

Human BK Polyomavirus Plasmid pBKV (34-2) (Dunlop) Contains Mutations Not Found in the Originally Published Sequences

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The plasmid pBKV (34-2) (ATCC 45025) contains the entire BK polyomavirus Dunlop genome. Sequencing revealed 12 point mutations compared to the GenBank sequence, but only 4 point mutations compared to the published sequence. The origin of these differences is unknown, but may impact virological as well as diagnostic research and development.

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The human BK polyomavirus (BKPyV) was first isolated in 1970 from the urine of a kidney transplant patient with ureteric stenosis shedding decoy cells (1). The isolate was passaged in cell culture and named after the senior author Susan Gardner and henceforth called the Gardner strain (2). In 1979, the whole-genome sequence of a derivative strain called Dunlop was reported (3). The circular double-stranded genome of 5,153 bp is divided into an early viral gene region (EVGR) encoding the regulatory proteins large, small, and truncated tumor antigen (LTag, sTag, and truncTag); a late viral gene region (LVGR) encoding the agnoprotein and the structural proteins VP1, VP2, and VP3; and a noncoding control region (NCCR) harboring the origin of replication and promoter-enhancer elements (4). The NCCR of BKPyV commonly found in urine is of a linear archetype architecture arbitrarily termed O₁₄₂-P₆₈-Q₃₉-R₆₃-S₆₃, where the subscript indicates the respective length in base pairs. Rearrangements of these blocks frequently arise in cell culture (5–7) and in patients (8, 9), giving rise to new viral strain variants. In contrast, the EVGR and the LVGR are rarely affected. Although the origin of Dunlop is not clear, the NCCR is identical to the one of the Gardner strain, except for one additional 44-bp deletion removing the Q-block (3, 10). The Dunlop genome is available as a plasmid, pBKV (34-2) (ATCC 45025), which is frequently used for research and diagnostic purposes (10–22). ATCC refers to the GenBank number J02038.1, which has been replaced by the number V01108 referring to the original publication (3). Since the backbone of pBKV (34-2) is the low-copy vector pBR322, a high-copy plasmid was generated by cloning of the BKPyV genome into a pGEM vector (22). Dye-terminator cycle sequencing of the plasmid using 20 primers in the BKPyV genome and M13R and T7 primers in the vector (23) revealed 12 point mutations compared to the GenBank sequence, causing 6 amino acid (aa) substitutions affecting the VP2 gene (aa 100 Lys to Arg and aa 103 Asp to Ser), the VP1 gene (aa 158 Glu to Asp, aa 171 Ser to Thr, and aa 219 Ala to Thr) and the LTag gene (aa 260 Ser to Asn). However, compared to the

originally published sequence (3), only 4 point mutations were found, resulting in 2 amino acid substitutions affecting the VP1 gene (aa 219 Ala to Thr) and the LTag gene (aa 260 Ser to Asn).

Sequencing of the original plasmid from ATCC gave identical results. The indicated changes in pBKV (34-2) may have accumulated over time, or alternatively, sequencing errors were already made during the original work. Since many laboratories use pBKV (34-2) for research and development, including as a positive control for real-time quantitative PCR (22), and since the GenBank sequence is frequently used to design primers and probes (22), the mutations detected may have consequences. The effect of these rather conservative amino acid substitutions on viral protein function is not known.

Nucleotide sequence accession number. The complete genome sequence of pBKV (34-2) is deposited in the GenBank under the accession no. [KP412983](https://www.ncbi.nlm.nih.gov/nuccore/KP412983).

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REFERENCES

- Gardner SD, Field AM, Coleman DV, Hulme B. 1971. New human papovavirus (B.K.) isolated from urine after renal transplantation. *Lancet* i:1253–1257. [http://dx.doi.org/10.1016/S0140-6736\(71\)91776-4](http://dx.doi.org/10.1016/S0140-6736(71)91776-4).
- Knowles WA. 2006. Discovery and epidemiology of the human polyomaviruses BK virus (BKV) and JC virus (JCV). *Adv Exp Med Biol* 577:19–45. http://dx.doi.org/10.1007/0-387-32957-9_2.
- Seif I, Khoury G, Dhar R. 1979. The genome of human papovavirus BKV. *Cell* 18:963–977. [http://dx.doi.org/10.1016/0092-8674\(79\)90209-5](http://dx.doi.org/10.1016/0092-8674(79)90209-5).
- Rinaldo CH, Tylden GD, Sharma BN. 2013. The human polyomavirus BK (BKPyV): virological background and clinical implications. *APMIS* 121:728–745. <http://dx.doi.org/10.1111/apm.12134>.
- Hanssen Rinaldo C, Hansen H, Traavik T. 2005. Human endothelial cells allow passage of an archetypal BK virus (BKV) strain—a tool for cultivation and functional studies of natural BKV strains. *Arch Virol* 150:1449–1458. <http://dx.doi.org/10.1007/s00705-005-0511-3>.

6. Rubinstein R, Harley EH. 1989. BK virus DNA cloned directly from human urine confirms an archetypal structure for the transcriptional control region. *Virus Genes* 2:157–165. <http://dx.doi.org/10.1007/BF00315259>.
7. Rubinstein R, Schoonakker BC, Harley EH. 1991. Recurring theme of changes in the transcriptional control region of BK virus during adaptation to cell culture. *J Virol* 65:1600–1604.
8. Olsen GH, Andresen PA, Hilmarsen HT, Bjørang O, Scott H, Midtvedt K, Rinaldo CH. 2006. Genetic variability in BK virus regulatory regions in urine and kidney biopsies from renal-transplant patients. *J Med Virol* 78:384–393. <http://dx.doi.org/10.1002/jmv.20551>.
9. Gosert R, Rinaldo CH, Funk GA, Egli A, Ramos E, Drachenberg CB, Hirsch HH. 2008. Polyomavirus BK with rearranged noncoding control region emerge *in vivo* in renal transplant patients and increase viral replication and cytopathology. *J Exp Med* 205:841–852. <http://dx.doi.org/10.1084/jem.20072097>.
10. Markowitz RB, Dynan WS. 1988. Binding of cellular proteins to the regulatory region of BK virus DNA. *J Virol* 62:3388–3398.
11. Bernhoff E, Gutteberg TJ, Sandvik K, Hirsch HH, Rinaldo CH. 2008. Cidofovir inhibits polyomavirus BK replication in human renal tubular cells downstream of viral early gene expression. *Am J Transplant* 8:1413–1422. <http://dx.doi.org/10.1111/j.1600-6143.2008.02269.x>.
12. Bernhoff E, Tylden GD, Kjerpeseth LJ, Gutteberg TJ, Hirsch HH, Rinaldo CH. 2010. Leflunomide inhibition of BK virus replication in renal tubular epithelial cells. *J Virol* 84:2150–2156. <http://dx.doi.org/10.1128/JVI.01737-09>.
13. Li R, Sharma BN, Linder S, Gutteberg TJ, Hirsch HH, Rinaldo CH. 2013. Characteristics of polyomavirus BK (BKPv) infection in primary human urothelial cells. *Virology* 440:41–50. <http://dx.doi.org/10.1016/j.virol.2013.01.024>.
14. Olsen GH, Hirsch HH, Rinaldo CH. 2009. Functional analysis of polyomavirus BK noncoding control region quasispecies from kidney transplant recipients. *J Med Virol* 81:1959–1967. <http://dx.doi.org/10.1002/jmv.21605>.
15. Rinaldo CH, Gosert R, Bernhoff E, Finstad S, Hirsch HH. 2010. 1-O-hexadecyloxypropyl cidofovir (CMX001) effectively inhibits polyomavirus BK replication in primary human renal tubular epithelial cells. *Antimicrob Agents Chemother* 54:4714–4722. <http://dx.doi.org/10.1128/AAC.00974-10>.
16. Sharma BN, Li R, Bernhoff E, Gutteberg TJ, Rinaldo CH. 2011. Fluoroquinolones inhibit human polyomavirus BK (BKV) replication in primary human kidney cells. *Antiviral Res* 92:115–123. <http://dx.doi.org/10.1016/j.antiviral.2011.07.012>.
17. Sharma BN, Marschall M, Henriksen S, Rinaldo CH. 2014. Antiviral effects of artesunate on polyomavirus BK replication in primary human kidney cells. *Antimicrob Agents Chemother* 58:279–289. <http://dx.doi.org/10.1128/AAC.01800-13>.
18. Moens U, Van Ghelue M, Johansen B, Seternes OM. 1999. Concerted expression of BK virus large T- and small t-antigens strongly enhances oestrogen receptor-mediated transcription. *J Gen Virol* 80:585–594.
19. Abend JR, Imperiale MJ. 2008. Transforming growth factor-beta-mediated regulation of BK virus gene expression. *Virology* 378:6–12. <http://dx.doi.org/10.1016/j.virol.2008.05.009>.
20. Jordan JA, Manley K, Dugan AS, O'Hara BA, Atwood WJ. 2010. Transcriptional regulation of BK virus by nuclear factor of activated T cells. *J Virol* 84:1722–1730. <http://dx.doi.org/10.1128/JVI.01918-09>.
21. Unterstab G, Gosert R, Leuenberger D, Lorentz P, Rinaldo CH, Hirsch HH. 2010. The polyomavirus BK agnoprotein colocalizes with lipid droplets. *Virology* 399:322–331. <http://dx.doi.org/10.1016/j.virol.2010.01.011>.
22. Dumoulin A, Hirsch HH. 2011. Reevaluating and optimizing polyomavirus BK and JC real-time PCR assays to detect rare sequence polymorphisms. *J Clin Microbiol* 49:1382–1388. <http://dx.doi.org/10.1128/JCM.02008-10>.
23. Henriksen S, Tylden GD, Dumoulin A, Sharma BN, Hirsch HH, Rinaldo CH. 2014. The human fetal glial cell line SVG p12 contains infectious BK polyomavirus. *J Virol* 88:7556–7568. <http://dx.doi.org/10.1128/JVI.00696-14>.