

Insulin: A pacesetter for the shape of modern biomedical science and the Nobel Prize



Jeffrey S. Flier^{a,*}, C. Ronald Kahn^b

ABSTRACT

Background: The 100th anniversary of the discovery of insulin in Toronto in 1921 is an important moment in medical and scientific history. The demonstration that an extract of dog pancreas reproducibly lowered blood glucose, initially in diabetic dogs and then in humans with type 1 diabetes, was a medical breakthrough that changed the course of what was until then a largely fatal disease. The discovery of the “activity”, soon named “insulin”, was widely celebrated, garnering a Nobel Prize for Banting and McLeod in 1923. Over the ensuing 100 years, research on insulin has advanced on many fronts, producing insights that have transformed our understanding of diabetes and our approach to its treatment.

Scope of Review: This paper will review research on insulin that had another consequence of far broader scientific significance, by serving as a pacesetter and catalyst to bioscience research across many fields. Some of this was directly insulin-related and was also recognized by the Nobel Prize. Equally important, however, was research stimulated by the discovery of insulin that has profoundly influenced biomedical research, sometimes also recognized by the Nobel Prize and sometimes without this recognition.

Major Conclusions: By reviewing some of the most notable examples of both insulin-related and insulin-stimulated research, it becomes apparent that insulin had an enormous and frequently under-appreciated impact on the course of modern bioscience.

© 2021 The Authors. Published by Elsevier GmbH. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Keywords Insulin; Discovery of insulin; Measurement of insulin; Sequence of insulin; Cloning of insulin; Insulin receptor

1. THE DISCOVERY OF INSULIN

The story of course begins with the discovery of insulin in Toronto in 1921. Since this epochal event has been so well researched and documented [for example, the excellent book “The Discovery of Insulin” by Michael Bliss [1]], our discussion of that discovery is limited. In fact, the “discovery” of insulin as a transformative therapeutic activity provided no information on the identity of “insulin” at a molecular level. Indeed, McLeod, who shared the Nobel Prize for insulin’s discovery, did not think it was a protein. Over the ensuing decades, however, multiple researchers sought to determine the precise identity of insulin, concluded that it was indeed a protein, identified insulin’s amino acid sequence, synthesized insulin using conventional protein synthesis, showed that this “man-made” version was actually biologically active, elucidated its crystal structure; developed a technique to measure its levels in biologic specimens, cloned its cDNA, and eventually — employing recombinant DNA technology — made insulin the first recombinant therapeutic protein. These, among many other advances, place insulin research at the center of modern bioscientific discovery.

Understanding the work leading to the discovery of insulin as an activity capable of reversing the metabolic dysfunction of diabetes involves consideration of two distinct but related periods. The first was between 1889 and 1920, involving the work of many international investigators; the second involved the activities of the Toronto group between 1920 and 1922.

In 1889, in Germany, Oskar Minkowski and Joseph von Mering at Strasbourg University reported that removal of the pancreas in experimental animals caused severe diabetes mellitus, allowing them to hypothesize that the pancreas contained a substance required for control of blood glucose. This powerful idea gained support from independent observations that diabetes was sometimes associated with pancreatic damage. In addition, the concept of internal endocrine secretions was beginning to develop, with the discovery of the first hormone, secretin, by Bayliss and Starling at University College London in 1902 [2]. As a result, the hypothesis that the pancreas produced an internal secretion that controlled carbohydrate metabolism became increasingly discussed.

Although the scientific community was vastly smaller and less connected than it is today, a number of investigators, starting with Oskar Minkowski, attempted to show that pancreatic extracts could reverse diabetes, mainly in animals, but some in humans. While there were many suggestions of activity in experiments carried out by Georg Zuelzer in Germany, Ernest Scott and Israel Kleiner in the US, and Nicolae Paulescu in Romania, these were accompanied by toxic reactions from contaminants in the crude extracts being tested, and none allowed a claim that the long-sought pancreatic activity had been found. In addition, the promising work of Paulescu was disrupted by World War I, and Kleiner, who was quite close to making the discovery, abandoned the work when he lost his position at Rockefeller University in 1919 and moved to what was to become New York Medical College, where he was unable to continue his research [3].

^aDepartment of Neurobiology and Medicine, Harvard Medical School, USA ^bJoslin Diabetes Center, Department of Medicine, Harvard Medical School, USA

*Corresponding author. Harvard Medical School, 220 Longwood Ave, Goldenson 542, Boston, MA 02115, USA. E-mail: Jeffrey_flier@hms.harvard.edu (J.S. Flier).

Received December 2, 2020 • Revision received February 12, 2021 • Accepted February 13, 2021 • Available online 18 February 2021

<https://doi.org/10.1016/j.molmet.2021.101194>

The discovery of insulin in Toronto in 1921 had an extremely unlikely provenance. It was not initiated by a researcher interested in or knowledgeable about metabolism or diabetes, but by Frederick Banting, who in 1920 was a 22-year-old orthopedic surgeon trying to establish a practice in London, Ontario, after returning from the war. The slow start to his practice led him to take on a part-time teaching role at London's Western University, where he was assigned to talk with students about carbohydrate metabolism. To learn about this subject, in which he had no expertise, he read the lead article in the November 1920 issue of the journal *Surgery, Gynecology and Obstetrics*, entitled "The Relation of the Islets of Langerhans to Diabetes with Special Reference to Cases of Pancreatic Lithiasis". Reading this article led Banting to develop an experimental hypothesis that by ligating the pancreatic ducts of dogs (analogous to the obstruction caused by a ductal stone) and then waiting for the acinar tissue to degenerate, he might then be able to extract the pancreatic factor capable of reversing diabetes and glycosuria. He documented these thoughts in a notebook he eventually left to the University of Toronto. Even more remarkable, the young surgeon decided to attempt to experimentally prove his hypothesis. He returned to his alma mater, the University of Toronto, met with the professor of physiology, J.J. Macleod, and tried to convince Macleod to provide him with facilities and supplies to attempt the work. Despite understandable skepticism about the likelihood of success, Macleod recognized Banting's ability to conduct the surgical component of the work and his obvious passion for the effort. In the summer of 1921, Macleod agreed to provide the facilities, several dogs, and a medical student assistant, Charles Best, to pursue this study (Figure 1).



Figure 1: Banting and Best and the experimental dog Marjorie on roof of medical building in Toronto, 1921.

Working with Best, under the supervision of Macleod, the work began in May 1921. Banting and Best performed duct ligations in one group of dogs and de-pancreatectomized others to create diabetes. By the end of July, with Macleod on vacation, they began to make saline extracts of chilled atrophied pancreas for injection into diabetic dogs. These produced variable effects, but on August 3, 1921, Banting and Best's crude extracts from the pancreas of a pancreatectomized dog named Marjorie showed reduced hyperglycemia. After reviewing their results, Macleod encouraged Banting and Best to add a visiting biochemist, J.B. Collip, to the team in an attempt to improve the approach to preparing the pancreatic extract. During this period, Banting also obtained beef pancreata from the slaughterhouse, to see if he could bypass the need to perform duct ligations in dogs, and when it became evident that this approach might also work, their progress accelerated.

On January 11, 1922, less than six months after the experiments in dogs, Leonard Thompson, a 14-year-old diabetic boy, became the first human to receive insulin when he was injected by doctors at Toronto General Hospital with 15 ml of a beef pancreatic extract prepared by Banting and Best. Unfortunately, there was only a minimal effect on blood and urine glucose, no effect on clinical status, and a sterile abscess resulted. However, on January 23, using an improved extract prepared by Collip, Thomson had an immediate response, with blood glucose becoming normal, ketones disappearing, and obvious clinical improvement. This was the first proof that a pancreatic extract could indeed reverse, at least briefly, the diabetic state. Success in other patients followed. The existence of an internal pancreatic secretion that regulated carbohydrate metabolism seemed secure [4] (Figure 2).

On May 3, 1922, armed with additional results, Macleod presented a summary of the Toronto research at a meeting of the Association of American Physicians in Washington, DC. The audience included many experts in the field, and he received a standing ovation for what was seen as one of the major achievements of medical history. Sadly, this achievement was accompanied by ongoing personal rivalries within the team. Banting and Best saw Collip and Macleod as wanting to take over the work and gain credit for it, refusing to recognize their important contributions. How this played out among the four individuals is well described in the historical work by Bliss [1].

A year and a half after this presentation, the 1923 Nobel Prize in Medicine or Physiology was awarded to Banting and Macleod. This enraged Banting, who had to be convinced to accept the Prize for the glory of Canada. He decided to share his half of the Prize with Best, prompting Macleod to share his with Collip, who Bliss thought should have been named a co-recipient by the Nobel Committee. Despite the Nobel going to Banting and Macleod, most people asked today who discovered insulin will say Banting and Best. Remarkably, none of the four conducted important insulin research again.

The University of Toronto, to which patents were assigned by Banting, Best, and Collip for \$1, made rapid commercial arrangements for insulin production by extraction of animal pancreas, first with Eli Lilly, then with Connaught Labs in Canada and the Nordisk Insulinlaboratorium, the Danish company founded by August and Marie Krogh, which eventually became Novo-Nordisk. As 1923 drew to a close, commercially produced beef and pork insulin was being used safely in most Western countries, a truly remarkable achievement.

2. THE MOLECULAR IDENTITY OF INSULIN

The discovery of insulin in Toronto provided no insights into the identity of the molecule (or molecules) responsible for producing the activity they called insulin. The field of endocrinology was very young, and no



Age 3
December 15, 1922
Weight 15 pounds

2 months later
February 15, 1923
Weight 29 pounds

Figure 2: Patient J.L., age 3, before and after insulin in 1923.

hormone had yet had its molecular identity revealed. In the 1920s and 1930s, several studies employing a variety of approaches supported the idea that insulin was most likely a protein. These included the crystallization of insulin by Abel in 1926, and reports that its biologic activity was lost after exposure to proteases. The industrial mass-production of insulin as a therapeutic, even if only partially purified by modern standards, provided scientists with substantial quantities of the hormone with which to help solve this problem.

Enter Fred Sanger. His interest in insulin and its commercial availability convinced Sanger to launch a major effort to determine whether insulin was a protein and what its amino acid sequence might be. By 1945, Sanger had developed a method for identifying and quantitatively measuring the terminal amino acids in insulin, making it possible to estimate the number and length of peptide chains in the protein. Using this, Sanger showed that bovine insulin contained two end-group amino acids, establishing that insulin was composed of two chains (A and B chains) linked together by cystine rather than 18 chains as had been hypothesized by others. Ultimately, sequences of these chains were published in 1951 and 1952, respectively [5,6].

To reach this conclusion, Sanger had to develop multiple new techniques. To determine insulin's amino acid composition, he employed a partition chromatography method initially used to study the amino acid composition of wool. The key to his novel approach was to develop a chemical reagent, 1-fluoro-2,4-dinitrobenzene (DNFB), now called Sanger's reagent, to label and then identify the N-terminal amino acids at one end of polypeptide chains. He hydrolyzed insulin into many short peptides using hydrochloric acid or trypsin, then fractionated these in two dimensions on filter paper. The fragments were identified by ninhydrin, revealing what he termed peptide "fingerprints". After hydrolysis, the N-termini were identified as the dinitrophenyl-amino

acids. By repeating these procedures multiple times with different hydrolysates, Sanger could deduce the complete sequence of the insulin molecule. This involved sequencing both the individual A and B chains, and then eventually identifying the two interchain and one intra-chain disulfide bonds. In this way, insulin was the first protein to have its amino acid sequence and chain composition identified, a truly profound achievement, considering that prior to this work, most assumed that proteins were somewhat amorphous in structure (Figure 3).

For this profound insight, in 1958, Sanger was awarded the first of his two Nobel Prizes in Chemistry. In 1980, he was recognized with his second for discovery of the "dideoxy" chain termination method for sequencing DNA, a prize he shared with Walter Gilbert and Paul Berg. The dideoxy method was eventually employed to sequence the human genome.

3. SYNTHESIS OF INSULIN

Although Sanger was awarded the Nobel Prize for developing the general technique for sequencing a protein, applied first to insulin, this work left many questions unanswered. How could we know whether Sanger had arrived at the right amino acid sequence, and would the molecule identified by Sanger actually evoke the biologic actions of insulin? There was then no independent standard against which the proposed amino acid sequence for insulin could be checked (cloning of the gene would not occur until many years later — see below). Furthermore, it was possible that some further modification of the primary structure might be required to render the hormone biologically active.

For these reasons, several groups in the emerging field of synthetic protein chemistry began work in the 1950s to synthesize insulin from

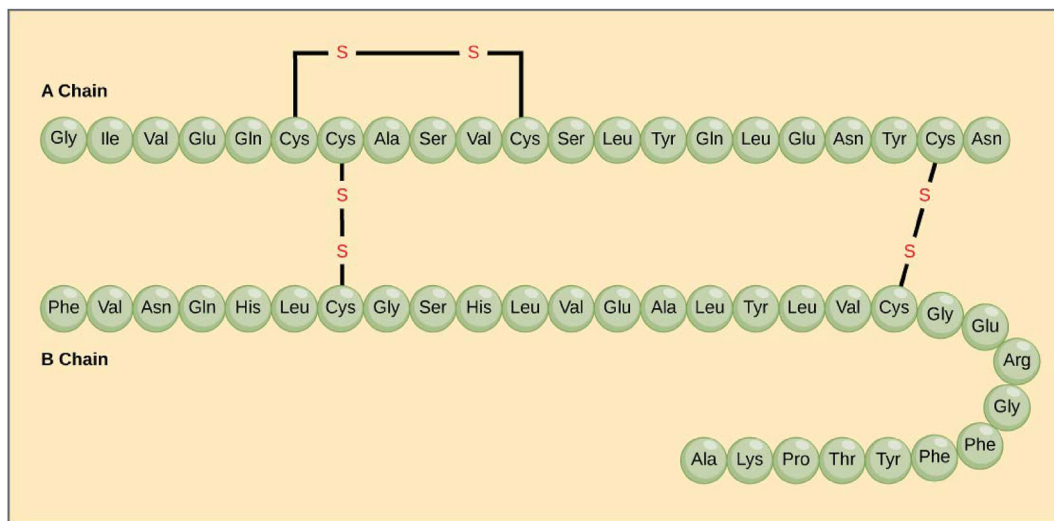


Figure 3: The amino acid sequence of insulin as determined by Sanger.

its component amino acids. Their goal was to test the fully synthetic insulin for biological activity, thereby confirming that the sequence “proposed” by Sanger was indeed correct. If their synthetic molecule was inactive in insulin bioassays, this could have several explanations: their synthesis might be in error, the sequence proposed by Sanger might be in error, or some unknown further modifications might be required for acquisition of biological activity. However, if the “Sanger insulin molecule” they synthesized was fully active, they would have confirmed that the proposed sequence was correct and demonstrated the capacity of a fully synthetic protein to act as insulin.

Three scientific teams pursued this work: one led by PG Katsyannis in the US, one led by Helmut Zahn in Germany, and a third group in China. At that time, synthetic techniques had already produced several small peptide hormones, specifically the posterior pituitary hormones oxytocin and vasopressin. This work was viewed as important, and Vincent Du Vigneaud, an American biochemist, was awarded the Nobel Prize for Chemistry in 1955 for that pathbreaking research. However, a protein as large and complex as insulin, with two chains and a total of 51 amino acids, was an enormous challenge.

The chemistry required to build the insulin A and B chains from individual amino acids was laborious, requiring multiple syntheses each with steps to protect amino and carboxyl groups then unblocking these protections, and finally condensing fragments to build the full chains. As this work proceeded, chemical techniques also needed to be developed to separate and then recombine A and B chains derived from animal insulin with sufficient yield and proper disulfide bond formation to create a bioactive molecule so that these techniques could be used to recombine the synthetic insulin chains. The demonstration that synthetic insulin built according to the Sanger model had full insulin biologic activity allowed the conclusion that the structure proposed by Sanger was indeed correct. The groups of Katsyannis and Zahn reported their successes almost simultaneously in 1963/4, and the Chinese group published its findings the following year [7,8].

These results were a major advance in the study of insulin and were simultaneously an advance for the broader field of protein biology. Katsyannis listed three reasons he considered the work to be important. First, as already stated, it confirmed that the Nobel Prize-winning insulin sequence published by Sanger was correct. Second, he speculated that, as diabetes increased in prevalence, it might be

necessary to supplement animal-derived insulins with synthetic human insulin as a therapeutic. This did not come to pass through protein chemistry, largely because recombinant human insulin eventually became the preferred option, as discussed below. Third, he correctly surmised that synthesis of insulin would enable creation of modified insulin analogs of known amino acid composition to examine the precise structural determinants of insulin action. The work Katsyannis, Zahn and their colleagues produced many such insulin analogs, providing new insights into insulin action and opening the door to future design of insulin analogs with improved characteristics, therapeutics that now dominate the insulin market. While some observers thought the synthesis of biologically active insulin might garner Katsyannis and Zahn another insulin-related Nobel Prize, that was not to be.

4. THE THREE-DIMENSIONAL STRUCTURE OF THE INSULIN MOLECULE

While Sanger was working to deduce the amino acid sequence of insulin, Dorothy Hodgkin was working to advance the field of X-ray crystallography as a means to determine the three-dimensional structure of biological molecules. Her initial success revealed the structures of penicillin and vitamin B12. The latter, published in 1955, brought her the Nobel Prize in Chemistry in 1964, but her work also added an important chapter in the insulin story, which began even before her award-winning work.

Hodgkin graduated from Oxford with a degree in chemistry in 1932 and was awarded a PhD from the University of Cambridge for pioneering studies of X-ray crystallography. Her PhD work was conducted with J.D. Bernal, a pioneer in the use of X-ray crystallography in molecular biology, including determining the x-ray structure of pepsin, the first biological substance successfully studied in this way. Her interest in insulin began very early, in 1934, when she was given a crystal of insulin. She used this to make an image of the insulin crystal with techniques available at the time, which were inadequate to assess a protein of insulin’s size and complexity. However, she remained fascinated by the biology of insulin, and after a career of pioneering technical advances and winning the Nobel Prize, she eventually returned to solving the structure of insulin. Working with a team at

Oxford University that included Thomas Blundell and Guy Dodson, they solved the structure of insulin and published the results in 1969 [9,10]. In the 1950s and 1960s, Dorothy Hodgkin also helped train Chinese scientists, who returned to China and almost simultaneously determined the structure of insulin. By 1971 and 1972, they obtained crystal structures of porcine insulin at resolutions of 2.5 and 1.8 Å, respectively, demonstrating that in this area of science, China had reached a first-class international level.

Thus, insulin was the first important protein to have its structure solved. Scientists wanted to work on insulin because it was a molecule known to have powerful biological effects, and their hope was that understanding its structure would facilitate deeper understanding of its mechanism of action.

5. THE MEASUREMENT OF INSULIN IN BLOOD

Endocrinology is the study of hormones, signaling molecules produced in a regulated fashion in endocrine cells in one tissue, then released into the blood, where they act by binding to receptors on or in one or more target tissues, often at a distance. A major step in understanding the biology of any hormone and its role in disease is, therefore, to measure its level in the blood and determine what makes its levels go up or down. In the case of insulin, understanding its levels in blood in response to glucose and in diseases, such as diabetes, was critical in understanding its physiology and pathophysiology. It was not until 1959, 38 years after its discovery, that insulin could be unambiguously detected and quantified in human serum. This was made possible by a technique first applied to insulin (though broadly applicable) called radioimmunoassay (RIA). RIA was invented by Berson and Yalow. Yalow won the Nobel Prize for this work in 1977, after Berson's death in 1972.

The first attempts to measuring insulin in plasma utilized a wide variety of bioassays. In these, extracts of plasma were administered to animals in which insulin could lower glucose levels or were added to *in vitro* preparations of fat or muscle in which insulin stimulated glucose uptake or metabolism. Such assays had some utility, but they were technically very challenging and provided widely variable results. Furthermore, their interpretation and specificity were limited both by the poor sensitivity of these approaches and the uncertainty as to whether actions were caused by insulin itself or by other molecules in the samples with insulin-like activity. Indeed, when insulin was added to plasma, it was often difficult to recover its full activity, suggesting that blood might also contain circulating antagonists which could modify insulin action, thus confounding interpretations.

Enter Berson and Yalow, a physician and a physicist, respectively, who worked together starting in 1950 to use radioisotopes to probe biological systems at the Bronx Veterans Hospital. Their research on insulin did not initially arise from a desire to measure the hormone, but a desire to measure its degradation. They became aware of research by I. Arthur Mirsky, who had hypothesized that maturity-onset diabetes was the consequence of insulin being rapidly degraded by a hepatic insulin-degrading enzyme or insulinase. They decided to use their facility with radioisotopes to test this hypothesis in quite a remarkable way. They iodinated (beef) insulin with ^{131}I and administered the tracer insulin to healthy humans and patients treated with insulin, following the course of radioactivity in blood after injection. If Mirsky was correct, they surmised that labeled insulin would disappear more rapidly. Instead, they observed that labeled insulin disappeared more slowly in insulin-treated subjects. Although there were many possible explanations, they hypothesized that these insulin-treated subjects had developed antibodies to administered insulin that delayed its clearance.

To further explore this hypothesis, they assessed the interaction of labeled insulin with plasma protein using chromatoelectrophoresis and other techniques. This demonstrated that labeled insulin bound to a globulin fraction in these patients. Further, they found that the binding was of high affinity and could be blocked by addition of unlabeled insulin. With this observation, they made the critical conceptual leap by realizing that the system could be used to detect and quantify the amount of insulin in a sample by comparing inhibition caused by the unknown sample to a standard curve using known insulin concentrations.

Initial efforts to measure insulin in human plasma using antibodies were unsuccessful, due to relatively low affinity of human insulin as a competitor, since the antibodies in human blood had been evoked to beef and pork insulin used as therapy, which differ from human insulin by three and one amino acid, respectively. However, they persevered and discovered that antibodies elicited by immunizing guinea pigs with beef insulin did cross-react with human insulin with sufficient affinity to permit measurement of human insulin in plasma. In a definitive paper in the *Journal of Clinical Investigation* in 1960, they reported that insulin measurements in human plasma were sensitive and reproducible, that insulin levels rose after glucose ingestion, and rose to higher levels in subjects with maturity onset diabetes who had never been treated with insulin [11] (Figure 4). They also reported increased insulin levels in patients with hypoglycemia due to islet cell tumors and patients with leucine-sensitive hypoglycemia.

Although it took several years from their initial publication, by the mid-1960s, the RIA approach was widely applied to the study of insulin physiology and pathophysiology, ushering in an era of many *in vitro* and *in vivo* investigations of insulin secretion and insulin action, none of which would have been possible without the ability to measure insulin with precision and sensitivity. The concept of RIA was not limited to insulin, of course. The technique was rapidly applied to the measurement of innumerable molecules of biologic interest, hormones and many others, essentially any molecule to which high affinity antibodies could be developed, transforming many fields of study. The principles of the RIA would also be eventually be applied to quantitating interactions between a labeled ligand (hormone, drug, etc.) and its receptor on cells or other bimolecular interactions of interest. Eventually, RIAs employing radioactively-labeled insulin were superseded by enzyme-linked immunosorbent assays (ELISAs) and related techniques, in which an analyte is quantitated by its binding to an antibody linked to an enzyme, which could be measured by its activity, thus eliminating the need for radioactivity. However, the place of RIA in the history of modern biomedical research is assured.

An interesting and initially undesired "side-effect" of RIA was to give no credence to the insulin-like bioactivity in blood which RIA did not measure and which could not be suppressed in bioassays by the addition of insulin antibodies. This non-suppressible insulin-like activity (NSILA), also called "atypical insulin", was initially discounted as being unimportant by Berson, Yalow, and many in the field, since it was not measured in the RIA for insulin. Thus, it was more than a decade later before this was recognized as being the activity of the insulin-like growth factors IGF-1 and IGF-2, which were also present in serum at even higher concentrations than insulin and circulated as high molecular weight complexes since they were bound to their many IGF binding proteins [12]. The purification, sequencing, and ultimate cloning of these insulin-like growth factors (IGFs) demonstrated that insulin was part of a larger family of hormones, the insulin-IGF family, with the IGFs having up to 50% sequence homology with insulin, but with their own receptors, very low affinities for insulin antibodies, and a very different balance between effects on metabolism, where insulin is

IMMUNOASSAY OF ENDOGENOUS PLASMA INSULIN IN MAN

BY ROSALYN S. YALOW AND SOLOMON A. BERSON

(From the Radioisotope Service, Veterans Administration Hospital, New York, N. Y.)

(Submitted for publication March 7, 1960; accepted March 22, 1960)

For years investigators have sought an assay for insulin which would combine virtually absolute specificity with a high degree of sensitivity, sufficiently exquisite for measurement of the minute insulin concentrations usually present in the circulation. Methods in use recently depend on the ability of insulin to exert an effect on the metabolism of glucose *in vivo* or in excised muscle or adipose tissue. Thus, the insulin concentration in plasma has been estimated: *a*) from the degree of hypoglycemia produced in hypophysectomized, adrenalectomized, alloxan-diabetic rats (1); *b*) from the augmentation of glucose uptake by isolated rat hemidiaphragm (2); or *c*) from the increased oxidation of glucose-1-C¹⁴ by the rat epididymal fat pad (3). Since there have been reports indicating the presence, in plasma, of inhibitors of insulin action (4) and of non-insulin substances capable of inducing an insulin-like effect (5, 6), these procedures, while yielding interesting information regarding the effects of various plasmas on glucose metabolism in tissues, are of doubtful specificity for the measurement of insulin *per se* (5).

Recently it has been shown (7, 8) that insulins from various species (pork, beef, horse and sheep) show quantitative differences in reaction and cross reaction with antisera obtained from human subjects treated with commercial insulin preparations (beef, pork insulin mixtures). An immunoassay method for beef insulin has been reported in which the insulin content is determined from the degree of competitive inhibition which the insulin offers to the binding of beef insulin-I¹²⁵ by human antisera (9-12). Although human insulin reacts with human antibeef, pork insulin antiserum and displaces beef insulin-I¹²⁵ by competitive inhibition (7, 8, 10), the reaction is too weak to permit measurement of the low insulin concentrations present in human plasma (7, 8, 11-13). In preliminary communications we have reported that the competitive inhibition by human insulin of binding of crystalline beef insulin-I¹²⁵ to guinea

pig antibeef insulin antibodies is sufficiently marked to permit measurement of plasma insulin in man (11, 12, 14), and to be capable of detecting as little as a fraction of a microunit of human insulin (12, 14). Preliminary data on insulin concentrations in man before and after glucose loading have been reported (12, 14, 15). The present communication describes in detail the methods employed in the immunoassay of endogenous insulin in the plasma of man, and reports plasma insulin concentrations during glucose tolerance tests in nondiabetic and in early diabetic subjects and plasma insulin concentrations in subjects with functioning islet cell tumors or leucine-sensitive hypoglycemia.

METHODS

Immunization of guinea pigs. Guinea pigs were injected subcutaneously at 1 to 4 week intervals with 5 to 10 units of either protamine zinc beef insulin (Squibb) or commercial regular beef insulin (Squibb) emulsified with mannide mono-oleate. Insulin-binding antibodies were detected in all animals after 2 to 3 injections. The antiserum employed in the present study (GP 49, serum 6-25-59) was obtained from a guinea pig immunized with protamine zinc beef insulin without adjuvant and was selected for its relatively high antibody concentration and other suitable characteristics described below.

Preparation of insulin-I¹²⁵. Because of the desirability of keeping the concentration of added insulin-I¹²⁵ as low as possible and yet assuring an adequate counting rate, it is necessary to prepare the insulin-I¹²⁵ with a high specific activity. The lots of insulin-I¹²⁵ employed in this study had specific activities of 75 to 300 mc per mg at the time of use. The preparation of such highly labeled preparations entails difficulties not encountered when the specific activity is very much lower. The Newerly modification (16) of the Pressman-Eisen method (17) was used for labeling with several further modifications designed to increase specific activity and to minimize damage to the insulin from irradiation and other causes. To approximately 0.3 ml chloroform in a 50 ml separatory funnel are added in turn, 0.2 ml of 2.5 N HCl, 20 μ l of 10⁻³ M KI, 30 to 80 mc I¹²⁵ (as iodide) and 1 drop of 1 M NaNO₂. Immediately after addition of the last reagent, the funnel is stoppered to prevent loss of I¹²⁵ into the atmosphere and is shaken vigorously for 2 to 3 minutes. The chloroform layer (bottom) is then drawn

1157

Figure 4: The measurement of insulin by radioimmunoassay, as reported by Berson and Yalow in the Journal of Clinical Investigation in 1960.

dominant, and effects on cell growth, where the IGF's are dominant. Indeed, in higher organisms, IGF-1 is the primary mediator of the action of growth hormone [13].

6. PROHORMONE PROCESSING — ANOTHER MODEL PARADIGM WITH INSULIN AS A LEADER

Application of the RIA to plasma also revealed that circulating insulin itself occurred in at least two forms, which differed in size and were simply

identified as "big" and "little" insulin, with the latter of the appropriate molecular weight to match the hormone [14]. In patients with islet cell tumors, the fraction of big insulin was increased. Using cell labeling and protein chemistry techniques, in 1967, Donald Steiner discovered that this heterogeneity was due to the fact that human insulin was produced as a single-chain precursor with the A and B chains connected by a 33- to 35-amino acid C-peptide. He termed this precursor proinsulin [15] and showed that proinsulin was cleaved in the beta-cell to release the two-chain insulin molecule and equimolar amounts of C-peptide (Figure 5).

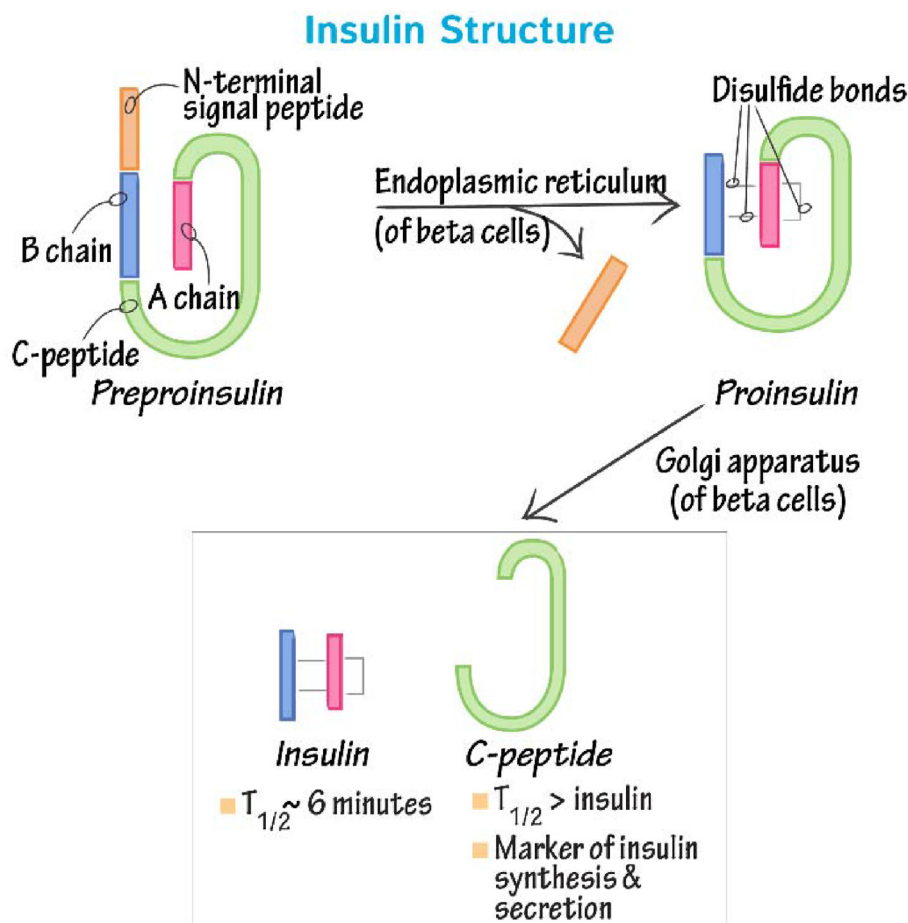


Figure 5: The processing of preproinsulin to proinsulin, and then to insulin and C-peptide, as determined by Steiner.

This insight solved the mystery of how the A and B chains of insulin could relatively easily become associated to form the insulin molecule and served as the first unambiguous demonstration of the post-translational processing of a polypeptide precursor into the mature functional form of the hormone by specialized proteolytic enzymes, a paradigm which is now well appreciated for many hormones.

Reduced efficiency of proinsulin processing in islet cell tumors, which led to the increase in “big” insulin in plasma, was the precursor for modern measurements of proinsulin in serum of patients with islet cell tumors, and the release of free C-peptide from the β -cell now serves as an independent marker for insulin secretion, especially in patients in whom insulin levels may be difficult to measure due to endogenous insulin antibodies. Steiner and his colleagues later discovered an even larger precursor of proinsulin, which they termed preproinsulin, i.e., proinsulin including its signal peptide targeting it for secretion [16]. The discovery of proinsulin and preproinsulin established the field of protein-precursor processing, paving the way to understanding how many other peptide hormones and neuropeptides are made and processed and was often discussed as another insulin-related discovery worthy of consideration for a Nobel Prize.

7. CLONING AND EXPRESSION OF THE INSULIN GENE, AND INSULIN AS THE FIRST RECOMBINANT HUMAN THERAPEUTIC

During the decades that insulin biology advanced through the application of techniques of protein chemistry, protein synthesis, X-ray

crystallography, and assay development, another biologic revolution was taking place that would also leave insulin’s mark on modern bioscience. That revolution was the understanding of DNA biochemistry, function, and techniques to manipulate it and created an ability to synthesize insulin and other proteins in a simpler way, thereby unlocking enormous therapeutic possibilities. In 1962, the Nobel Prize for Physiology or Medicine was awarded to Francis Crick, James Watson, and Maurice Wilkins “for their discoveries concerning the molecular structure of nucleic acids and its significance for information transfer in living material”. Over the ensuing years, many scientists focused their attention on understanding the function and regulation of DNA and RNA, and by the early 1970s, recombinant DNA technology emerged as a powerful tool to combine, amplify and express DNA from different species.

In 1973/4, Stanley Cohen of Stanford and Herbert Boyer of UCSF collaborated to develop a process for joining and replicating DNA from different species, and a patent based on this work submitted by Stanford and UCSF was granted by the US Patent Office in 1980 [17]. Many scientists saw the implications of recombinant DNA for development of recombinant therapeutic proteins. It soon became evident that insulin was the first and most important molecule on which to focus, since by that time the Eli Lilly Company was using pancreata from 56 million pigs and cattle each year just to supply the growing US insulin market, and even with only 1 and 3 amino acid differences, respectively, some patients developed significant antibodies to insulin, reducing its potency in vivo.

At least three major groups set out to win the scientific and commercial race to make recombinant human insulin in a commercially viable manner, seeking to replace the animal-derived insulins then available. The dominant groups in the race were at UCSF (led by Bill Rutter and Howard Goodman), Harvard (led by Wally Gilbert), and the newly formed company, Genentech, which had licensed the earlier Cohen and Boyer patents from Stanford and UCSF and which bet its future on their ability to win the race. In 1977, the UCSF and Harvard groups published results showing their distinct abilities to clone the rat insulin cDNAs, the former using mRNA of pancreatic islets, the latter using mRNA from a rat insulinoma [18,19]. The Harvard effort was further complicated by the fact that Cambridge, Massachusetts where Harvard is located had passed a city ordinance banning work on recombinant DNA, thus requiring Gilbert and his team to go to England's Porton Down facility to try and isolate human insulin. Rather than cDNA cloning, Genentech took the alternative approach of creating the human insulin-encoding sequence with synthetic DNA using technology developed by Riggs and Itakura at City of Hope Medical Center. Their

approach was to separately clone and express the human insulin A and B chains and then recombine them to make intact insulin, as done earlier by insulin chemists like Katsoyannis and Zahn [20]. The Genentech group was the first to reach a commercially viable level of human insulin production, enabling them to sign an agreement with Lilly to perform clinical trials of recombinant human insulin, which proved to be highly effective and less immunogenic. Humulin was approved by the FDA in 1982 and marketed as the first recombinant human therapeutic in 1983, a landmark event.

The 1980 Nobel Prize in Chemistry was split, with half of the Prize awarded to Paul Berg for his contributions to recombinant DNA technology and the other half shared by Wally Gilbert and Fred Sanger for DNA sequencing. Although this Prize was not directly for cloning the insulin gene, it was certainly linked to insulin, since during the several years before his Nobel Prize, Gilbert's primary focus was on the cloning and expression of the insulin gene, and ultimately, insulin was the first human therapeutic derived using recombinant DNA technology.

Proc. Nat. Acad. Sci. USA
Vol. 68, No. 8, pp. 1833-1837, August 1971

Insulin Receptors in the Liver: Specific Binding of [¹²⁵I]Insulin to the Plasma Membrane and Its Relation to Insulin Bioactivity

(insulins and derivatives)

PIERRE FREYCHET*, JESSE ROTH, AND DAVID M. NEVILLE, JR.†

Section on Diabetes and Intermediary Metabolism, Clinical Endocrinology Branch, National Institute of Arthritis and Metabolic Diseases; and †Section on Physical Chemistry, Laboratory of Neurochemistry, National Institute of Mental Health, National Institutes of Health, Bethesda, Maryland 20014

Communicated by Robert W. Berliner, May 14, 1971

ABSTRACT With [¹²⁵I]insulin at 7×10^{-10} M, 25% of the radioactivity was bound to plasma membranes purified from rat liver. 20% of the [¹²⁵I]insulin binding was inhibited by unlabeled insulin at 10^{-7} M (6 ng/ml), equivalent to insulin concentrations in hepatic portal blood; inhibition of [¹²⁵I]insulin binding was 80% at 10^{-7} M and 90% at 10^{-6} M. Eight insulins and derivatives with biological potencies that differed over a 100-fold range inhibited the binding of [¹²⁵I]insulin to liver membranes in direct proportion to their ability to stimulate glucose oxidation in isolated fat cells. Inactive insulin chains, as well as glucagon, ACTH, and human growth hormone were without effect. The binding of [¹²⁵I]insulin increased 55-fold as plasma membrane was purified from crude homogenate. Binding was time- and temperature-dependent, and addition of excess insulin produced rapid dissociation of [¹²⁵I]insulin. This study demonstrates directly the binding of insulin to its biologically important receptors.

The first step in the action of polypeptide hormones appears to be rapid, reversible interaction between hormone in the extracellular medium and specific recognition sites on the plasma membrane of target cells (1-3). In previous studies (3), using [¹²⁵I]ACTH, we have demonstrated directly the binding of ACTH to its biologically important receptors. Others have studied binding of [¹²⁵I]angiotensin to several tissues (4) and of radioiodinated glucagon to the plasma membrane fraction from rat liver (5, 6).

The binding of radioactively labeled insulin to tissues has been measured in many studies, including one reported recently (7) in which binding to the plasma membrane fraction from rat liver was studied, but uncertainties about the bioactivity of the iodinated molecules, the site of the binding, and proportion of "nonspecific" binding have limited the scope of the conclusions (8-15). We developed a method for the preparation of moniodoinsulin that is as active as unlabeled insulin in stimulating glucose oxidation in isolated fat cells (16, 17). [¹²⁵I]insulin so prepared was bound to isolated fat cells and to purified liver membranes (16, 17). We describe here the biological specificity and other characteristics of [¹²⁵I]insulin binding to its receptors in the liver.

MATERIALS AND METHODS

Insulins and other hormones

Human (258-1025B-88), bovine (795372), and porcine (PJ 5589) insulins, porcine proinsulin (615-984B-99-C), and

* Permanent address: Groupe U.55 (I.N.S.E.R.M.), Hotel-Dieu, 1 Place du Parvis-Notre Dame, Paris 4, France.

beef-pork glucagon were gifts of Eli Lilly. Bonito fish insulin was kindly supplied by Dr. R. S. Yalow (gift of Shimizu Seiyako Co., Ltd., Japan); guinea pig insulin (65-3-GPB) by Dr. L. F. Smith; desoctapeptide (JG-I-65-8) and desalanine-desasparagine (JG-II-21-10) bovine insulins by Dr. H. Carpenter. Carboxymethylated A and B chains of porcine insulin were purchased from Mann Research and porcine ACTH from Sigma. Human growth hormone (HGH) was kindly supplied by the National Pituitary Agency, Endocrinology Study Section, NIH.

Insulin assays

Biological activity of insulins and insulin derivatives was measured as the stimulation of glucose oxidation in isolated fat cells (18), with porcine insulin as standard. Except that 5-10 mg of fat cells was used per flask, conditions were as described elsewhere (19). The immunoreactivity of insulins and insulin derivatives was measured by radioimmunoassay, with the use of porcine [¹²⁵I]insulin and guinea pig anti-porcine insulin serum (gift of S. M. Glick and A. Kagan), and with porcine insulin as the standard (19, 20).

Binding of [¹²⁵I]insulin to liver membranes

[¹²⁵I]insulin (porcine) was prepared (16) at specific activities of 25-100 Ci/g for studies of binding to liver membranes. Plasma membranes from rat liver parenchymal cells (21) were kept at -70°C until use. Electron microscopy shows the purified plasma membranes to be composed mainly of paired membrane sheets (22) with intercellular junctions and bile canalicular structures (23), which identify them as originating from hepatic parenchymal cells; studies with marker enzymes have shown that contamination with microsomes, lysosomes, and mitochondria account for less than 10% of the total membrane protein (22). Protein concentrations were determined (24) with bovine albumin as the standard. Immediately after thawing, the purified (step 15, ref. 21) plasma membranes (160 μg of membrane protein) were diluted to 100 μl with 1 mM KHCO_3 and incubated with or without unlabeled insulin. After 20 min at 30°C , [¹²⁵I]insulin was added to incubation tubes in 50 μl of Krebs-Ringer phosphate (Ca^{2+} -free) buffer, pH 7.5, that contained 3% bovine albumin (Fraction V, Armour), to give a final [¹²⁵I]insulin concentration of 0.6-3.0 nM (see legends to Figures). After 40 more minutes of incubation, duplicate 60- μl aliquots were removed and layered onto 250 μl of Krebs-Ringer phosphate buffer, pH 7.5, that contained albumin at 10 mg/ml, in plastic microtubes as

1833

Figure 6: First report of specific high-affinity insulin receptor binding by Freychet, Roth, and Neville in 1971.

8. HOW INSULIN BRINGS ABOUT ACTIONS ON TARGET CELLS

In the 1960s, understanding of insulin's actions in target cells was extremely limited. The ability of insulin to increase glucose uptake and metabolism in fat and muscle was established, as was the ability of insulin to alter activity of several intracellular enzymes, but how this was accomplished was unknown and highly debated. Some scientists thought insulin, and other peptide hormones, entered cells and bound directly to enzymes it regulated (perhaps through formation of disulfide bonds), while others thought a membrane receptor binding step coupled to signal generation might be involved. The term "signal transduction", which is widely used today, was 20 years in the future. However, this concept was greatly aided by studies of the mechanism of action of the hormone epinephrine by the group of Earl Sutherland, Jr. who showed that this beta-adrenergic agonist generated an intracellular mediator or second messenger named cyclic AMP, for which Sutherland was awarded the 1971 Nobel Prize in Physiology or Medicine [21]. The first step by which epinephrine interacted with cells to generate cAMP, however, was unknown.

Enter Jesse Roth, a physician scientist and Solomon Berson's first research fellow, and Pedro Cuatrecasas, a Spanish born and U.S. educated MD, both of whom came to the National Institutes of Health (NIH) in the early 1960s. This was a golden era at NIH, with both of these investigators doing truly ground-breaking work. Cuatrecasas, who was pioneering affinity chromatography, showed that insulin bound to sepharose beads was biologically active, suggesting it must work through a receptor on the surface of the cell. Likewise, Roth and his collaborators, showed that thyroid-stimulating hormone (TSH) also likely worked through a surface receptor on thyroid cells, since its activity could be reversed by addition of anti-TSH antibodies. Both aimed to devise methods to identify plasma membrane receptors for peptide hormones, using I^{131} -labeled hormones. Roth's first effort involved ACTH, and a 1970 paper first authored by Robert Lefkowitz (who also went on to win a Nobel Prize for his work on adrenergic receptors) demonstrated high affinity and specific binding sites for labeled ACTH in adrenal membranes [22]. Roth soon turned his attention to insulin, and the 1971 paper from his lab characterizing high affinity and specific insulin binding to sites in rat liver membranes was the first demonstration of what soon became known as "insulin receptors" [23] (Figure 6). Cuatrecasas showed similar insulin receptor binding on adipocyte membranes, another major tissue of insulin action [24]. Over the next decade, both labs, but especially that of Jesse Roth, went on to further characterize insulin receptor binding kinetics, provide the evidence for physiologic regulation of their expression and their role in disorders of insulin resistance, and provide initial insights into their structure [25,26]. Similar methodologies were soon applied to studies of many other peptide hormones, thus creating a whole new field of membrane receptor biology. All of these insights occurred long before the receptor was purified, or its cDNA cloned [27]. One final link to Nobel history was that Stanley Cohen received the Nobel Prize for Physiology or Medicine in 1986 for his discovery of epidermal growth factor, which he showed to be the first receptor tyrosine kinase in 1982 [28], the year before Kahn and co-workers showed that the insulin receptor was a tyrosine kinase [29].

Additional biochemical studies of what is now known as "signal transduction" and the additional analysis that became possible following the cloning of the receptor cDNA's have transformed both our understanding of insulin actions and provided new insights into parallel signaling systems used by insulin and the IGFs and the truly massive

field of signal transduction from membrane receptors to numerous cellular pathways. Once again, insulin was at the heart of this burgeoning field of study.

This also opened the field to many other important aspects of insulin and its actions, many of which are covered in more detail in other articles in this issue. These include understanding the insulin molecule in evolution, defining the complimentary relationships between insulin and IGFs, understanding insulin and IGF-1 receptor signaling systems, including the role of insulin action in tissues not previously thought to be insulin sensitive, to the development of new insulin analogs to improve treatment of type 1 diabetes, and understanding the role of insulin resistance in type 2 diabetes and metabolic syndrome.

From the breadth of these topics, it should be clear that since its discovery 100 years ago, understanding the insulin molecule, how it works, and what goes wrong in diabetes have been important drivers in defining many paradigms underlying contemporary biology, biochemistry, physiology and medicine and many discoveries recognized by the Nobel Prize or worthy of that recognition.

CONFLICT OF INTEREST

None declared.

REFERENCES

- [1] Bliss, M., 1982. *The discovery of insulin*. Chicago: University of Chicago Press. p. 304, 16 p. of plates.
- [2] Bayliss, W.M., Starling, E.H., 1902. *The mechanism of pancreatic secretion*. *The Journal of Physiology* 28(5):325–353.
- [3] Friedman, J., 2015. *Discovery, interrupted*. In: Harpers.
- [4] Banting, F.G., Best, C.H., Collip, J.B., Campbell, W.R., Fletcher, A.A., 1922. *Pancreatic extracts in the treatment of diabetes mellitus*. *Canadian Medical Association Journal* 12(3):141–146.
- [5] Sanger, F., Tuppy, H., 1951. *The amino-acid sequence in the phenylalanyl chain of insulin. 2. The investigation of peptides from enzymic hydrolysates*. *Biochemical Journal* 49(4):481–490.
- [6] Sanger, F., Thompson, E.O., 1952. *The amino-acid sequence in the glycyl chain of insulin*. *Biochemical Journal* 52(1):iii.
- [7] Meienhofer, J., Schnabel, E., Bremer, H., Brinkhoff, O., Zabel, R., Sroka, W., et al., 1963. *[Synthesis of insulin chains and their combination to insulin-active preparations]*. *Zeitschrift für Naturforschung B* 18:1120–1121.
- [8] Katsoyannis, P.G., 1964. *The synthesis of the insulin chains and their combination to biologically active material*. *Diabetes* 13:339–348.
- [9] Adam, M.G., Collier, L., Hodgkin, D.C., Dodson, G.G., 1966. *X-ray crystallographic studies on zinc insulin crystals*. *The American Journal of Medicine* 40(5):667–671.
- [10] Blundell, T.L., Dodson, G.G., Dodson, E., Hodgkin, D.C., Vijayan, M., 1971. *X-ray analysis and the structure of insulin*. *Recent Progress in Hormone Research* 27:1–40.
- [11] Yalow, R.S., Berson, S.A., 1960. *Immunoassay of endogenous plasma insulin in man*. *Journal of Clinical Investigation* 39:1157–1175.
- [12] Rinderknecht, E., Humbel, R.E., 1978. *The amino acid sequence of human insulin-like growth factor I and its structural homology with proinsulin*. *Journal of Biological Chemistry* 253(8):2769–2776.
- [13] Roberts Jr., C.T., Leroith, D., 1988. *Molecular aspects of insulin-like growth factors, their binding proteins and receptors*. *Bailliere's Clinical Endocrinology and Metabolism* 2(4):1069–1085.

Review

- [14] Roth, J., Gorden, P., Pastan, I., 1968. "Big insulin": a new component of plasma insulin detected by immunoassay. *Proceedings of the National Academy of Sciences of the United States of America* 61(1):138–145.
- [15] Steiner, D.F., Cunningham, D., Spigelman, L., Aten, B., 1967. Insulin biosynthesis: evidence for a precursor. *Science* 157(3789):697–700.
- [16] Lomedico, P.T., Chan, S.J., Steiner, D.F., Saunders, G.F., 1977. Immunological and chemical characterization of bovine preproinsulin. *Journal of Biological Chemistry* 252(22):7971–7978.
- [17] Cohen, S.N., Chang, A.C., Boyer, H.W., Helling, R.B., 1973. Construction of biologically functional bacterial plasmids in vitro. *Proceedings of the National Academy of Sciences of the United States of America* 70(11):3240–3244.
- [18] Ullrich, A., Shine, J., Chirgwin, J., Pictet, R., Tischler, E., Rutter, W.J., et al., 1977. Rat insulin genes: construction of plasmids containing the coding sequences. *Science* 196(4296):1313–1319.
- [19] Villa-Komaroff, L., Broome, S., Naber, S.P., Efstratiadis, A., Lomedico, P., Tizard, R., et al., 1980. The synthesis of insulin in bacteria: a model for the production of medically useful proteins in prokaryotic cells. *Birth Defects: Original Article Series* 16(1):53–68.
- [20] Goeddel, D.V., Kleid, D.G., Bolivar, F., Heyneker, H.L., Yansura, D.G., Crea, R., et al., 1979. Expression in *Escherichia coli* of chemically synthesized genes for human insulin. *Proceedings of the National Academy of Sciences of the United States of America* 76(1):106–110.
- [21] Sutherland, E.W., Robison, G.A., 1966. The role of cyclic-3',5'-AMP in responses to catecholamines and other hormones. *Pharmacological Reviews* 18(1):145–161.
- [22] Lefkowitz, R.J., Roth, J., Pricer, W., Pastan, I., 1970. ACTH receptors in the adrenal: specific binding of ACTH-125I and its relation to adenyl cyclase. *Proceedings of the National Academy of Sciences of the United States of America* 65(3):745–752.
- [23] Freychet, P., Roth, J., Neville Jr., D.M., 1971. Insulin receptors in the liver: specific binding of (125I)insulin to the plasma membrane and its relation to insulin bioactivity. *Proceedings of the National Academy of Sciences of the United States of America* 68(8):1833–1837.
- [24] Jacobs, S., Cuatrecasas, P., 1981. Insulin receptor: structure and function. *Endocrine Reviews* 2(3):251–263.
- [25] Flier, J.S., Kahn, C.R., Roth, J., 1979. Receptors, antireceptor antibodies and mechanisms of insulin resistance. *New England Journal of Medicine* 300(8):413–419.
- [26] Kahn, C.R., Baird, K.L., Flier, J.S., Grunfeld, C., Harmon, J.T., Harrison, L.C., et al., 1981. Insulin receptors, receptor antibodies, and the mechanism of insulin action. *Recent Progress in Hormone Research* 37:477–538.
- [27] Ullrich, A., Bell, J.R., Chen, E.Y., Herrera, R., Petruzzelli, L.M., Dull, T.J., et al., 1985. Human insulin receptor and its relationship to the tyrosine kinase family of oncogenes. *Nature* 313(6005):756–761.
- [28] Cohen, S., Fava, R.A., Sawyer, S.T., 1982. Purification and characterization of epidermal growth factor receptor/protein kinase from normal mouse liver. *Proceedings of the National Academy of Sciences of the United States of America* 79(20):6237–6241.
- [29] Kasuga, M., Fujita-Yamaguchi, Y., Blithe, D.L., Kahn, C.R., 1983. Tyrosine-specific protein kinase activity is associated with the purified insulin receptor. *Proceedings of the National Academy of Sciences of the United States of America* 80(8):2137–2141.