Research Article **Tanshinone IIA Alleviates the AD Phenotypes in APP and PS1 Transgenic Mice**

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Therapeutic approach for Alzheimer's disease (AD) is still deficient. To find active compounds from herbal medicine is of interest in the alleviation of AD symptoms. This study aimed to investigate the protective effects of Tanshinone IIA (TIIA) on memory performance and synaptic plasticity in a transgenic AD model at the early phase. 25–100 mg/kg TIIA (intraperitoneal injection, *i.p.*) was administered to the six-month-old APP and PS1 transgenic mice for 30 consecutive days. After treatment, spatial memory, synaptic plasticity, and related mechanisms were investigated. Our result showed that memory impairment in AD mice was mitigated by 50 and 100 mg/kg TIIA treatments. Hippocampal long-term potentiation was impaired in AD model but rescued by 100 mg/kg TIIA treatment. Mechanically, TIIA treatment reduced the accumulations of beta-amyloid 1–42, C-terminal fragments (CTFs), and p-Tau in the AD model. TIIA did not affect basal BDNF but promoted depolarization-induced BDNF synthesis in the AD mice. Taken together, TIIA repairs hippocampal LTP and memory, likely, through facilitating the clearance of AD-related proteins and activating synaptic BDNF synthesis. TIIA might be a candidate drug for AD treatment.

1. Introduction

Alzheimer's disease (AD) is an aging-related disease hallmarked by the decline of memory performance and other cognitive abilities. It is considered as the most severe neurodegenerative disease, affecting an estimated 25 million people globally [1]. Effective treatment is still lacking in AD due to the complexity of the disease pathology [2–4]. Impairment of synaptic plasticity and pyramid cell loss are thought to render the disease progression. Recent available drugs for AD treatment are mainly working through their activities in promoting cerebral-vascular blood flow [5]. However, those drugs are either too expensive or unsatisfactory for symptoms alleviation.

At the early phase of AD, synaptic transmission and plasticity are affected without loss of pyramid neurons. Cerebral atrophy or cell death is responsible for the cognitive impairment at the later stage of AD [6]. Early prevention or treatment is relatively effective to interfere with AD. A network of signaling pathways regulate synaptic transmission or plasticity. Protein kinase A (PKA), extracellular signalregulated kinase (ERK), and brain derived neurotrophic factor- (BDNF-) Tropomyosin receptor kinase B (TrkB) signaling pathways are prominent [7–9]. According to previous publications, blockage or deletion of those important pathways leads to impairment of synaptic plasticity and memory [10, 11].

Chinese herbal medicines are popularized for their multitarget effects and low toxic effects [12, 13]. Among the documented herbal medicines, Salvia miltiorrhiza Bunge (Danshen) is beneficial for AD treatment [14-16]. Danshen belongs to the Labiatae family of the plant kingdom. Based on the theory of traditional Chinese medicine (TCM), Danshen has the function of activating blood circulation and removing blood stasis, targeting the "heart," "pericardium," and "liver" channels [17, 18]. Clinically, Danshen has been widely used to treat various circulatory disturbance-related diseases for its special pharmacological actions, including vasodilatation, anticoagulation, anti-inflammation, and free radical scavenging [19]. Tanshinone IIA (TIIA) is an active lipophilic component extracted from the root of Salvia miltiorrhiza Bunge and possesses pharmacological activities of anti-inflammatory, antioxidative, and cytotoxic activity as well as inducing apoptosis. TIIA displayed neuroprotective effects on β -amyloid-induced toxicity in rat cortical neurons [20]. Tanshinone, especially, influences the levels of nitric oxide synthase and acetylcholinesterase in the brain of AD rats [21]. However, the effects of TIIA on synaptic plasticity in AD model and related mechanisms are not reported. In this study, an AD transgenic mice model (APPswe, PSEN1dE9) was used to investigate the effects of TIIA on memory and its mechanisms.

2. Materials and Methods

2.1. Animals. APP/PS1 mice (B6C3-Tg) were purchased from Jackson Lab and bred in our own colony in Weifang Medical University. The offspring was genotyped by primers for APP and PS1. APP/PS1 mice have accelerated the AD phenotype characterized by increased $A\beta$ deposits and behavioral deficits as young as 13–16 weeks [22]. The 6-month-old male mice were used in the experiments and housed in a 12 h light/dark cycle with food and water *ad libitum*. Age-and sex-matched C57BL/6J mice (Animal Center of Chinese Academy of Sciences, China) were used as Controls. All the experimental procedures were approved by Weifang Medical University.

2.2. Tanshinone IIA Treatment. Tanshinone IIA (Santa Cruz, USA) by intraperitoneal injection (*i.p.*) (25 mg/kg, 50 mg/kg, and 100 mg/kg) was chronically administered to the mice once each day for 30 consecutive days. This dose range was chosen based on previous publications [23, 24]. TIIA (16 mg/kg) readily penetrated the blood brain barrier reaching a peak concentration of 0.41 nmol/g brain wet weight 60 minutes after intraperitoneal injection [25]. TIIA was dissolved in DMSO and diluted in saline. Control and AD mice received the similar volume of vehicle (DMSO diluted in saline). Diet, drink, and body weight were monitored during the drug administration. 30 days after administration, behavioral, electrophysiological, and biochemical experiments were carried out.

2.3. Electrophysiological Experiments. Mice were anesthetized by ether and decapitated. The brain was isolated quickly on ice. After that, acute hippocampal slices (400 μ m) were prepared in cutting solution (124 NaCl, 26 NaHCO₃, 10 Dglucose, 3 KCl, 1.25 KH₂PO₄, 5 MgSO₄, and 3.4 CaCl₂). The slices were then transferred to an interface recording chamber and exposed to a warm, humidified atmosphere of 95% $O_2/5\%$ CO₂ and continuously perfused with oxygenated and preheated (32±0.5°C) artificial cerebrospinal fluid (aCSF) (in mM) [110 NaCl, 5 KCl, 2.5 CaCl₂, 1.5 MgSO₄, 1.24 KH₂PO₄, 10 D-glucose, and 27.4 NaHCO₃] with a speed of 1.5 mL/min. The field EPSP was elicited by stimulating Schaffer collateral pathway with twisted nichrome wires. The input-output and paired-pulse facilitation at 30 ms, 50 ms, and 100 ms intervals were tested. Long-term potentiation was triggered by thetaburst stimulation (TBS, 10 bursts of four pulses at 100 Hz delivered at 5 Hz interval).

2.4. ELISA. Beta-amyloid 1–42 was quantified by ELISA method. The hippocampus from different groups were homogenized in homogenization buffer (5 M guanidine HCl/ 50 mM Tris–HCl) and centrifuged. Protein concentrations of supernatants were determined using a BCA kit (Thermo Fisher Scientific, USA). Supernatant fractions were analyzed by beta-amyloid 1–42 ELISA kit (KHB3441, Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocol. Absorbance was determined for each well at 450 nm using a microplate reader (Thermo Scientific, USA).

2.5. Real-Time PCR. Total RNA was extracted from hippocampus using Trizol reagent (Invitrogen, Carlsbad, USA). Reverse transcription was carried out using Moloney murine leukemia virus reverse transcriptase (Promega, Madison, USA). Real-time PCR was performed for the quantification of APP in hippocampus with a quantitative thermal cycler (Mastercycler ep realplex, Eppendorf, Germany). Relative expression values were calculated as the ratio of target cDNA to β -actin. The primers used in real-time PCR were listed as follows:

> APP: sense primer 5'-TGCTGGCAGAACCCC-AGATCG-3'; antisense primer 5'-TTCTGGATG-GTCACTGGCTGG-3';

> β -actin: sense primer 5-ATGAGGTAGTCTGTC-AGGT-3; antisense primer 5-ATGGATGACGAT-ATCGCT-3.

2.6. Biochemical Experiments. Acute hippocampal slices were incubated with normal aCSF. One hour after recovery, slices from different groups were depolarized by KCl (90 mM, 3 min). The concentration of NaCl in KCl-aCSF was reduced making the composition of KCl-aCSF as follows: 37.5 mM NaCl, 90 mM KCl, 1.25 mM NaH₂PO₄, 25 mM NaHCO₃, 2 mM CaCl₂, 1 mM MgCl₂, and 25 mM glucose. One hour after depolarization, the slices were collected in dry ice and kept at -80° C until use.

Hippocampus homogenates were obtained and lysed. Protein concentrations were measured using BCA protein assay kit (Thermo, US). Equivalent amounts of proteins were processed for SDS-PAGE and western blot. The primary antibodies used were BDNF (1:1000, Millipore), Actin (1:10000, Millipore), p-Tau (1:3000, Cell Signaling), and CTFs (1:3000, Cell Signaling).

2.7. Morris Water Maze. Morris water maze was conducted in a circular pool with a 150 cm diameter filled with a depth of 22 cm water ($25\pm2^{\circ}$ C). A circular Plexiglass platform (8 cm diameter) was placed 2 cm below the water level. Distinctive visual cues were set. A video camera was positioned above the water maze. The swim was tracked, digitized, and stored for later behavioral analysis using EthoVision 3.1 (Noldus). The water maze was divided into four logical quadrants that served as starting positions for the rats.

The spatial learning task consisted of a 5-day acquisition using the hidden platform. This was followed by a probe trial on the sixth day without the platform. The platform was fixed



FIGURE 1: Tanshinone IIA improved the spatial memory in AD mice. (a) Changes of escape latency to reach the hidden platform during the 5 d acquisition trails; (b) the time spent in the target quadrant 24 h after the last acquisition trial. The data were presented as Mean and SEM. In each group, there were 10 animals. High dose (100 mg/kg), medium dose (50 mg/kg), and low dose of Tanshinone IIA (25 mg/kg) were administered to the mice for 30 consecutive days, respectively. ** P < 0.01 compared with Control and $^{#}P < 0.05$ compared with AD.

in the middle of the west quadrant, 45 cm from the maze wall. During the first 5 days, four swim trials were given per day, in which each animal was released from a different quadrant in each trial. This was done in a pseudorandom manner and the start quadrant used was varied across the sessions. A maximum of 60 s was allowed for each trial. If the rat did not find the platform within 60 s, it was guided to the platform and allowed to remain there for 10 s. The latency to escape onto the platform was recorded.

2.8. Statistical Analyses. Data are presented as means \pm SEM. All the statistical analyses were performed by one-way ANOVA with GraphPad Prism 6.0. Bonferroni correction for post hoc *t*-test was performed to compare the differences between groups. *P* value less than 0.05 was considered statistically significant.

3. Results

3.1. TIIA Ameliorates Spatial Memory in AD Mice. The diet, drink, and weight were not affected during TIIA treatment. After a period of 30-day treatment, spatial memory was evaluated. Within the five-day training section, 100 mg/kg and 50 mg/kg TIIA remarkably decreased the latency to find the platform (Figure 1(a)). On the sixth day, the probe was removed. As shown in Figure 1(b), the time spent in the targeted quadrant was significantly decreased in AD mice when compared to Control mice (P < 0.05). By contrast, TIIA treatment significantly reversed the deficit (P < 0.05). These data suggested that TIIA mitigated the impaired spatial memory in the AD model. 3.2. TIIA Reverses the Deficit of Long-Term Potentiation in AD Mice. As evidenced by previous study, hippocampal LTP was also impaired at the early phase of the AD model [6]. To confirm the effect of TIIA on hippocampal synaptic transmission and plasticity, fEPSP at Schaffer collateral-CA1 synapses was measured. TBS-induced LTP was impaired in the slice obtained from the AD mice (P < 0.05 versus Control) (Figures 2(a) and 2(b)). However, after TIIA treatment, TBS-LTP was mitigated (P < 0.05 versus AD model). As shown in Figures 2(c) and 2(d), TIIA did not affect the basal synaptic transmission, including input-output and paired-pulse facilitation. These results suggest that TIIA could ameliorate the synaptic deficit at the early phase of AD.

3.3. TIIA Attenuates AD-Related Protein Expression. We detected APP expression in hippocampus by RT-PCR. Compared to wild type mice, APP expression in mRNA level was significantly elevated in AD mice (P < 0.05 versus Control). By contrast, TIIA did not alter the APP expression (Figure 3(a)). We also detected CTFs expression. In the model mice, CTFs expression was significantly increased (P < 0.05versus Control); however, it was decreased by TIIA treatment (P < 0.05 versus AD model) (Figure 3(c)). As compared with wild type mice, p-Tau expression increased significantly in model mice (P < 0.05 versus Control). By contrast, TIIA treatment also reduced this protein level (P < 0.05 versus AD model) (Figure 3(c)). In addition, TIIA treatment apparently reduced beta-amyloid 1–42 level in AD mice (P < 0.05versus Control) (Figure 4). These results suggested that TIIA treatment mitigated the accumulation of AD-related protein expression in AD mice.



FIGURE 2: Tanshinone IIA reversed the impairment of long-term potentiation in AD mice. (a) Impairment of TBS-induced LTP in AD model was improved by Tanshinone IIA administration. (b) LTP level at the 90th min after TBS. (c) Input-output was not affected by Tanshinone IIA application. (d) Paired-pulse facilitation was not affected by Tanshinone IIA application. The data were presented as Mean and SEM. 5–10 slices from five animals were included in each group. *P < 0.05 compared with WT and *P < 0.05 compared with AD.

3.4. TIIA Improves Synaptic Activation-Induced BDNF Synthesis in AD Mice. We also detected synaptic related protein expression. BDNF expression was not remarkably altered in basal level of this AD model. Moreover, TIIA treatment did not promote the protein expression. We analyzed the synaptic activation-induced BDNF synthesis. High concentration of KCl was applied to trigger BDNF synthesis [26]. As shown in Figure 5, BDNF level was significantly elevated after KCl incubation in Control mice. By contrast, KCl-induced BDNF synthesis was blocked in the AD mice. After TIIA treatment, KCl-induced BDNF synthesis was significantly improved (P < 0.05 versus AD model).

4. Discussion

We demonstrated in this study that TIIA treatment improved the memory and hippocampal LTP in AD model. ADrelated protein expressions were effectively reduced and synaptic activation-induced BDNF was improved after TIIA treatment.

Danshen has the pharmacological activities to improve memory in different disease models. Compound Danshen ameliorated cognitive deficit in A β 25–35 peptide-induced rat model of Alzheimer's disease [14, 27]. Salvia miltiorrhiza injection protects against memory impairments in



FIGURE 3: Tanshinone IIA downregulated CTFs and p-Tau expression. (a) APP expression in mRNA level was increased in AD mice. (b) CTFs expression was increased in AD mice, while it was decreased by Tanshinone IIA treatment. Representative blots of CTFs and Actin are inserted above. (c) p-Tau expression was increased in AD mice, while it was decreased by Tanshinone IIA treatment. Representative blots of p-Tau and total Tau are inserted above. The data were presented as Mean and SEM from five animals in each group. *P < 0.05, **P < 0.01 compared with Control and *P < 0.05 compared with AD.

streptozotocin-induced diabetic rats [28, 29]. In addition, the formula HX106N with Danshen as the main component was also reported to ameliorate memory deficit in AD model [30]. These consistent reports revealed the potential activity of Danshen in the treatment of AD. As the major active compound in Danshen, TIIA might be one of the active substances exerting the memory improving function. Lam et al. reported that TIIA was detected in blood within 5 minutes after intraperitoneal injection and also detected in the brain 5 minutes after injection, showing that the drug was able to penetrate the blood brain barrier. They also revealed that TIIA (16 mg/kg) readily penetrated the blood brain barrier reaching a peak concentration of 0.41 nmol/g brain wet weight 60 minutes after intraperitoneal injection [25]. In our study, we chose a dose range from 25 to 100 mg/kg and revealed that 50-100 mg/kg of TIIA could effectively improve the memory in AD mice.

The "amyloid hypothesis" indicates that amyloid clearance is an effective strategy for the treatment or prevention of AD. Inhibitors of secretase prohibit the overall A β production. However, preclinical study indicated that semagacestat, an inhibitor of secretase, did not show any significant slowing of AD phenotypes [31]. In APP/PS1 transgenic mice, APP was overexpressed, leading to increase of amyloid 1-42 accumulation in hippocampus. In addition, the AD protein p-Tau and CTFs were also enhanced at 6-month-old AD model. In TIIAtreated mice, APP expression was not affected. However, p-Tau, CTFs, and amyloid 1-42 accumulation in hippocampus were significantly reduced. In previous reports, estrogenmediated PI3K/Akt signaling promotes α -secretase cleavage of APP and inhibits the production of A β [32, 33]. As a new identified member of the phytoestrogen family [34, 35], TIIA possibly mediates PI3K/AKT signaling pathway to cleavage of secretase and contributes to the decrease of $A\beta$ production. Besides the direct effects on A β production, TIIA also likely affects the degradation of A β . The elevation of activity of ubiquitin proteasome system by parkin overexpression facilitates the A β -related protein clearance [6]. It is possible



FIGURE 4: Tanshinone IIA downregulated beta-amyloid 1–42 level in AD model. The data were presented as Mean and SEM from five animals in each group. **P < 0.01, *P < 0.05 compared with Control and *P < 0.05 compared with AD.



FIGURE 5: Tanshinone IIA activates KCl-induced BDNF synthesis. The data were presented as Mean and SEM from five animals in each group. *P < 0.05 compared with Control and $^{\#}P < 0.05$ compared with AD.

that TIIA treatment increased the proteasome activity to facilitate $A\beta$ clearance [36].

At the early phase of AD, impairment of synaptic plasticity rather than cell loss contributes to the memory deficit. Although it is difficult to ascertain the direct AD stage displayed by the transgenic AD mice (6-7 months), it is conceivable that the mice were still at the early phase of AD progression, as we did not observe cell loss or impairment of input-output of the basal synaptic transmission. By contrast, hippocampal LTP was impaired in the AD mice. TIIA treatment reversed the impairment LTP but did not affect basal synaptic transmission. Amyloid plague is supposed to be the detrimental toxin contributing to the impairment of hippocampal synaptic plasticity and hippocampal cell death [37–39].

A lot of signaling pathways are involved in the modulation of hippocampal synaptic plasticity [7]. BDNF-TrkB pathway is especially important for the formation of memory and LTP consolidation [10]. BDNF-TrkB was impaired to affect memory in many disease models [8, 40]. Moreover, not only the basal BDNF level but also synaptic activationinduced BDNF synthesis was reduced [41]. In our study, the basal BDNF level was not affected at the early phase of AD mice. However, KCl-induced BDNF synthesis was impaired in the AD model. After TIIA treatment, the abnormality was rescued. In combination with the effects of TIIA on AD-related protein expression, TIIA possibly interferes with the protein accumulation to reverse the synaptic activation-induced BDNF synthesis.

5. Conclusion

In this study, we provided data revealing the memory improving effect after TIIA treatment in a transgenic AD model. The clearance of AD-related protein and activation of BDNF-TrkB pathway might possibly contribute to the effect of TIIA on hippocampal LTP and memory. These results implicated that TIIA is a potential memory improver in AD model.

Competing Interests

The authors declare that they have no competing interests.

References

- C. Qiu, M. Kivipelto, and E. von Strauss, "Epidemiology of Alzheimer's disease: occurrence, determinants, and strategies toward intervention," *Dialogues in Clinical Neuroscience*, vol. 11, no. 2, pp. 111–128, 2009.
- [2] S. Salloway, "Current and future treatments for Alzheimer's disease," CNS Spectrums, vol. 14, no. 8, supplement 7, pp. 4–18, 2009.
- [3] K. G. Yiannopoulou and S. G. Papageorgiou, "Current and future treatments for Alzheimer's disease," *Therapeutic Advances in Neurological Disorders*, vol. 6, no. 1, pp. 19–33, 2013.
- [4] P. S. Aisen, J. Cummings, and L. S. Schneider, "Symptomatic and nonamyloid/tau based pharmacologic treatment for Alzheimer disease," *Cold Spring Harbor Perspectives in Medicine*, vol. 2, no. 3, Article ID a006395, 2012.
- [5] A. Corbett, G. Williams, and C. Ballard, "Drug repositioning in Alzheimer's disease," *Frontiers in Bioscience-Scholar*, vol. 7S, no. 1, pp. 184–188, 2015.
- [6] X. Hong, J. Liu, G. Zhu et al., "Parkin overexpression ameliorates hippocampal long-term potentiation and β-amyloid load in an Alzheimer's disease mouse model," *Human Molecular Genetics*, vol. 23, no. 4, Article ID ddt501, pp. 1056–1072, 2014.
- [7] M. Baudry, G. Zhu, Y. Liu, Y. Wang, V. Briz, and X. Bi, "Multiple cellular cascades participate in long-term potentiation and in hippocampus-dependent learning," *Brain Research*, vol. 1621, pp. 73–81, 2015.
- [8] G. Zhu, Y. Liu, Y. Wang, X. Bi, and M. Baudry, "Different patterns of electrical activity lead to long-term potentiation by activating different intracellular pathways," *The Journal of Neuroscience*, vol. 35, no. 2, pp. 621–633, 2015.
- [9] Z. Amtul and R. Atta Ur, "Neural plasticity and memory: molecular mechanism," *Reviews in the Neurosciences*, vol. 26, no. 3, pp. 253–268, 2015.
- [10] J. J. An, K. Gharami, G.-Y. Liao et al., "Distinct role of long 3/ UTR BDNF mRNA in spine morphology and synaptic plasticity in hippocampal neurons," *Cell*, vol. 134, no. 1, pp. 175–187, 2008.
- [11] H. Kang, A. A. Welcher, D. Shelton, and E. M. Schuman, "Neurotrophins and time: different roles for TrkB signaling in

hippocampal long-term potentiation," *Neuron*, vol. 19, no. 3, pp. 653–664, 1997.

- [12] I.-C. Hung, S.-S. Chang, P.-C. Chang, C.-C. Lee, and C. Y.-C. Chen, "Memory enhancement by traditional Chinese medicine?" *Journal of Biomolecular Structure and Dynamics*, vol. 31, no. 12, pp. 1411–1439, 2013.
- [13] B. H. May, C. Lu, Y. Lu, A. L. Zhang, and C. C. L. Xue, "Chinese herbs for memory disorders: a review and systematic analysis of classical herbal literature," *Journal of Acupuncture and Meridian Studies*, vol. 6, no. 1, pp. 2–11, 2013.
- [14] M. Liu, H. Guo, C. Li et al., "Cognitive improvement of compound danshen in an Aβ25–35 peptide-induced rat model of Alzheimer's disease," *BMC Complementary and Alternative Medicine*, vol. 15, article 382, 2015.
- [15] H. M. Hügel, "Brain food for alzheimer-free ageing: focus on herbal medicines," *Advances in Experimental Medicine and Biology*, vol. 863, pp. 95–116, 2015.
- [16] H. Hu, D. S. Zhou, and R. Yu, "Effects of compound danshen tablet on the expression of Abeta in transgenic cell model of Alzheimer's disease," *Zhongguo Zhong Xi Yi Jie He Za Zhi*, vol. 32, no. 12, pp. 1663–1666, 2012.
- [17] X.-Y. Yu, S.-G. Lin, X. Chen et al., "Transport of cryptotanshinone, a major active triterpenoid in Salvia miltiorrhiza Bunge widely used in the treatment of stroke and Alzheimer's disease, across the blood-brain barrier," *Current Drug Metabolism*, vol. 8, no. 4, pp. 365–377, 2007.
- [18] H.-Q. Li, J.-J. Wei, W. Xia et al., "Promoting blood circulation for removing blood stasis therapy for acute intracerebral hemorrhage: a systematic review and meta-analysis," *Acta Pharmacologica Sinica*, vol. 36, no. 6, pp. 659–675, 2015.
- [19] E.-W. Hao, J.-G. Deng, Z.-C. Du et al., "Experimental study on two-way application of drugs with neutral property for promoting blood circulation and removing blood stasis on cold and heat blood stasis syndromes II," *Zhongguo Zhongyao Zazhi*, vol. 38, no. 4, pp. 605–610, 2013.
- [20] T. Liu, H. Jin, Q.-R. Sun, J.-H. Xu, and H.-T. Hu, "The neuroprotective effects of tanshinone IIA on β-amyloid-induced toxicity in rat cortical neurons," *Neuropharmacology*, vol. 59, no. 7-8, pp. 595–604, 2010.
- [21] Y. Yin, L. Huang, Y. Liu et al., "Effect of tanshinone on the levels of nitric oxide synthase and acetylcholinesterase in the brain of Alzheimer's disease rat model," *Clinical and Investigative Medicine*, vol. 31, no. 5, pp. E248–E257, 2008.
- [22] L. Holcomb, M. N. Gordon, E. Mcgowan et al., "Accelerated Alzheimer-type phenotype in transgenic mice carrying both mutant amyloid precursor protein and presenilin 1 transgenes," *Nature Medicine*, vol. 4, no. 1, pp. 97–100, 1998.
- [23] C. Liu, Y. Wu, S. Zha et al., "Treatment effects of tanshinone IIA against intracerebroventricular streptozotocin induced memory deficits in mice," *Brain Research*, vol. 1631, pp. 137–146, 2016.
- [24] P. Jiang, C. Li, Z. Xiang, and B. Jiao, "Tanshinone IIA reduces the risk of Alzheimer's disease by inhibiting iNOS, MMP-2 and NF-κBp65 transcription and translation in the temporal lobes of rat models of Alzheimer's disease," *Molecular Medicine Reports*, vol. 10, no. 2, pp. 689–694, 2014.
- [25] B. Y. H. Lam, A. C. Y. Lo, X. Sun, H. W. Luo, S. K. Chung, and N. J. Sucher, "Neuroprotective effects of tanshinones in transient focal cerebral ischemia in mice," *Phytomedicine*, vol. 10, no. 4, pp. 286–291, 2003.
- [26] C. Maharana, K. P. Sharma, and S. K. Sharma, "Feedback mechanism in depolarization-induced sustained activation of

extracellular signal-regulated kinase in the hippocampus," *Scientific Reports*, vol. 3, article 1103, 2013.

- [27] Y. Teng, M. Q. Zhang, W. Wang et al., "Compound danshen tablet ameliorated a β 25–35-induced spatial memory impairment in mice via rescuing imbalance between cytokines and neurotrophins," *BMC Complementary and Alternative Medicine*, vol. 14, article 23, 2014.
- [28] H. Cai, L. Lian, Y. Wang, Y. Yu, and W. Liu, "Protective effects of salvia miltiorrhiza injection against learning and memory impairments in streptozotocin-induced diabetic rats," *Experimental and Therapeutic Medicine*, vol. 8, no. 4, pp. 1127– 1130, 2014.
- [29] T. Wang, F. Fu, B. Han, L. Zhang, and X. Zhang, "Danshensu ameliorates the cognitive decline in streptozotocin-induced diabetic mice by attenuating advanced glycation end productmediated neuroinflammation," *Journal of Neuroimmunology*, vol. 245, no. 1-2, pp. 79–86, 2012.
- [30] D. S. Lee, J. Choi, S.-H. Kim, and S. Kim, "Ameliorating effects of HX106N, a water-soluble botanical formulation, on Aβ25– 35-induced memory impairment and oxidative stress in mice," *Biological and Pharmaceutical Bulletin*, vol. 37, no. 6, pp. 954– 960, 2014.
- [31] A. Extance, "Alzheimer's failure raises questions about diseasemodifying strategies," *Nature Reviews Drug Discovery*, vol. 9, no. 10, pp. 749–750, 2010.
- [32] C. Shi, N. Na, X. Zhu, and J. Xu, "Estrogenic effect of ginsenoside Rg1 on APP processing in post-menopausal platelets," *Platelets*, vol. 24, no. 1, pp. 51–62, 2013.
- [33] C. Shi, D.-D. Zheng, L. Fang, F. Wu, W. H. Kwong, and J. Xu, "Ginsenoside Rgl promotes nonamyloidgenic cleavage of APP via estrogen receptor signaling to MAPK/ERK and PI3K/Akt," *Biochimica et Biophysica Acta—General Subjects*, vol. 1820, no. 4, pp. 453–460, 2012.
- [34] G.-W. Fan, X.-M. Gao, H. Wang et al., "The anti-inflammatory activities of Tanshinone IIA, an active component of TCM, are mediated by estrogen receptor activation and inhibition of iNOS," *Journal of Steroid Biochemistry and Molecular Biology*, vol. 113, no. 3–5, pp. 275–280, 2009.
- [35] G. Fan, Y. Zhu, H. Guo, X. Wang, H. Wang, and X. Gao, "Direct vasorelaxation by a novel phytoestrogen tanshinone IIA is mediated by nongenomic action of estrogen receptor through endothelial nitric oxide synthase activation and calcium mobilization," *Journal of Cardiovascular Pharmacology*, vol. 57, no. 3, pp. 340–347, 2011.
- [36] M. Xu, F. Cao, L. Liu et al., "Tanshinone IIA-induced attenuation of lung injury in endotoxemic mice is associated with reduction of hypoxia-inducible factor 1α expression," *American Journal of Respiratory Cell and Molecular Biology*, vol. 45, no. 5, pp. 1028– 1035, 2011.
- [37] J. Carter and C. F. Lippa, "β-Amyloid, neuronal death and alzheimer's disease," *Current Molecular Medicine*, vol. 1, no. 6, pp. 733–737, 2001.
- [38] H. Oakley, S. L. Cole, S. Logan et al., "Intraneuronal β-amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: potential factors in amyloid plaque formation," *The Journal of Neuroscience*, vol. 26, no. 40, pp. 10129–10140, 2006.
- [39] P. F. Chapman, G. L. White, M. W. Jones et al., "Impaired synaptic plasticity and learning in aged amyloid precursor protein transgenic mice," *Nature Neuroscience*, vol. 2, no. 3, pp. 271–276, 1999.

- [40] G. Zhu, Y. Wang, J. Li, and J. Wang, "Chronic treatment with ginsenoside Rg1 promotes memory and hippocampal longterm potentiation in middle-aged mice," *Neuroscience*, vol. 292, pp. 81–89, 2015.
- [41] G. Zhu, J. Li, L. He, X. Wang, and X. Hong, "MPTP-induced changes in hippocampal synaptic plasticity and memory are prevented by memantine through the BDNF-TrkB pathway," *British Journal of Pharmacology*, vol. 172, no. 9, pp. 2354–2368, 2015.