria for the behavioural variant of frontotemporal dementia, Brain 134 (2011) 2456–2477.
- [6] M. Omoto, S. Suzuki, T. Ikeuchi, T. Ishihara, T. Kobayashi, et al., Autosomal dominant tauopathy with parkinsonism and central hypoventilation, Neurology 78 (10) (2012 Mar) 762–764.
- [7] R.B. Firat, Opening the "Black Box": functions of the frontal lobes and their implications for sociology, Front. Sociol. 4 (3) (2019 Feb).
- [8] M. Bocchetta, J.E. Iglesias, D.M. Cash, J.D. Warren, J.D. Rohrer, Amygdala subnuclei are differentially affected in the different genetic and pathological forms of frontotemporal dementia, Alzheimers Dement. 11 (2019 Dec) 136–141.
- [9] T. Umeda, T. Yamashita, T. Kimura, K. Ohnishi, H. Takuma, et al., Neurodegenerative disorder FTDP-17-related tau intron 10 +16C → T mutation increases tau exon 10 splicing and causes tauopathy in transgenic mice, Am. J. Pathol. 183 (1) (2013 Jul) 211–225.
- [10] Z. Ahmed, E.H. Bigio, H. Budka, D.W. Dickson, I. Ferrer, et al., Globular glial tauopathies (GGT): consensus recommendations, Acta Neuropathol. 126 (4) (2013 Oct) 537–544.

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