

Article

# EPR Spectroscopy: A Powerful Tool to Analyze Supramolecular Host•Guest Complexes of Stable Radicals with Cucurbiturils

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Abstract: Stable organic free radicals are increasingly studied compounds due to the multiple and unusual properties imparted by the single electron(s). However, being paramagnetic, classical methods such as NMR spectroscopy can hardly be used due to relaxation and line broadening effects. EPR spectroscopy is thus better suited to get information about the immediate surroundings of the single electrons. EPR has enabled obtaining useful data in the context of host•guest chemistry, and a classical example is reported here for the stable (2,2,6,6-tetramethyl-4-oxo-piperidin-1-yl)oxyl or 4-oxo-TEMPO nitroxide (TEMPONE) inside the macrocycle host cucurbit[7]uril (CB[7]). Generally and also observed here, a contraction of the spectrum is observed as a result of the reduced nitrogen coupling constant due to inclusion complexation in the hydrophobic cavity of the host. Simulations of EPR spectra allowed determining the corresponding binding constant pointing to a weaker affinity for CB[7], compared to TEMPO with CB[7]. We complement this work by the results of EPR spectroscopy of a biradical: bis-TEMPO-bis-ketal (bTbk) with cucurbit[8]uril (CB[8]). Initial investigations pointed to very weak effects on the spectrum of the guest and incorrectly led us to conclude an absence of binding. However, simulations of EPR spectra combined with NMR data of reduced bTbk allowed showing inclusion complexation. EPR titrations were performed, and the corresponding binding constant was determined. <sup>1</sup>H NMR spectra with reduced bTbk suggested a shuttle mechanism, at nearly one equivalent of CB[8], for which the host moves rapidly between two stations.

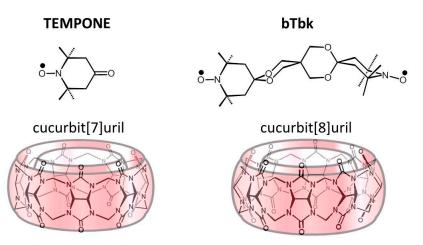
**Keywords:** EPR spectroscopy; supramolecular chemistry; host•guest chemistry; nitroxides; cucurbiturils

## 1. Introduction

Since the discovery of the first organic stable radical by Gomberg in 1900 [1], radical chemistry has dramatically expanded with initial intense focus on mastering the structures [2] and the life times [3] of transient species before using transient or stable free radicals in applications ranging from living polymerization [4], to batteries [5], or more recently, for dynamic nuclear polarization (DNP) [6,7]. Free radicals are intimately related to EPR spectroscopy since this technique is specially focused on investigating the single electron(s) carried by several types of compounds. EPR spectra contain a



wealth of information, which, correctly extracted, can give users access to crucial information such as stability, structure, or dynamics, in various liquid or solid environments [8]. Supramolecular chemistry principally features diamagnetic compounds, but paramagnetic ones are expected to enable access to new types of applications including molecular magnetic switches [9], dynamic covalent systems [10], or tracers for imaging by stabilizing free radicals in reducing conditions. In the context of host•guest chemistry, the guests are almost exclusively [11] the radicals, and they are included in macrocyclic hosts. Several reviews [12–14] have already documented host•guest complexes featuring free radicals and several families of hosts including calixarenes, cyclodextrins, and cucurbiturils. This latter family of pumpkin-shape macrocycles [15–19] possess unique properties, in a broader context, such as ultrahigh binding [20,21], gas adsorption [22], or drug encapsulation and release [23,24]. With cucurbit[n]urils (CB[*n*]), the main radicals studied are nitroxides [25], in the majority of cases stable ones. In a seminal work, Lucarini and coworkers studied the cucurbit[7]uril (CB[7]) cavity by EPR spectroscopy using several nitroxides [26]. Later, Kaifer and coworkers showed how CB[n] could modulate the extent of spin exchange between covalently linked nitroxides [27]. We also discovered in 2009 [28], thanks to EPR spectroscopy, the formation of triangular 3:3 host:guest assemblies of 1:1 CB[8]:nitroxide complexes, at the same time as Lucarini and coworkers [29] and the teams of Ottaviani, Ramamurthy, and Turro [30]. EPR spectroscopy was key in this finding since it gave a clear spectral signature of the triangular assembly that could hardly be found by other techniques. We recently showed that, provided the guest possesses the right structure, diamagnetic triangles of CB[8] could also be produced and hence studied by NMR [31]. Since CB[n] are almost exclusively soluble in water, nitroxide guest recognition is monitored using EPR by comparing the spectrum of the radical alone to that in the presence of the host in aqueous solutions. Often, a contraction of the spectrum is observed as a result of the guest experiencing a more hydrophobic environment upon inclusion in CB[n] (reduced nitrogen coupling constant  $a_N$ ), and the high-field line is broadened due to reduced tumbling (i.e., reduced mobility because of a larger molecular weight) [12–14,26]. This way, inclusion of the nitroxide function carrying the single-electron in the host cavity is believed to be important to obtain significant changes in EPR spectra. Here, we illustrate this concept using the 4-oxo-TEMPO (or TEMPONE, Scheme 1) nitroxide with CB[7].



Scheme 1. Structures of nitroxides TEMPONE and bis-TEMPO-bis-ketal (bTbk), and of cucurbiturils CB[7] and CB[8].

This host•guest system has been briefly investigated before [32] in the context of improved resistance to bioreduction provided by CB[n] and unusual rotational dynamics, but with a simple mention for the order of magnitude of the binding constant (~10<sup>3</sup> M<sup>-1</sup>). We describe here the EPR titration and simulations of EPR spectra allowing determining the binding constant. On the other hand, dinitroxide biradicals exhibiting spin exchange or not have started to be widely investigated for applications in DNP [6,7], and cyclodextrins proved to be interesting macrocycles to modulate

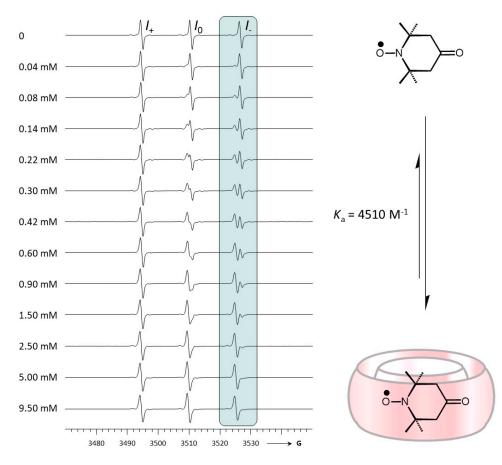
polarization transfer [33]. In this context, we wanted to see if inclusion complexation could occur between a cucurbituril and a representative dinitroxide used for DNP: the stable dinitroxide of TEMPO, **bTbk** [34], to enhance its water solubility for perspective work in DNP.

#### 2. Results

The first case investigated is the one of the nitroxide **TEMPONE** for which the corresponding EPR spectrum is known to exhibit three sharp lines.

## 2.1. Complexation between TEMPONE and CB[7]

The EPR spectrum of **TEMPONE** in water is characterized by three main lines with a width at half-height (Wahh) of ~0.8 G (Figure 1). Two small <sup>13</sup>C satellite lines can be seen just next to each of the three main lines as a result of a weak coupling with isotopically rare <sup>13</sup>C (nuclear spin = 1/2).



**Figure 1.** EPR titration of **TEMPONE** (0.1 mM) with CB[7] (concentration on the left) in water and proposed inclusion complexation (right).

Twenty-five spectra were recorded with increased concentrations of CB[7] illustrating the gradual inclusion of **TEMPONE** in CB[7] (Figure 1 and Figures S1 to S25). The presence of additional lines especially at high field for intermediate host:guest ratios indicated slow exchange with respect to the EPR timescale. Above 5 mM of CB[7], no further change could be observed, and the spectrum was that of the included radical featuring reduced  $a_N$  (from ~16.0 G for **TEMPONE** alone to ~15.3 G in the complex) typical of a switch to a more hydrophobic surrounding. These results were in line with previous reports of TEMPO in CB[7] suggesting perpendicular position of the nitroxide group with respect to the  $C_7$  axis of CB[7] [26,28]. All the spectra were then simulated thanks to a 2D EPR program [35] (Figures S1 to S28) enabling testing several hypotheses of binding. The simplest case

assuming a host:guest 1:1 binding afforded a binding constant  $K_a = 4510 \text{ M}^{-1}$  (Table 1) with a regression coefficient of 0.9918.

|                         | TEMPONE                       | TEMPONE•CB[7]                 |
|-------------------------|-------------------------------|-------------------------------|
| g                       | 2.0055 <sup>a</sup>           | 2.0060 <sup>a</sup>           |
| $a_{\rm N}/{\rm G}$     | $15.99 \pm 0.05$ <sup>b</sup> | $15.29 \pm 0.05$ <sup>b</sup> |
| a <sub>C</sub> /G       | $6.0 \pm 0.5$ <sup>b</sup>    | $4.7 \pm 0.5$ <sup>b</sup>    |
| Wahh/G                  | $0.82 \pm 0.05$ <sup>a</sup>  | $0.96 \pm 0.06$ <sup>a</sup>  |
| $I_{-}/I_{0}$           | 0.92 <sup>a</sup>             | 0.85 <sup>a</sup>             |
| $K_{\rm a}/{ m M}^{-1}$ | -                             | $4510 \pm 500^{\text{ b}}$    |

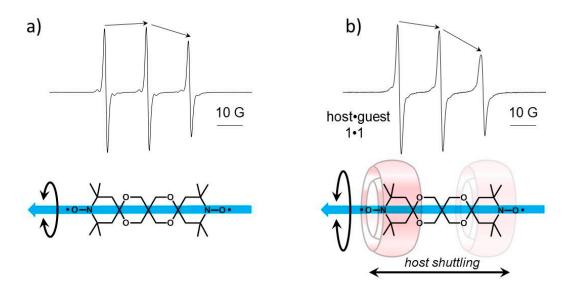
 Table 1. EPR parameters for TEMPONE and the host:guest 1:1 complex. Wahh, width at half-height.

<sup>a</sup> From experimental spectra. <sup>b</sup> From simulations.

Controls using reduced **TEMPONE** with CB[7] by <sup>1</sup>H NMR afforded a similar binding constant of  $K_a = 3960 \text{ M}^{-1}$  (slightly weaker due to the more polar feature expected for the hydroxylamine of reduced **TEMPONE**, Figure S29). However, simulated EPR spectra at high CB[7] concentrations showed better agreement with experimental ones when a 2:1 complex was considered (with two CB[7]). Previous reports mentioned the possibility for cucurbiturils to aggregate in water [28–31,36–40], but there is only a slight improvement in the agreement between simulated spectra and experimental ones (regression coefficient of 0.9955) assuming this model; other techniques would be necessary to confirm this possibility. As mentioned above, a decrease in  $a_N$  was associated with a more hydrophobic surrounding for the nitroxide group, while the observed decrease in  $a_C$  also indicated a redistribution of spin densities on the corresponding ring, together with a probable freeze in dihedral angles due to the inclusion.

#### 2.2. Complexation between bTbk and CB[8]

In **bTbk**, the two TEMPO residues carrying the single electrons are too far for spin exchange, and they thus behave magnetically independently, giving rise to EPR spectra with three lines as for TEMPO alone (Figure 2a). **bTbk** is still an excellent polarizing agent for DNP-enhanced applications, but is hardly soluble in water [34]. Cyclodextrins have previously been used to improve the solubility of **bTbk** for DNP applications [33], and we reasoned that CB[7] or CB[8] could be another interesting option for this purpose.



**Figure 2.** EPR spectra of **bTbk** (50  $\mu$ M) without (**a**) and with CB[8] (1.1 mM, (**b**) and the proposed host position and shuttling based on EPR and NMR (see the text).

However, **bTbk** is a larger molecule compared to **TEMPONE**, and a different binding mode is expected for its association with CB[7] or CB[8]. While CB[7] showed almost undetectable effects, the addition of CB[8] slightly impacted the EPR spectrum of **bTbk** in water. The  $a_N$  coupling constant was slightly reduced (by an ~0.17 G average value, Table 2), in stark contrast with the usual behaviors (i.e.,  $\Delta a_N = 0.7$  G for **TEMPONE** with CB[7]).

**Table 2.** EPR parameters for **bTbk** and for the **bTbk**•CB[8] complex. Binding constants based on simulations of EPR spectra (see the text).

|  | bTbk                          | bTbk•CB[8]                                      |
|--|-------------------------------|---|
| g  | 2.0055 <sup>a</sup>           | 2.0056 <sup>a</sup>                             |
| $a_{\rm N}/{ m G}$                       | $16.59 \pm 0.05$ <sup>b</sup> | $16.25\pm0.05$ and $16.59\pm0.05$ $^{\rm b}$    |
| $a_{\rm C}/{\rm G}$                      | $5.8 \pm 0.5$ <sup>b</sup>    | $6.0\pm0.5$ and $5.8\pm0.5$ $^{ m b}$           |
| Wahh/G                                   | $1.18 \pm 0.05^{a}$           | $1.44 \pm 0.10^{a}$                             |
| I_/I_0                                   | 0.78 <sup>a</sup>             | 0.67 <sup>a</sup>                               |
| $K_{a \ 1:1 \ model}/M^{-1}$             | -                             | $36\ 600\ \pm 5000\ \mathrm{M}^{-1,\mathrm{b}}$ |
| $K_{\rm a shuttling model}/{\rm M}^{-1}$ | -                             | $20\ 900 \pm 5000\ M^{-1,b}$                    |

<sup>a</sup> From experimental spectra. <sup>b</sup> From simulations (see Supporting Information).

This small reduction of  $a_N$  upon CB[n] addition initially led us to conclude an absence of binding or a weak external binding. However, the slight broadening of the high-field line was significant as the  $L/I_0$  ratio (Table 2), but we could not explain it until we realized that the inclusion of **bTbk** in CB[8] would leave the EPR responsive N-O• bonds significantly bulk exposed, because the TEMPO fragments cannot be included as for **TEMPONE** in the **TEMPONE**•CB[7] complex. Because **bTbk** is too large to be fully included in a CB, the guest has to orientate such that the TEMPO fragments of **bTbk** bind in the CB cavity with the nitroxide group parallel to the  $C_8$  symmetry axis of the host. In this geometry (Figure 2b), the N-O• bonds are not facing the cavity walls anymore and become able to interact with water molecules, even if the guest is included in the host. In this particular case, the best evidence comes from the broader high-field line of the EPR spectrum (Figure 2b), suggesting a slower molecular tumbling caused by a larger molecular weight.

By analyzing a series of 36 EPR spectra of **bTbk** with increasing host concentrations (Figures S30 to S68) by the same 2D simulation program as that used for TEMPONE [35], we could determine the respective concentrations of each species at relevant host concentrations and evaluate the corresponding binding constant ( $K_a$  1:1 model in Table 2). In this first model, averaged effects were assumed (two TEMPO groups with the same parameters). However, <sup>1</sup>H NMR spectra of reduced **bTbk** suggested a shuttling process for CB[8] (Figure S69). Indeed, <sup>1</sup>H NMR spectroscopy of **bTbk** (0.5 mM) with excess ascorbic acid (used to reduce the N-O• bonds into diamagnetic N-OH bonds) showed that, with 1.25 equiv. of CB[8], the signals of all methyl groups were shifted upfield by 0.29 ppm (Figure S69). This was in line with the slightly reduced  $a_N$  value of **bTbk** observed with CB[8] ( $\langle \Delta a_N \rangle = 0.17$  G). For binding the two reduced TEMPO fragments, one would expect a necessary quantity of two CB[8]. Because only 1.25 equiv. of CB[8] was enough to shield the signals of all methyl groups, we proposed a 1:1 binding mode in which CB[8] would shuttle between two TEMPO stations (Figure 2b). <sup>1</sup>H NMR spectra at 2 equiv. of CB[8] were identical to those at 1.25 equiv. of CB[8] (Figure S69). Hence, even if with 1 equiv. of CB[8], a shuttle movement of CB[8] was suspected; with 2 equiv. of the host, a 1:2 complex was possible (Figure S69). However, in the conditions used to record EPR spectra ([**bTbk**] = 0.05 mM), 1:1 complexes mainly formed.

We thus had to consider another model for simulations of EPR spectra. In a second model, the first TEMPO group had EPR parameters fixed and corresponding to non-included TEMPO, and the second TEMPO group had its parameters fitted (complexation by CB[8], overall regression parameter 0.9959). With the second "shuttle" model (Table 2), the obtained binding constant of 20 900  $M^{-1}$  was consistent with the value obtained by NMR. Indeed, the NMR spectrum at ~1.1 equiv. of CB[8] (Figure

S69) showed signals for methyl groups assigned to free guest and the 1:1 complex in a ratio of  $\sim$ 5/19 respectively (79% of complexed guest). Assuming that the binding constant of reduced **bTbk** was similar to that of **bTbk** ( $K_a \approx 20\ 900\ M^{-1}$ ), this would give 77% respectively of 1:1 complex in the conditions of NMR, while the value determined from <sup>1</sup>H NMR was 79%. The proposed binding constant of **bTbk** toward CB[8] determined by EPR was consistent with <sup>1</sup>H NMR spectra of reduced **bTbk**. Controls by EPR of **bTbk** with CB[8] using a competitor (amantadine hydrochloride) that was known to have a very high affinity for CB[8] showed free **bTbk** in solution (Figure S70), supporting an associative mechanism of **bTbk** with CB[8] in the absence of competitor. Finally, simulations provided relaxation parameters  $\alpha$ ,  $\beta$ , and  $\gamma$  (see Supporting Information), which were related to the tumbling of the molecule. They were determined from the time dependent perturbation theory [41,42] from the anisotropy of g and  $a_N$  hyperfine tensors.  $\alpha$  depends on the anisotropy of g,  $\beta$  on the product of g and  $a_N$  tensors, while  $\gamma$  is determined from the anisotropy of the  $a_N$  tensor [41,42]. The rotational tumbling can average the tensors, but the scatter of resonance position can produce line broadening expressed by the relaxation parameters. If the rotational tumbling becomes asymmetric, averaging of tensors will produce different values, which strongly influences the broadening effect. In this case, compared to the free guest (or free TEMPO group), relaxation parameters for the uncomplexed TEMPO group did not change much except for  $\alpha$ , which was about doubled. However, significant differences were found for the complexed TEMPO group. First, a decrease in  $a_N$  of 0.34 G was observed, typical of a nitroxide inclusion in CB[8] with the N-O• bond relatively bulk exposed [31]. Then, while the  $\alpha$ value increased by ~50%,  $\beta$  and  $\gamma$  were multiplied by a factor of ~5, suggesting a tumbling on this side, largely different from the uncomplexed side. The presence of CB[8] surrounding the complexed TEMPO group expectedly hampered movements on this side of **bTbk**, while the tumbling remained about similar on the other side (totally bulk-exposed TEMPO group).

#### 3. Discussion

The association of **TEMPONE** with CB[7] is typical of the "general case" we have frequently observed for which the nitrogen coupling constant is reduced as a result of the nitroxide bond experiencing a more hydrophobic surrounding. We observed this behavior for a variety of nitroxides in the presence of several types of cyclodextrins [43–45] and of cucurbiturils [28,31,39]. Besides kinetic [46] and thermodynamic [47] parameters, which can sometimes be extracted from EPR experiments, binding constants are important parameters to assess the affinity of a guest for a host. The binding constant of **TEMPONE** for CB[7] ( $K_a = 4510 \text{ M}^{-1}$ ) was lower than that of TEMPO for CB[7], which was ~10<sup>4</sup> M<sup>-1</sup> [26,28]. As both guests bonded in CB[7] with the nitroxide groups perpendicular to the  $C_7$  axis of the host, the lower value determined for **TEMPONE** could be ascribed to the ketone function, which may be more difficult to desolvate to enter the cavity of CB[7] compared to the corresponding -CH<sub>2</sub>-group of TEMPO.

Contrary to **TEMPONE**, **bTbk** is a much larger molecule that cannot be entirely complexed by CB[7] or CB[8]. When one TEMPO fragment enters a sufficiently large CB cavity (at least that of CB[8]), the remaining bis-ketal-TEMPO part remains bulk-exposed, but not only (Figure 2b). The N-O• bond of the included TEMPO fragment is also exposed to water because, due to the bulkiness of the bis-ketal-TEMPO remaining group, the included N-O• bond cannot be placed like that of **TEMPONE** in CB[7] (i.e., facing the host cavity walls). This was the reason for the only slight effects observed in EPR spectra. The largest changes in these cases were expected to be due to reduced  $a_N$  coupling constants, but these were very sensitive to solvent polarity. If the two N-O• bonds remained bulk-exposed, then no large changes could be expected besides an increase in molecular weight and lower tumbling. This indicated that, slight effects on EPR spectra of paramagnetic guests after the addition of relevant hosts did not necessarily mean a lack of binding, and researchers should be cautious when analyzing EPR spectra. The solubility of **bTbk** in water was around 50  $\mu$ M, and CB[8] did not increase it very largely. However, these investigations allowed finding that molecular shuttles using cucurbiturils

were not limited to CB[6] [48] or CB[7] [49–51] and that CB[8] could also behave as a functional ring in water-soluble molecular shuttles provided the guest had the right structure.

## 4. Materials and Methods

The methods and protocols used in this work are mentioned in this paper and referred to in the References section. More details can be found in the Supporting Information.

## 5. Conclusions

The first case investigated (binding of **TEMPONE** in CB[7]) was efficiently monitored by EPR spectroscopy and allowed to determine a previously undescribed binding constant of  $K_a = 4510 \text{ M}^{-1}$ . Additionally, EPR allowed identifying a new supramolecular complex between **bTbk** and CB[8], initially thought to be weak or not forming, due to the very slight changes observed on EPR spectra upon host addition. EPR titrations and simulations of EPR spectra allowed proposing a binding constant  $K_a = 20 900 \text{ M}^{-1}$ . <sup>1</sup>H NMR of the reduced radical (reduced **bTbk**) suggested a shuttling mechanism, which was accounted for in simulations of EPR spectra. Besides the modulation of the properties imparted by the single electron of the N-O• bonds, we believe that this kind of complex could have relevance for investigating the dynamics of molecular machines [52,53] such as supramolecular rotors and shuttles.

Supplementary Materials: Simulations of EPR spectra and <sup>1</sup>H NMR spectra are available online.

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#### References

- 1. Gomberg, M. An instance of trivalent carbon: Triphenylmethyl. J. Am. Chem. Soc. 1900, 22, 757–771. [CrossRef]
- 2. Tidwell, T.T. Triarylmethyl and related radicals. In *Stable Radicals: Fundamentals and Applied Aspects of Odd-Electron Compounds*, 1st ed.; Hicks, R.G., Ed.; John Wiley & Sons Ltd.: Chichester, UK, 2010; pp. 1–31.
- 3. Keana, J.F. Newer aspects of the synthesis and chemistry of nitroxide spin labels. *Chem. Rev.* **1978**, *78*, 37–64. [CrossRef]
- 4. Nicolas, J.; Guillaneuf, Y.; Lefay, C.; Bertin, D.; Gigmes, D.; Charleux, B. Nitroxide-mediated polymerization. *Prog. Polym. Sci.* 2013, *38*, 63–235. [CrossRef]
- 5. Nakahara, K.; Oyaizu, K.; Nishide, H. Organic radical battery approaching practical use. *Chem. Lett.* **2011**, 40, 222–227. [CrossRef]
- Wisser, D.; Karthikeyan, G.; Lund, A.; Casano, G.; Karoui, H.; Yulikov, M.; Menzildjian, G.; Pinon, A.C.; Purea, A.; Engelke, F.; et al. BDPA-Nitroxide Biradicals Tailored for Efficient Dynamic Nuclear Polarization Enhanced Solid-State NMR at Magnetic Fields up to 21.1 T. *J. Am. Chem. Soc.* 2018, 140, 13340–13349. [CrossRef] [PubMed]
- Casano, G.; Karoui, H.; Ouari, O. Polarizing agents: Evolution and outlook in free radical development for DNP. *eMagRes* 2018, 7, 195–208.

- Altenbach, C.; López, C.J.; Hideg, K.; Hubbell, W.L. Exploring Structure, Dynamics, and Topology of Nitroxide Spin-Labeled Proteins Using Continuous-Wave Electron Paramagnetic Resonance Spectroscopy. In *Methods in Enzymology*; Academic Press: Cambridge, MA, USA, 2015; Volume 564, Part B, pp. 59–100. [CrossRef]
- 9. Buck, A.T.; Paletta, J.T.; Khindurangala, S.A.; Beck, C.L.; Winter, A.H. A Noncovalently Reversible Paramagnetic Switch in Water. *J. Am. Chem. Soc.* **2013**, *135*, 10594–10597. [CrossRef]
- 10. Schulte, B.; Tsotsalas, M.; Becker, M.; Studer, A.; De Cola, L. Dynamic Microcrystal Assembly by Nitroxide Exchange Reactions. *Angew. Chem. Int. Ed.* **2010**, *49*, 6881–6884. [CrossRef]
- 11. Nakabayashi, K.; Ozaki, Y.; Kawano, M.; Fujita, M. A Self-Assembled Spin Cage. *Angew. Chem. Int. Ed.* 2008, 47, 2046–2048. [CrossRef]
- 12. Lucarini, M. Supramolecular Radical Chemistry. In *Encyclopedia of Radicals in Chemistry, Biology and Materials;* John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2012; pp. 1–20. [CrossRef]
- Lucarini, M.; Mezzina, E. EPR investigations of organic non-covalent assemblies with spin labels and spin probes. In *Electron Paramagnetic Resonance*; RSC publishing: London, UK, 2011; Volume 22, pp. 41–70. [CrossRef]
- 14. Bardelang, D.; Hardy, M.; Ouari, O.; Tordo, P. Spin Labels and Spin Probes. In *Encyclopedia of Radicals in Chemistry, Biology and Materials*; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2012; pp. 1–51. [CrossRef]
- Lagona, J.; Mukhopadhyay, P.; Chakrabarti, S.; Isaacs, L. The Cucurbit[n]uril Family. *Angew. Chem. Int. Ed.* 2005, 44, 4844–4870. [CrossRef]
- 16. Lee, J.W.; Samal, S.; Selvapalam, N.; Kim, H.-J.; Kim, K. Cucurbituril Homologues and Derivatives: New Opportunities in Supramolecular Chemistry. *Acc. Chem. Res.* **2003**, *36*, 621–630. [CrossRef] [PubMed]
- 17. Barrow, S.J.; Kasera, S.; Rowland, M.J.; del Barrio, J.; Scherman, O.A. Cucurbituril-Based Molecular Recognition. *Chem. Rev.* 2015, 115, 12320–12406. [CrossRef] [PubMed]
- Assaf, K.I.; Nau, W.M. Cucurbiturils: From synthesis to high-affinity binding and catalysis. *Chem. Soc. Rev.* 2015, 44, 394–418. [CrossRef] [PubMed]
- 19. Masson, E.; Ling, X.; Joseph, R.; Kyeremeh-Mensah, L.; Lu, X. Cucurbituril chemistry: A tale of supramolecular success. *RSC Adv.* **2012**, *2*, 1213–1247. [CrossRef]
- 20. Cao, L.; Sekutor, M.; Zavalij, P.Y.; Mlinaric-Majerski, K.; Glaser, R.; Isaacs, L. Cucurbit[7]uril·Guest Pair with an Attomolar Dissociation Constant. *Angew. Chem. Int. Ed.* **2014**, *53*, 988–993. [CrossRef]
- 21. Rekharsky, M.V.; Mori, T.; Yang, C.; Ko, Y.H.; Selvapalam, N.; Kim, H.; Sobransingh, D.; Kaifer, A.E.; Liu, S.; Isaacs, L.; et al. A synthetic host-guest system achieves avidin-biotin affinity by overcoming enthalpy–entropy compensation. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 20737–20742. [CrossRef]
- 22. Kim, H.; Kim, Y.; Yoon, M.; Lim, S.; Park, S.M.; Seo, G.; Kim, K. Highly Selective Carbon Dioxide Sorption in an Organic Molecular Porous Material. *J. Am. Chem. Soc.* **2010**, *132*, 12200–12202. [CrossRef]
- 23. Yin, H.; Wang, R. Applications of Cucurbit[*n*]urils (*n* = 7 or 8) in Pharmaceutical Sciences and Complexation of Biomolecules. *Isr. J. Chem.* **2018**, *58*, 188–198. [CrossRef]
- 24. Walker, S.; Oun, R.; McInnes, F.J.; Wheate, N.J. The Potential of Cucurbit[*n*]urils in Drug Delivery. *Isr. J. Chem.* **2011**, *51*, 616–624. [CrossRef]
- 25. Ouari, O.; Bardelang, D. Nitroxide Radicals with Cucurbit[*n*]urils and Other Cavitands. *Isr. J. Chem.* **2018**, *58*, 343–356. [CrossRef]
- 26. Mezzina, E.; Cruciani, F.; Pedulli, G.F.; Lucarini, M. Nitroxide Radicals as Probes for Exploring the Binding Properties of the Cucurbit[7]uril Host. *Chem. Eur. J.* **2007**, *13*, 7223–7233. [CrossRef] [PubMed]
- 27. Yi, S.; Captain, B.; Ottaviani, M.F.; Kaifer, A.E. Controlling the Extent of Spin Exchange Coupling in 2,2,6,6-Tetramethylpiperidine-1-oxyl (TEMPO) Biradicals via Molecular Recognition with Cucurbit[*n*]uril Hosts. *Langmuir* **2011**, *27*, 5624–5632. [CrossRef] [PubMed]
- Bardelang, D.; Banaszak, K.; Karoui, H.; Rockenbauer, A.; Waite, M.; Udachin, K.; Ripmeester, J.A.; Ratcliffe, C.I.; Ouari, O.; Tordo, P. Probing Cucurbituril Assemblies in Water with TEMPO like Nitroxides: A Trinitroxide Supraradical with Spin–Spin Interactions. J. Am. Chem. Soc. 2009, 131, 5402–5404. [CrossRef] [PubMed]
- Mileo, E.; Mezzina, E.; Grepioni, F.; Pedulli, G.F.; Lucarini, M. Preparation and Characterisation of a New Inclusion Compound of Cucurbit[8]uril with a Nitroxide Radical. *Chem.-Eur. J.* 2009, 15, 7859–7862. [CrossRef] [PubMed]

- Jayaraj, N.; Porel, M.; Ottaviani, M.F.; Maddipatla, M.V.S.N.; Modelli, A.; Da Silva, J.P.; Bhogala, B.R.; Captain, B.; Jockusch, S.; Turro, N.J.; et al. Self Aggregation of Supramolecules of Nitroxides@Cucurbit[8]uril Revealed by EPR Spectra. *Langmuir* 2009, 25, 13820–13832. [CrossRef]
- 31. Combes, S.; Tran, K.T.; Ayhan, M.M.; Karoui, H.; Rockenbauer, A.; Tonetto, A.; Monnier, V.; Charles, L.; Rosas, R.; Viel, S.; et al. Triangular Regulation of Cucurbit[8]uril 1:1 Complexes. *J. Am. Chem. Soc.* **2019**, 141, 5897–5907. [CrossRef]
- Casano, G.; Poulhès, F.; Tran, T.K.; Ayhan, M.M.; Karoui, H.; Siri, D.; Gaudel-Siri, A.; Rockenbauer, A.; Jeschke, G.; Bardelang, D.; et al. High binding yet accelerated guest rotation within a cucurbit[7]uril complex. Toward paramagnetic gyroscopes and rolling nanomachines. *Nanoscale* 2015, 7, 12143–12150. [CrossRef]
- 33. Mao, J.; Akhmetzyanov, D.; Ouari, O.; Denysenkov, V.; Corzilius, B.; Plackmeyer, J.; Tordo, P.; Prisner, T.F.; Glaubitz, C. Host–Guest Complexes as Water-Soluble High-Performance DNP Polarizing Agents. *J. Am. Chem. Soc.* **2013**, *135*, 19275–19281. [CrossRef]
- 34. Matsuki, Y.; Maly, T.; Ouari, O.; Karoui, H.; Le Moigne, F.; Rizzato, E.; Lyubenova, S.; Herzfeld, J.; Prisner, T.; Tordo, P.; et al. Dynamic Nuclear Polarization with a Rigid Biradical. *Angew. Chem. Int. Ed.* **2009**, *48*, 4996–5000. [CrossRef]
- 35. Rockenbauer, A.; Szabo-Planka, T.; Arkosi, Z.; Korecz, L. A Two-Dimensional (Magnetic Field and Concentration) Electron Paramagnetic Resonance Method for Analysis of Multispecies Complex Equilibrium Systems. Information Content of EPR Spectra. J. Am. Chem. Soc. 2001, 123, 7646–7654. [CrossRef]
- 36. Wheate, N.J.; Anil Kumar, P.G.; Torres, A.M.; Aldrich-Wright, J.R.; Price, W.S. Examination of Cucurbit[7]uril and Its Host-Guest Complexes by Diffusion Nuclear Magnetic Resonance. *J. Phys. Chem. B* 2008, 112, 2311–2314. [CrossRef]
- 37. Grant, M.P.; Wheate, N.J.; Aldrich-Wright, J.R. Diffusion Coefficient of Cucurbit[*n*]urils (*n* = 6 or 7) at Various Concentrations, Temperatures, and pH. *J. Chem. Eng. Data* **2009**, *54*, 323–326. [CrossRef]
- Guagnini, F.; Antonik, P.M.; Rennie, M.L.; O'Byrne, P.; Khan, A.R.; Pinalli, R.; Dalcanale, E.; Crowley, P.B. Cucurbit[7]uril-Dimethyllysine Recognition in a Model Protein. *Angew. Chem. Int. Ed.* 2018, 57, 7126–7130. [CrossRef] [PubMed]
- Bardelang, D.; Casano, G.; Poulhès, F.; Karoui, H.; Filippini, J.; Rockenbauer, A.; Rosas, R.; Monnier, V.; Siri, D.; Gaudel-Siri, A.; et al. Spin exchange monitoring of the strong positive homotropic allosteric binding of a tetraradical by a synthetic receptor in water. *J. Am. Chem. Soc.* 2014, 136, 17570–17577. [CrossRef] [PubMed]
- Hwang, I.; Jeon, W.S.; Kim, H.-J.; Kim, D.; Kim, H.; Selvapalam, N.; Fujita, N.; Shinkai, S.; Kim, K. Cucurbit[7]uril: A Simple Macrocyclic, pH-Triggered Hydrogelator Exhibiting Guest-Induced Stimuli-Responsive Behavior. *Angew. Chem. Int. Ed.* 2007, 46, 210–213. [CrossRef] [PubMed]
- 41. Freed, J.H.; Fraenkel, G.K. Theory of Linewidths in Electron Spin Resonance Spectra. *J. Chem. Phys.* **1963**, *39*, 326–348. [CrossRef]
- 42. Freed, J.H. Generalized Cumulant Expansions and Spin-Relaxation Theory. J. Chem. Phys. **1968**, 49, 376–391. [CrossRef]
- Bardelang, D.; Rockenbauer, A.; Karoui, H.; Finet, J.-P.; Biskupska, I.; Banaszak, K.; Tordo, P. Inclusion complexes of EMPO derivatives with 2,6-di-O-methyl-β-cyclodextrin: Synthesis, NMR and EPR investigations for enhanced superoxide detection. *Org. Biomol. Chem.* **2006**, *4*, 2874–2882. [CrossRef]
- Bardelang, D.; Rockenbauer, A.; Finet, J.-P.; Karoui, H.; Tordo, P. Inclusion complexes of PBN-type nitrone spin traps and their superoxide spin adducts with cyclodextrin derivatives: Parallel determination of the association constants by NMR titrations and 2D-EPR simulations. *J. Phys. Chem. B* 2005, 109, 10521–10530. [CrossRef]
- 45. Karoui, H.; Rockenbauer, A.; Pietri, S.; Tordo, P. Spin trapping of superoxide in the presence of β-cyclodextrins. *Chem. Commun.* **2002**, *24*, 3030–3031. [CrossRef]
- 46. Franchi, P.; Lucarini, M.; Pedulli, G.F.; Sciotto, D. An EPR Investigation of the Kinetics of Inclusion of a Persistent Radical in Water-Soluble Calix[4]arenes. *Angew. Chem. Int. Ed.* **2000**, *39*, 263–266. [CrossRef]
- Kirilyuk, I.; Polovyanenko, D.; Semenov, S.; Grigorev, I.; Gerasko, O.; Fedin, V.; Bagryanskaya, E. Inclusion Complexes of Nitroxides of Pyrrolidine and Imidazoline Series with Cucurbit[7]uril. *J. Phys. Chem. B* 2010, 114, 1719–1728. [CrossRef]
- 48. Kolman, V.; Khan, M.S.A.; Babinsky, M.; Marek, R.; Sindelar, V. Supramolecular Shuttle Based on Inclusion Complex between Cucurbit[6]uril and Bispyridinium Ethylene. *Org. Lett.* **2011**, *13*, 6148–6151. [CrossRef]

- 49. Sindelar, V.; Silvi, S.; Kaifer, A.E. Switching a molecular shuttle on and off: Simple, pH-controlled pseudorotaxanes based on cucurbit[7]uril. *Chem. Commun.* **2006**, 2185–2187. [CrossRef] [PubMed]
- 50. Gong, Y.; Chen, H.; Ma, X.; Tian, H. A Cucurbit[7]uril Based Molecular Shuttle Encoded by Visible Room-Temperature Phosphorescence. *ChemPhysChem* **2016**, *17*, 1934–1938. [CrossRef] [PubMed]
- Chernikova, E.Y.; Berdnikova, D.V.; Fedorov, Y.V.; Fedorova, O.A.; Maurel, F.; Jonusauskas, G. Light-induced piston nanoengines: Ultrafast shuttling of a styryl dye inside cucurbit[7]uril. *Phys. Chem. Chem. Phys.* 2017, 19, 25834–25839. [CrossRef] [PubMed]
- 52. Mezzina, E.; Manoni, R.; Romano, F.; Lucarini, M. Spin-Labelling of Host-Guest Assemblies with Nitroxide Radicals. *Asian J. Org. Chem.* **2015**, *4*, 296–310. [CrossRef]
- Trabolsi, A.; Khashab, N.; Fahrenbach, A.C.; Friedman, D.C.; Colvin, M.T.; Coti, K.K.; Benitez, D.; Tkatchouk, E.; Olsen, J.-C.; Belowich, M.E.; et al. Radically enhanced molecular recognition. *Nat. Chem.* 2010, 2, 42–49. [CrossRef]

Sample Availability: Samples of the compounds are available from the authors.



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