

Review

Open Access

## Potential involvement of the immune system in the development of endometriosis

Cleophas M Kyama<sup>1,2</sup>, Sophie Debrock<sup>1</sup>, Jason M Mwenda<sup>2</sup> and Thomas M D'Hooghe\*<sup>1,2</sup>

Address: <sup>1</sup>Leuven University Fertility Centre, Department of Obstetrics & Gynaecology, University Hospital Gasthuisberg, Leuven, Belgium and <sup>2</sup>Division of Reproductive Biology, Institute of Primate Research, Karen, Nairobi, Kenya

Email: Cleophas M Kyama - [kcleophas@yahoo.com](mailto:kcleophas@yahoo.com); Sophie Debrock - [sophie.debrock@uz.kuleuven.ac.be](mailto:sophie.debrock@uz.kuleuven.ac.be); Jason M Mwenda - [jmmwenda@arcc.or.ke](mailto:jmmwenda@arcc.or.ke); Thomas M D'Hooghe\* - [thomas.dhooghe@uz.kuleuven.ac.be](mailto:thomas.dhooghe@uz.kuleuven.ac.be)

\* Corresponding author

Published: 02 December 2003

Received: 26 August 2003

*Reproductive Biology and Endocrinology* 2003, 1:123

Accepted: 02 December 2003

This article is available from: <http://www.rbej.com/content/1/1/123>

© 2003 Kyama et al; licensee BioMed Central Ltd. This is an Open Access article: verbatim copying and redistribution of this article are permitted in all media for any purpose, provided this notice is preserved along with the article's original URL.

### Abstract

This article presents an overview of immunological factors and their role in the development of endometriosis, with emphasis on inflammatory cytokines, growth and adhesion factors. Although retrograde menstruation is a common phenomenon among women of reproductive age, not all women who have retrograde menstruation develop endometriosis. The development of endometriosis is hypothesised to be a complex process, which may be facilitated by several factors, including the quantity and quality of endometrial cells in peritoneal fluid (PF), increased inflammatory activity in PF, increased endometrial-peritoneal adhesion and angiogenesis, reduced immune surveillance and clearance of endometrial cells, and increased production of autoantibodies against endometrial cells. Potential biomarkers like cytokines and autoantibodies upregulated during development of endometriosis may be useful in the development of a non-surgical diagnostic tool. Although endometriosis can be treated using hormonal suppression, there is need for non-hormonal drugs, which can inhibit the development of endometriosis and alleviate pain or infertility without inhibition of ovulation. New molecules that modulate immune function in endometriosis should be the targets for future research.

### Introduction

Endometriosis is a gynaecological disorder characterised by the presence and growth of endometrial tissues in the ectopic site. The endometrial deposits are mostly found in the pelvis (ovaries, peritoneum, uterosacral ligaments, pouch of Douglas and rectovaginal septum). The prevalence of endometriosis among asymptomatic women ranges from 2–22%, while in women with dysmenorrhoea, the incidence of endometriosis is 40% to 60% [1]. It is the most common cause of pelvic pain and occurs in 13%–33% of women with infertility [2]. Although endometriosis stands as one of the most investigated dis-

orders of gynaecology [3], our current understanding of aetiology and pathophysiology of the disease remains elusive. Retrograde menstruation is a widely accepted and proposed mechanism that may explain mostly the presence of endometrial cells in ectopic sites [4]. However, it does not account for the fact that these misplaced cells survive in women with endometriosis and not in healthy women. An immunological/inflammatory aetiology has been conjectured, as demonstrated by increased concentrations of activated macrophages, cytokines, T cells and B cells [2]. The endometrial fragments desquamated during menstruation and deposited into peritoneal cavity,

implant, proliferate and develop into endometriotic lesions. The development of endometriosis may be influenced by quantity and quality of endometrial cells in the PF, and by immune factors, including increased inflammatory activity in PF, impaired immune recognition and clearance of ectopic endometrial cells, and formation of autoantibodies. These factors will be discussed in this review.

### **Retrograde Menstruation**

Several theories exist that attempt to explain the mechanisms involved in development of endometriosis, but Sampson's theory [4] of retrograde menstruation has gained most supportive evidence. Retrograde menstruation is the reflux of menses through fallopian tube to ectopic site especially the peritoneal cavity. Viable endometrial cells have been found in the peritoneal cavity during nonmenstrual phases of the cycle [5] and during menses (reviewed by D'Hooghe et al. [2]). Although retrograde menstruation occurs in 70–90% of women [6] and 83% in baboons [7], endometriosis is diagnosed in at least 10% of the former and 25% of the latter. If only a small percentage of all women who have reverse menstruation actually develop the disease, the puzzling enigma is why the pelvic environment becomes receptive to allow implantation and proliferation of endometrial cells and development of endometriosis? Other factors must be involved allowing retrogradely displaced endometrial tissue to implant and develop into endometriotic lesions. It has been hypothesized that the quantity of endometrial cells deposited into the peritoneal cavity during menstruation could be higher among women who develop endometriosis, as suggested by a positive correlation between the number of menstrual cycles and the prevalence, cumulative incidence and progression of spontaneous endometriosis in baboons (reviewed by D'Hooghe and Debrock [8]). Similarly, it is well known that women with short cycles and long duration of menstrual flow are more likely to develop endometriosis [9]. Furthermore, outflow obstruction of menstrual effluent, resulting in excessive retrograde menstruation has been associated with endometriosis both in humans [9,10] and in baboons [11]. It may be possible that both dysmenorrhoea (painful menstruation) and endometriosis are manifestations of outflow obstruction [9].

### **Role of increased inflammatory activity in the development of endometriosis**

Peritoneal fluid (PF) in women with endometriosis is marked by increased inflammation, including increased volume of PF, increased concentration of white blood cells and macrophages, and increased activation status of these macrophages (reviewed by D'Hooghe and Hill [12]). These activated peripheral mononuclear cells as well as endometriotic cells *in situ* are hypothesised to

secrete various cytokines with pleiotropic biological activities. Cytokines are low molecular weight proteins or glycoproteins typically synthesised by peritoneal macrophages, lymphocytes, ectopic endometrial implants or mesothelial cells of the peritoneum [13,14]. Usually, inflammatory cytokines and growth factors are secreted and culminate in recruitment of numerous cell types to the peritoneal cavity [15]. Aberrant expression of several cytokines by activated macrophages, such as interleukin (IL)-1, IL-6, IL-8 and TNF- $\alpha$  in peritoneal fluid of women with endometriosis compared to controls [16] may contribute to a peritoneal microenvironment, which favours the implantation of endometrial cells and the establishment of endometriosis [17]. Indeed, as reviewed in the next section, cytokines like IL-8 and TNF-alpha are known to promote endometrial cell proliferation, endometrial adhesion and angiogenesis. Not only peritoneal macrophages, but also endometriotic lesions and mesothelial cells of peritoneal origin may secrete cytokines such as Tumor Necrosis Factor-alpha (TNF- $\alpha$ ) and Interleukin-1 (IL-1) in women with endometriosis. These cytokines in turn modulate the stimulation of other cytokines and chemokines such as Interleukin-8 (IL-8) and RANTES (Regulated upon activation, normal T-cell expressed and secreted). RANTES is a potent attractant and activator of macrophages, T-lymphocytes and eosinophils [18,19], while IL-8 promotes angiogenesis [20]. In one study [21], a positive correlation was found between the rAFS stages of endometriosis and the concentration of TNF- $\alpha$  in PF. The concentration of TNF- $\alpha$  in PF was significantly higher in patients with stage III/IV disease (168 pg/ml) than in women with stage I/II disease (60.2 pg/ml) or control patients (3.3 pg/ml) [21]. TNF- $\alpha$  and IL-8 concentrations in peritoneal fluid have also been reported to correlate with the size and number of active lesions [22]. The increased concentration of TNF- $\alpha$  reflects enhanced secretory activity of the peritoneal macrophages and not just the mere increase in the number of peritoneal macrophages [20].

### **Are changes in immunological/inflammatory mediators a cause or consequence of endometriosis?**

Does endometriosis lead to inflammation caused by an inappropriate and exaggerated immune response to ectopic endometrial debris? Or is endometriosis caused by peritoneal inflammation? Obviously, these cause-effect relationships cannot be studied in women for ethical reasons. In baboons, current evidence suggests that peritoneal inflammation is a consequence, not a cause of endometriosis.

Firstly, in baboons, both spontaneous retrograde menstruation and experimental intrapelvic injection of endometrium are associated with intrapelvic

inflammation (increased PF volume and increased PF concentration of white blood cells and inflammatory cytokines) [23,24]. This peritoneal inflammatory effect is observed within one month after intrapelvic injection of endometrium (24), but disappears after 2 to 3 months later [25]. Secondly, it has been reported that the WBC concentration and proportion of macrophages and cytotoxic T cells is increased in the PF of baboons with spontaneous endometriosis [25,26]. Thirdly, the percentage of CD4+ and IL2R+ cells has been shown to be increased in the peripheral blood of baboons with stage II to IV endometriosis, (both spontaneous long term endometriosis and induced) when compared to those with recent spontaneous endometriosis (Stage I) or a normal pelvis.

Even if peritoneal inflammation is a consequence rather than a cause of endometriosis [Fig. 1], the coexistence of endometriosis and peritoneal inflammation may offer new anti-inflammatory therapeutic options in the treatment of endometriosis. In an earlier study in baboons, a high dose of immunosuppression with azathioprin and methylprednisolone during 3 months did not affect the incidence of spontaneous endometriosis, the extent of induced endometriosis, and had a marginal stimulatory effect on the progression of spontaneous endometriosis [27], suggesting that overall immunosuppression does not have much effect on the incidence, prevalence or degree of endometriosis. Similarly, there is no evidence that the prevalence of endometriosis is higher in women using long term immunosuppression. However, more specific anti-inflammatory agents may affect the development of endometriosis. For instance drugs suppressing macrophage activation, such as verapamil (calcium channel blocking agent) and pentoxifylline, have been tested in hamster and mice respectively [28,29]. Other drugs have also been tested in rodents and nonhuman primates, as is reported in the last section of this paper.

### **Role of the immune system in endometrial-peritoneal adhesion**

#### **Quality of viable endometrial cells**

It has been hypothesised that the quality of endometrial cells in PF of women with endometriosis is different from women with normal pelvis. Viable endometrial cells from human endometriotic biopsies but not from human endometrial biopsies are invasive in an *in vitro* collagen invasion assay, probably because they have a higher proportion of potentially invasive E-cadherin-negative epithelial cells [30]. Inflammatory cytokines (TNF-alpha, IL-8 and IL-6) produced by endometrial cells probably contribute to this adhesion process [31-33]. IL-8 has been shown to stimulate the adhesion of endometrial cells to fibronectin [31]. TNF- $\alpha$  has been reported to also promote endometrial stromal cell proliferation *in vitro* [32] and endometrial stromal cell adhesion to extracellular

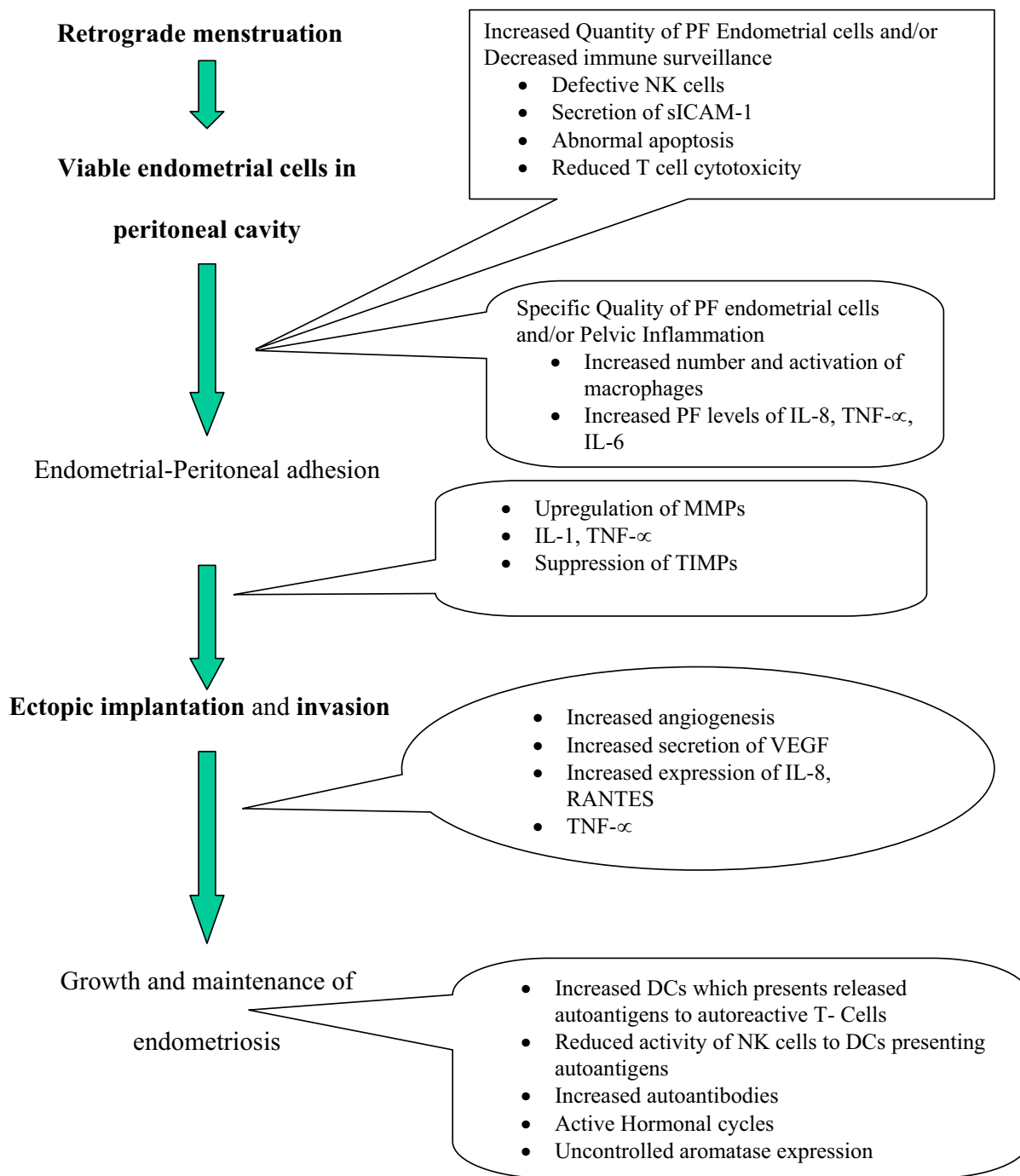
matrix components [33]. TNF- $\alpha$  may induce IL-8 gene and IL-8 protein expression in a dose-dependent manner, and the stimulating effect of TNF- $\alpha$  on endometrial stromal cell proliferation can be reversed by adding anti-IL8 antibodies [30].

Does endometriosis then only occur among women with a high degree of endometrial-peritoneal adhesion? This is unknown at present, since it is impossible to study this process in women *in vivo*. Debrock and colleagues [34] reported a 80–100% success rate of endometrial-peritoneal adhesion in cultured explants after 48 hours, regardless of the presence or absence of endometriosis. Witz *et al* [35] showed endometrial adhesion occurs within 1 hour and transmesothelial invasion occurs within 18 hours. However, all these assays are merely descriptive and there is a need to develop a quantitative *in vitro* assay to measure endometrial-peritoneal adhesion.

Endometrial quality can also be affected by local estrogen production in eutopic/ectopic endometrium. Indeed, the expression of uncontrolled aromatase mRNA in endometriotic lesions [36] suggests that a local estrogenic milieu is important in the development of endometriosis. It is possible that persistent expression of aromatase and 17 $\beta$ -hydroxysteroid dehydrogenase in endometriotic lesions may also be driven by a T-like autoantibody response [Table. 1]. Indeed, autoantibodies recognising T-like antigens have been reported to be upregulated in endometriosis and may trigger the synthesis of cytokines such as IL-1, TNF- $\alpha$  and IL-6, which in turn may induce the expression of aromatase and 17 $\beta$ -hydroxysteroid dehydrogenase in endometriotic lesions [37].

#### **Increased Angiogenesis**

Macrophage derived cytokines like TGF-beta and VEGF are suggested to contribute to the development of endometriosis by promoting neovascularisation of endometrial cells attached to the peritoneum [16,38]. Increased angiogenesis is reported to be common around the peritoneal explants and increased angiogenic activity has been observed in PF from women with endometriosis [39]. For instance, vascular endothelial growth factor (VEGF) has been detected in high concentration in PF from women with moderate to severe endometriosis [40-42], and is also secreted in endometriotic lesions, possibly as a downstream consequence of proinflammatory cytokine IL-1 $\beta$  activity [40,41]. VEGF is involved in the development of blood vessels, that are critical in the growth, and maintenance of ectopic endometrial implants.



**Figure 1**

A simplified view of the role of immune system in the development and maintenance of endometriosis. It illustrates the sequence of events leading to survival of endometrial cells in peritoneum, adherence, implantation, invasion and progression to disease.

**Table 1: Aberrantly regulated immune/inflammatory factors associated with the development of endometriosis**

Factors	Reference
Increased TNF- $\alpha$ , IL-8, IL-6, IL-1, TGF- $\beta$	[31,32]
Increased RANTES, VEGF	[19,40,41]
Increased Aromatase, 17 $\beta$ -hydroxysteroid dehydrogenase	[37]
Increased B cell function	[47]
Increased T-like autoantibodies against hemopexin	[37].
Increased MMPs	[44]
Decreased TIMPs	[45]
Increased IL-4, IL-10, sICAM-1	[38,51,57,58]

### **Upregulation of matrix metalloproteinases (MMPs) and potential involvement of auto-antibodies**

After attachment, endometrial cells invade the extracellular matrix, a process influenced by matrix metalloproteinases (MMPs). MMPs are a group of enzymes important for the control of extracellular matrix turnover [43]. MMPs are upregulated by TNF- $\alpha$  and IL-1, which could contribute to the invasiveness of endometrial fragments in women with endometriosis [44]. TNF- $\alpha$  may also contribute to the decreased expression of endogenous tissue inhibitors of MMPs (TIMPs) under *in vitro* conditions [45]. Both the downregulation of TIMP and increased expression of MMPs may support the invasive growth of endometriotic explants (Fig. 1). It has been postulated that the hemopexin domain expressed by most MMPs is involved in this process of MMP upregulation. Hemopexin is a Thomsen-Friedenreich antigen bearing serum protein that binds to jacalin. A hemopexin domain with high sequence homology to plasma hemopexin is expressed by MMPs except MMP-7 and can be recognised by T-like autoantibodies in women with endometriosis [37]. It has been postulated that the binding of T-like autoantibodies to the hemopexin domain may lead to dysregulation of the expression of MMPs and TIMPs in ectopic lesions, leading to increased invasiveness of these lesions in women with endometriosis (37). However, the role of autoantibodies in the development of endometriosis remains an area of great interest and ignorance, as reviewed before [46], even though new reports have suggested that in women with endometriosis B-cell activity is altered accompanied by increased incidence of autoantibodies [47,48].

### **Altered immune function in women with endometriosis**

#### **Defective natural killer cells activity?**

Retrograde menstruation occurs to some degree in all women. Therefore, it has been hypothesised that the clearance of endometrial cells within the pelvic cavity could be decreased among women with endometriosis. A defective cellular immunity (Fig. 1), especially impaired natural

killer (NK) cell function may contribute to the survival and ectopic implantation of sloughed endometrial cells. Wilson and colleagues [49] reported decreased NK cells cytotoxicity in women with endometriosis. D'Hooghe and colleagues [50] documented no difference in lymphocyte-mediated cytotoxicity and NK cell activity between baboons with and without endometriosis. A dysfunction of two subclasses of NK cells may nurture autoimmunity associated with endometriosis [51]. One subset, NK T cells, is characterised by the capacity to kill cell target and secrete cytokines, such as IL4 and IL-10, which are important in the regulation of autoimmunity [52]. Another subset of NK cells, CD16/CD56 NK cells kills autologous dendritic cells (DCs) presenting self-antigens to autoreactive T cells. The inability of NK cells in eliminating autologous DCs expressing endometrial self-antigens, may allow their presentation to autoreactive T cells and the production of autoantibodies [51]. The failure of NK cells to scavenge autologous endometrial cells may allow development of endometriosis. It has been hypothesized that natural killer cells in the endometrium can attack the implantation site of the embryo as it tries to attach to the uterine wall and that autoantibodies can play a role in this process [53]. However, a recent review has shown that there is no proven relationship between abortion, recurrent abortion and endometriosis [54], and at present there is no proof that endometriosis-associated subfertility is caused by impaired embryo implantation.

#### **Decreased T Lymphocyte Cytotoxicity**

CD4 T cells are divided into type1 (Th1) helper T cells which secrete interleukin (IL)-2, IL-12 and interferon  $\gamma$  and type2 (Th2) helper T cells, which secrete IL-4, 5, 6 10, 13 [38]. Cell-mediated immunity, including T-cell-mediated cytotoxicity is activated or suppressed by cytokines produced by Th1 and Th2 cells, respectively. Under normal conditions, there is a tightly regulatory control mechanism between Th1 and Th2 cells. For instance, Th1 cells secrete IL-12, which activates cytotoxic NK cell activity, whereas Th2 cells may reduce NK cell activity by producing IL-10 [38]. In women with endometriosis, Th2 helper

cells from PF are reported to aberrantly suppress cell-mediated immunity by upregulating IL-4 and 10 secretions in PF from women with endometriosis [38,55]. As a result, decreased T cell cytotoxicity may allow implantation of endometrial cells in peritoneum.

#### **Impaired immune-surveillance and abnormal apoptosis**

The failure of immune cells to transmit death signals to endometrial cells, and/or the ability of endometrial fragments to avoid cell death may be associated with the development of endometriosis. Indeed, in women with endometriosis, it has been hypothesised that endometrial cells in the peritoneal fluid avoid immunosurveillance and implant into peritoneum [56]. It has been speculated that lymphocytes can adhere to endometrial cells through the Lymphocyte Function-Associated Antigen-1 (LFA-1) – Intercellular adhesion molecule-1 (ICAM-1) dependent pathway and present them as a target to NK cells. Soluble forms of ICAM-1 (s-ICAM-1) secreted by PF endometrial cells/endometriotic lesions can also bind to LFA-1 presenting lymphocytes and could prevent the recognition of endometrial cells by these lymphocytes and prevent subsequent NK cell-mediated cytotoxicity [57,58]. Furthermore, IL-6 secreted by endometriotic cells in concert with interferon- $\gamma$  may upregulate sICAM-1 production by macrophages of patients with endometriosis [59]. As a result, increased secretion of sICAM-1 may allow endometrial fragments to evade immunosurveillance, survive and implant.

Another major pathway in programmed cell death, Fas-Fas Ligand (FasL) system, could also be abnormal in women with endometriosis [60,61]. It has been speculated that the expression of FasL by viable endometrial cells induces apoptosis of T cells through ligation of Fas, allowing endometrial fragments to escape cell death, implant and develop to endometriotic lesions [62]. Interestingly, Garcia-Velasco *et al* [62] showed that macrophage-conditioned media might stimulate Fas-Fas ligand (FasL) expression by endometrial cells.

#### **Relevance in clinical research**

##### *Biomarkers to predict endometriosis non-surgically*

Presently, the diagnosis of endometriosis can be made only by laparoscopy and biopsy of suspicious lesions with subsequent histological confirmation of endometrial tissue and there is no non-invasive way to diagnose this condition. Laparoscopy is minimally invasive procedure, but requires general anaesthesia and surgical skills with potential complications and procedural costs. Hence, a non-surgical diagnostic tool would be of paramount benefit to both physicians and patients.

Efforts to evaluate the diagnostic value of endometrial markers for endometriosis have been hampered by the

lack of easy, reliable and quantitative techniques to assess the expression levels of these markers in sample material. Emerging proteomic techniques offer new approaches to identifying biomarkers for the early detection and follow-up of endometriosis.

Aromatase P450 mRNA has been identified as a candidate diagnostic marker but low sensitivity and specificity impair its application in clinical practice (Reviewed by Brosens *et al* [63]). In a recent study [64], the measurement of serum IL-6 levels and PF TNF- $\alpha$  levels could discriminate between patients with endometriosis and those without the disease. Endometriosis could be diagnosed if TNF-alpha levels in PF were higher than 15 pg/ml (100% sensitivity and 89% specificity) and if IL-6 levels in serum were above 2 pg/ml (90% sensitivity and 67% specificity) [64]. Potentially, the quantitation of autoantibodies against endometrial cells could also provide a novel method for the non-invasive diagnosis for endometriosis [65]. However, more studies are needed to confirm that this approach may be clinically useful. The role of Transvaginal Ultrasonography (TVU) and Magnetic resonance Imaging (MRI) in diagnosis and follow up of small endometriotic lesions is limited at present, but represents an interesting area of research [63].

##### *New treatment options and future management of endometriosis*

Endometriosis causes pelvic pain and infertility and can be treated by surgery and by hormonal suppression (progestins, continuous use of oral contraceptives, danazol, GnRH agonists), as reviewed recently [66]. A drug like danazol also works as an immunosuppressive agent (reviewed by D'Hooghe and Hill [66]). Indeed, the immunologic effects of danazol have been studied in women with endometriosis and adenomyosis and include a decrease in serum immunoglobulins [67,68], a decrease in serum C3, a rise in serum C4 levels [68], decreased serum levels of autoantibodies against various phospholipid antigens [67,68], and decreased serum levels of CA125 during treatment [69-72]. Danazol inhibits peripheral blood lymphocyte proliferation in cultures activated by T-cell mitogens but does not affect macrophage-dependent T-lymphocyte activation of B lymphocytes [73]. Danazol inhibits interleukin-1 (IL1) and TNF production by monocytes in a dose-dependent manner [74] and suppresses macrophage/monocyte-mediated cytotoxicity of susceptible target cells in women with mild endometriosis [75]. These immunological findings may be important in the remission of endometriosis with danazol treatment and may offer an explanation of the effect of danazol in the treatment of a number of autoimmune diseases, including hereditary angioedema [76], autoimmune hemolytic anemia [77], systemic lupus erythematosus, and idiopathic thrombocytopenic purpura [78,79].

Overall, medical treatment of endometriosis is limited by cost, side effects and recurrence of endometriosis after the cessation of treatment. Therefore, there is need for new drugs that treat endometriosis-associated pain and infertility without inhibition of ovulation. Future potential targets in the treatment or management of endometriosis may be inflammatory cytokines, MMPs, adhesion and growth factors [80,38]. Pentoxifylline has been shown to reduce endometriotic implant growth without inducing hypoestrogenism in both humans and hamsters [38]. In a recent study, Hornung and colleagues [81] demonstrated that Thiazolidinedione (TZD) significantly reduced leukocyte infiltration in the mouse model with endometriosis. Inhibition of TNF-alpha activity has also been a new target in the prevention and treatment of endometriosis. Experimental endometriosis in rats was treated with recombinant human tumour necrosis factor-binding protein-1 (r-hTBP-1), a soluble form of tumour necrosis factor- $\alpha$  receptor type-1 [82]. It was demonstrated that r-hTBP-1 could reduce the size of endometriotic-like peritoneal lesions by 64% [82]. Similarly, a study carried out in baboons showed that r-hTBP-1 effectively inhibited the development of endometriosis and endometriosis-related adhesions [83]. The potential of etanercept, a soluble TNF receptor (TNFR) fusion protein, for the treatment of endometriosis is also being considered [84].

## Conclusion

Understanding the involvement of the immune system in the development of endometriosis may help to understand the pathogenesis and spontaneous evolution of this condition. At present, most evidence suggests that pelvic inflammation and other immunological changes are a consequence of endometriosis. The development of a non-invasive diagnostic tools based on cytokines and autoantibodies could be of great benefit in the clinical management of endometriosis. Therapeutic strategies to eliminate the inflammatory reaction associated with endometriosis could lead to new treatment options for endometriosis.

## References

- Farquhar CM: **Extracts from the "clinical evidence". Endometriosis.** *BMJ* 2000, **320**:1449-1452.
- D'Hooghe TM, Debrock S, Hill JA, Meuleman C: **Endometriosis and subfertility: is the relationship resolved?** *Semin Reprod Med* 2003, **21**:243-254.
- Guarnaccia M, Olive DL: **The structure and future of endometriosis research.** *Obstet Gynecol Clin North Am* 1997, **24**:455-465.
- Sampson JA: **Peritoneal endometriosis due to menstrual dissemination of endometrial tissue into the peritoneal cavity.** *Am J Obstet Gynecol* 1927, **14**:422-469.
- Bartosik D, Jacobs SL, Kelly LJ: **Endometrial tissue in peritoneal fluid.** *Fertil Steril* 1986, **46**:796-800.
- Blumenkrantz MJ, Gallagher N, Bashore RA, Tenckhoff H: **Retrograde menstruation in women undergoing chronic peritoneal dialysis.** *Obstet Gynecol* 1981, **142**:890-895.
- D'Hooghe TM, Bambra CS, Raeymaekers BM, Koninckx PR: **Increased incidence and recurrence of retrograde menstruation in baboons with spontaneous endometriosis.** *Hum Reprod* 1996, **11**:2022-2025.
- D'Hooghe TM, Debrock S: **Endometriosis, retrograde menstruation and peritoneal inflammation in women and baboons.** *Human Reproduction Update* 2002, **8**:84-88.
- Cramer DW, Missmer SA: **The epidemiology of endometriosis.** *Ann N Acad Sci* 2002, **955**:11-22.
- Pinsonneault O, Goldstein DP: **Obstructing malformations of the uterus and vagina.** *Fertil Steril* 1985, **44**:241-247.
- D'Hooghe TM, Bambra CS, Suleman MA, Dunselman GA, Evers HL, Koninckx PR: **Development of a model of retrograde menstruation in baboons (*Papio anubis*).** *Fertil Steril* 1994, **62**:635-638.
- D'Hooghe TM, Hill JA: **Immunobiology of endometriosis.** In: *Immunology of reproduction* Edited by: Bronston R, Anderson DJ. Cambridge, MA: Blackwell Scientific; 1996:322-356.
- Betjes MG, Tuk CV, Struijk DG, Krediet RT, Arisz L, Hart M, Beelen RH: **Interleukin-8 production by human peritoneal mesothelial cells in response to tumor necrosis factor-alpha, interleukin-1, and medium conditioned by macrophages cocultured with *Staphylococcus epidermidis*.** *J Infect Dis* 1993, **168**:1202-1210.
- Akoum A, Lemay A, Paradis I, Rheault N, Maheux R: **Secretion of interleukin-6 by human endometriotic cells and regulation by proinflammatory cytokines and sex steroids.** *Hum Reprod* 1996, **11**:2269-2275.
- D'Hooghe TM, Debrock S, Meuleman C, Hill JA, Mwenda JM: **Future directions in endometriosis research.** *Obstet Gynecol Clin North Am* 2003, **30**:221-244.
- Wieser F, Fabjani G, Tempfer C, Schneeberger C, Zeillinger R, Huber JC, Wenzl R: **Tumor necrosis factor-alpha promotor polymorphisms and endometriosis.** *J Soc Gynecol Invest* 2002, **9**:313-318.
- Pizzo A, Salmeri FM, Ardita FV, Sofo V, Tripepi M, Marsico S: **Behaviour of cytokine levels in serum and peritoneal fluid of women with endometriosis.** *Gynecol Obstet Invest* 2002, **54**:82-87.
- Altman GB, Gown AM, Luchtel DL, Baker C: **RANTES production by cultured primate endometrial epithelial cells.** *Am J Reprod Immunol* 1999, **42**:168-174.
- Hornung D, Fujii E, Lim KH, Vigne JL, McMaster MT, Taylor RN: **Histocompatibility leukocyte antigen-G is not expressed by endometriosis or endometrial tissue.** *Fertil Steril* 2001, **75**:814-817.
- Bullimore DW: **Endometriosis is sustained by tumour necrosis factor-alpha.** *Med Hypotheses* 2003, **60**:84-88.
- Rana N, Braun DP, House R, Gebel H, Rotman C, Dmowski WP: **Basal and stimulated secretion of cytokines by peritoneal macrophages in women with endometriosis.** *Fertil Steril* 1996, **65**:925-930.
- Harada T, Enatsu A, Mitsunari M, Nagano Y, Ito M, Tsudo T, Tanaguchi F, Iwabe T, Takinawa M, Terakawa N: **Role of cytokines in progression of endometriosis.** *Gynecol Obstet Invest* 1999, **Suppl 47**:34-39.
- D'Hooghe TM, Bambra CS, Raeymaekers BM, Hill JA: **Pelvic inflammation induced by diagnostic laparoscopy in baboons.** *Fertil Steril* 1999, **72**:1134-1141.
- D'Hooghe TM, Bambra CS, Ling X, Peixe K, Hill JA: **The effect of menstruation and intrapelvic injection of endometrium on peritoneal fluid parameters in the baboon.** *Am J Obstet Gynecol* 2001, **184**:917-925.
- D'Hooghe TM, Hill JA, Oosterlynck DJ, Koninckx PR, Bambra CS: **Effect on endometriosis and the menstrual cycle on white blood cell subpopulations in the peripheral blood and peritoneal fluid of baboons.** *Hum Reprod* 1996, **11**:1736-1740.
- D'Hooghe TM: **Clinical relevance of the baboon as a model for the study of endometriosis.** *Fertil Steril* 1997, **68**:613-625.
- D'Hooghe TM, Bambra CS, Raeymaekers BM, De Jonge I, Hill JA, Koninckx PR: **The effects of immunosuppression on development and progression of endometriosis in baboons (*Papio anubis*).** *Fertil Steril* 1995, **6**:172-178.
- Steinleitner A, Lambert H, Suarez M, Serpa N, Robin B, Canter B: **Periovarian calcium channel blockade enhances reproductive performance in an animal model for endometriosis-associated subfertility.** *Am J Obstet Gynecol* 1991, **164**:949-952.
- Steinleitner A, Lambert H, Roy S: **Immunomodulation with pentoxifylline abrogates macrophage-mediated infertility in an vivo model: a paradigm for a novel approach to the treat-**

- ment of endometriosis-associated infertility. *Fertil Steril* 1991, **55**:26-31.
30. Gaetje R, Kotzian S, Herrmann G, Baumann R, Starzinski-Powitz A: **Nonmalignant epithelial cells, potentially invasive in human endometriosis, lack the tumor suppressor molecule E-cadherin.** *Am J Pathol* 1997, **150**:461-467.
  31. Berkkanoglu M, Arici A: **Immunology and endometriosis.** *Am J Reprod Immunol* 2003, **50**:48-59.
  32. Iwabe T, Harada T, Tsudo T, Nagano Y, Yoshida S, Tanikawa M, Terakawa N: **Tumor necrosis factor-alpha promotes proliferation of endometriotic stromal cells by inducing interleukin-8 gene and protein expression.** *J Clin Endocrinol Metab* 2000, **85**:824-829.
  33. Witz CA, Montoya-Rodriguez IA, Schenken RS: **Whole explants of peritoneum and endometrium: a novel model of the early endometriosis lesion.** *Fertil Steril* 1999, **71**:56-60.
  34. Debrock S, Vander Perre S, Meuleman C, Moerman P, Hill JA, D'Hooghe TM: **In-vitro adhesion of endometrium to autologous peritoneal membranes: effect of the cycle phase and the stage of endometriosis.** *Hum Reprod* 2002, **17**:2523-2528.
  35. Witz CA, Thomas MR, Montoya-Rodriguez IA, Nair AS, Centonze VE, Schenken RS: **Short-term culture of peritoneum explants confirms attachment of endometrium to intact peritoneal mesothelium.** *Fertil Steril* 2001, **75**:385-390.
  36. Nothnick WB, D'Hooghe TM: **Medical Management of Endometriosis: Novel Targets and Approaches towards the Development of Future Treatment Regimes.** *Gynecol Obstet Invest* 2003, **55**:189-198.
  37. Yeaman GR, Collins JE, Lang GA: **Autoantibody responses to carbohydrate epitopes in endometriosis.** *Ann N Y Acad Sci* 2002, **955**:174-182.
  38. Harada T, Iwabe T, Tarakawa N: **Role of cytokines in endometriosis.** *Fertil Steril* 2001, **76**:1-10.
  39. Nothnick WB: **Treating endometriosis as an autoimmune disease.** *Fertil Steril* 2001, **7**:223-231.
  40. McLaren J, Prentice A, Charnock-Jones DS, Smith SK: **Vascular endothelial growth factor (VEGF) concentrations are elevated in peritoneal fluid of women with endometriosis.** *Hum Reprod* 1996, **11**:220-223.
  41. McLaren J: **Vascular endothelial growth factor and endometriotic angiogenesis.** *Hum Reprod Update* 2000, **6**:45-55.
  42. Kats R, Collette T, Metz CN, Akoum A: **Marked elevation of macrophage migration inhibitory factor in the peritoneal fluid of women with endometriosis.** *Fertil Steril* 2002, **78**:69-76.
  43. Bruner KL, Eisenberg E, Gorstein F, Osteen KG: **Progesterone and transforming growth factor-beta coordinately regulate suppression of endometrial matrix metalloproteinases in a model of experimental endometriosis.** *Steroids* 1999, **64**:648-653.
  44. Sillem M, Prifti S, Koch A, Neher M, Jauckus J, Runnebaum B: **Regulation of matrix metalloproteinases and their inhibitors in uterine endometrial cells of patients with and without endometriosis.** *Eur J Obstet Gynecol Reprod Biol* 2001, **95**:167-174.
  45. Gottschalk C, Malberg K, Arndt M, Schmitt J, Roessner A, Schultze D, Kleinstein J, Ansgore S: **Matrix metalloproteinases and TACE play a role in the pathogenesis of endometriosis.** *Adv Exp Med Biol* 2000, **477**:483-486.
  46. D'Hooghe TM, Hill JA: **Is endometriosis an autoimmune disease?** In: *Immunology of reproduction* Edited by: Kurpisz M, Fernandez N. Oxford University Press; 1995:133-162.
  47. Chishima F, Hayakawa S, Hirata Y, Nagai N, Kanaeda T, Tsubata K, Satoh K: **Peritoneal and peripheral B-1-cell populations in patients with endometriosis.** *J Obstet Gynaecol Res* 2000, **26**:141-149.
  48. Startseva NV: **Clinical immunological aspects of genital endometriosis.** *Akush Ginekolog (Mosk)* 1980, **3**:23-26.
  49. Wilson TJ, Hertzog PJ, Angus D, Munnery L, Wood EC, Kola I: **Decreased natural killer cell activity in endometriosis patients: relationship to disease pathogenesis.** *Fertil Steril* 1994, **62**:1086-1088.
  50. D'Hooghe TM, Scheerlinck JP, Koninckx PR, Hill JA, Bamba CS: **Anti-endometrial lymphocytotoxicity and natural killer cell activity in baboons (*Papio anubis* and *Papio cynocephalus*) with endometriosis.** *Hum Reprod* 1995, **10**:558-562.
  51. Matarese G, De Placido G, Nikas Y, Alviggi C: **Pathogenesis of endometriosis: natural immunity dysfunction or autoimmune disease?** *Trends Mol Med* 2003, **9**:223-228.
  52. Wilson MT, Van Kaer L: **Natural killer T cells as targets for therapeutic intervention in autoimmune diseases.** *Curr Pharm Des* 2003, **9**:201-220.
  53. Dmowski WP, Rana N, Michalowska J, Friberg J, Papierniak C, el-Roeiy A: **The effect of endometriosis, its stage and activity, and of autoantibodies on in vitro fertilization and embryo transfer success rates.** *Fertil Steril* 1995, **63**:555-562.
  54. Vercammen EE, D'Hooghe TM: **Endometriosis and recurrent pregnancy loss.** *Semin Reprod Med* 2000, **18**:363-368.
  55. Hsu CC, Yang BC, Wu MH, Huang KE: **Enhanced interleukin-4 expression in patients with endometriosis.** *Fertil Steril* 1997, **67**:1059-1064.
  56. Somigliana E, Vigano P, Gaffuri B, Guarneri D, Busacca M, Vignali M: **Human endometrial stromal cells as a source of soluble intercellular adhesion molecule (ICAM)-1 molecules.** *Hum Reprod* 1996, **11**:1190-1194.
  57. Vigano P, Gaffuri B, Somigliana E, Busacca M, Di Blasio AM, Vignali M: **Expression of intercellular adhesion molecule (ICAM)-1 mRNA and protein is enhanced in endometriosis versus endometrial stromal cells in culture.** *Mol Hum Reprod* 1998, **4**:1150-1156.
  58. Becker JC, Termeer C, Schmidt RE, Brocker EB: **Soluble intercellular adhesion molecule-1 inhibits MHC-restricted specific T cell/tumor interaction.** *J Immunol* 1993, **151**:7224-7232.
  59. Fukaya T, Sugawara J, Yoshida H, Murakami T, Yajima A: **Intercellular adhesion molecule-1 and hepatocyte growth factor in human endometriosis: original investigation and a review of literature.** *Gynecol Obstet Invest* 1999, **47**:11-16.
  60. Lebovic DI, Mueller M, Taylor R: **Immunobiology of endometriosis.** *Fertil Steril* 2001, **75**:1-10.
  61. Garcia-Velasco JA, Arici A: **Apoptosis and the pathogenesis of endometriosis.** *Semin Reprod Med* 2003, **21**:165-172.
  62. Garcia-Velasco JA, Arici A, Zreik T, Noftolin F, Mor G: **Macrophage derived growth factors modulate Fas ligand expression in cultured endometrial stromal cells: a role in endometriosis.** *Mol Hum Reprod* 1999, **5**:642-650.
  63. Brosens J, Timmerman D, Starzinski-Powitz A, Brosens I: **Noninvasive diagnosis of endometriosis: the role of imaging and markers.** *Obstet Gynecol Clin North Am* 2003, **30**:95-114.
  64. Bedaiwy MA, Falcone T, Sharma RK, Goldberg JM, Attaran M, Nelson DR, Agarwal A: **Prediction of endometriosis with serum and peritoneal fluid markers: a prospective controlled trial.** *Hum Reprod* 2002, **17**:426-431.
  65. Inagaki J, Sugiura-Ogasawara M, Nomizu M, Nakatsuka M, Ikuta K, Suzuki N, Kaihara K, Kobayashi K, Yasuda T, Shoenfeld Y, Aoki K, Matsuura E: **An association of IgG anti-laminin-1 autoantibodies with endometriosis in infertile patients.** *Hum Reprod* 2003, **18**:544-549.
  66. D'Hooghe TM, Hill JA: **Chapter 25. Endometriosis.** In: *Novak's Gynecology* 13th edition. Edited by: Williams and Wilkins, JS Berek. Philadelphia, USA; 2002:931-972.
  67. El-Roeiy A, Dmowski WP, Gleicher N, Radwanska E, Harlow L, Binor Z et al.: **Danazol but not gonadotropin-releasing hormone agonists suppresses autoantibodies in endometriosis.** *Fertil Steril* 1988, **50**:864-871.
  68. Ota H, Maki M, Shidara Y, Kodama H, Takahashi H, Hayakawa M et al.: **Effects of danazol at the immunologic level in patients with adenomyosis, with special reference to autoantibodies: a multi-center cooperative study.** *Am J Obstet Gynecol* 1992, **167**:481-486.
  69. Takahashi K, Yoshino K, Kusakari M, Katoh S, Shibukawa T, Kitao M: **Prognostic potential of serum CA125 levels in danazol-treated patients with external endometriosis: a preliminary study.** *Int J Fertil* 1990, **35**:226-229.
  70. Franssen AMHW, van der Heijden PFM, Thomas CMG, Doesburg VH, Willemsen WNP, Rolland R: **On the origin and significance of serum CA125 concentrations in 97 patients with endometriosis before, during, and after buserelin acetate, nafarelin, or danazol.** *Fertil Steril* 1992, **57**:974-979.
  71. Bischof P, Galfetti MA, Seydoux J, von Hospenenthal JU, Campana A: **Peripheral CA125 levels in patients with uterine fibroids.** *Hum Reprod* 1992, **7**:35-38.



72. Acien P, Shaw RW, Irvine L, Burford G, Gardner R: **CA125 levels in endometriosis patients before, during and after treatment with danazol or LHRH agonists.** *Eur J Obstet Gynecol* 1989, **32**:241-246.
73. Hill JA, Barbieri RL, Anderson DJ: **Immunosuppressive effects of danazol in vitro.** *Fertil Steril* 1987, **48**:414-418.
74. Mori H, Nakagawa M, Itoh N, Wada K, Tamaya T: **Danazol suppresses the production of interleukin-1 $\beta$  and tumor necrosis factor by human monocytes.** *Am J Reprod Immunol* 1990, **24**:45-50.
75. Braun DP, Gebel H, Rotman C, Rana N, Dmowski WP: **The development of cytotoxicity in peritoneal macrophages from women with endometriosis.** *Fertil Steril* 1992, **1203**:1203-1210.
76. Gelfand JA, Sherins RJ, Alling DW, Frank MM: **Treatment of hereditary angioedema with danazol.** *N Engl J Med* 1976, **295**:1444-1448.
77. Ahn YS, Harrington WJ, Mylvaganam R, Ayub J, Pall LM: **Danazol therapy for autoimmune hemolytic anemia.** *Ann Intern Med* 1985, **102**:298-301.
78. Agnello V, Pariser K, Gell J, Gelfand J, Turksoy RN: **Preliminary observations on danazol therapy of systemic lupus erythematosus: effect on DNA antibodies, thrombocytopenia and complement.** *J Rheumatol* 1983, **10**:682-687.
79. Mylvaganam R, Ahn YS, Harrington WJ, Kim CI: **Immune modulation by danazol in autoimmune thrombocytopenia.** *Clin Immunol Immunopathol* 1987, **42**:281-287.
80. D'Hooghe TM: **Immunomodulators and aromatase inhibitors: are they the next generation of treatment for endometriosis?** *Curr Opin Obstet Gynecol* 2003, **15**:243-249.
81. Hornung D, Chao VA, Vigne JL, Wallwiener D, Taylor RN: **Thiazolidinedione inhibition of peritoneal inflammation.** *Gynecol Obstet Invest* 2003, **55**:20-24.
82. D'Antonio M, Martelli F, Peano S, Papoian R, Borrelli F: **Ability of recombinant human TNF binding protein-1 (r-hTBP-1) to inhibit the development of experimentally-induced endometriosis in rats.** *J Reprod Immunol* 2000, **48**:81-98.
83. D'Hooghe TM, Nugent N, Cuneo S, Chai D, Deer F, Debrock S, Mwenda J: **Recombinant human TNF binding protein (r-hTBP-1) inhibits the development of endometriosis in baboons: a prospective, randomised, placebo- and drug controlled study. Presented at the Annual Meeting of the American Society for Reproductive Medicine, Orlando, USA, October 22<sup>nd</sup>-24<sup>th</sup> 2001 [abstract].** *Fertil Steril* 2001, **76**:O-1-S-1.
84. Pugsley MK: **Etanercept. Immunex.** *Curr Opin Investig Drugs* 2001, **2**:1725-1731.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

