Review

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Serotonin as a New Therapeutic Target for Diabetes Mellitus and Obesity

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Serotonin (5-hydroxytryptamine [5-HT]) is a monoamine that has various functions in both neuronal and non-neuronal systems. In the central nervous system, 5-HT regulates mood and feeding behaviors as a neurotransmitter. Thus, there have been many trials aimed at increasing the activity of 5-HT in the central nervous system, and some of the developed methods are already used in the clinical setting as anti-obesity drugs. Unfortunately, some drugs were withdrawn due to the development of unwanted peripheral side effects, such as valvular heart disease and pulmonary hypertension. Recent studies revealed that peripheral 5-HT plays an important role in metabolic regulation in peripheral tissues, where it suppresses adaptive thermogenesis in brown adipose tissue. Inhibition of 5-HT synthesis reduced the weight gain and improved the metabolic dysfunction in a dietinduced obesity mouse model. Genome-wide association studies also revealed genetic associations between the serotonergic system and obesity. Several genetic polymorphisms in tryptophan hydroxylase and 5-HT receptors were shown to have strong associations with obesity. These results support the clinical significance of the peripheral serotonergic system as a therapeutic target for obesity and diabetes.

Keywords: Diabetes mellitus; Obesity; Serotonin

INTRODUCTION

Serotonin (5-hydroxytryptamine [5-HT]) has diverse functions in neuronal and non-neuronal systems. It acts as a hormone and a mitogen, as well as a neurotransmitter. At the time of its discovery (1918), 5-HT was thought to be a vasoconstrictor stored in platelets, so it was named serotonin after the Latin word serum and the Greek word tonic [1,2]. In 1937, 5-HT was found in the enterochromaffin (EC) cells of the gastrointestinal tract and named enteramine because it was thought to cause smooth muscle contraction in the gut [3]. It was not until 1952 that scientists reported 5-HT's role as a neurotransmitter [4]. It has since been discovered that 5-HT is a neurotransmitter associated with mood, behavior, sleep cycles, and appetite [2].

The synthesis of 5-HT is initiated with the hydroxylation of tryptophan by tryptophan hydroxylase (TPH), which is the rate-limiting enzyme for 5-HT synthesis. Thus, 5-HT production is regulated by the activity of TPH and the availability of tryptophan. The biological actions of 5-HT are mediated through the membrane-bound 5-HT receptors and terminated by its uptake into cells through a 5-HT transporter (SERT, Slc6a4) [5]. There are two isoforms of TPH. TPH1 is predominantly expressed in the peripheral tissues including the pineal gland, whereas TPH2 is exclusively expressed in the brain and the enteric nervous system [6,7]. Because 5-HT cannot cross the blood-brain barrier, the peripheral 5-HT system is functionally separate from the central 5-HT system.

Within the central and enteric nervous systems, 5-HT is

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synthesized and stored in the presynaptic neurons. Most 5-HT in the periphery is synthesized in EC cells [8] and platelets are the major storage site for circulating 5-HT. Several studies recently reported that 5-HT can be synthesized in other peripheral tissues (e.g., the heart, pancreas, and adipose tissue) and plays a role in a cell-autonomous manner [9-11].

In this review, we will discuss the recent findings on the roles of central/peripheral 5-HT in metabolism and its clinical implications for the treatment of obesity.

THE 5-HT RECEPTOR FAMILIES

5-HT regulates various physiological and pathological processes, which are mediated through numerous 5-HT receptors. The 5-HT receptors are phylogenetically ancient and present in the lowest invertebrates as well as the higher mammals [12]. More than fourteen 5-HT receptors in seven families are known to mediate the diverse effects of 5-HT (Table 1) [13,14]. All 5-HT receptors are G-protein-coupled receptors except for the 5-HT3 receptor, which is a ligand-gated cation channel [12,15].

Table 1. The 5-HT receptors, their functions and the specific receptor agonists/antagonists

Receptor subtype	Structure	Transduction system			
5-HT1A	GPCR	↓cAMP			
		G-protein coupled- K+ current			
5-HT1B	GPCR	↓cAMP			
5-HT1D	GPCR	↓cAMP			
5-HT1E	GPCR	↓cAMP			
5-HT1F	GPCR	↓cAMP			
5-HT2A	GPCR	↑PCR			
5-HT2B	GPCR	↑PCR			
5-HT2C	GPCR	↑PCR			
5-HT3	LGIC	Ion conductance (K+, Na+, Ca2+			
5-HT4	GPCR	↑cAMP			
5-HT5A	GPCR	↓cAMP			
		Ca ²⁺ mobilization			
		K ⁺ current			
5-HT5B	GPCR	Not known			
5-HT6	GPCR	↑cAMP			
5-HT7	GPCR	↑cAMP			

5-HT, 5-hydroxytryptamine; GPCR, G protein coupled receptor; cAMP, cyclic adenosine monophosphate; PCR, phosphocreatine; LGIC, ligand-gated ion channel.

The 5-HT1 receptor family consists of five receptors: the 5-HT1A, 5-HT1B, 5-HT1D, 5-HT1E, and 5-HT1F receptors. All these receptors are Gαi/o protein-coupled receptors that inhibit adenyl cyclase and reduce the cyclic adenosine monophosphate levels. The 5-HT2 receptor family consists of the 5-HT2A, 5-HT2B, and 5-HT2C receptors. These receptors are Gαg/11 protein-coupled receptors that mediate excitatory neurotransmission. The 5-HT2A and 5-HT2C receptors are mainly expressed in the central nervous system (CNS). The 5-HT2B receptor is expressed in peripheral tissues including the liver, kidney, heart, and stomach. It is associated with cardiac function, valvular heart disease, and heart morphogenesis. The 5-HT3 receptor is a member of the Cys-loop ligand-gated ion channel family. This channel is formed by a homopentamer of 5-HT3A subunits or heteropentamer of 5-HT3A subunits with 5-HT3B-E subunits. Opening of the channel in response to 5-HT activates an inward current and depolarizes the membrane.

REGULATION OF APPETITE BY CENTRAL 5-HT

Appetite is regulated by hypothalamic feeding circuits. Briefly, anorexigenic proopiomelanocortin (POMC) neurons release α -melanocyte-stimulating hormone, the endogenous ligand for the melanocortin 4 receptor (MC4R), to reduce appetite and food intake. Orexigenic neuropeptide Y/agouti-related peptide (NPY/AgRP) neurons increase appetite and food intake by releasing the endogenous MC4R antagonist, AgRP, and they suppress POMC neurons by releasing γ -aminobutyric acid (GABA).

It has long been known that 5-HT in the hypothalamus plays an important role in the control of appetite and energy balance. Central 5-HT has suppressive effects on food intake and body weight. Fenfluramine, which enhances 5-HT release, is a prototypical agent that leads to serotonergic suppression of feeding [16]. 5-HT reuptake inhibitors and monoamine oxidase inhibitors, which can increase the central serotonergic activity, also reduce food intake [17,18]. In contrast, inhibition of central 5-HT synthesis by intraventricular injection of a TPH inhibitor induced hyperphagia and obesity in rats [19].

Studies using receptor-specific knock-out (KO) mice have provided more precise insight into the roles of 5-HT in appetite regulation. The involvement of central 5-HT1B and 5-HT2C receptors in the regulation of appetite was initially suggested by the anorectic effects of a central serotonergic agonist (m-chlorophenylpiperazine [mCPP]) [20]. A major contribution of



Table 2. Anti-obesity drugs that work by regulating the central serotonergic system

Name or code	Company	Action	Current status
Lorcaserin	Arena Pharma	5-HT2C receptor agonist	FDA approved.
ATHX-105	Athersys	5-HT2C receptor agonist	Withdrawn
BVT.74316	Biovitrum	5-HT6 receptor antagonist	Withdrawn
PRX-07034	EPIX Pharma	5-HT6 receptor antagonist	Phase II
Tesofensine	NeuroSearch	5-HT/DA/NA reuptake inhibitor	Phase III
Dov-21,947	Dov Pharmaceuticals	5-HT/DA/NA reuptake blocker	Phase ll
Sibutramine		5-HT/NA reuptake inhibitor	FDA approved (1997)
			Withdrawn due to cardiovascular complication (2010)
Fenfluramine/phentermine		5-HT releasing agent/NA stimulant	FDA approved (1973)
			Withdrawn due to cardiovascular complication (1997)

Adapted from ClinicalTrials.gov [25].

5-HT, 5-hydroxytryptamine; FDA, Food and Drug Administration; DA, dopamine; NA, noradrenaline.

5-HT2C receptor to feeding control was confirmed by a study using Htr2c KO mice [21]. Htr2c KO mice are hyperphagic and obese, and the anorectic effects of mCPP do not occur in Htr2c KO mice [22]. Furthermore, studying the effects of 5-HT on the hypothalamic feeding circuit revealed that 5-HT activates POMC neurons through the 5-HT2C receptor and inhibits NPY/AgRP neurons through the 5-HT1B receptor [23]. Thus, central 5-HT inhibits food intake by modulating the hypothalamic feeding circuits. Many drugs targeting 5-HT2C receptor have been produced and some are currently being used in the clinical setting. However, others were discontinued because of severe complications such as pulmonary hypertension and valvular heart disease (Table 2) [24,25]. Recently, a selective 5-HT2C receptor agonist (Belviq, lorcaserin hydrochloride; Arena, Zofingen, Switzerland) was officially approved for use in the treatment of obesity [26].

Given that TPH2 is exclusively expressed in the brain, *Tph2* KO mice were expected to be hyperphagic and obese. However, *Tph2* KO mice do not show an obese phenotype although their central 5-HT level is decreased [27,28]. In addition, *Htr1b* KO mice do not become obese although they are hyperphagic [29]. There have also been reports supporting that central 5-HT increases the sympathetic tone and activates brown adipose tissue (BAT) [30,31]. Taken together with the known adverse effects of anti-obesity drugs with regard to their increasing the 5-HT activity in the brain, these controversial data indicate that increasing the 5-HT activity in the brain may not be a good strategy for anti-obesity treatment.

ROLES OF 5-HT IN PERIPHERAL TISSUES

Although TPH1 is widely expressed in the peripheral tissues, over 90% of the total body 5-HT is produced in the gut [32]. The majority of the peripheral 5-HT is stored in platelets and controls the hemodynamics upon the activation of platelets. 5-HT is also present in other peripheral tissues and acts autonomously on many cells in the cardiovascular, gastrointestinal, hematopoietic, and immune systems, as well as in the bone, liver, and placenta [33].

Gut

EC cells are dispersed throughout the intestinal mucosa and are the main source of 5-HT in the gut. The major targets of the gut-derived 5-HT (GDS) are the mucosal projections of primary afferent neurons. These include extrinsic nerves and afferent neurons. The extrinsic nerves transmit sensations of nausea and discomfort to the CNS. There are two types of intrinsic primary afferent neurons (IPANs). The submucosal IPAN regulate the peristaltic and secretory reflexes, whereas myenteric IPAN regulate giant migrating contractions. The 5-HT secreted from myenteric neurons mediates fast and slow excitatory neurotransmission and regulates gastrointestinal motility.

Many 5-HT receptor agonists/antagonists are already used to regulate gut motility in the clinical setting (Table 3). Pressure, acid, and/or cancer therapeutic agents are known to release 5-HT from EC cells, which provokes abdominal discomfort and nausea [34,35]. 5-HT3 receptor antagonists, such as



Table 3. Effects of 5-HT receptor agonists/antagonists in the gut

Class	Compound	Effect		
5-HT3 receptor antagonist	Ondansetron Granisetron	Slower small bowel transit Decreased intestinal secretion		
	Alosetron	Inhibition of colonic response to feeding		
	Cilansetron	Slower colonic transit		
5-HT3 receptor agonist	MKC-733	Slower emptying of liquids Faster small bowel transit		
		Stimulation of interdigestive phase 3		
5-HT4 receptor agonist	Tegaserod Prucalopride	Faster gastric emptying Enhanced gastric accommodation		
	Renzapride	Faster small bowel and colonic transit		
	Cisapride	Enhanced intestinal secretion		
5-HT1 receptor agonist	Sumatriptan	Enhanced gastric accommodation		
	Buspirone	Slower gastric emptying		
	R-137696	Stimulation of interdigestive phase 3		

5-HT, 5-hydroxytryptamine.

ondansetron (Zofran; GSK, London, UK), granisetron (Kytril; Roche Laboratories, Basel, Switzerland), or alosetron (Lotronex; GSK, London, UK), alleviate the 5-HT-induced discomfort that extrinsic sensory nerves convey to the CNS. Irritable bowel syndrome (IBS) is a disease linked to the serotonergic system. The persistent visceral hypersensitivity of the bowel in IBS patients can be blunted by the administration of a 5-HT4 receptor agonist (Tegaserod; Novartis, Basel, Switzerland) [36]. Thus, drugs that inhibit the 5-HT3 receptor or enhance the 5-HT4 receptor have been used to treat IBS.

Many studies suggest that 5-HT plays a role in the pathogenesis of gastrointestinal diseases by affecting the production of proinflammatory mediators and the immune system [37]. 5-HT receptors have been identified on various immune cells such as lymphocytes, monocytes, macrophages, and dendritic cells [38,39]. The T lymphocyte-derived 5-HT may act as an autocrine factor and has demonstrated a fundamental role as an intrinsic cofactor in T-cell activation and function through the 5-HT7 receptor [40].

Liver

5-HT is known to regulate liver regeneration. Partial hepatec-

tomy has been widely used as an experimental model to understand the role of 5-HT in liver regeneration. The number of platelets (filled with 5-HT) strongly correlates with the hepatocyte proliferative capacity [41]. In contrast, several studies using SERT-deficient rats reported that the regenerative process is not influenced when the platelet 5-HT level is reduced [42]. Thus, the mechanism underlying liver regeneration is more complex than has previously been thought, and more studies are needed to elucidate the main source of 5-HT to regulate liver regeneration. To directly examine the function of 5-HT in liver regeneration, partial hepatectomy was performed on Tph1 KO and wild type (WT) control mice. All markers of hepatocyte proliferation were reduced in Tph1 KO mice, suggesting that 5-HT is involved in the induction of hepatocyte proliferation after a major loss of hepatic tissue [43]. It has also been demonstrated that 5-HT can promote liver regeneration after liver injury through the 5-HT2 receptor or 5-HT7 receptor [44,45].

Crane et al. [46] reported a role of peripheral 5-HT in the liver after exposure to a high fat diet (HFD). When fed a HFD, WT mice become obese and develop fatty liver disease. In contrast, *Tph1* KO mice do not develop a fatty liver after exposure to a HFD and have reduced fat accumulation in the liver. 5-HT increases hepatic glucose uptake and fat content in liver [47,48]. The duodenal 5-HT content increases in *ob/ob* mice and treatment with a 5-HT3 receptor antagonist caused the reduction of the elevated 5-HT levels and an increase in SERT in the duodenum. In these mice, the 5-HT3 receptor antagonist also reduced the fat content, inflammation, and necrosis of the liver [48].

With regard to insulin resistance, Sumara et al. [8] reported that inhibition of GDS improves the glucose tolerance in mice fed a HFD. In hepatocytes, GDS signaling through the 5-HT2B receptor promotes gluconeogenesis. In addition, GDS prevents glucose uptake into hepatocytes in a glucose transporter 2-dependent manner. Therefore, gut-specific *Tph1* KO mice and liver-specific *Htr2b* KO mice showed improved glucose tolerance compared to WT mice [8].

Pancreatic β -cells and the impact of 5-HT on diabetes mellitus

It has long been known that 5-HT is present in the same vesicle with insulin in pancreatic β -cell after preloading of 5-hydroxytryptophan, a precursor of 5-HT, and 5-HT has been associated with the regulation of the blood glucose concentration [49,50]. Recently, 5-HT has been identified as a down-



stream molecule of placental lactogen that mediates the adaptation of β -cells to pregnancy [10]. In response to the lactogens produced during pregnancy, the expression of Tph1 rises sharply in pancreatic β -cells, resulting in massive production of 5-HT in β -cells [10]. Islet 5-HT acts in an autocrine/paracrine manner through the 5-HT3 receptor to increase the glucose responsiveness of β -cells and thereby increases the overall islet glucose-stimulated insulin secretion (GSIS) [51,52]. In addition, the expression of the 5-HT2B receptor increases during mid-gestation to stimulate β -cell proliferation and increase the β -cell mass, whereas 5-HT1D receptor expression increases at the end of gestation to reduce the β -cell mass [10].

5-HT also regulates insulin secretion in a diet-induced insulin-resistant state. The β -cell-specific Tph1 KO mice and Htr3a KO mice developed glucose intolerance compared to WT mice after exposure to an HFD. The GSIS was impaired in islets isolated from HFD-fed Htr3a KO and β -cell-specific Tph1 KO mice, and 5-HT treatment improved the insulin secretion from the islets of β -cell-specific Tph1 KO mice, but not from the Htr3a KO islets [51]. In addition, 5-HT production has been observed in the embryonic pancreas. In particular, TPH1 expression is upregulated in newborn β -cells during the perinatal period while β -cells are actively proliferating [53]. Given that β -cell produce 5-HT under two physiological conditions of β -cell proliferation, the perinatal period and pregnancy, 5-HT is thought to be an important regulator of β -cell proliferation as well as insulin secretion [10,51,52,54].

Adipose tissues

The biological functions of peripheral 5-HT in the regulation of energy homeostasis have been extensively studied using chemical agonists and antagonists since the 1960s. By taking advantage of mouse genetic studies, it has become possible to better understand the precise roles of 5-HT in energy metabolism. SERT KO mice exhibit an obese phenotype although they are expected to be slim due to the anorexigenic effects of central 5-HT [55]. Body weight is reduced in Tph1 and Tph2 double KO mice, as well as Tph1 KO mice [27,28]. However, the body weights of gut-specific Tph1 KO mice are comparable to those of WT control mice [8]. These data suggest that 5-HT, other than GDS, may play a role in regulating systemic energy homeostasis. Recently, two independent studies have highlighted the role of adipocyte-derived 5-HT in energy storage in the white adipose tissue (WAT) and energy expenditure in the BAT [11,46]. In a diet-induced mouse model of obesity, the

Tph1 expression and tissue 5-HT concentrations were elevated in adipose tissues [11,46]. Tph1 KO mice were protected from obesity and the related metabolic dysfunctions [46]. Tph1 KO mice gained significantly less weight and had lower adiposity when fed an HFD. The glycemic control is also improved in Tph1 KO mice, although the glucose uptake was similar in the muscle, liver and heart, indicating that the BAT make a major contribution to the increase in the basal metabolic rate [46]. Indeed, energy expenditure was enhanced in Tph1 KO mice when they were fed an HFD. The BAT activity increased in a Ucp1-dependent manner in Tph1 KO mice. The obesogenic actions of peripheral 5-HT were also confirmed using a peripheral TPH inhibitor (LP-533401) [11,46]. Furthermore, a cell autonomous effect of 5-HT in adipose tissue has been shown in adipocyte-specific Tph1 KO mice [11]. The adipocyte-specific Tph1 KO induced Ucp1 and Dio2 expression in the BAT and subcutaneous WAT [11].

Regarding brown fat thermogenesis, the 5-HT3 receptor plays a major role in diet-induced thermogenesis [11]. Diet-induced thermogenesis was robustly increased in the BAT of Htr3a KO mice fed a HFD [11]. In addition to the role of 5-HT in the BAT, in vitro experiments using 3T3-L1 adipocytes provided a hint about the role of 5-HT in adipogenesis [11,56]. It was known that 5-HT can increase the lipid accumulation in human and mouse fat cells [57]. It was also known that 5-HT2A receptor expression is increased in the hypertrophied 3T3L1 adipocytes and WAT of db/db mice, and the activation of the 5-HT2A receptor reduces adiponectin expression in hypertrophied 3T3L1 adipocytes [58]. Treatment with a 5-HT2A receptor antagonist increased the lipid accumulation and circulating adiponectin levels [11,59]. An antagonist for the 5-HT2A receptor inhibited adipogenesis [56]. Thus, 5-HT increases energy storage in the WAT through 5-HT2A receptor and inhibits energy expenditure in the BAT through 5-HT3 receptor. Taken together, these recent findings suggest that 5-HT negatively regulates the sensitivity of BAT to β -adrenergic stimulation.

GENETICS OF 5-HT AND OBESITY

To understand the genetic basis of obesity, many genome-wide association studies (GWAS) have been conducted. Several GWAS revealed genetic associations between the serotonergic system and obesity. The first association between TPH1 and obesity was identified through a GWAS in Koreans [60]. A genetic variant of TPH1 (rs623580) was associated with the body



Table 4. The results of genome-wide association studies on the	serotonergic system and obesity
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Study	Ethnicity	Sample size, n	SNP ID	Gene	Chromosome	Allele	Association
Kwak et al. (2012) [60]	Korean	8,842	rs623580	TPH1	11	A/T	BMI and WC
Rosmond et al.(2002) [61]	Swedish	284	rs6311	HTR2A	13	C/T	BMI, WHR
Halder et al. (2007) [62]	European	934	rs6311 rs2070040	HTR2A HTR2A	13 13	C/T A/G	Central obesity, metabolic syndrome
Li et al. (2014) [63]	European	1,982	rs912127	HTR2A	13	A/G	BMI
	American		rs1745837	HTR2A	13	C/T	
Opgen-Rhein et al. (2010) [64]	German	128	rs498207	HTR2C	X	C/T	Weight gain
Suviolahti et al. (2003) [65]	Finnish,	2,104	rs2011162	SLC6A14	X	C/G	BMI
	Swedish		rs2071877	SLC6A14	X	C/T	
Corpeleijn et al. (2010) [66]	European	722	rs2011162	SLC6A14	X	C/G	BMI, fat oxidation in women
Chen et al. (2013) [67]	Chinese	478	rs1018079	SLC18A1	8	A/G	BMI
			rs11214769	HTR3B	11	A/G	
			rs2224721	HTR2A	13	A/C	
			rs4911871	HTR2C	X	A/G	

SNP, single nucleotide polymorphism; BMI, body mass index; WC, waist circumference; WHR, waist to hip ratio.

mass index in a GWAS comprising 8,842 subjects (P=0.0047), and single nucleotide polymorphisms (SNPs) of TPH1 were significantly associated with weight gain during pregnancy in women with gestational diabetes mellitus. Several GWAS revealed associations between SNPs of the 5-HT receptors and obesity (Table 4) [60-67]. SNPs of HTR2A had strong associations with obesity in both European and Asian ethnic groups [61-63,67]. SNPs in the promoter (rs6311, -1438 [G/A]) and intron (rs2070040, 2416 [C/T]) were associated with metabolic syndrome in a study of 934 European subjects. Central obesity (a high waist circumference) showed a significant association with the -1438 GG genotype (odds ratio [OR], 1.57; *P*= 0.003), and the -1438 GG and 2416 TT genotypes were associated with an increased risk of metabolic syndrome (OR, 1.71; P=0.016) [62]. In 478 healthy Chinese subjects, 12 SNPs in seven genes (MAOB, SLC18A1, HTR2A, HTR2C, HTR3B, HTR4, and HTR7) were significantly associated with the body mass index [67]. These genetic studies indicate that the identification of serotonergic system defects can be used for risk prediction and to develop individualized treatments for obesity and metabolic syndrome.

CONCLUSIONS

The human body has two separate serotonergic systems: the

central 5-HT and peripheral 5-HT systems (Fig. 1). With regard to the central 5-HT system, increasing the 5-HT signaling has been utilized therapeutically to reduce body weight by reducing appetite. In 2010, the U.S. Food and Drug Administration approved a new selective 5-HT2C receptor agonist (Belvig, lorcaserin hydrochloride) for use in the treatment of obesity [26]. This drug was also approved by the Ministry of Food and Drug Substances in Korea. In peripheral tissues, suppressing 5-HT signaling might represent a new target for anti-obesity treatment by increasing energy expenditure and improving insulin resistance [11,46]. Systemic TPH1 inhibitors and a peripheral TPH1 inhibitor (LP-533401) have already been patented for treating diabetes and obesity [68,69]. However, there have not yet been any clinical trials using these drugs to treat obesity. Recent GWAS results support the clinical importance of serotonergic regulation in human metabolism, but most of these studies have small sample sizes and have not been validated. Therefore, to enhance our understanding of the role of 5-HT in metabolism and to provide effective therapeutic applications of 5-HT, more clinical studies are needed on drugs that can regulate the serotonergic system, and genetic studies are needed to investigate the associations between 5-HT and metabolic diseases.



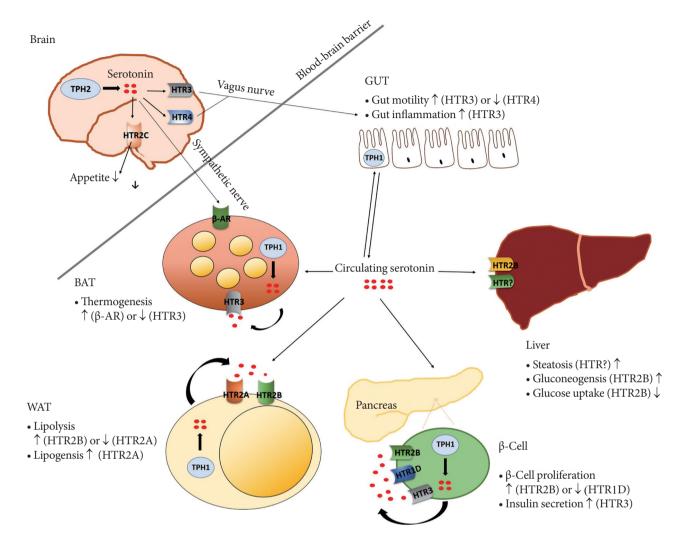


Fig. 1. Roles of 5-hydroxytryptamine (5-HT) on metabolic organs. Central 5-HT regulates appetite and controls metabolic organs through sympathetic/parasympathetic nervous system. Peripheral 5-HT regulates glucose metabolism, insulin resistance and energy expenditure in an autocrine/paracrine or endocrine manner. TPH, tryptophan hydroxylase; HTR, 5-HT receptor; β -AR, β -adrenergic receptor; BAT, brown adipose tissue; WAT, white adipose tissue.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

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REFERENCES

- 1. Janeway TC, Richardson HB, Park EA. Experiments on the vasoconstrictor action of blood serum. Arch Intern Med 1918; 21:565-603.
- Zucker MB. A study of the substances in blood serum and platelets which stimulate smooth muscle. Am J Physiol 1944; 142:12-26.
- Erspamer V, Asero B. Identification of enteramine, the specific hormone of the enterochromaffin cell system, as 5-hydroxytryptamine. Nature 1952;169:800-1.



- 4. Brodie BB, Shore PA. A concept for a role of serotonin and norepinephrine as chemical mediators in the brain. Ann N Y Acad Sci 1957;66:631-42.
- Wade PR, Chen J, Jaffe B, Kassem IS, Blakely RD, Gershon MD. Localization and function of a 5-HT transporter in crypt epithelia of the gastrointestinal tract. J Neurosci 1996;16:2352-64.
- Malek ZS, Dardente H, Pevet P, Raison S. Tissue-specific expression of tryptophan hydroxylase mRNAs in the rat midbrain: anatomical evidence and daily profiles. Eur J Neurosci 2005;22:895-901.
- Li Z, Chalazonitis A, Huang YY, Mann JJ, Margolis KG, Yang QM, Kim DO, Cote F, Mallet J, Gershon MD. Essential roles of enteric neuronal serotonin in gastrointestinal motility and the development/survival of enteric dopaminergic neurons. J Neurosci 2011;31:8998-9009.
- 8. Sumara G, Sumara O, Kim JK, Karsenty G. Gut-derived serotonin is a multifunctional determinant to fasting adaptation. Cell Metab 2012;16:588-600.
- 9. Ponicke K, Gergs U, Buchwalow IB, Hauptmann S, Neumann J. On the presence of serotonin in mammalian cardiomyocytes. Mol Cell Biochem 2012;365:301-12.
- 10. Kim H, Toyofuku Y, Lynn FC, Chak E, Uchida T, Mizukami H, Fujitani Y, Kawamori R, Miyatsuka T, Kosaka Y, Yang K, Honig G, van der Hart M, Kishimoto N, Wang J, Yagihashi S, Tecott LH, Watada H, German MS. Serotonin regulates pancreatic beta cell mass during pregnancy. Nat Med 2010;16:804-8.
- 11. Oh CM, Namkung J, Go Y, Shong KE, Kim K, Kim H, Park BY, Lee HW, Jeon YH, Song J, Shong M, Yadav VK, Karsenty G, Kajimura S, Lee IK, Park S, Kim H. Regulation of systemic energy homeostasis by serotonin in adipose tissues. Nat Commun 2015;6:6794.
- 12. Peroutka SJ, Howell TA. The molecular evolution of G protein-coupled receptors: focus on 5-hydroxytryptamine receptors. Neuropharmacology 1994;33:319-24.
- 13. Mohammad-Zadeh LF, Moses L, Gwaltney-Brant SM. Serotonin: a review. J Vet Pharmacol Ther 2008;31:187-99.
- 14. Barnes NM, Neumaier JF. Neuronal 5-HT receptors and SERT. Tocris Biosci Sci Rev Ser 2011;34:1-16.
- 15. Reeves DC, Lummis SC. The molecular basis of the structure and function of the 5-HT3 receptor: a model ligand-gated ion channel (review). Mol Membr Biol 2002;19:11-26.
- Lam DD, Heisler LK. Serotonin and energy balance: molecular mechanisms and implications for type 2 diabetes. Expert Rev Mol Med 2007;9:1-24.
- 17. Heal DJ, Aspley S, Prow MR, Jackson HC, Martin KF,

- Cheetham SC. Sibutramine: a novel anti-obesity drug. A review of the pharmacological evidence to differentiate it from d-amphetamine and d-fenfluramine. Int J Obes Relat Metab Disord 1998;22 Suppl 1:S18-28.
- 18. Heisler LK, Kanarek RB, Gerstein A. Fluoxetine decreases fat and protein intakes but not carbohydrate intake in male rats. Pharmacol Biochem Behav 1997;58:767-73.
- 19. Breisch ST, Zemlan FP, Hoebel BG. Hyperphagia and obesity following serotonin depletion by intraventricular p-chlorophenylalanine. Science 1976;192:382-5.
- Kennett GA, Curzon G. Evidence that hypophagia induced by mCPP and TFMPP requires 5-HT1C and 5-HT1B receptors: hypophagia induced by RU 24969 only requires 5-HT1B receptors. Psychopharmacology (Berl) 1988;96:93-100.
- Tecott LH, Sun LM, Akana SF, Strack AM, Lowenstein DH, Dallman MF, Julius D. Eating disorder and epilepsy in mice lacking 5-HT2c serotonin receptors. Nature 1995;374:542-6.
- Nonogaki K, Strack AM, Dallman MF, Tecott LH. Leptin-independent hyperphagia and type 2 diabetes in mice with a mutated serotonin 5-HT2C receptor gene. Nat Med 1998;4:1152-6.
- Heisler LK, Cowley MA, Tecott LH, Fan W, Low MJ, Smart JL, Rubinstein M, Tatro JB, Marcus JN, Holstege H, Lee CE, Cone RD, Elmquist JK. Activation of central melanocortin pathways by fenfluramine. Science 2002;297:609-11.
- 24. Smith SR, Weissman NJ, Anderson CM, Sanchez M, Chuang E, Stubbe S, Bays H, Shanahan WR; Behavioral Modification and Lorcaserin for Overweight and Obesity Management (BLOOM) Study Group. Multicenter, placebo-controlled trial of lorcaserin for weight management. N Engl J Med 2010;363: 245-56.
- CinicalTrials.gov: A service of the U.S. National Institutes of Health. Available from: https://www.clinicaltrials.gov (cited 2016 Jan 11).
- U.S. Food and Drug Administration. FDA approves Belviq to treat some overweight or obese adults. Home Healthc Nurse 2012;30:443-4.
- 27. Savelieva KV, Zhao S, Pogorelov VM, Rajan I, Yang Q, Cullinan E, Lanthorn TH. Genetic disruption of both tryptophan hydroxylase genes dramatically reduces serotonin and affects behavior in models sensitive to antidepressants. PLoS One 2008; 3:e3301.
- 28. Gutknecht L, Araragi N, Merker S, Waider J, Sommerlandt FM, Mlinar B, Baccini G, Mayer U, Proft F, Hamon M, Schmitt AG, Corradetti R, Lanfumey L, Lesch KP. Impacts of brain serotonin deficiency following Tph2 inactivation on develop-



- ment and raphe neuron serotonergic specification. PLoS One 2012;7:e43157.
- 29. Bouwknecht JA, van der Gugten J, Hijzen TH, Maes RA, Hen R, Olivier B. Male and female 5-HT(1B) receptor knockout mice have higher body weights than wildtypes. Physiol Behav 2001;74:507-16.
- Le Feuvre RA, Aisenthal L, Rothwell NJ. Involvement of corticotrophin releasing factor (CRF) in the thermogenic and anorexic actions of serotonin (5-HT) and related compounds. Brain Res 1991;555:245-50.
- 31. Sakaguchi T, Bray GA. Effect of norepinephrine, serotonin and tryptophan on the firing rate of sympathetic nerves. Brain Res 1989;492:271-80.
- 32. Berger M, Gray JA, Roth BL. The expanded biology of serotonin. Annu Rev Med 2009;60:355-66.
- Amireault P, Sibon D, Cote F. Life without peripheral serotonin: insights from tryptophan hydroxylase 1 knockout mice reveal the existence of paracrine/autocrine serotonergic networks. ACS Chem Neurosci 2013;4:64-71.
- 34. Gershon MD. Nerves, reflexes, and the enteric nervous system: pathogenesis of the irritable bowel syndrome. J Clin Gastroenterol 2005;39(5 Suppl 3):S184-93.
- Gregory RE, Ettinger DS. 5-HT3 receptor antagonists for the prevention of chemotherapy-induced nausea and vomiting. A comparison of their pharmacology and clinical efficacy. Drugs 1998;55:173-89.
- 36. Liang LX, Zhang Q, Qian W, Hou XH. Antinociceptive property of tegaserod in a rat model of chronic visceral hypersensitivity. Chin J Dig Dis 2005;6:21-5.
- Manocha M, Khan WI. Serotonin and GI disorders: an update on clinical and experimental studies. Clin Transl Gastroenterol 2012;3:e13.
- 38. O'Connell PJ, Wang X, Leon-Ponte M, Griffiths C, Pingle SC, Ahern GP. A novel form of immune signaling revealed by transmission of the inflammatory mediator serotonin between dendritic cells and T cells. Blood 2006;107:1010-7.
- 39. Ahern GP. 5-HT and the immune system. Curr Opin Pharmacol 2011;11:29-33.
- 40. Leon-Ponte M, Ahern GP, O'Connell PJ. Serotonin provides an accessory signal to enhance T-cell activation by signaling through the 5-HT7 receptor. Blood 2007;109:3139-46.
- 41. Murata S, Ohkohchi N, Matsuo R, Ikeda O, Myronovych A, Hoshi R. Platelets promote liver regeneration in early period after hepatectomy in mice. World J Surg 2007;31:808-16.
- 42. Matondo RB, Punt C, Homberg J, Toussaint MJ, Kisjes R, Kor-

- poraal SJ, Akkerman JW, Cuppen E, de Bruin A. Deletion of the serotonin transporter in rats disturbs serotonin homeostasis without impairing liver regeneration. Am J Physiol Gastrointest Liver Physiol 2009;296:G963-8.
- 43. Lesurtel M, Graf R, Aleil B, Walther DJ, Tian Y, Jochum W, Gachet C, Bader M, Clavien PA. Platelet-derived serotonin mediates liver regeneration. Science 2006;312:104-7.
- 44. Papadimas GK, Tzirogiannis KN, Panoutsopoulos GI, Demonakou MD, Skaltsas SD, Hereti RI, Papadopoulou-Daifoti Z, Mykoniatis MG. Effect of serotonin receptor 2 blockage on liver regeneration after partial hepatectomy in the rat liver. Liver Int 2006;26:352-61.
- 45. Tzirogiannis KN, Kourentzi KT, Zyga S, Papalimneou V, Tsironi M, Grypioti AD, Protopsaltis I, Panidis D, Panoutsopoulos GI. Effect of 5-HT7 receptor blockade on liver regeneration after 60-70% partial hepatectomy. BMC Gastroenterol 2014;14: 201.
- 46. Crane JD, Palanivel R, Mottillo EP, Bujak AL, Wang H, Ford RJ, Collins A, Blumer RM, Fullerton MD, Yabut JM, Kim JJ, Ghia JE, Hamza SM, Morrison KM, Schertzer JD, Dyck JR, Khan WI, Steinberg GR. Inhibiting peripheral serotonin synthesis reduces obesity and metabolic dysfunction by promoting brown adipose tissue thermogenesis. Nat Med 2015;21:166-72.
- 47. Moore MC, DiCostanzo CA, Dardevet D, Lautz M, Farmer B, Neal DW, Cherrington AD. Portal infusion of a selective serotonin reuptake inhibitor enhances hepatic glucose disposal in conscious dogs. Am J Physiol Endocrinol Metab 2004;287: E1057-63.
- 48. Haub S, Ritze Y, Ladel I, Saum K, Hubert A, Spruss A, Trautwein C, Bischoff SC. Serotonin receptor type 3 antagonists improve obesity-associated fatty liver disease in mice. J Pharmacol Exp Ther 2011;339:790-8.
- 49. Gershon MD, Ross LL. Location of sites of 5-hydroxytryptamine storage and metabolism by radioautography. J Physiol 1966;186:477-92.
- Ekholm R, Ericson LE, Lundquist I. Monoamines in the pancreatic islets of the mouse. Subcellular localization of 5-hydroxytryptamine by electron microscopic autoradiography. Diabetologia 1971;7:339-48.
- Kim K, Oh CM, Ohara-Imaizumi M, Park S, Namkung J, Yadav VK, Tamarina NA, Roe MW, Philipson LH, Karsenty G, Nagamatsu S, German MS, Kim H. Functional role of serotonin in insulin secretion in a diet-induced insulin-resistant state. Endocrinology 2015;156:444-52.
- 52. Ohara-Imaizumi M, Kim H, Yoshida M, Fujiwara T, Aoyagi K,



- Toyofuku Y, Nakamichi Y, Nishiwaki C, Okamura T, Uchida T, Fujitani Y, Akagawa K, Kakei M, Watada H, German MS, Nagamatsu S. Serotonin regulates glucose-stimulated insulin secretion from pancreatic beta cells during pregnancy. Proc Natl Acad Sci U S A 2013;110:19420-5.
- 53. Ohta Y, Kosaka Y, Kishimoto N, Wang J, Smith SB, Honig G, Kim H, Gasa RM, Neubauer N, Liou A, Tecott LH, Deneris ES, German MS. Convergence of the insulin and serotonin programs in the pancreatic beta-cell. Diabetes 2011;60:3208-16.
- 54. Berger M, Scheel DW, Macias H, Miyatsuka T, Kim H, Hoang P, Ku GM, Honig G, Liou A, Tang Y, Regard JB, Sharifnia P, Yu L, Wang J, Coughlin SR, Conklin BR, Deneris ES, Tecott LH, German MS. Galphai/o-coupled receptor signaling restricts pancreatic beta-cell expansion. Proc Natl Acad Sci U S A 2015;112: 2888-93.
- 55. Murphy DL, Lesch KP. Targeting the murine serotonin transporter: insights into human neurobiology. Nat Rev Neurosci 2008;9:85-96.
- 56. Kinoshita M, Ono K, Horie T, Nagao K, Nishi H, Kuwabara Y, Takanabe-Mori R, Hasegawa K, Kita T, Kimura T. Regulation of adipocyte differentiation by activation of serotonin (5-HT) receptors 5-HT2AR and 5-HT2CR and involvement of microRNA-448-mediated repression of KLF5. Mol Endocrinol 2010;24:1978-87.
- 57. Gres S, Canteiro S, Mercader J, Carpene C. Oxidation of high doses of serotonin favors lipid accumulation in mouse and human fat cells. Mol Nutr Food Res 2013;57:1089-99.
- 58. Uchida-Kitajima S, Yamauchi T, Takashina Y, Okada-Iwabu M, Iwabu M, Ueki K, Kadowaki T. 5-Hydroxytryptamine 2A receptor signaling cascade modulates adiponectin and plasminogen activator inhibitor 1 expression in adipose tissue. FEBS Lett 2008;582:3037-44.
- 59. Nomura S, Shouzu A, Omoto S, Nishikawa M, Iwasaka T. 5-HT2A receptor antagonist increases circulating adiponectin in patients with type 2 diabetes. Blood Coagul Fibrinolysis 2005;16:423-8.
- 60. Kwak SH, Park BL, Kim H, German MS, Go MJ, Jung HS, Koo BK, Cho YM, Choi SH, Cho YS, Shin HD, Jang HC, Park KS. Association of variations in TPH1 and HTR2B with gestational weight gain and measures of obesity. Obesity (Silver Spring)

- 2012;20:233-8.
- 61. Rosmond R, Bouchard C, Bjorntorp P. Increased abdominal obesity in subjects with a mutation in the 5-HT(2A) receptor gene promoter. Ann N Y Acad Sci 2002;967:571-5.
- 62. Halder I, Muldoon MF, Ferrell RE, Manuck SB. Serotonin receptor 2A (HTR2A) gene polymorphisms are associated with blood pressure, central adiposity, and the metabolic syndrome. Metab Syndr Relat Disord 2007;5:323-30.
- 63. Li P, Tiwari HK, Lin WY, Allison DB, Chung WK, Leibel RL, Yi N, Liu N. Genetic association analysis of 30 genes related to obesity in a European American population. Int J Obes (Lond) 2014;38:724-9.
- 64. Opgen-Rhein C, Brandl EJ, Muller DJ, Neuhaus AH, Tiwari AK, Sander T, Dettling M. Association of HTR2C, but not LEP or INSIG2, genes with antipsychotic-induced weight gain in a German sample. Pharmacogenomics 2010;11:773-80.
- 65. Suviolahti E, Oksanen LJ, Ohman M, Cantor RM, Ridderstrale M, Tuomi T, Kaprio J, Rissanen A, Mustajoki P, Jousilahti P, Vartiainen E, Silander K, Kilpikari R, Salomaa V, Groop L, Kontula K, Peltonen L, Pajukanta P. The SLC6A14 gene shows evidence of association with obesity. J Clin Invest 2003;112: 1762-72.
- 66. Corpeleijn E, Petersen L, Holst C, Saris WH, Astrup A, Langin D, MacDonald I, Martinez JA, Oppert JM, Polak J, Pedersen O, Froguel P, Arner P, Sorensen TI, Blaak EE. Obesity-related polymorphisms and their associations with the ability to regulate fat oxidation in obese Europeans: the NUGENOB study. Obesity (Silver Spring) 2010;18:1369-77.
- 67. Chen C, Chen W, Chen C, Moyzis R, He Q, Lei X, Li J, Wang Y, Liu B, Xiu D, Zhu B, Dong Q. Genetic variations in the serotoninergic system contribute to body-mass index in Chinese adolescents. PLoS One 2013;8:e58717.
- 68. Karsenty G, Sumara G, Sumara O, inventors; The Trustees of Columbia University in the City of New York, assignee. Methods of preventing and treating diabetes by inhibiting serotonin synthesis. United States patent US 9,150,521. 2013 Aug 8.
- Khan W, Steinberg G, Palanivel R, inventors; Mcmaster University, assignee. A method of treating obesity. Canada patent CA 2900413A1. 2014 Aug 21.