

Effector Memory T cells Are Associated With Atherosclerosis in Humans and Animal Models

Enrico Ammirati, MD; Domenico Cianflone, MD; Viviana Vecchio, BSc; Michela Banfi, BSc; Anna C. Vermi, MD; Monica De Metrio, MD; Liliana Grigore, MD; Fabio Pellegatta, MD; Angela Pirillo, PhD; Katia Garlaschelli, BSc; Angelo A. Manfredi, MD; Alberico L. Catapano, PhD; Attilio Maseri, MD; Alessio G. Palini, MSc; Giuseppe D. Norata, PhD

Background—Adaptive T-cell response is promoted during atherogenesis and results in the differentiation of naïve CD4⁺T cells to effector and/or memory cells of specialized T-cell subsets. Aim of this work was to investigate the relationship between circulating CD4⁺T-cell subsets and atherosclerosis.

Methods and Results—We analyzed 57 subsets of circulating CD4⁺T cells by 10-parameter/8-color polychromatic flow cytometry (markers: CD3/CD4/CD45RO/CD45RA/CCR7/CCR5/CXCR3/HLA-DR) in peripheral blood from 313 subjects derived from 2 independent cohorts. In the first cohort of subjects from a free-living population ($n=183$), effector memory T cells (T_{EM} : CD3⁺CD4⁺CD45RA⁻CD45RO⁺CCR7⁻ cells) were strongly related with intima-media thickness of the common carotid artery, even after adjustment for age ($r=0.27$; $P<0.001$). Of note, a significant correlation between T_{EM} and low-density lipoproteins was observed. In the second cohort ($n=130$), T_{EM} levels were significantly increased in patients with chronic stable angina or acute myocardial infarction compared with controls. HLA-DR⁺ T_{EM} were the T_{EM} subpopulation with the strongest association with the atherosclerotic process ($r=0.37$; $P<0.01$). Finally, in animal models of atherosclerosis, T_{EM} (identified as CD4⁺CD44⁺CD62L⁻) were significantly increased in low-density lipoprotein receptor and apolipoprotein E deficient mice compared with controls and were correlated with the extent of atherosclerotic lesions in the aortic root ($r=0.56$; $P<0.01$).

Conclusions—Circulating T_{EM} cells are associated with increased atherosclerosis and coronary artery disease in humans and in animal models and could represent a key CD4⁺T-cell subset related to the atherosclerotic process. (*J Am Heart Assoc* 2012;1:27-41.)

Key Words: atherosclerosis • C-c chemokine receptor type 7 • chemokines • coronary artery disease • effector memory T cells

T cells play a key role in the immune response observed during atherogenesis.^{1,2} In the arterial wall, cholesterol

accumulation followed by vascular inflammation promotes adaptive T-cell response,³ and results in the differentiation of naïve CD4⁺T cells to effector and/or memory cells of specialized T-cell subsets in secondary lymphoid organs and in the chemokine-driven recruitment of specific T-cell and monocyte subsets into the atherosclerotic plaque.⁴⁻⁷

Most of T cells found in human atherosclerotic lesions are activated effector and/or memory CD4⁺ that predominate over CD8⁺T cells,⁸⁻¹⁰ with a specific antigenic response.^{6,11,12} The activation of inflammatory pathways in atherosclerotic

Editorial on p 3

lesions is reflected also in neutrophils, monocytes, and T lymphocytes present in peripheral blood,¹³⁻¹⁶ supporting the concept of a systemic process with local infiltration in the arterial wall. In animal models, a CCR7-dependent recirculation of T cells and monocytes between secondary lymphoid organs and inflamed tissues and egression from atherosclerotic plaques to blood was observed,¹⁷⁻¹⁹ suggesting a dynamic process that involves local factors (ie, oxidative and shear stress) and systemic factors (ie, traditional cardiovascular risk factors,

From the Clinical Cardiovascular Biology Centre (E.A., D.C., M.B.); the Flow cytometry Resource Analytical Cytology Technical Applications Laboratory (V.V., A.G.P.); the Clinical Immunology Unit (A.A.M.); San Raffaele Scientific Institute and the Università Vita-Salute San Raffaele, Milan, Italy; Heart Transplantation Division, Ospedale Niguarda Ca' Granda (E.A.); Centro Cardiologico Monzino, Istituto Di Ricovero e Cura a Carattere Scientifico (IRCCS), Milan, Italy; Department of Cardiovascular Sciences, University of Milan, Milan, Italy (M.D.M.); Center for the Study of Atherosclerosis, Italian Society for the Study of Atherosclerosis Lombardia Chapter, Bassini Hospital Cinisello Balsamo Milan, Italy (L.G., F.P., K.G., S.T., G.D.N.); Department of Pharmacological Sciences, Università degli Studi di Milano Milan, Italy (A.L.C., G.D.N.); Multimedita IRCCS, S.S. Giovanni Milan (L.G., A.L.C.); and Heart Care Foundation Florence, Italy (E.A., A.M.).

Correspondence to: Enrico Ammirati, MD, San Raffaele Scientific Institute and the Università Vita-Salute San Raffaele, via Olgettina 58, 20132 Milan, Italy. E-mail ammirati.enrico@hsr.it Or Giuseppe Danilo Norata, PhD, Department of Pharmacological Sciences, University of Milan, Via Balzaretti 9, 20133 Milan, Italy. E-mail Danilo.Norata@unimi.it

Received December 7, 2011; accepted December 21, 2011.

© 2012. The Authors. Published on behalf of the American Heart Association, Inc., by Wiley-Blackwell. This is an Open Access article under the terms of the Creative Commons Attribution Noncommercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

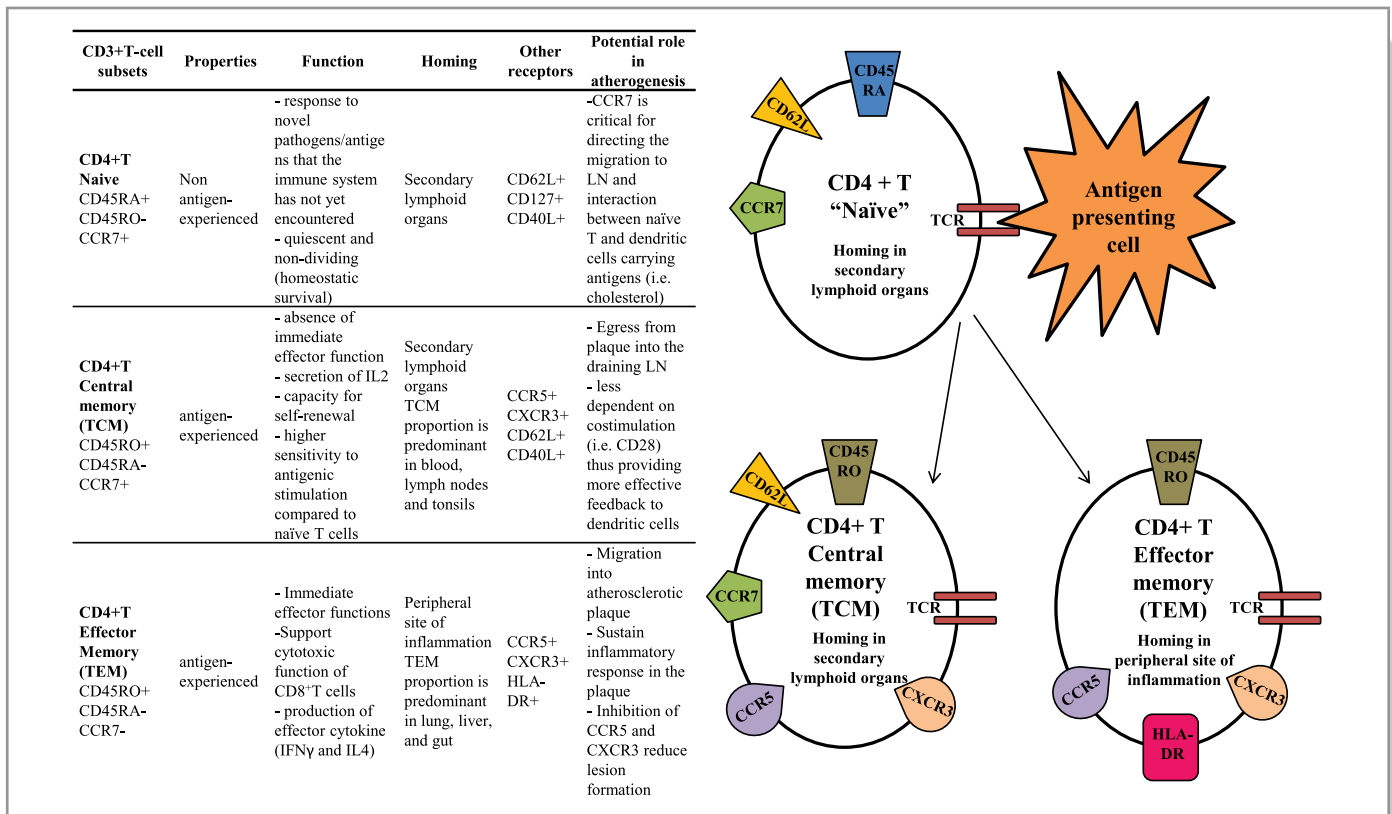


Figure 1. CD4+T cells subsets: properties and potential role in atherosclerosis.

activation of platelets, and coagulation system).^{20,21} Of note, inhibition of chemokine receptors that are involved in the recruitment of circulating T cells into the plaque such as CCR5 and CXCR3,^{22,23} attenuates atherosclerotic lesion formation in animal models.^{24–27}

In spite of the strong experimental evidence supporting a role for T cells in murine atherosclerosis, their clinical significance in humans remains limited.²⁸ Therefore, the characterization of circulating CD3⁺CD4⁺T-cell subsets in patients with different stages of atherosclerosis and with various manifestations of coronary artery disease (CAD) could contribute to clarify the role of these cells on atherosclerotic plaque formation and progression. In humans, forefront technologies such as polychromatic flow cytometry, revealed the presence of hundreds of phenotypically distinct leukocytes in the peripheral blood with specialized profiles and programmed functions that may reflect specific activation in the immune response, even in the absence of an overall alteration in total T-cell number.²⁹

Adaptive immune response during atherogenesis involves the expansion of memory T cells, antigen-experienced T cells, which (1) elicit stronger and more sustained immune response upon antigen reexposure and (2) support the cytotoxic function of CD8⁺T cells. Because antigens associated with cholesterol-rich lipoproteins promote adaptive immune response and atherosclerosis in animal models,⁵ we hypothesized that in subjects with atherosclerotic disease, alterations

in the levels of circulating T-cell subsets that occur, can be appreciated in peripheral blood.

During atherogenesis, antigens generated directly or indirectly as a consequence of hypercholesterolemia and presented by antigen-presenting cells,⁷ are likely to stimulate the development of central memory T cells (T_{CM}) and effector memory T cells (T_{EM}), from naïve (T_N) cells. T_N, T_{CM}, and T_{EM} have specific homing capacities and effector functions, and are identifiable by different expression of surface receptors (see Figure 1).^{7,30} Through their expression of CCR7 and CD62L, T_N and T_{CM} preferentially home to T-cell areas of secondary lymphoid organs and display little immediate effector functions.^{7,30} On the other hand, T_{EM} which have lost the constitutive expression of CCR7, express tissue homing receptors associated with inflammation (ie, CCR5 and CXCR3) and display more readily effector functions.^{7,30}

Aim of this study was to investigate the relationships between atherosclerosis and 7 main CD4⁺T-cell subsets with 50 derived T-cell subpopulations, resulting from the expression of specific markers (CD3/CD4/CD45RO/CD45RACCR7/CCR5/CXCR3/HLA-DR) investigated by 10-parameter/8-color polychromatic flow cytometry. We first investigated the association between levels of specific T-cell subsets with a surrogate marker of atherosclerosis, that is, the intima-media thickness (IMT) of the common carotid artery in a free-living population. Next, we examined the levels of specific T-cell

subsets in patients with stable stenotic coronary atherosclerosis (ie, chronic stable angina [CSA]) and acute unstable manifestations (ie, acute myocardial infarction [AMI]). These analyses identified circulating T_{EM} as the best T-cell subset associated with atherosclerotic disease characterized by a significant correlation with lipid profile. We sought to confirm these findings in animal models by evaluating the correlation of T_{EM} levels with atherosclerosis extent in the aortic root of apolipoprotein E and low-density lipoprotein (LDL) receptor deficient animals.

Methods

Study Populations

Institutional Ethics Committees approved the study and informed written consent was obtained from all participating subjects. The study was conducted according to the standards of the Declaration of Helsinki and Good Clinical Practice. Blood samples from 313 subjects from two independent cohort were analyzed.

Carotid study: This cohort was composed by 183 consecutively enrolled subjects from a free-living population during 3 months (September to November 2008) in the context of a larger observational study named PLIC (Progressione Lesioni Intimali Carotidae), previously described.³¹ The clinical and anthropometrical characteristics of this cohort are presented in Table 1.

For the study in patients with CAD blood samples were obtained from patients on admission to San Raffaele Scientific Institute and Centro Cardiologico Monzino, both in Milan from December 2007 to June 2009. The control group was made of 40 control subjects age and sex matched without clinical and electrocardiographic signs of CAD. Ninety patients with CAD were divided in a CSA group including 30 patients with effort angina (lasting more than 3 months and without previous history of unstable angina or AMI) and angiographic evidence of coronary artery stenosis (>50%) and an AMI group including 60 patients. AMI samples were obtained before coronary angiography and early after the onset of symptoms when the elevation of troponin I were still minimal (median: 0.4 ng/mL), thus minimizing the possible confounding effect of myocardial necrosis.³² A second sample was also obtained from 10 AMI patients after 24 hours from the acute event to check for variation of T-cell subsets during the acute phase. The analysis showed similar levels of these cell subsets (data not shown) suggesting that, at least in the early hours from the onset of AMI, perturbations attributable at myocardial necrosis were limited in the T-cell subsets under investigation. Clinical characteristics and biological parameters of patients with CAD and control subjects are reported in Table 2.

Biochemical Parameters and IMT Measurement

Measurement of biochemical parameters and clinical outcome in the patients of the PLIC study has been described

Table 1. Clinical Characteristics and Biological Parameters of Subjects From the General Population (PLIC Study)

	Carotid Study (all subjects)
No.	183
Age (y)	54±14
Male Sex—no. (%)	93 (50.5)
Current smoker no. (%)	43 (23.4)
Hypertension no. (%)	45 (24.8)
Diabetes mellitus no. (%)	14 (7.6)
Hypercholesterolemia no. (%)	31 (16.8)
Systolic blood pressure (mm Hg)	124±18
Diastolic blood pressure (mm Hg)	76±9
BMI (kg/m ²)	25.9±3.7
Glicemia (mg/dL)	100±22
Total cholesterol (mg/dL)	217±42
LDL-cholesterol (mg/dL)	142±38
HDL-cholesterol (mg/dL)	56±13
Tryglicerides (mg/dL)	100±55
Creatinine (mg/dL)	0.88±0.17
IMT (mm)	0.67±0.14
WBC (10 ⁹ /L)	6.6±1.2
Lymphocytes (10 ⁹ /L)	2.3±0.4

Data are presented as mean±standard deviation. BMI indicates body mass index; HDL, high-density lipoprotein; IMT, intima-medial thickness; LDL, low-density lipoprotein; WBC, whole blood count.

elsewhere.^{31,33} For IMT, briefly, ultrasound scanning and reading of carotid arteries were performed by a single expert sonographer, using an 8-MHz transducer (Biosound 2000 II sa, Indianapolis, IN) with an axial and lateral resolution of 0.385 and 0.500 mm, respectively. The sonographer was blinded to the subject's identity. B-mode evaluations are obtained from captures of the far wall in the first centimeter of common carotid arteries, proximal to the bulb dilation, in lateral projection. Five standardized points 5, 10, 20, 25, and 30 mm from bulb were measured in both arteries and averaged to calculate the mean IMT (IMT_m) for each subject. In two scans performed on 75 subjects by the same operator, the mean difference in IMT_m was 0.005±0.002 mm and the variation coefficient equal to 1.93%. The correlation between two scans was significant with $r=0.96$ ($P<0.0001$).

Polychromatic Flow Cytometry

Whole blood from each subject was collected in EDTA anticoagulated vacutainer tube. Samples were stained and fixed within the day of collection. We verified that there were no significant differences in the investigated marker levels in samples stained immediately after collection in comparison to samples

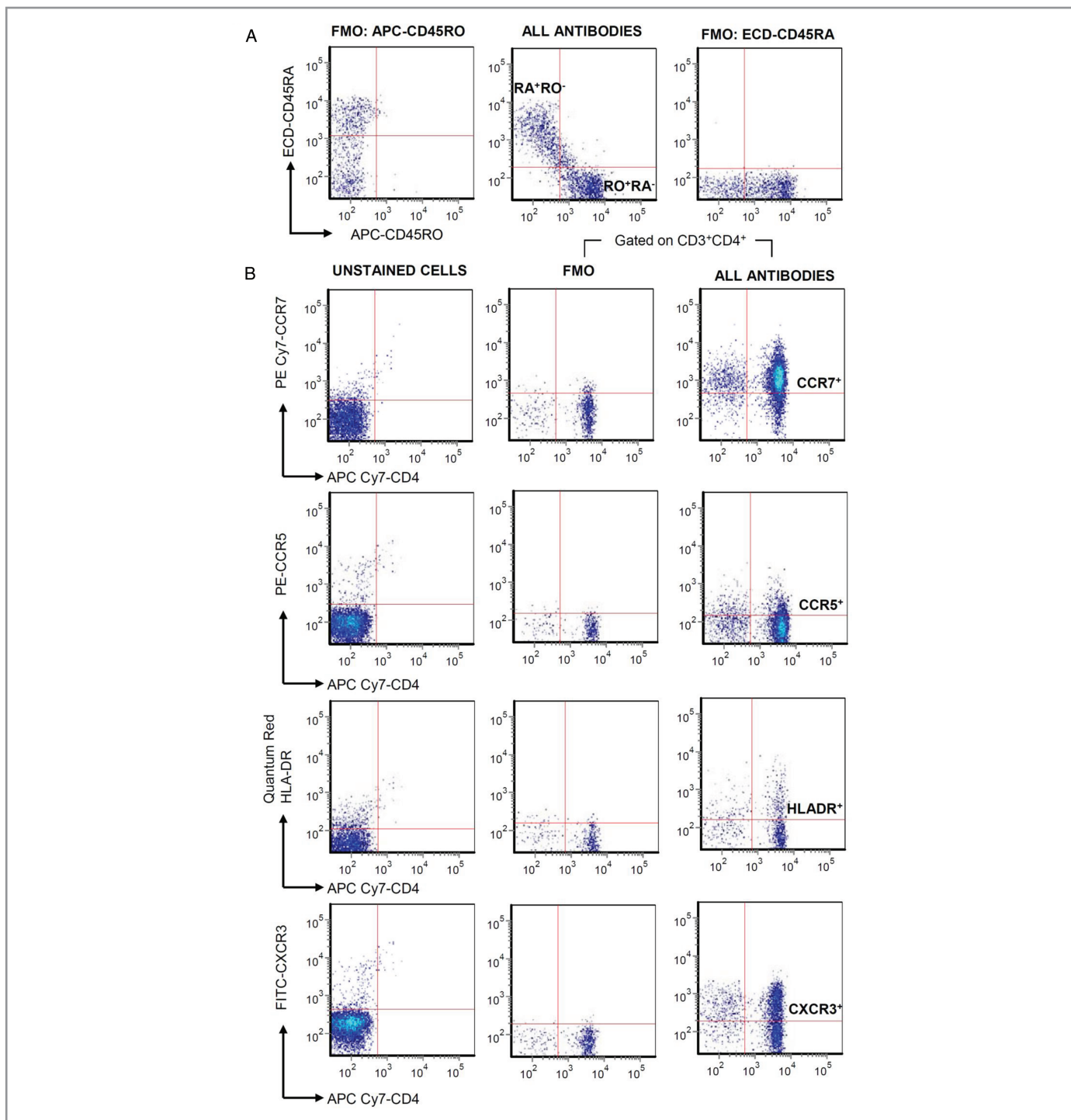


Figure 2. Identification of positive stained cells using the Fluorescence Minus One (FMO) strategy. To appropriately identify positive stained cells and differentiate them from background autofluorescence for gate inclusion, we used the FMO strategy. The background nonspecific fluorescence is collected in the detector assigned to the missing antibody in FMO control. FMO controls are samples that include all the conjugated antibodies but one. The channel for the missing conjugated antibody is that of the FMO gating control. FMO controls are important for setting thresholds in cell populations that express a continuous spectrum of numbers of receptors (such as CD45RO, CD45RA, CCR7, CCR5, HLA-DR, and CXCR3) and allows to better define the positive cell population in comparison with unstained cells. In fact, although unstained cell populations are centered close to zero, their width can vary depending on autofluorescence and spillover corrections in different channels. In (A), the plot on the left the antibody CD45RO-APC conjugated is missing and in the plot on the right the antibody CD45RA-ECD conjugated is missing. In this way, it is possible to correctly define the thresholds of CD45RO positive and CD45RA positive cells in the plot in the center. In (B), the same procedure for the other antibodies used in our experiments. The unstained populations are presented to assess the difference with the positive threshold identified with FMO.

Table 2. Clinical Characteristics and Biological Parameters of Patients with coronary artery disease (CAD) and Matched Controls

	Controls	CSA	AMI	P-Value
No.	40	30	60	
Age(y)	61 (±9)	64 (±11)	64 (±11)	0.37
Male Sex—no. (%)	32 (80)	22 (73)	46 (77)	0.81
Family history of CAD	17 (43)	12 (40)	20 (33)	0.95
Current smoker	14 (35)	7 (23)	25 (42)	0.23
Hypertension	20 (50)	22 (73)	36 (60)	0.14
Diabetes mellitus	0 (0)	3 (10)	5 (8)	0.14
Hypercholesterolemia	22 (55)	19 (63)	29 (48)	0.40
Troponin I (ng/mL)	—	—	0.4 (0.1–1.1)	—
WBC (10 ⁹ /L)	6.5±1.4	7.9±1.4	11.1±4.0*	<0.0001
Lymphocytes (10 ⁹ /L)	2.2±0.5	2.2±0.4	2.4±0.6	0.11
Monovessel disease	—	11 (37)	22 (37)	0.42
Bivessel disease	—	7 (23)	21 (35)	
Trivessel disease	—	12 (40)	17 (28)	
Aspirin	2 (5)	22 (73)*	36 (60)*	<0.0001
Thienopyridine	0 (0)	8 (27) [†]	14 (23) [†]	0.003
β-blockers	1 (3)	14 (47)*	16 (40)*	<0.0001
Statins	1 (3)	14 (47)*	10 (17) [†]	<0.0001
Ca-blockers	4 (10)	8 (27)	5 (8)	0.04
ACE inhibitors/ARBs	9 (23)	12 (40)	23 (38)	0.19
Nitrates	0 (0)	11 (37)*	24 (40)*	<0.0001
Diuretics	5 (13)	4 (13)	6 (10)	0.87

* <0.001 vs controls after Bonferroni post hoc test.

† <0.01 vs controls after Bonferroni post hoc test.

ACE indicates angiotensin converting enzyme; ARBs, angiotensin receptor blockers; CAD, coronary artery disease; WBC, whole blood count.

stained up to a maximum 24-hour time interval postsampling and stored at room temperature. In order to reduce cellular loss and analysis sampling bias in the specimen, the no wash, whole blood lyses technique was used. For each specimen, 50 μ L of a mixture of 8 antibodies was added to 100 μ L of whole blood followed by a 20-minute incubation in the dark at room temperature. After staining, the red blood cells were lysed and fixed with the Immune-prep System (Beckman Coulter). White cells, diluted in 1 mL total volume were analyzed on a LSR II Flow Cytometer (BD Biosciences) equipped with four lasers and standard optics. The antibodies were selected to minimize spectral overlap. Furthermore to reduce the nonspecific fluorescence background and optimize the fluorescence signal, we used appropriately titred directly conjugated monoclonal antibodies.³⁴ The panel used, consisted of the subsequent cellular surface markers: CD3 (Pacific Blue-labeled, clone HIT3a, BD Pharmingen), CD4 (APC-Cy7, SK3, BD Biosciences), CCR5/CD195 (PE, 2D7, BD Pharmingen), CXCR3/CD183 (FITC, 49801, R&D Systems Inc), CCR7/CD197 (PE-Cy7, 3D12, BD Pharmingen),

HLA-DR (Quantum Red, HK14, Sigma-Aldrich), CD45RO (APC, UCHL1, Caltag), and CD45RA (ECD, 2H4, IOTest, Beckman Coulter). To appropriately identify positive stained cells and differentiate them from background autofluorescence for gate inclusion, we used the Fluorescence Minus One strategy. The channel for the missing conjugated antibody is that of the Fluorescence Minus One gating control (Figure 2). Fluorescence intensity for each signal measured was standardized using multiple peak Rainbow calibration particles (Code RCP-30-5A, Spherotech) to allow reproducible and comparable median fluorescence intensity throughout the study period, as previously described.³³ We identified 7 principal T-cell subsets and 50 T-cell subpopulations derived by principal subsets by means of the combination of the surface markers. All data were acquired in FCS format using FACSDiva Software 5.0 (BD Biosciences). Lymphocytes were identified and electronically gated on forward and orthogonal light scatter signals. The fluorescent signals for phenotype analyses were accumulated for the gated lymphocytes. The instrument raw

data were stored electronically to a server for archiving and data processing. Data were processed and analyzed using FCS Express V3 Research edition (De Novo Software, Inc; <http://www.denovosoftware.com>). Cell viability was >99%, assessed using the Molecular Probes Patented LIVE/DEAD Viability (Invitrogen) according to the manufacturer instructions.

Identification of T-Cell Subsets in Animal Models of Atherosclerosis by Flow Cytometry

T cell subsets were investigated in peripheral blood collected from male mice with apolipoprotein E or LDL receptor deficiency (two animal models of atherosclerosis) and control animals with the same C57Bl6/J background.³⁵ The investigation conformed to the European Commission Directive 86/609/EEC and was approved by the local committee (Progetto di Ricerca 2009/3). Briefly, at 8 weeks of age, animals were fed ad libitum with western-type diet (21% fat, 0.15% cholesterol, and 19.5% casein, Harlan, Bresso, Italy) for 16 weeks except for the group 1 (C57Bl6/J) animals that continued at chow diet). Mice were euthanized with an overdose of Avertin 2.5% (Aldrich Chemical Co), followed by cervical dislocation. Blood samples were collected in EDTA tubes immediately before death and plasma was separated by low-speed centrifugation at 4°C. The measurement of plasma lipids was performed by enzymatic techniques (ABX for Cobas Mira Plus, Montpellier, France)³⁵ and atherosclerosis was quantified at the aortic sinus as described.³⁶ For flow-cytometry analysis, the following panel of cellular surface markers was used: CD4 (Pacific Blue-labeled, RPA-T4, BD Pharmingen), CD44 (FITC, IM7, BD Pharmigen), and CD62L (APC, mMEL-14, BD Pharmigen).

Monitoring T-Cell Subsets in Subjects Who Underwent Tetanus Vaccination

For this study, blood samples from three control subjects that performed booster shots of tetanus vaccine for personal reasons (travel to a zone at increased risk for tetanus) were taken. The subjects signed an informed consent. Subjects were sampled (4 mL of whole blood in EDTA each time) the day of the vaccination (before exposure), and 2, 7, 14, and 21 days after exposure. All three subjects were previously vaccinated. They received a single dose of 40 International Units of tetanus toxoid (ANATETALL, Novartis Vaccines & Diag) by intramuscular injection.

Statistical Analysis

Results are reported as mean±standard deviation (SD) or median, first to third interquartile on the basis of normal or nonnor-

mal distribution respectively (Shapiro-Wilk normality test was performed). Group differences in continuous variables were determined by using Student's *t* test or one-way analysis of variance with Bonferroni's multiple comparison test for normally distributed values and Mann-Whitney U test or Kruskal-Wallis test with Dunn's multiple comparison test as appropriate. Analysis of variance univariate analysis was performed between all variables and IMT. Multiple stepwise regression analysis was performed with IMT as the dependent variable, and by entering the independent variable with the highest partial correlation coefficient at each step. Associations are summarized using correlation coefficients, Pearson correlation coefficient is shown for variables normally distributed while Spearman rank correlation coefficient is shown for variables that were not normally distributed. Group differences or correlations with $P<0.05$ were deemed as statistically significant. GraphPad Prism 4 and SPSS Statistics 17.0 softwares were used for analysis.

Results

Identification of Seven Main CD3⁺CD4⁺T-Cell Subsets and 50 Derived T-Cell Subpopulations

We identified seven principal T-cell subsets by 10-parameter/8-color flow cytometry analysis: naïve T cells (called T_N defined as CD3⁺CD4⁺CD45RA⁺CD45RO⁻CCR7⁺), memory T cells (T_M, defined as CD3⁺CD4⁺CD45RA⁻CD45RO⁺), central memory T cells (T_{CM}, defined as CD3⁺CD4⁺CD45RA⁻CD45RO⁺CCR7⁺) and effector memory T cells (T_{EM}, defined as CD3⁺CD4⁺CD45RA⁻CD45RO⁺CCR7⁻), HLA-DR⁺T cells (CD3⁺CD4⁺HLA-DR⁺), CCR5⁺T cells (CD3⁺CD4⁺CCR5⁺), and CXCR3⁺T cells (CD3⁺CD4⁺CXCR3⁺) (Figure 3). Then, we described 50 T-cell subpopulations derived from these principal T cell subsets (see Table 3 for the list of all analyzed subpopulations). Principal T-cell subsets and subpopulations were expressed as percentage of total CD3⁺CD4⁺T cells.

Antigen Reexposure Can Modify the Composition of Memory T-Cell Compartment in Humans

As proof of principle in a limited number of subjects, we analyzed changes in the composition of memory T-cell compartment after antigen reexposure. We demonstrated that our approach was sensitive enough to clearly identify these modifications. In particular, antigen reexposure led to a temporally limited T_{EM} increase in the peripheral blood of control subjects vaccinated with tetanus toxoid (For details also concerning T_M and T_{CM} modifications see Figure 4).

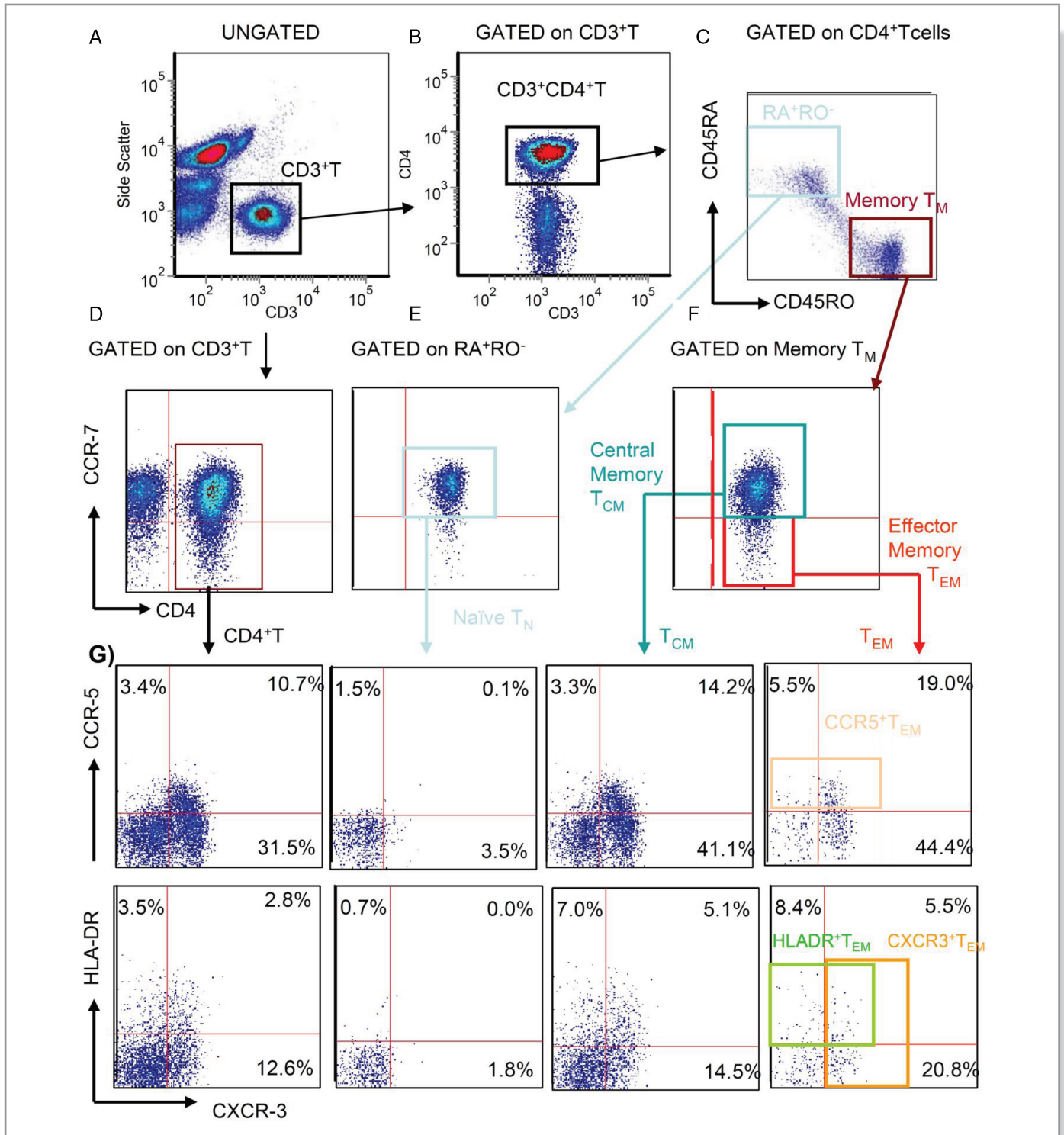


Figure 3. Gating strategy to identify the principal CD4⁺T-cell subsets, and some of the T-cell subpopulations. Color dot plot of a representative subject. Lymphocytes were identified and electronically gated on orthogonal light scatter signals and CD3 immunopositivity (approximately 30.000 events on CD3 for each sample) (A). Then CD3⁺CD4⁺T cells were identified (B). Gating on CD3⁺CD4⁺T cells, memory T cells (T_M) were identified as CD45RA negative and CD45RO positive (C). In (D), the expression of the chemokine receptor CCR-7 was shown in CD3⁺T cells. In (E), the expression of CCR7 defined the naïve T cells (T_N) as CD45RA⁺CD45RO⁻ and CCR7 positive. In (F), the expression of CCR7 is used to define central memory T cells (T_{CM}; CCR7 positive) and effector memory T cells (T_{EM}; CCR7 negative). In (G), it is shown the different expression of chemokine receptors CXCR3, CCR5, and marker of activation HLA-DR in CD3⁺CD4⁺CCR7⁺ T cells, in T_N, in T_{CM} and in T_{EM}. T_{EM} have a relative increased expression of markers of activation and chemokine receptors in comparison with other T-cell subsets. In G are shown some of the T-cell subpopulations that are considered in further analysis (such as CXCR3⁺T_{EM}, CCR5⁺T_{EM}, HLA-DR⁺T_{EM}).

Table 3. Description of All T-Cells Subpopulation Analyzed

CD3 ⁺ CD4 ⁺ T Subsets	Subpopulations	Levels (% of CD3 ⁺ CD4 ⁺)
Naïve CD45RA ⁺ RO ⁻ CCR7 ⁺		
	CXCR3 ⁺	1.1 (1.1)
	CCR5 ⁺	0.9 (0.8)
	HLA-DR ⁺	0.2 (0.2)
	CXCR3 ⁺ CCR5 ⁺	>0.1*
	CXCR3 ⁺ CCR5 ⁻	0.1 (0.1)
	CXCR3 ⁻ CCR5 ⁺	>0.1*
	CXCR3 ⁺ HLA-DR ⁺	>0.1*
	CXCR3 ⁺ HLA-DR ⁻	1.0 (1.0)
	CXCR3 ⁻ HLA-DR ⁺	>0.1*
CD45RA ⁺ RO ⁻ CCR7 ^{-†}		2.4 (1.7)
	CXCR3 ⁺	0.1 (0.1)
Central memory T CD45RA ⁻ RO ⁺ CCR7 ⁺		
	CXCR3 ⁺	20.1 (7.3)
	CCR5 ⁺	5.2 (3.6)
	HLA-DR ⁺	3.8 (1.9)
	CXCR3 ⁺ CCR5 ⁺	3.1 (2.4)
	CXCR3 ⁺ CCR5 ⁻	17.1 (6.1)
	CXCR3 ⁻ CCR5 ⁺	2.2 (1.7)
	CXCR3 ⁺ HLA-DR ⁺	1.8 (1.1)
	CXCR3 ⁺ HLA-DR ⁻	18.4 (6.9)
CXCR3 ⁻ HLA-DR ⁺	2.0 (1.2)	
Effector memory T CD45RA ⁻ RO ⁺ CCR7 ⁻		
	CXCR3 ⁺	5.2 (2.5)
	CCR5 ⁺	0.8 (0.5)
	HLA-DR ⁺	1.0 (0.7)
	CXCR3 ⁺ CCR5 ⁺	2.0 (1.9)
	CXCR3 ⁺ CCR5 ⁻	3.9 (2.1)
	CXCR3 ⁻ CCR5 ⁺	0.6 (0.5)
	CXCR3 ⁺ HLA-DR ⁺	0.4 (0.3)
	CXCR3 ⁺ HLA-DR ⁻	4.2 (2.3)
CXCR3 ⁻ HLA-DR ⁺	0.6 (0.4)	
CD45RA ⁺ RO [‡]		5.6 (3.5-8.5)
CD45RA ⁺ RO ⁺ CCR7 ⁺		5.2 (3.1-7.8)
	CXCR3 ⁺	1.1 (0.8-1.5)
	CCR5 ⁺	0.5 (0.3-0.7)
	HLA-DR ⁺	0.1 (0.1)

Continued

Table 3. Continued

CD3 ⁺ CD4 ⁺ T Subsets	Subpopulations	Levels (% of CD3 ⁺ CD4 ⁺)
	CXCR3 ⁺ CCR5 ⁺	0.4 (0.2-0.5)
	CXCR3 ⁺ CCR5 ⁻	0.7 (0.5-1.1)
	CXCR3 ⁻ CCR5 ⁺	0.1 (0.04-0.2)
	CXCR3 ⁺ HLA-DR ⁺	>0.1*
	CXCR3 ⁺ HLA-DR ⁻	1.0 (0.7-1.4)
	CXCR3 ⁻ HLA-DR ⁺	>0.1*
CD45RA ⁺ RO ⁺ CCR7 ⁻		0.4 (0.2-0.7)
	CXCR3 ⁺	0.1 (0.04-0.13)
	CCR5 ⁺	>0.1*
	HLA-DR ⁺	>0.1*
	CXCR3 ⁺ CCR5 ⁺	>0.1*
	CXCR3 ⁺ CCR5 ⁻	0.1 (0.03-0.11)
	CXCR3 ⁻ CCR5 ⁺	>0.1*
	CXCR3 ⁺ HLA-DR ⁺	>0.1*
	CXCR3 ⁺ HLA-DR ⁻	0.1 (0.04-0.11)
	CXCR3 ⁻ HLA-DR ⁺	>0.1*

Data are presented as mean±standard deviation or median (25th quartile to 75th quartile) as appropriate.

*We did not report precise values in case of small subpopulations with frequencies below 0.1% as median or mean value.

†CD45RA⁺RO⁻CCR7⁻ is a naïve subpopulation that had lost the expression of CCR7. This subpopulation is small and non-well characterized.

‡CD45RA⁺RO⁺ represents a subpopulation of transition from a naïve to memory phenotype that coexpresses both CD45RA and CD45RO.

Correlations Between Circulating T-Cell Subsets and Cardiovascular Risk Factors

First, we investigated the clinical variables that can affect the distribution of T-cell subsets under investigation. Overall, men and women had similar CD3⁺CD4⁺T cells (61.9±9.1% versus 59.5±11.5%, $P=0.13$; expressed as percentage of total CD3⁺ lymphocytes). Male subjects had lower T_N (30.0±12.1% versus 34.6±12.9%, $P=0.01$; percentage of CD3⁺CD4⁺T cells), but higher T memory cells (62.7±12.1% versus 57.8±14.4%, $P=0.01$) including higher T_{EM} (11.9±4.9% versus 10.3±3.7%, $P=0.01$) and T_{CM} (50.9±10.1% versus 47.6±12.3%, $P=0.04$) compared with female (Table 4). T_N were inversely correlated with age (Spearman $r=-0.31$, $P<0.01$), while T_M (Spearman $r=0.32$, $P<0.01$) including T_{EM} (Spearman $r=0.31$, $P<0.01$) and T_{CM} (Spearman $r=0.22$, $P<0.01$) or HLA-DR⁺T cells (Spearman $r=0.20$, $P<0.01$) were directly correlated with age.^{37,38}

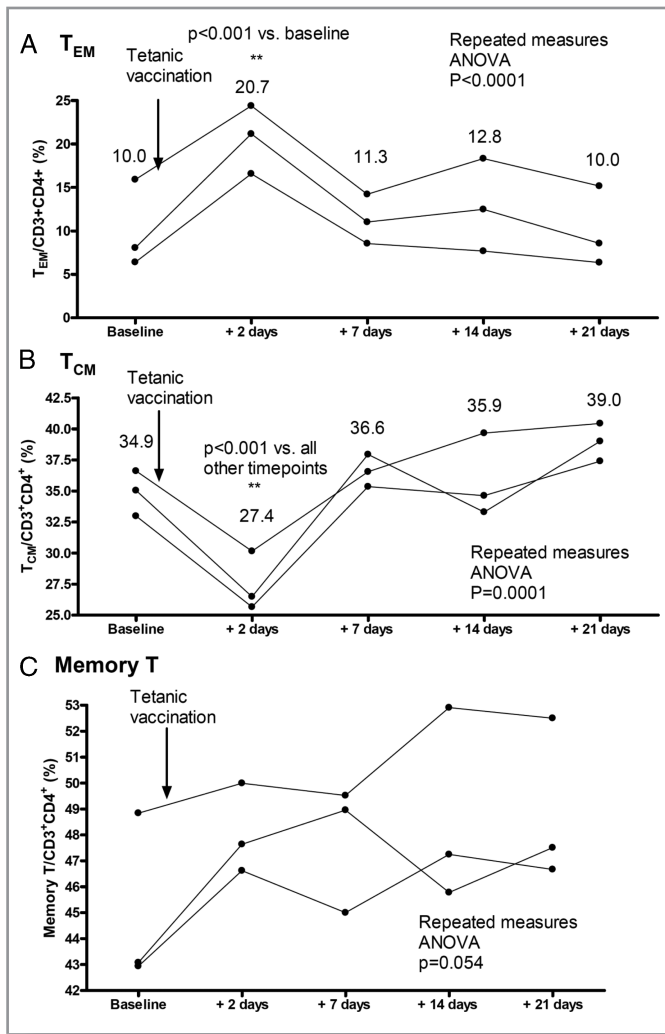


Figure 4. Effector memory T cell (T_{EM}), central memory T cell (T_{CM}), and memory T cell variations at several time points after tetanic vaccination in three healthy subjects. Dots represent subject and continuous lines show the temporal changes between the first sampling (before the tetanic vaccination) and the following ones at several time point after the antigen exposure (after 2, 7, 14, and 21 days). We observed a temporally limited increase in levels of T_{EM} and a reciprocal decrease in levels of T_{CM} at 2 days after antigen exposure with a subsequent decrease of T_{EM} and increase of T_{CM} to prevaccination levels after 1 and 3 weeks after the exposure. Means percentage are reported and referred to the total number of $CD3^+CD4^+$ T cells (A, B). A trend vs a significant increase in total memory T cells have been observed after 2 to 3 weeks from tetanic vaccination (C).

Hypercholesterolemic subjects showed increased levels of T_{EM} (mean±SD 12.9±1.0% versus 10.7±0.3%, $P=0.02$) and HLA-DR⁺ T cells (6.2±0.7% versus 4.6±0.2%, $P=0.03$). The presence of hypertension was associated with decreased levels of T_N (27.2±1.9% versus 33.6±1.4%, $P<0.01$) and increased levels of T_M (66.3±2.0% versus 58.7±1.2%, $P<0.01$), including T_{CM} (53.5±1.6% versus 43.2±1.4%, $P=0.01$) and T_{EM} (12.9±0.8% versus 10.6±0.4%, $P<0.01$). Smoking habits were not associated with increased TEM levels (data not shown).

Table 4. T-Cell Principal Subpopulation Distribution According to the Gender

T lymphocytes	n 183	♂ 93	♀ 91
CD3+CD4+			
CCR7+ (%)	85.8±4.8	85.1±5.5	86.6±4.1
CCR5+ (%)	6.9±4.1	7.3±4.4	6.6±3.8
CXCR3+ (%)	26.4±9.1	27.4±10.0	25.5±8.0
HLA-DR+ (%)	4.9±2.4	5.0±2.0	4.8±2.7
Naïve CD45RA+RO-CCR7+ (%)	32.3±12.7	29.9±12.1	34.6±12.9
Memory CD45RA-RO+ (%)	60.3±13.4	62.8±12.1	57.8±14.4
Central memory (%)	49.3±11.4	50.9±10.1	47.6±12.3
Effector memory (%)	11.1±4.4	11.9±4.9	10.3±3.7

Data are presented as mean±standard deviation.

Different correlations between T-cell subsets and cardiometabolic risk factors were observed. Upon age-adjustment, T_{EM} were significantly and directly correlated with plasma total cholesterol, triglycerides, LDL cholesterol, glycemia, body mass index, and systolic blood pressure. Furthermore, subjects with metabolic syndrome or diabetes had significantly higher levels of T_{EM} (14.1±4.8% and 13.8±5.1%, respectively) compared with subjects without metabolic syndrome (10.8±1.2%, $P=0.02$) or without diabetes (10.8±1.3%, $P=0.03$). No significant correlation was observed between T_{CM} and cardiometabolic risk factors, while T_N were inversely correlated only with plasma triglyceride levels. Among other main subsets analyzed, HLA-DR⁺ T cells showed a direct correlation with total cholesterol, triglycerides, and LDL cholesterol, and an inverse correlation with HDL cholesterol.³⁹

Correlations Between Circulating T-Cell Subsets and Carotid IMT

Second, we investigated whether specific T-cell subsets are associated with preclinical atherosclerosis, defined by IMT at the common carotid levels. Table 5 summarizes the correlations between IMT with clinical, lipid, and metabolic variables, and T-cell subsets. IMT correlated positively with age, systolic blood pressure, total, and LDL cholesterol, triglyceride levels, body mass index, and glycemia, whereas IMT inversely correlated with high-density lipoprotein cholesterol levels. As expected, several correlations were lost after age-adjustment of the IMT (Table 5). The analysis of $CD3^+CD4^+$ T-cell subsets revealed that T_M , T_{EM} , and HLA-DR⁺ T cells significantly correlate with IMT, whereas T_N inversely correlates with IMT. As age is the known main determinant of carotid IMT, and several $CD3^+CD4^+$ T-cell subsets and cardio-metabolic risk factors correlated with age, the analysis of the correlation between

Table 5. Unadjusted and Age-Adjusted Correlation Between Intima-media thickness (IMT) and Clinical, Lipid, Metabolic and Inflammatory Parameters, and T-Cell Subsets (Pearson Correlation Coefficients are Shown, $n = 183$)

	IMT		Age-adjusted IMT	
	<i>r</i>	<i>P</i>	<i>r</i>	<i>P</i>
Age	0.63	<0.001	-	-
Clinical variables				
Systolic blood pressure	0.33	<0.001	0.07	0.38
Diastolic blood pressure	0.12	0.14	-0.04	0.61
Lipid profile				
Total cholesterol	0.15	0.04	-0.11	0.12
LDL cholesterol	0.19	0.01	-0.07	0.33
HDL cholesterol	-0.26	0.001	-0.23	0.03
Triglyceridemia	0.25	0.001	0.06	0.42
Metabolic profile				
Body mass index	0.26	<0.001	0.07	0.36
Creatinine	0.34	<0.001	0.15	0.05
Glycemia	0.27	<0.001	0.14	0.07
Inflammatory profile				
CRP	0.08	0.43	-0.02	0.84
T-lymphocytes profile				
CD3 ⁺ CD4 ⁺	-0.08	0.28	-0.11	0.11
Naïve CD45RA ⁺ RO ⁻ CCR7 ⁺	-0.19	0.01	0.10	0.18
Memory CD45RA ⁻ RO ⁺	0.18	0.02	-0.05	0.54
Central memory T	0.06	0.42	-0.10	0.20
Effector memory T	0.40	<0.001	0.27	<0.001
CCR5 ⁺	0.08	0.30	0.10	0.18
CXCR3 ⁺	0.14	0.05	0.04	0.63
HLA-DR ⁺	0.28	<0.001	0.14	0.07

LDL indicates low-density lipoprotein; HDL, high-density lipoprotein; CRP, C-reactive protein.

T-cell subsets and IMT was performed after age-adjustment. A significant direct correlation remained between age-adjusted IMT and T_{EM} ($P < 0.001$), whereas only a trend for correlation was observed between age-adjusted IMT and HLA-DR⁺ ($P = 0.07$).

In the multiple regression analysis, age, creatinine, and T_{EM} cells emerged as independent predictors of IMT (Table 6). We cannot exclude that the relatively limited number of subjects studied could have underestimated the impact of the lipid profile on the IMT.

Table 6. Overall Multiple Regression Analysis Between Intima-Media Thickness (IMT; dependent variable) and Age, Gender, Smoking Habits, Clinical Parameters, and T-Cell Subsets that Correlate with IMT in the Single Model

Independent variables	Model	
	β	<i>P</i>
Age	0.56	<0.01
Gender	0.05	0.67
Smoking habits	-0.03	0.98
Systolic Blood Pressure	-0.02	0.82
Total cholesterol	-0.12	0.13
LDL cholesterol	-0.11	0.11
HDL cholesterol	-0.10	0.09
Triglyceridemia	0.01	0.89
Body mass index	-0.05	0.51
Creatinine	0.19	<0.01
Glycemia	0.05	0.55
CD3 ⁺ CD4 ⁺		
Naïve T cells	-0.12	0.19
Central memory T cells	0.11	0.56
Effector memory T cells	0.22	<0.01
CCR5 ⁺	-0.07	0.34
CXCR3 ⁺	-0.06	0.45
HLA-DR ⁺	0.09	0.26

BMI indicates body mass index; HDL, high-density lipoprotein; LDL, low-density lipoprotein.

T-Cells Subset Levels in Controls and Patients With CAD

To further confirm the association of T_{EM} with the extent of atherosclerosis, we studied the levels of T_{EM} in relation to symptomatic atherosclerosis in the coronary district. Levels of CD3⁺CD4⁺T-cell subsets were investigated in a second independent cohort of patients with stable and unstable manifestations of CAD compared with age- and sex-matched controls. While T_N and T_{CM} levels were not different among patients with CSA, AMI, and controls; T_{EM} resulted in significant increase in both CSA and AMI patients compared with controls ($P < 0.01$ for both) with similar levels between CSA and AMI patients (Figure 5 and Table 7). Similarly, HLA-DR⁺T cells were significantly increased in patients with CSA and AMI compared with controls ($P < 0.01$ for both), without differences between CSA and AMI patients. CCR5⁺T cells and CXCR3⁺T cell levels did not differ among controls, and patients with CSA and AMI (Table 7).

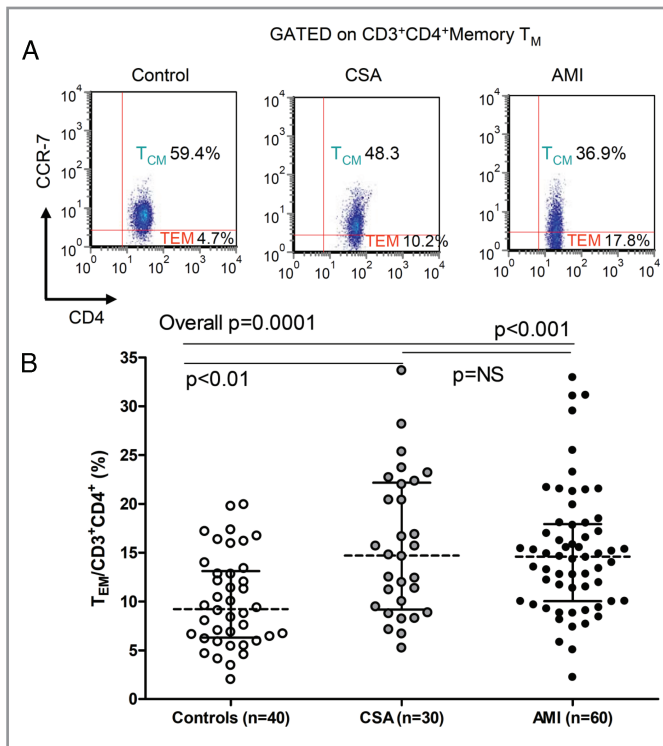


Figure 5. Effector memory T cells (T_{EM}) levels are increased in patients with coronary artery disease (CAD). Representative color dot plots from a control and patients with different CAD manifestations: CSA, chronic stable angina; AMI, acute myocardial infarction (A). Significant increases in T_{EM} levels were observed in patients with CSA and AMI in comparison with controls. There were no significant difference in levels of T_{EM} between CSA and AMI. Kruskal-Wallis and Dunn's test were performed. Dots represent individual patient data; dashed lines show median value and continuous lines show 25th and 75th percentiles (B).

Circulating T_{EM}-Cell Subpopulations in Subclinical Atherosclerosis and in CAD

Given the potential association between T_{EM} and atherosclerosis and to further investigate whether any of the T_{EM} subpopulations could better explain this finding, the correlation between CXCR3⁺T_{EM}, CCR5⁺T_{EM}, HLA-DR⁺T_{EM} subpopulations, and IMT was investigated. Compared with all T_{EM} that showed a strong correlation with IMT (Spearman $r=0.27$, $P<0.01$), HLA-DR⁺T_{EM} showed the highest degree of correlation (Spearman $r=0.37$, $P<0.01$), with CCR5⁺T_{EM} showing a degree of correlation similar to T_{EM} (Spearman $r=0.25$, $P<0.01$) while a lower but still significant correlation was observed also for T_{EM} CXCR3⁺T_{EM} and IMT (Spearman $r=0.15$, $P<0.05$) (Figure 6A). HLA-DR⁺T_{EM} and CXCR3⁺T_{EM} resulted the independent covariates associated with IMT (Table 8). These data were supported with the findings in patients with CAD. Indeed HLA-DR⁺T_{EM} subpopulation resulted significantly increased in both CSA and AMI patients compared with controls ($P<0.01$ for both) and, again, HLA-DR⁺T_{EM} levels were not different in CSA and AMI. Also CXCR3⁺T_{EM} were significantly increased in CSA and AMI patients compared with controls (overall $P=0.002$), whereas no differences were observed for CCR5⁺T_{EM} (Figure 6B). These data suggest that a specific T_{EM} subpopulation, namely HLA-DR⁺T_{EM}, which express the activation marker HLA-DR, could better reflect the association with the atherogenic process.

T_{EM} Cells Were Increased in Animal Model of Atherosclerosis and Correlate With the Extent of Atherosclerosis

To further characterize the presence of T_{EM}, T_{CM}, or T_N cells in relation to the extent of atherosclerosis, the presence of CD4⁺CD44⁻CD62L⁺ (T_N), CD4⁺CD44⁺CD62L⁻ (T_{EM}),

Table 7. Circulating T-Cell Subset Levels in Controls, Patients With Chronic Stable Angina (CSA) and Acute Myocardial Infarction (AMI)

	Controls (n = 40)	CSA (n = 30)	AMI (n = 60)
T-lymphocytes profile CD3 ⁺ CD4 ⁺			
Naïve CD45RA ⁺ RO ⁻ CCR7 ⁺	29.3 (18.3–35.2)	24.0 (17.9–33.8)	24.4 (19.7–32.8)
Memory CD45RA ⁻ RO ⁺	59.5 (15.9)	65.0 (12.4)	62.8 (12.6)
Central memory T	49.3 (14.5)	48.9 (10.9)	46.6 (11.7)
Effector memory T	9.2 (6.3–13.1)	14.7 (9.2–22.2)*	14.6 (10.0–17.9) [†]
CCR5 ⁺	9.1 (5.9–12.3)	7.0 (5.2–17.4)	9.3 (5.4–18.3)
CXCR3 ⁺	27.5 (22.1–33.0)	28.9 (23.7–35.4)	28.2 (21.3–31.5)
HLA-DR ⁺	2.2 (1.3–4.0)	5.8 (3.7–8.4) [†]	5.9 (4.0–9.9) [†]

* <0.01 and [†] <0.001 vs controls. Overall $P=0.0001$ for Effector Memory T (T_{EM}), and overall $P=0.002$ for HLA-DR⁺ (Kruskal-Wallis test). Results are expressed as percentage of total CD3⁺CD4⁺T cells. Values are presented as mean (standard deviation) or as median (1^o to 3^o quartile) as appropriate.

Table 8. Relative Contribution of Each Effector Memory T-Cell Subset to IMT Prediction. Data From Forced Entry Multiple Regression Analysis With IMT as Dependent Variable and T_{EM} , T_{EM} -HLA-DR+, T_{EM} -CCR5+, and T_{EM} -CXCR3+ as Covariates Are Shown

Model		Standardized Coefficient	<i>t</i>	Significance	Confidence Intervals 95.0% for B	
		Beta			Lower limit	Upper limit
1	(Constant)		21.554	0.000	0.528	0.634
	EM	-0.227	-1.333	0.184	-0.018	0.003
	TEM_CCR5	-0.028	-0.381	0.704	-0.048	0.032
	TEM_CXCR3	0.370	2.569	0.011	0.005	0.038
	TEM_HLA_DR	0.432	4.425	<0.001	0.072	0.189

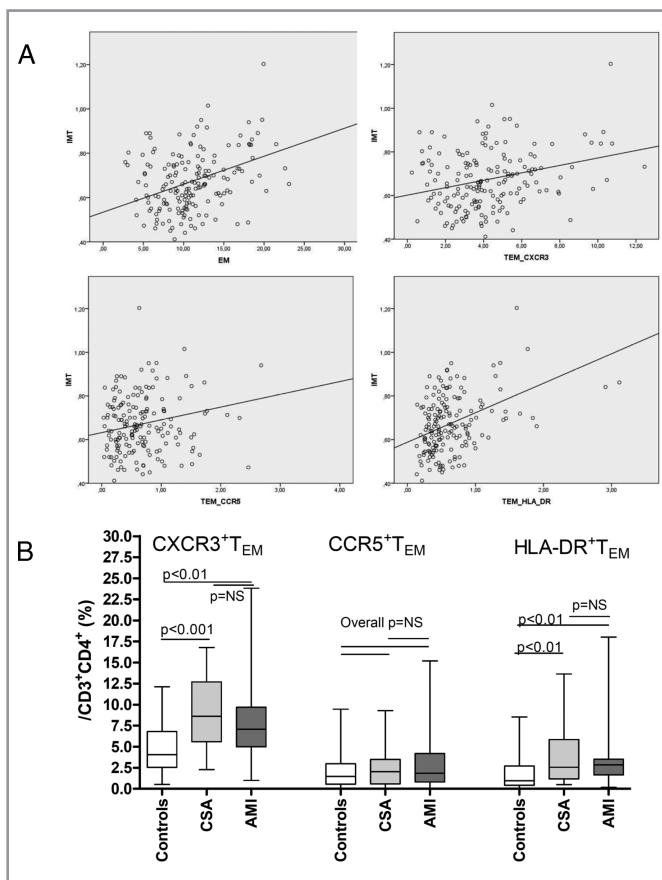


Figure 6. Effector memory T-cell (T_{EM}) subpopulations correlate with intima-media thickness (IMT) and are increased in patients with coronary artery disease (CAD). The correlation between IMT and T_{EM} , $CXCR3^+T_{EM}$, $HLA-DR^+T_{EM}$, and $CCR5^+T_{EM}$ is shown in A to D (see Methods Section for details). (E) The percentage of $CXCR3^+T_{EM}$, $HLA-DR^+T_{EM}$, and $CCR5^+T_{EM}$ in patients with chronic stable angina (CSA, $n = 30$) or acute myocardial infarction (AMI, $n = 60$) and controls ($n = 40$). The Kruskal-Wallis test with Dunn's comparison for all groups was used.

and of $CD4^+CD44^+CD62L^+$ (T_{CM}) was investigated in animal models of atherosclerosis fed an atherogenic diet (Figure 7A).⁴⁰ Of note, the percentage of T_N significantly decreased while that of T_{EM} significantly increased in LDL re-

ceptor knockout and apolipoprotein E knockout animals compared with control animals (Figure 7B). T_{CM} were significantly increased in LDL receptor knockout mice (Figure 7B). Of interest, the percentage of T_N was inversely correlated with the extent of atherosclerotic lesions in the aortic sinus of the animals ($r = -0.57$; $P < 0.01$) while that of T_{EM} was directly correlated ($r = 0.56$; $P < 0.01$; Figure 7C), further supporting the link between T_{EM} and atherosclerosis also in animal models.

Discussion

Several studies have suggested a role of $CD4^+$ T cells in the pathogenesis of atherosclerosis.^{2,5,41} The present study is, to date, the largest one which investigated circulating $CD4^+$ T-cell subsets in relation to atherosclerotic disease in humans. The simultaneous assessment of 8 independent membrane T-cell molecules by polychromatic flow cytometry allowed to identify more accurately and in relatively unbiased manner the phenotype of circulating T cells.

T_{EM} emerged as the T-cell subset with the strongest association with atherosclerosis in carotid and coronary vascular districts at different stages of the process. T_{EM} were significantly and directly correlated with plasma total cholesterol and LDL cholesterol, although the association between T_{EM} and carotid atherosclerosis was independent of the classical cardiovascular risk factors, supporting the relevance of adaptive immune response in cardiovascular disorders.^{28,41}

Partial characterizations of circulating T cells, which had been previously performed, are consistent with our results: indeed increased levels of $CD28^{\text{null}}$ or TCR^{dim} T cells and decreased levels of CCR7 have been reported in patients with CAD,^{16,27,42-44} whereas an expansion of the overall memory T-cell population has been associated with the extent of IMT in elderly Japanese subjects.⁴⁵

T_{CM} and T_{EM} cells persist in the memory pool once the antigen that elicited an immune response has been eliminated. They keep memory of (1) antigen specificity, (2) array of cytokines they had produced, and (3) the site where their

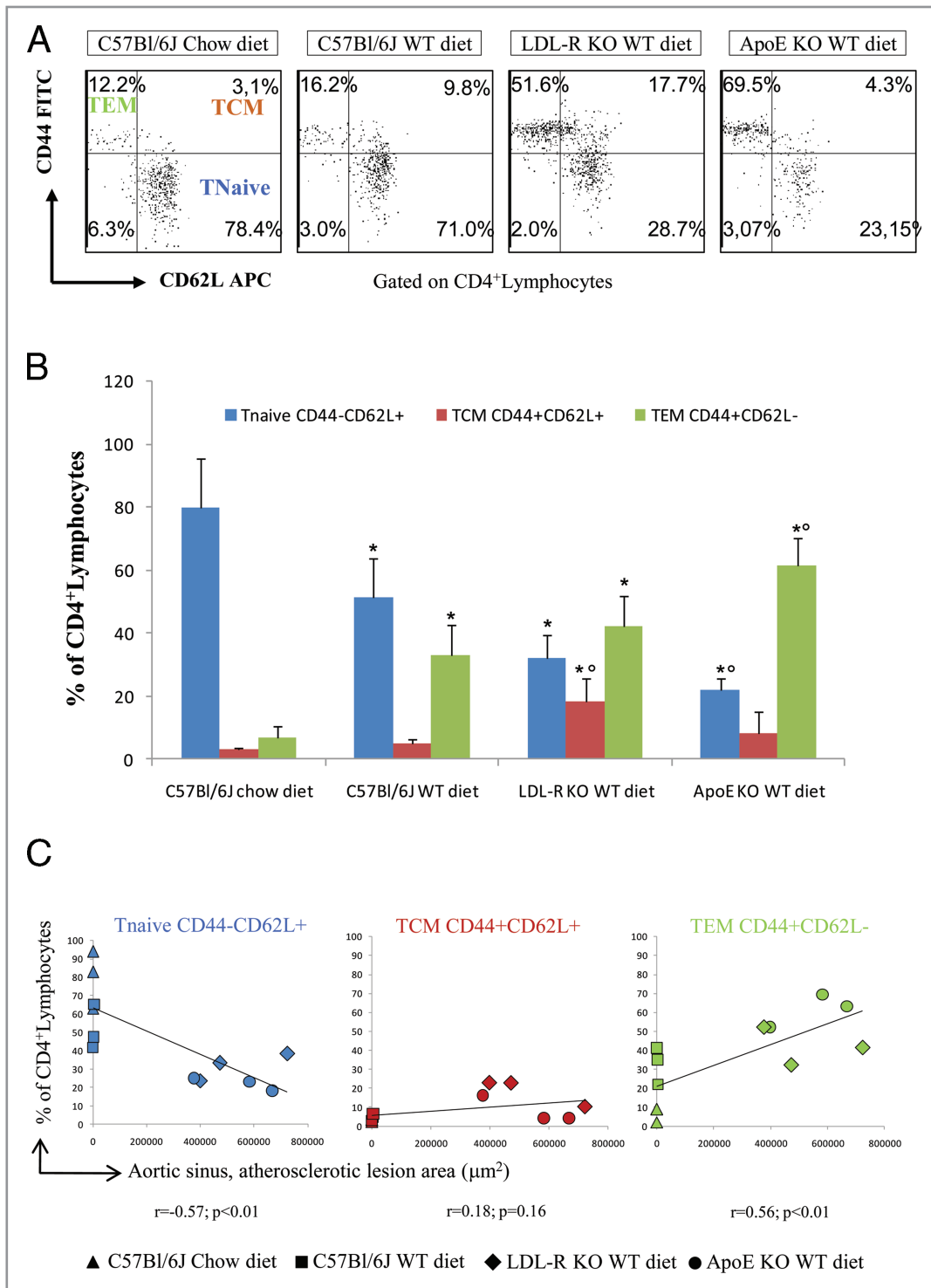


Figure 7. Effector memory (T_{EM}) levels are increased in animal models of atherosclerosis. Representative color dot plots from wild-type (C57BL/6J) animals fed a chow diet, a cholesterol rich (western type, WT) diet and from LDL receptor (LDL-R), and apolipoprotein E (Apo-E) knockout animals are shown in (A), while the percentages of circulating naïve T cells (T_N: in mouse defined as CD62L⁺CD44⁻), central memory T cells (T_{CM}: CD62L⁺CD44⁺), and memory effector T cells (T_{EM}: CD62L⁻CD44⁺) are shown in (B) (n = 12; 3 for each group) (*P<0.05 vs C57Bl/6J chow diet; °P<0.05 vs C57Bl/6J WT diet, pairwise test with Bonferroni correction). The correlations between circulating levels of T_N, T_{CM}, and T_{EM} and atherosclerotic plaque area at the aortic sinus are shown in panel C (n = 12; 3 for each group, Spearman correlation coefficients are shown).

effector function is needed. Upon antigen reexposure, T_{EM} display immediate effector functions in inflamed peripheral tissues, whereas T_{CM}, activated by dendritic cells in secondary lymphoid organs, generate successive waves of effector cells.^{7,30} CD4⁺T_{EM} are normally excluded from resting lymph nodes and migrate in a CD62P-dependent fashion into reactive lymph nodes or to inflamed tissue, mainly by the expression of CCR5 and CXCR3. The expression of HLA-DR constitutes a marker of effector function acquisition.⁴⁰ The function of T cells depends on their ability to exploit integrins, selectins and chemokine receptors to extravasate and migrate to sites where antigen is present. Of note, increased levels of T_{EM} have been previously described in vasculitis and chronic graft versus host disease.^{46,47} Also in these conditions, altered levels of human T_{EM} were not accompanied by significant changes in T_N and T_{CM} pools.^{46,47}

Potential Role of T_{EM} in the Atherosclerotic Process

In hypercholesterolemic animal models it has been observed that CCR7 knockout attenuates ath-

therosclerotic plaque development.¹⁷ This finding stresses the relevance of the differentiation process of naïve CD4⁺T cells to effector and/or memory cells of specialized T-cell subsets (T_{CM} and T_{EM}), and the CCR7-dependent T-cell and monocyte migration during atherogenesis. However, as CCR7 is expressed on several lymphocytes, including cells that express adhesion molecules required for homing to nonlymphoid tissues,⁴⁸ the deficiency of CCR7 could also impair the trafficking of cells other than CD4 lymphocytes during atherosclerosis. Therefore, the identification of a correlation between a specific memory T-cell subset that lost CCR7 receptor (T_{EM}) with atherosclerosis and at the same time, the association with LDL cholesterol levels, support the intriguing interaction between LDL cholesterol, expansion of T_{EM}, and atherosclerosis.⁵ Indeed, experimental models have shown that CD4⁺T cells recognize epitopes on native ApoB100 protein. Furthermore, blocking T-cell receptor-dependent antigen recognition by these T cells protects against atherosclerosis.^{5,49} Our data demonstrate that animal models where atherosclerosis derives from impaired lipid metabolism and dyslipidemia, have increased levels of T_{EM} compared with controls that are directly associated with the extent of the atherosclerotic lesions. While these results can support the hypothesis of a causal link between LDL cholesterol and T_{EM} increase, further studies are warranted in patients to investigate whether a sustain antigen stimulation, possibly mediated by LDL cholesterol, could explain the expansion of T_{EM}. In this context, although smoking is a strong determinant of total leukocytes count and differential leukocytes subsets, smoking habits were not associated with increased T_{EM} levels. This might suggest other mechanisms associated with smoking deleterious effects on atherosclerosis.

Of note, within the T_{EM} subsets, we have identified subpopulations with a strong correlation with the extent of atherosclerosis, such as those expressing HLA-DR, CXCR3, and CCR5. These receptors modulate the recruitment of T cells into the atherosclerotic plaques,^{25,26} and could thus represent an asset to facilitate T_{EM} patrolling of atherosclerotic plaques at distant sites in the organism. We further confirmed previous reports that had identified the expression of HLA-DR as a T-cell subset associated with CAD, and we identified T_{EM} as the CD4⁺T-cell subset in which HLA-DR is specifically increased.^{14,50}

Conclusion

Among CD4⁺T cell, T_{EM} and related T_{EM} subpopulations are those strongly correlated with the extent of atherosclerosis in carotid and coronary districts. As T_{EM} are antigen-experienced and long-surviving cells that have lost CCR7 and express HLA-DR, CXCR3, and CCR5, this finding strengthens the concept that the understanding of the inflammatory pathogenesis of

atherosclerosis requires careful cellular subphenotyping. Furthermore, the presence of increased levels of circulating T_{EM} in humans and the association with the extent of aortic lesions in animal models, suggest the possibility of targeting plaque chemotaxis and/or antigen encountering as emerging antiatherosclerotic strategies on the top of cardiovascular risk factors control.^{25,26,49}

Acknowledgments

We thank Ivan Muradore for technical support (Flow cytometry Resource Analytical Cytology Technical Applications Laboratory, San Raffaele Scientific Institute, Milan, Italy). We thank Elena Tragni and Manuela Casula for statistical support (SEFAP, Centro Interuniversitario di Epidemiologia, Università degli Studi di Milano).

Sources of Funding

E.A. received financial support from the “Giovane Ricercatore 2009 Grant” from Italian Health Ministry (project code GR-2009-1608780) and from the Heart Care Foundation, Florence, Italy. G.D.N. received financial support from the Cariplo Foundation and from the Italian Society for the Study of Atherosclerosis, Lombardia Chapter. A.L.C. received financial support from the Cariplo Foundation.

Disclosures

None.

References

- Libby P, Ridker PM, Maseri A. Inflammation and atherosclerosis. *Circulation*. 2002;105:1135–1143.
- Hansson GK, Hermansson A. The immune system in atherosclerosis. *Nat Immunol*. 2011;12:204–212.
- Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med*. 2005;352:1685–1695.
- Zernecke A, Weber C. Chemokines in the vascular inflammatory response of atherosclerosis. *Cardiovasc Res*. 2010;86:192–201.
- Hermansson A, Ketelhuth DF, Strothoff D, Wurm M, Hansson EM, Nicoletti A, Paulsson-Berne G, Hansson GK. Inhibition of T cell response to native low-density lipoprotein reduces atherosclerosis. *J Exp Med*. 2010;207:1081–1093.
- Stemme S, Faber B, Holm J, Wiklund O, Witztum JL, Hansson GK. T lymphocytes from human atherosclerotic plaques recognize oxidized low density lipoprotein. *Proc Natl Acad Sci U S A*. 1995;92:3893–3897.
- Sallusto F, Geginat J, Lanzavecchia A. Central memory and effector memory T cell subsets: function, generation, and maintenance. *Annu Rev Immunol*. 2004;22:745–763.
- Stemme S, Holm J, Hansson GK. T lymphocytes in human atherosclerotic plaques are memory cells expressing CD45RO and the integrin VLA-1. *Arterioscler Thromb*. 1992;12:206–211.
- Hosono M, de Boer OJ, van der Wal AC, van der Loos CM, Teeling P, Piek JJ, Ueda M, Becker AE. Increased expression of T cell activation markers (CD25, CD26, CD40L and CD69) in atherectomy specimens of patients with unstable angina and acute myocardial infarction. *Atherosclerosis*. 2003;168:73–80.
- Hansson GK, Holm J, Jonasson L. Detection of activated T lymphocytes in the human atherosclerotic plaque. *Am J Pathol*. 1989;135:169–175.
- De Palma R, Del Galdo F, Abbate G, Chiariello M, Calabro R, Forte L, Cimmino G, Papa MF, Russo MG, Ambrosio G, Giombolini C, Tritto I, Notaristefano S, Berrino L, Rossi F, Golino P. Patients with acute coronary syndrome show oligoclonal T-cell recruitment within unstable plaque: evidence for a local, intracoronary immunologic mechanism. *Circulation*. 2006;113:640–646.

12. Sun J, Hartvigsen K, Chou MY, Zhang Y, Sukhova GK, Zhang J, Lopez-Illasaca M, Diehl CJ, Yakov N, Harats D, George J, Witztum JL, Libby P, Ploegh H, Shi GP. Deficiency of antigen-presenting cell invariant chain reduces atherosclerosis in mice. *Circulation*. 2010;122:808–820.
13. Buffon A, Biasucci LM, Liuzzo G, D'Onofrio G, Crea F, Maseri A. Widespread coronary inflammation in unstable angina. *N Engl J Med*. 2002;347:5–12.
14. Caligiuri G, Paulsson G, Nicoletti A, Maseri A, Hansson GK. Evidence for antigen-driven T-cell response in unstable angina. *Circulation*. 2000;102:1114–1119.
15. Methe H, Brunner S, Wiegand D, Nabauer M, Koglin J, Edelman ER. Enhanced T-helper-1 lymphocyte activation patterns in acute coronary syndromes. *J Am Coll Cardiol*. 2005;45:1939–1945.
16. Ammirati E, Vermi AC, Cianflone D, Banfi M, Foglieni C, Godino C, Airoldi F, Ferri LA, Gorman CL, Manfredi AA, Maseri A, Cope AP, Monaco C. Expansion of T-cell receptor zeta dim effector T cells in acute coronary syndromes. *Arterioscler Thromb Vasc Biol*. 2008;28:2305–2311.
17. Luchtfeld M, Grothausen C, Gagalik A, Jagavelu K, Schuett H, Tietge UJ, Pabst O, Grote K, Drexler H, Forster R, Schieffer B. Chemokine receptor 7 knockout attenuates atherosclerotic plaque development. *Circulation*. 2010;122:1621–1628.
18. Feig JE, Pineda-Torra I, Sanson M, Bradley MN, Vengrenyuk Y, Bogunovic D, Gautier EL, Rubinstein D, Hong C, Liu J, Wu C, van Rooijen N, Bhardwaj N, Garabedian M, Tontonoz P, Fisher EA. LXR promotes the maximal egress of monocyte-derived cells from mouse aortic plaques during atherosclerosis regression. *J Clin Invest*. 2010;120:4415–4424.
19. Bromley SK, Thomas SY, Luster AD. Chemokine receptor CCR7 guides T cell exit from peripheral tissues and entry into afferent lymphatics. *Nat Immunol*. 2005;6:895–901.
20. Feig JE, Rong JX, Shamir R, Sanson M, Vengrenyuk Y, Liu J, Rayner K, Moore K, Garabedian M, Fisher EA. HDL promotes rapid atherosclerosis regression in mice and alters inflammatory properties of plaque monocyte-derived cells. *Proc Natl Acad Sci U S A*. 2011;108:7166–7171.
21. Lievens D, Zerneck A, Seijkens T, Soehnlein O, Beckers L, Munnix IC, Wijnands E, Goossens P, van Kruchten R, Thevissen L, Boon L, Flavell RA, Noelle RJ, Gerdes N, Biessen EA, Daemen MJ, Heemskerk JW, Weber C, Lutgens E. Platelet CD40L mediates thrombotic and inflammatory processes in atherosclerosis. *Blood*. 2010;116:4317–4327.
22. Mach F, Sauty A, Iarossi AS, Sukhova GK, Neote K, Libby P, Luster AD. Differential expression of three T lymphocyte-activating CXC chemokines by human atheroma-associated cells. *J Clin Invest*. 1999;104:1041–1050.
23. van Loosdregt J, van Oosterhout MF, Bruggink AH, van Wichen DF, van Kuik J, de Koning E, Baan CC, de Jonge N, Gmelig-Meyling FH, de Weger RA. The chemokine and chemokine receptor profile of infiltrating cells in the wall of arteries with cardiac allograft vasculopathy is indicative of a memory T-helper 1 response. *Circulation*. 2006;114:1599–1607.
24. Braunersreuther V, Steffens S, Arnaud C, Pelli G, Burger F, Proudfoot A, Mach F. A novel RANTES antagonist prevents progression of established atherosclerotic lesions in mice. *Arterioscler Thromb Vasc Biol*. 2008;28:1090–1096.
25. van Wanrooij EJ, Happe H, Hauer AD, de Vos P, Imanishi T, Fujiwara H, van Berkel TJ, Kuiper J. HIV entry inhibitor TAK-779 attenuates atherogenesis in low-density lipoprotein receptor-deficient mice. *Arterioscler Thromb Vasc Biol*. 2005;25:2642–2647.
26. van Wanrooij EJ, de Jager SC, van Es T, de Vos P, Birch HL, Owen DA, Watson RJ, Biessen EA, Chapman GA, van Berkel TJ, Kuiper J. CXCR3 antagonist NBI-74330 attenuates atherosclerotic plaque formation in LDL receptor-deficient mice. *Arterioscler Thromb Vasc Biol*. 2008;28:251–257.
27. Zhang X, Niessner A, Nakajima T, Ma-Krupa W, Kopecky SL, Frye RL, Goronzy JJ, Weyand CM. Interleukin 12 induces T-cell recruitment into the atherosclerotic plaque. *Circ Res*. 2006;98:524–531.
28. Libby P, Ridker PM, Hansson GK. Progress and challenges in translating the biology of atherosclerosis. *Nature*;473:317–325.
29. Perfetto SP, Chattopadhyay PK, Roederer M. Seventeen-colour flow cytometry: unravelling the immune system. *Nat Rev Immunol*. 2004;4:648–655.
30. Sallusto F, Lenig D, Forster R, Lipp M, Lanzavecchia A. Two subsets of memory T lymphocytes with distinct homing potentials and effector functions. *Nature*. 1999;401:708–712.
31. Norata GD, Garlaschelli K, Ongari M, Raselli S, Grigore L, Benvenuto F, Maggi FM, Catapano AL. Effect of the Toll-like receptor 4 (TLR-4) variants on intima-media thickness and monocyte-derived macrophage response to LPS. *J Intern Med*. 2005;258:21–27.
32. Cristell N, Cianflone D, Durante A, Ammirati E, Vanuzzo D, Banfi M, Calori G, Latib A, Crea F, Marenzi G, De Metrio M, Moretti L, Li H, Uren NG, Hu D, Maseri A. High-sensitivity C-reactive protein is within normal levels at the very onset of first ST-segment elevation acute myocardial infarction in 41% of cases a multiethnic case-control study. *J Am Coll Cardiol*. 2011;58:2654–2661.
33. Ammirati E, Cianflone D, Banfi M, Vecchio V, Palini A, De Metrio M, Marenzi G, Panciroli C, Tumminello G, Anzuini A, Pallosi A, Grigore L, Garlaschelli K, Tramontana S, Tavano D, Airoldi F, Manfredi AA, Catapano AL, Norata GD. Circulating CD4+CD25hiCD127lo regulatory T-Cell levels do not reflect the extent or severity of carotid and coronary atherosclerosis. *Arterioscler Thromb Vasc Biol*. 2010;30:1832–1841.
34. Perfetto SP, Ambrozak D, Nguyen R, Chattopadhyay P, Roederer M. Quality assurance for polychromatic flow cytometry. *Nat Protoc*. 2006;1:1522–1530.
35. Norata GD, Marchesi P, Pulakazhi Venu VK, Pasqualini F, Anselmo A, Moalli F, Pizzitola I, Garlanda C, Mantovani A, Catapano AL. Deficiency of the long pentraxin PTX3 promotes vascular inflammation and atherosclerosis. *Circulation*. 2009;120:699–708.
36. Tibolla G, Norata GD, Meda C, Arnaboldi L, Ubaldi P, Piazza F, Ferrarese C, Corsini A, Maggi A, Vegeto E, Catapano AL. Increased atherosclerosis and vascular inflammation in APP transgenic mice with apolipoprotein E deficiency. *Atherosclerosis*. 2010;210:78–87.
37. Goronzy JJ, Lee WW, Weyand CM. Aging and T-cell diversity. *Exp Gerontol*. 2007;42:400–406.
38. Provinciali M, Moresi R, Donnini A, Lisa RM. Reference values for CD4 +and CD8+ T lymphocytes with naive or memory phenotype and their association with mortality in the elderly. *Gerontology*. 2009;55:314–321.
39. Norata GD, Pirillo A, Ammirati E, Catapano AL. Emerging role of high density lipoproteins as a player in the immune system. *Atherosclerosis*. 2012;220:11–21.
40. Martin-Fontecha A, Baumjohann D, Guarda G, Reboldi A, Hons M, Lanzavecchia A, Sallusto F. CD40L+ CD4+ memory T cells migrate in a CD62P-dependent fashion into reactive lymph nodes and license dendritic cells for T cell priming. *J Exp Med*. 2008;205:2561–2574.
41. Lahoute C, Herbin O, Mallat Z, Tedgui A. Adaptive immunity in atherosclerosis: mechanisms and future therapeutic targets. *Nat Rev Cardiol*. 2011;8:348–358.
42. Liuzzo G, Goronzy JJ, Yang H, Kopecky SL, Holmes DR, Frye RL, Weyand CM. Monoclonal T-cell proliferation and plaque instability in acute coronary syndromes. *Circulation*. 2000;101:2883–2888.
43. Raines EW. Antigen-independent targeting of long-lived CD4 +cytolytic T effector cells to lesions of atherosclerosis. *Circ Res*. 2006;98:434–436.
44. Damas JK, Smith C, Oie E, Fevang B, Halvorsen B, Waehre T, Boullier A, Breland U, Yndestad A, Ovchinnikova O, Robertson AK, Sandberg WJ, Kjekshus J, Tasken K, Froland SS, Gullestad L, Hansson GK, Quehenberger O, Aukrust P. Enhanced expression of the homeostatic chemokines CCL19 and CCL21 in clinical and experimental atherosclerosis: possible pathogenic role in plaque destabilization. *Arterioscler Thromb Vasc Biol*. 2007;27:614–620.
45. Tanigawa T, Kitamura A, Yamagishi K, Sakurai S, Nakata A, Yamashita H, Sato S, Ohira T, Imano H, Shimamoto T, Iso H. Relationships of differential leukocyte and lymphocyte subpopulations with carotid atherosclerosis in elderly men. *J Clin Immunol*. 2003;23:469–476.
46. Abdulahad WH, van der Geld YM, Stegeman CA, Kallenberg CG. Persistent expansion of CD4+ effector memory T cells in Wegener's granulomatosis. *Kidney Int*. 2006;70:938–947.
47. Yamashita K, Choi U, Woltz PC, Foster SF, Sneller MC, Hakim FT, Fowler DH, Bishop MR, Pavletic SZ, Tamari M, Castro K, Barrett AJ, Childs RW, Illei GG, Leitman SF, Malech HL, Horwitz ME. Severe chronic graft-versus-host disease is characterized by a preponderance of CD4(+) effector memory cells relative to central memory cells. *Blood*. 2004;103:3986–3988.
48. Campbell JJ, Murphy KE, Kunkel EJ, Brightling CE, Soler D, Shen Z, Boisvert J, Greenberg HB, Vierra MA, Genovese MC, Wardlaw AJ, Butcher EC, Wu L. CCR7 expression and memory T cell diversity in humans. *J Immunol*. 2001;166:877–884.
49. Hermansson A, Johansson DK, Ketelhuth DF, Andersson J, Zhou X, Hansson GK. Immunotherapy with tolerogenic apolipoprotein B-100-loaded dendritic cells attenuates atherosclerosis in hypercholesterolemic mice. *Circulation*. 2011;123:1083–1091.
50. Neri Serneri GG, Prisco D, Martini F, Gori AM, Brunelli T, Poggesi L, Rostagno C, Gensini GF, Abbate R. Acute T-cell activation is detectable in unstable angina. *Circulation*. 1997;95:1806–1812.