KF-1 ubiquitin ligase: an anxiety suppressor

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Anxiety is an instinct that may have developed to promote adaptive survival by evading unnecessary danger. However, excessive anxiety is disruptive and can be a basic disorder of other psychiatric diseases such as depression. The KF-1, a ubiquitin ligase located on the endoplasmic reticulum (ER), may prevent excessive anxiety; kf-1^{-/-} mice exhibit selectively elevated anxiety-like behavior against light or heights. It is surmised that KF-1 degrades some target proteins, responsible for promoting anxiety, through the ER-associated degradation pathway, similar to Parkin in Parkinson's disease (PD). Parkin, another ER-ubiquitin ligase, prevents the degeneration of dopaminergic neurons by degrading the target proteins responsible for PD. Molecular phylogenetic studies have revealed that the prototype of kf-1 appeared in the very early phase of animal evolution but was lost, unlike *parkin*, in the lineage leading up to *Drosophila*. Therefore, kf-1^{-/-} mice may be a powerful tool for elucidating the molecular mechanisms involved in emotional regulation, and for screening novel anxiolytic/antidepressant compounds.

Keywords: depression, ERAD pathway, Parkinson's disease, Alzheimer's disease, animal evolution

INTRODUCTION

Anxiety, or learned fear, is not necessarily harmful to everyday life but, rather, is a natural ability that may have arose to evade unnecessary dangers. However, excessive anxiety is debilitating or disadvantageous for life as it reduces behavioral activities necessary for adaptation. Moreover, anxiety can be a core symptom of various mental/ behavioral disorders, such as major depressive disorders, obsessive-compulsive disorders, panic disorder, adaptive disorder, post-traumatic stress disorder, social withdrawal disorder, and various phobias. Patients with anxiety/depression interpret circumstantial incidences, including episodes, comments, and expressions, in a negative way. The interpretation leads individuals to enhanced capture or delusions caused by

potential signs of danger. This suggests that a system of negative and positive regulation of the emotional expression may have developed under the evolutionary constraint. Such a system would be most apparent in highly social animals with relatively little reproduction per generation, like humans (Darwin, 1872). Indeed, there is evidence that the amygdala is responsible for the expression of anxiety or fear, and the prefrontal cortex plays a role in fear extinction by regulating the amygdala-mediated expression of fear (see Bishop, 2007). Although the molecular mechanisms underlying negative and positive regulation of the anxiety are not fully understood, many genes have been reported to affect anxiety or fear (Chen et al., 1994; Gogos et al., 1998; Heisler et al., 1998; Holmes et al., 2003; Miyakawa et al., 1994,

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2003; Nakajima et al., 2008; Yamasaki et al., 2008). Among these, the genes related to the serotonergic system are seen to be of special interest. For example, human genomic studies of anxiety/ depression have focused on genes related to the monoaminergic neurotransmission of serotonin receptors (5-HT_{1A}) and transporters (5-HTT) (see Levinson, 2006; Uher and McGuffin, 2008). Genetic studies using gene-targeting techniques have revealed that knockout mice lacking either 5-ht, or 5-htt exhibit significantly increased anxiety-like behaviors (Heisler et al., 1998; Holmes et al., 2003). However, proteins that interact directly with neurotransmitters seem to function downstream, rather than upstream, of the serotonergic pathway.

Anxiety/depression is the most common psychiatric disease seen in patients irrespective of nations, societies, and religions. Consequently, pharmaceutical companies have made extensive worldwide efforts to develop anxiolytic/ antidepressant drugs, particularly serotonergic compounds such as selective serotonin reuptake inhibitor (SSRI) and serotonin noradrenalin reuptake inhibitor (SNRI) based on the monoamine hypothesis. The hypothesis assumes that the pathogenesis of depression is caused by the depletion of monoamines such as serotonin in the brain. This assumption has however not been proven in the last 60 years. Testing systems have been developed for rodents to measure behavioral despair and to screen serotonergic drugs. These include the forced swim test and the tail suspension test. Compounds that elevate monoamine levels in the brain reduce the despair-like behavior or immobility time of animals under fearful conditions (Porsolt et al., 1978; Steru et al., 1985). However, approximately one-third of the patients with anxiety/depression do not respond to the serotonergic drugs, suggesting that despair-like behavior in rodents does not precisely represent anxiety/depression in humans, from a pharmacological point of view. Therefore, it is desirable to have some genetic animal models that display excessive anxiety-like behavior specifically without affecting despair-like behavior, as observed in the kf-1^{-/-} mice (Tsujimura et al., 2008), to look for novel anxiolytic/antidepressant drugs that are effective in patients who do not respond to serotonergic drugs.

IDENTIFICATION AND NATURE OF KF-1

The gene for KF-1 was originally discovered in connection with Alzheimer's disease (AD). Genetic studies of familial AD (FAD) made significant progress in 1990s. In this period, three genes encoding β -amyloid precursor protein, pressenilin-1 (PS1), and presenilin-2 (PS2) were identified as the causative genes for FAD (see Hashimoto-Gotoh et al., 2006; Lundkvist and Näslund, 2007), and two other genes were found expressed more frequently in the frontal cortex of an AD patient than a non-AD subject (Yasojima et al., 1997). One of the later two genes, gfap, was a known gene encoding glial fibrillary acidic protein, and the other was a novel gene, named kf-1. The *kf-1* gene is expressed most prominently in the brain and more or less throughout the entire body, but least, if any, in the liver, which implies that kf-1 may not be a housekeeping gene. Human kf-1 has four exons ranging over approximately 30 kb, and is mapped to chromosome 2p11.2. The protein structure deduced from the cDNA sequences has revealed that human KF-1 protein (GenBank Acc No. BAA19739) consists of 685 amino acids, and contains a possible leader peptide (amino acid positions 1-19), two membranespanning segments (326-345 and 366-380), and a RING-H2 finger motif (621-662) close to the C-terminus (Figure 1A). KF-1-like proteins have been found in other animals including fish (Danio rerio), lancelet (Blanchistoma floridae), sea urchin (Strongylocentrotus purpuratus), sea anemone (Nematostella vectensis), and tablet animals (Trichoplax adhaerens) (Figure 1A). Molecular phylogenetic studies suggest that genes for the animal and vertebral proteins may be kf-1 orthologues (Figures 1B,C, respectively). Unexpectedly, KF-1 homologues do not exist in insects (Drosophila melanogaster) and thread worms (Caenorhabditis elegans, and C. briggsae), even though it is likely that the prototype gene appeared in the very early phase of animal evolution, before the separation of Placozoa and Eumetazoa (Miller and Ball, 2008; Srivastava et al., 2008). Homologues of the prototype do not exist in prokaryotes, plants, or fungi, but are found first in one of the most primitive multicellular animals, such as tablet animals (Schubert, 1993) (Figure 1D). The results may be consistent with the report based on the EST analysis that some genes, formerly thought to be vertebrate inventions, must have been present in the common metazoan ancestor (Kortschak et al., 2003). Choanoflagellates (Monosiga brevicollis, and M. ovata), supposedly one of the most closely related unicellular protists to animals (King et al., 2008), do not possess kf-1 (Figure 1D).

The KF-1 protein is an E3 ubiquitin ligase that may modulate the cellular protein levels of its unknown substrate(s) (Lorick et al., 1999). The expression of kf-1 is also increased in the frontal cortex and hippocampus after chronic administration of SSRI in rats (Yamada et al., 2000). Furthermore, rat kf-1 expression is elevated

RING-H2 finger motif

A type of zinc binding domains, similar to RING finger motif containing a C3HC4 amino acid motif for binding to two zinc ions. RING-H2 finger motif contains two histidine residues as in C3H2C3 motif. Many of the RING finger domains function as ubiquitin ligases.

Α					

Hs	MWLKLFFLLVFVLARFFE-AIVWYETGIFATQLVDPVALSFKKLKTILECRGLGYSGLPEKKDVRELVEKSGDLMEGELYSALKEE-EASESVSSTNFSGEMHFYELV	111			
Dr	MWW KLFFLLYFFILFILARFFE-AIVWYETGIFATQLVDPVTLSFKKLKTILECRGLGYSGLAEKRDVRELVENSGELMQGELYSALKNEKEQAGSDSSTTFSGEMHFYELV	112			
Bf	$\texttt{MLV} = \texttt{MLV} \\ \texttt$	109			
Sp	${\tt MFM}{=}{-}{-}{-}{\tt KIVFLLVYATLLFLLARMLE}{-}{\tt YIPWYQTGLLMMKLIDPVSLSVKKLKSLLDGRGLSYEGVIDKAELTQLVEESGHVMEGEVLMMEQDASEREEEEPTTTTFSSHA}{-}{-}{-}{-}{-}{\tt KIVFLLVYATLLFLLARMLE}{-}{\tt YIPWYQTGLLMMKLIDPVSLSVKKLKSLLDGRGLSYEGVIDKAELTQLVEESGHVMEGEVLMMEQDASEREEEEPTTTTFSSHA}{-}{-}{-}{-}{-}{\tt KIVFLLVYATLLFLARMLE}{-}{\tt YIPWYQTGLLMMKLIDPVSLSVKKLKSLLDGRGLSYEGVIDKAELTQLVEESGHVMEGEVLMMEQDASEREEEEPTTTTFSSHA}{-}{-}{-}{-}{-}{\tt KIVFLLVYATLLFLARMLE}{-}{\tt KIVFLVYATLLFLARMLE}{-}{\tt KIVFLARMLE}{-}{\tt KIVFLARMLE}{-}{{\tt KIVFL$	109			
Nv	$\texttt{MLT}{}\texttt{KLLLLL}\texttt{VYFFLIFLTTRFLELTASWFEAGCIASQLFDPLSLSVRKLKAILDQRGVSYNGVVEKSELADLVEVSGAVTDPESALTAQGSNDNEQNSDEFTFKGASHFFEEV}$	113			
Ta	MTAGWFLTAIKIILLIVYLICVLLSCKYWN-VQLWKVGDKAARLLLDPATFNLKELIEIIDYRGVSDLNLHDRTNLSYMVNASGLMSEEEKWQSAIIM-AQSKKREAINFTAENYLRAEI 1:				
CS	M LY W DP L RG V SG E				
	***************************************	005			
HS Dec	EDINDGINUVQIANDROP-LVGRINNERMVKVSRFGIRIGINGSDPRICKRGWVKSILINSVFGISISKVMUKEVSGE-LEVENIPKWIIANASKKV-IIINAELLKE	225			
Dr	EDINDOLWUVQUIAQUKKP-LLSIANWKKPUQKSQFGIKIGIPACSDSKTCHKKQWKSILLIBSVFQITASKGVMLKEINGK-KLEIEHIFKWIAHVASKIK-IVKISQLMD				
Sp	EDINDSYWLYQYIFEDRIF-LLCYQQWRSUYAVSDRGINIGIACQUDRALCWRAGWURSJALAALYAGQAAGHISQQYFHIFCAAGHILHUHUQLSSRH-SJLSFAQQQ				
Nv	QUESTINGT ELEMENT TO THE AND T	210			
Та	EDNTFGTWLLRTDTRNSSKEDILVDNLWDDVVPKLYTLGVOSGTIRCRTCPTICRKNNWTKSDILLRTASDSDWSKGTLYYPGRLANANSSSILLWMRHLUSHINHFSTRIPPOLRE	238			
CS	W K G G C C W W				
	360				

Hs	$\label{eq:entropy} ewident evidence of the the the transformed of transformed of the transformed of transformed of the transformed of the transformed of transformed of the transformed of the transformed of the transformed of transformed of the transformed of transf$	339			
Dr	DWYQMEKQPVKMFLFARLLQPPAFFSALSIKFTGRIEFIFVDVRNWDNNTCLEEIGVQQMPSYILKTPEGIYRYGNSTGEFISLHAMDTFLRSVQPEVNDLFILSLVMVNLMAWWILMAWWWILMAWWWILMAWWWILMAWWWILMAWWWILMAWWWILMAWWWILMAWWWILMAWWWILMAWWWILMAWWWWWWWWWW	340			
Bf	$\label{eq:discrete} DWLMKNRTHPVQVVFFSRLKQPPMFYSALSVKFTGRVKFGYMRLNNSRNRLDISGREKIPGILVITPERRYWYGTGKGELLNLQSMQTYLRTMQPEVNDIFLVCVVVNLMAV and the second statement of the $	335			
Sp	$\label{eq:construction} DWD KLESN RDV AVKVVLFSR NQEPPVFFSALNLKYSG RVKFVFVSDSKTFYVLDNR-AQKYWLPSYIIVTPEGKKVYGEN NGEYCTYSALDLYLQILSPEANDIFVLTFINVNAICFFSALNLKYSG RVKFVFVSDSKTFYVLDNR-AQKYWLPSYIIVTPEGKKVYGEN NGEYCTYSALDLYLQILSPEANDIFVLTFINVNAICFFSALNLKYSG RVKFVFVSDSKTFYVLDNR-AQKYWLPSYIIVTPEGKKVYGEN NGEYCTYSALDLYLQILSPEANDIFVLTFINVNAICFFSALNLKYSG RVKFVFVSDSKTFYVLDNR-AQKYWLPSYIIVTPEGKKVYGEN NGEYCTYSALDLYLQILSPEANDIFVLTFINVNAICFFSALNLKYSG RVKFVFVSDSKTFYVLDNR-AQKYWLPSYIIVTPEGKKVYGEN NGEYCTYSALDLYLQILSPEANDIFVLTFINVNAICFFSALNLKYSG RVKFVFVSDSKTFYVLDNR-AQKYWLPSYIIVTPEGKKVYGEN NGEYCTYSALDLYLQILSPEANDIFVLTFINVNAICFFSALNLKYSG RVKFVFVSDSKTFYVLDNR-AQKYWLPSYIIVTPEGKKVYGEN NGEYCTYSALDLYLQILSPEANDIFVLTFINVNAICFFSALNTAFFSAL$	331			
Nv	ILQSDRDPNYIYVVYHSTLTEPPMFLSSLSIKFTGRVKFVYCRSHLKHRKEDIN-FDGFKVPSLFVITPERRVLFGLKKGEIYDYSSLELYLRTLHPEVNDLFLAALVITNLCCM	340			
Ta	TIKKQSGIHVILFSTLTVAPTFISSLAIKFSGRINFSMVTIKTINDTIQNLFANEFNVEKLPTYRIFTPEKNFTYGNRHGEYYGYHCHHEFLTSLYPATNDIFVAIVIGINLHCL	353			
CS	PFSLKGRF PTPEGGELPNDFN				
		455			
HS	MDLFITQGATLKKFVVLLSTLGTYNSLLIISWLPVLGFLQLFYLDSFYLFYSLKLLKKYSNITTLASWVRAD-WMFYSSHPALFLSTYLGHGLLDYFEKKKKKNNNNDEVNANNLEWL-	455			
Dr	MDLFITQGATIKRFVVLISTLGTVNSLLIISWLFILGFLQLPYLDSFYEYSLKLIKVADTTTTIASWVRAD-WTFYSSHPALFLSTYLAHGLLIDYFEKKRRGSNEDQ-NANNLEWL- 45-				
DI Cn	LEVELCGQUVGVGCLRLLWAVGKYRCVLLWVCLPVVGGEQLECMEGVVQAGTTALKNISSSGLVAQARQD-WLLYSSHRPFLVGTFLLYSMAVGTVASKWKS-GEEMASETTPTEQAGT- WKLETLOGCTAVETCGELUTTCKYNTTILLULULULULULULULULULULULULULULULULULU				
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CS	G VN P P R R R				
	****** 600				
Hs	SSLWDWYTSYLFHPIASFQNFPVESDWDEDPDLFLERLAFPDLWLHPLIPTDYIKNLPMWRFKCL	520			
Dr	SSLWDWYTSYLVHPIASFQNFESDWDDDPNFLLERLAFPDLWLRPLVPIDYIKNLPTWKFRLL	517			
Bf	${\tt NGWWSSFMNYLFQPISTFNHTRPPNLIGLEDGLDYLIERLAVPDLWLHPVIPTDYVRNLPVWLYKGKMPLVRKVCSKCCGWGGSADQAAMVMKLPTVGHTESNRVMSLPPSPRRPGHRPPICAUCAUCAUCAUCAUCAUCAUCAUCAUCAUCAUCAUCAUC$				
Sp	ASLVDFR-SLLLRPSPSFRNGLPRN-GHTEEGLDLLIDQMGVPDLWLHPLIRDDYIKDLLTWRYSCQ				
Nv	TQDLNYF-SRILQSLSHWQPQIHHTTSGFEDGFEMLVRRLAVPDLWLHPIIPTDYIKQLPTWNFCCK	524			
Ta	SLSSYQSSLYSILTYRSACERYSREDIEEGIDILIEHFAIPKLYVQPLVPMDYIQSLPTFTYGDK55				
CS	PL P DY L				
	720				
HS	GVUSEEEMSEGSUDTENDS-ESENTDILESSEKEVFEDKU-SVLINSPCTASHCDAEACSCARKYCUTSPCERKGRSYGSYNTNEDMEPDWLTWP 612				
Dr					
Sn	SPSKHPSQRHKOKK IKS IDJ IPLS ISDJNISSDQRAK I I GEKSKGASDHGW YGHDV MVPSKHNNSPSGRHPPP VMPI PSLDKPCI GUNPQU ISSAGG				
Nv					
Ta		567			
CS					

Hs	ADMLHCTECVVCLENFENGCLLMGLPCGHVFHQNCIVMWLAGGRHCCPVCRWPSYKKKQPYAQHQPLSNDVPS 685 100.0%				
Dr	CGMLHCTECVVCLENFETDCLVMGLPCGHVFHQQCIVVWLAGGRHCCPVCRWPSYKKRPVRQRATEQLDPE 656 69.8%				
Bf	LGILPCEDCAICLEEYEVGCSLLGLPCGHSFHERCIMMWLSAGNHCCPVCRWPAFKFKPA-LHLHSE737 39.1%				
Sp	EGILYDAQCAICIEAYTNGAELCGLPCGHAYHQQCIVAWLNNGNHVCPICRWPAYKKKGSKLSKHME 679 37.5%				
Nv	-WMIPCGECVICLDEFKPGCTLLGLPCGHSFHQHCIEVWLAGDNTAPHHCCPNCRWPAYRAKSH-VH617 34.9%				
Ta	-YNCDDNQCSICLTNYINDDYLCCLPCSHVFHHDCIVQWLSIGTINT-CRCPLCRWPAYRSYLQPSSACHATSITSNEDSS 646 24.7%				
CS	G G LPC H H CI WL CP CRW				
Figure	1 Molecular phylogenetic studies of the <i>kf-1</i> gene. (A) The alignment The alignment positions, including gaps, are given on the top right of each	row.			
ot K⊦-1	protein sequences. The sequence identities (%) given at the ends were Hs, Dr, Bt, Sp, Nv, and Ta denote Homo sapiens, Danio rerio, Blanchistoma	a			
uptaine	motif namely after amino acid position 621 in human KE-1. Actoriske show the adhacrone respectively. The gape identification (CI) support are 1045615				

motif, namely after amino acid position 621 in human KF-1. Asterisks show the alignment sites used for inferring the **molecular phylogenetic tree** in **(B)**. The two transmembrane regions of human KF-1 are indicated by '*z*' and the RING-H2 finger motif for zinc-binding is indicated by '*#*'. The number given at the right side indicates the amino acid position of the C-terminal amino acid in each line.

The alignment positions, including gaps, are given on the top right of each row. Hs, Dr, Bf, Sp, Nv, and Ta denote *Homo sapiens, Danio rerio, Blanchistoma floridae, Strongylocentrotus purpuratus, Nematostella vectensis* and *Trichoplax adhaerens*, respectively. The gene identification (GI) numbers are 1945615, 52219054, 210102052, 115660659, 156406687, and 196009364 in the respective species. Blast *Expect* values (blastp with BLOSUM62 for Matrix) of Dr, Bf, Sp, Nv, and Ta against Hs are $E < 10^{-130}$, $E = 2 \times 10^{-130}$, $E = 3 \times 10^{-117}$, $E = 3 \times 10^{-104}$, and $E = 3 \times 10^{-57}$, respectively. CS denotes consensus amino acid. (*continued*)



after physical antidepressant treatments, such as electroconvulsive therapy (Nishioka et al., 2003), and repetitive transcranial magnetic stimulation (Kudo et al., 2005). This implies that the upregulation of kf-1 expression is associated with some physiological responses to antidepressant treatments rather than being caused by a chemical reaction to serotonergic compounds such as SSRIs. It was, however, not clear whether this was a result, cause, or coincidental side effect of antidepressive processes.

As a possible correlation between KF-1 and AD has been suggested (Yasojima et al., 1997), and as depression can occur early in the course of AD in *ps1* mutation carriers unaware of their genetic status (Ringman et al., 2004), the first working hypothesis was that presenilins could be KF-1 substrates in the endoplasmic reticulum (ER) associated degradation (ERAD) pathway (see Carvalho et al., 2006; Lorick et al., 2006). Therefore, the intracellular localization of KF-1 was examined in comparison to that of presenilins known to be

located on ER (Kovacs et al., 1996). The results have revealed that KF-1 is co-localized with both PS1 and PS2 (**Figures 2A,B** and Tsujimura et al., 2008), which implies that KF-1 may in fact be an ER-ubiquitin ligase. The co-localization of KF-1 with other ER markers, such as Der-1 and VCP, has also been observed (Maruyama et al., 2008).

SELECTIVE EXPRESSION OF INCREASED ANXIETY-LIKE BEHAVIORS IN kf-1^{-/-} MICE

Although mouse *kf-1* is expressed in many tissues, particularly in the brain, the lack of *kf-1* does not lead to any abnormalities in appearance or behaviors, including body weight, reproductive capability, exploratory locomotion, nociception, social behavior, motor coordination, behavioral despair, spatial working memory, and context memory (**Table 1**).

The exception is that kf- $1^{-/-}$ mice display a pronounced increase in anxiety-like behavior in the light/dark transition test (stay time in light compartment: p = 0.0004; number of

Molecular phylogenetic tree

A diagram showing the evolutionary relationship or history of organisms, genes or proteins. It is inferred by a phylogentic method such as neighbor-joining, maximum parsimony, or maximum likelihood, based on the nucleotide or amino acid sequences of various species.





Figure 2 | Immunostaining of *Xenopus* KF-1 tagged with the Myc epitope and PS α or PS β co-expressed in COS-1 cells. (A) Proteins were stained with anti-Myc antibody probed with rhodamin conjugated anti-mouse IgG for KF-1 (red) and anti-PS α (*Xenopus* PS1 homologue) probed with fluorescein isothiocyanate (FITC) conjugated to anti-rabbit IgG for PS α (green). Images of 'anti Myc (KF-1)' and 'anti PS α ' are merged ('merge'). **(B)** The same as **(A)** except PS β (*Xenopus* PS2 homologue) was used instead of PS α . KF-1 is co-localized with either PS α or PS β as indicated in yellow or orange ('merge').

Table 1 | Behavioral phenotypes of kf- $f^{-/-}$ mice compared to the behavioral phenotypes of kf- $f^{+/+}$ littermates.

Tests	Measurements	Phenotypes ^a
General health examination	Whisker, coat, reflexes	=
Physical test	Body temperature	=
	Body weight	=
	Wire hanging time	Ŷ
	Grip strength	=
	Auditory capacity	=
Light/dark transition test	Anxiety	Ŷ
Open field test	Exploratory locomotion	=
Elevated plus maze test	Anxiety	\uparrow
Hot plate test	Pain sensitivity (latency time)	=
Social interaction test	Total duration of contacts	=
	Number of contacts	=
	Total duration of active contacts	=
	Mean duration/contact	=
	Distance traveled	=
Rotarod test	Motor coordination	=
Prepulse inhibition (PPI) test	Sensorimotor gating	\uparrow
Porsol forced swimming test	Immobility time (behavioral despair)	=
T-maze test	Spatial working memory	=
Cued and contextual fear conditioning test	Immediate freezing during conditioning phase	\uparrow
	Contextual testing conducted after conditioning	=
	Cued test with altered context	=
Tail suspension test	Immobility time (behavioral despair)	=

The testing order is from the top to the bottom of this table. Further details are presented in Tsujimura et al. (2008). a =, no significant difference; \uparrow , increased in kf-1^{-/-} compared to kf-1^{+/+}. The raw data of the results are available at https://behav.hmro.med.kyoto-u.ac.jp/.

SNP

DNA sequence variation, called single nucleotide polymorphism, occurring on a single nucleotide in the genome among individual members of a species or between paired chromosomes within an individual. 'SNP (pronounced *snip*)' was defined initially as that with a minor allele frequency of $\geq 1\%$, but this definition is rather artificial and not meaningful.

Ubiquitination

Ubiquitination (or ubiquityation) is the post-translational tagging reaction mediated by E3 ubiquitin ligase to a protein with one or more highly conserved peptide molecules called 'ubiquitin' consisting of 76 amino acids. Ubiquitination targets the substrate protein for proteasomal degradation.

transitions: p = 0.0392) (Tsujimura et al., 2008). Consistently, the mice also show 'timidity-like' responses under stressful situations, such as prolonged wire-hanging time, enhanced immediate freezing with an aversive foot shock, and decreased locomotor activity at heights (Table 1 and Tsujimura et al., 2008). As significant differences are not observed in body weight and grip strength, and in general exploratory locomotion, the 'timidity-like' responses are likely to be related to psychological but not physical alterations in *kf-1^{-/-}* mice. This implies that KF-1 plays a role in emotional control by suppressing anxiety at least in mice. In this context, it is of particular interest to note that a number of SNPs are reported in the coding and non-coding exonic regions of human kf-1 according to the NCBI's SNP database, and some are supposed to substantially modulate KF-1 activity in homozygous or dual heterozygous carriers; for example, rs35921467 resulting in a frame shift at amino acid position 180, rs17857046 and rs17853383 in critical amino acid substitutions (S to P and P to H) at 251 and 502, respectively, and rs11695337 in a termination codon at 626.

As KF-1 is an E3 ubiquitin ligase located on ER, KF-1 may be responsible for conducting the ERAD pathway, in which some factors promoting anxiety may be targeted. Therefore, the absence or reduction of KF-1 activity may result in 'enhanced anxiety' by increasing putative anxiety promoters (**Figure 3A**). Regarding serotonergic metabolisms, it is not clear at the moment whether or how the ERAD pathway directed by KF-1 is associated with the serotonergic pathway, and it has to be elucidated in future. However, KF-1's targets for proteasomal degradation should be neither the serotonin receptor (5-HT_{1A}) nor the transporter (5-HTT), because a lack of these proteins increases anxiety-like behaviors (Heisler et al., 1998; Holmes et al., 2003).

KF-1 AND PARKIN AS ER-BASED E3 UBIQUITIN LIGASES

Ubiquitination plays an essential regulatory role in all critical eukaryotic cellular processes. Proteasomal degradation through the ERAD pathway is not an exception. It has been well established that these processes play an important role in a variety of human somatic diseases, ranging from cancer, viral infection, diabetes, and inflammation to muscle wastage and neurodegenerative disorders (see Kostova et al., 2007; Petroski, 2008). However, there are no reports on the animal specific ubiquitin ligase, which is responsible for emotional control or mental disorders.



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PPI

Prepulse inhibition (PPI) is a neurological phenomenon where a weaker pre-stimulus inhibits the reaction of animals to a subsequent strong startling stimulus.

Sensorimotor gating

The brain's ability to filter out the unnecessary information.

Concerning neurodegenerative disorders, Parkin, another ER-ubiquitin ligase, plays a key role in preventing the degeneration of dopaminergic neurons in Parkinson's disease (PD). A recessive mutation in *parkin* is responsible for the neurodegenerative disorder known as the autosomal recessive juvenile form of Parkinsonism, the most common form of familial PD (Kitada et al., 1998). Human parkin with 12 exons ranging over approximately 1.5 Mb is located on chromosome 6q25.2.27, and encodes an E3 ubiquitin ligase, consisting of 465 amino acids, with two RING finger motifs. Parkin substrates have been identified being degraded through the ERAD pathway, such as Pael-R, CDCrel-1, α-Syn, and synphilin-1 (Yang et al., 2003). The accumulation of these proteins in dopaminergic neurons

leads to ER stress-induced apoptosis, resulting in PD (see von Coelln et al., 2004). Promoting their degradation would aid PD patients by preventing neurodegeneration. In fact, the neuronal synthesis of the Parkin substrate Pael-R has been shown to cause a loss of dopaminergic cells in transgenic *Drosophila* models. The co-expression of *parkin* results in the degradation of Pael-R and nearly completely blocks neurodegeneration, whereas interfering with the function of endogenous *Drosophila* Parkin promotes Pael-R accumulation and augments its toxicity (Yang et al., 2003). Hence, the over-expression of *parkin*

has emerged as a powerful approach to PD with

complementary effects to approaches described for the use of neurotrophic factors against PD (Ulusoy and Kirik, 2008). An analogy could be made by having such Drosophila models to study the role of KF-1 in anxiety/depression. However, the analogy may not be feasible as no evidence is available for the involvement of neuronal cell death in anxiety/depression, and kf-1 is absent in Drosophila (Figure 1D). It should be noted that *kf-1^{-/-}* mice have increased **PPI** values (**Table 1**), which implies that at least one of the KF-1 target proteins is involved in regulation of sensorimotor gating. In such cases, over-expression of kf-1 might cause serious side effects as the PPI values are usually reduced in schizophrenic patients. Despite this, one cannot rule out the possibility of kf-1 up-regulation as an anxiolytic/antidepressant treatment.

Instead, kf- $1^{-/-}$ mice provide a powerful tool for some pharmacological approaches to anxiety/ depression. For example, by using the mice, one can identify the putative anxiety promoters that are degraded through the ERAD pathway mediated by KF-1 and are responsible for the sensitivity to potential signs of danger or to stressful situations (**Figures 3A,B**). KF-1 substrates acting as anxiety promoter may be found by the computed differential screening in 2-dimensional gel electrophoresis (see Marengo et al., 2008) of cell homogenates derived from the frontal cortex or hippocampus of kf- $1^{-/-}$ and kf- $1^{+/+}$ mice. As the



expression of kf-1 is up-regulated in the frontal cortex and hippocampus after antidepressant treatments, it is likely that the anxiolytic/antidepressive effect of KF-1 functions primarily in these tissues and that their inhibitory signals are transmitted to the amygdala to prevent the manifestation of anxiety or fear (**Figure 4**). This proposal may be highly hypothetical and hence has to be elucidated in future studies. To conclude, examining kf-1^{-/-} mice in a simple light/dark transition test is an effective method of screening novel anxiolytic/antidepressant drugs that inhibit the putative anxiety promoters.

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