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# Preweaned heifer management on US dairy operations: Part V. Factors associated with morbidity and mortality in preweaned dairy heifer calves

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# ABSTRACT

The objective of this study was to evaluate morbidity and mortality in preweaned dairy heifer calves based on different health, feeding, and management practices, as well as environmental factors. This study was conducted as part of the calf component of the National Animal Health Monitoring System's Dairy 2014 study, which included 104 dairy operations in 13 states. The calf component was an 18-mo longitudinal study focused on dairy heifer calves from birth to weaning; data were collected on 2,545 calves. The percentage morbidity for all calves enrolled in the study was 33.9%. Backward elimination model selection was used after univariate screening to determine which management practices and environmental factors significantly affected morbidity and mortality. The final morbidity model included birth weight, serum IgG concentration, ventilation type, and average temperature-humidity index (THI) during the preweaning period. After controlling for other independent variables in the model, calves born at a higher birth weight had a lower predicted risk of morbidity than calves with a lower birth weight. An increase in serum IgG concentration was associated with decreased morbidity. Calves housed in positive- or cross-ventilated systems had a 2.2 times higher odds of developing disease compared with calves housed in natural ventilation systems. Average THI during the preweaning period was inversely correlated with morbidity; as THI increased, the predicted morbidity risk decreased. The percent mortality for all calves enrolled in the study was 5.0%. The final mortality model included birth weight, serum IgG concentration, amount of fat/day in the liquid diet, and morbidity. After controlling for other independent variables in the model, calves born at a higher birth weight had a lower risk of mortality. An increase in serum IgG concentration decreased the risk of mortality. The odds of mortality were 3.1 times higher in calves fed <0.15 kg of fat/d in the liquid diet compared with calves fed >0.22 kg of fat/d. The odds of mortality were 4.7 times higher in calves that experienced any disease throughout the preweaning period than in calves with no disease. In summary, morbidity and mortality were both associated with birth weight and serum IgG concentration. Additionally, morbidity was associated with ventilation type and average monthly THI, and mortality was associated with amount of fat per day in the liquid diet and morbidity.

**Key words:** preweaned calves, morbidity, treatment, mortality

# INTRODUCTION

The preweaning phase of dairy heifer calves represents a critical period in the life of replacement females, as they are very susceptible to disease and death. Calves are at the greatest risk of dying during the first 21 d of life (Waltner-Toews et al., 1986; USDA, 1994; Wells et al., 1996, 1997). Overall, the health status of a preweaned dairy heifer can greatly affect lifelong production, including growth, reproductive efficiency, and milk production. An association between calfhood bovine respiratory disease complex and decreased milk production, reproduction, and early culling has been demonstrated (Stanton et al., 2012). Furthermore, morbidity, disease treatment, and mortality in preweaned calves result in a forfeiture of genetic potential for future herd improvement (Wells et al., 1997; Swali and

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Wathes, 2006). The economic loss of replacing a dairy heifer due to disease and death is significant. Although raising a dairy heifer is expensive, this cost generally does not surpass the cost of purchasing a springing heifer (McGuirk, 2008). Therefore, understanding relationships between management practices and calf health is essential for minimizing morbidity and mortality and enhancing future production.

Few recently published studies have examined risk factors for morbidity and mortality in preweaned dairy heifer calves. The most recent estimates of national morbidity and mortality in preweaned dairy heifers were reported in 1992 and 2007 (USDA, 1994, 2010). The percentage morbidity in the 1992 National Dairy Heifer Evaluation project was 36.1% and the percentage mortality was 8.4% (USDA, 1994). These numbers stayed relatively consistent with the USDA NAHMS Dairy 2007 study, which reported a morbidity of 38.5% and a mortality of 7.8%. Both studies had morbidity and mortality percentages above the Dairy Calf and Heifer Association (2010) target rates for preweaned dairy heifers, which are 25 and 5% for morbidity and mortality, respectively.

The objectives of this prospective study were to (1) estimate current national morbidity and mortality rates, (2) describe common clinical signs and treatment methods currently used in preweaned dairy heifers, and (3) examine risk factors via inferential modeling for morbidity and mortality.

# MATERIALS AND METHODS

## Study Design

The USDA's National Animal Health Monitoring System (**NAHMS**) conducts national surveys to collect information on the health, management, and productivity of domestic livestock species (USDA, 2016). In 2014, a nationwide survey was conducted to collect information about the US dairy industry and included an 18-mo longitudinal preweaned heifer calf study. All participation in NAHMS studies is voluntary. However, to participate in the longitudinal calf study, operations had to have completed both NAHMS Dairy 2014 surveys, and agreed to provide information on enrolled calves, whether raised onsite or offsite.

The calf component was part of the NAHMS Dairy 2014 study, and it consisted of a convenience sample of 104 dairy operations, including both conventional and organic operations. These operations were located in 13 states, including California, Colorado, and Washington in the West region, and Iowa, Michigan, Minnesota, Missouri, New York, Ohio, Pennsylvania, Vermont, Virginia, and Wisconsin in the East region. Dairy operations were categorized based on the number of mature cows, as small (30 to 99 cows), medium-sized (100 to 499 cows), and large (500 or more cows). Figures describing the study sample and operations can be found in Urie et al. (2018a).

Data collection for the calf component of the study occurred from March 2014 to September 2015. Data collectors were trained on data and sample collection. Each operation was instructed to enroll 24 heifer calves over a 1-yr period, or an average of 2 calves/mo. Farm personnel selected which calves to enroll in the study. However, a calf must have been alive at 24 h of age to be enrolled. Because fewer operations participated than originally planned, the target number enrolled per operation was increased to 48 calves. Additionally, because enrollment of farms did not occur as quickly as anticipated, the study encompassed 18 mo instead of the 12-mo period that was planned.

# Heifer Calf Health Card

Each calf enrolled in the study had a Heifer Calf Health Card ("Calf Card") filled out to record information on events that occurred from birth to weaning (https://www.aphis.usda.gov/animal\_health/nahms/ dairy/downloads/dairy14ques/CalfHealth.pdf). The Calf Card contained questions in both English and Spanish and was filled out by farm personnel, a veterinary medical officer, extension personnel, veterinarians, or a combination of people involved with calf raising. The Calf Card included birth data (e.g., birth date, weight, and calving ease), colostrum feeding data (including timing, amount, and method of colostrum feeding), preweaning housing and procedures data (e.g., housing, ventilation, bedding, navel disinfection, and dehorning), milk feeding (including type of liquid diet fed, any additives, and method of feeding), milk consumption record (volume and frequency of feedings), preweaning growth record (hip height and heart girth recordings every 2 wk), biologic sampling record (including serum collection and fecal sampling dates), vaccinations, disease incidence and treatment, weaning data (weaning date, primary weaning criteria), and any additional notes. Starter feed labels and milk replacer labels, if applicable, were also collected.

Producers were provided with the Calf Health Scoring Chart (https://www.vetmed.wisc.edu/dms/fapm/ apps/chs.htm) to guide the reporting of clinical signs. The goal of the study was to collect recognized clinical signs and causes of death, not to standardize clinical signs and causes of death across operations. All clinical signs and causes of death were reviewed, edited, and categorized, as described in the Statistical Analysis section.

Because of the variation observed in the quantity and quality of the liquid diet fed per day, the amounts (kg) of protein and fat fed per day were calculated for each calf. The amount of liquid diet fed per feeding and the frequency of feedings per day were recorded on the Calf Card and used to calculate the total amount of liquid diet fed. For calves fed milk replacer, the percent protein and percent fat were obtained from the producer-reported values or the milk replacer label. For calves fed whole or waste milk, percent protein was set at 3.1% and percent fat as 3.8% (as-fed basis; Chester-Jones and Hoffman, 2003). The total amounts of protein and fat were calculated and then divided by the number of days fed a liquid diet to determine the average kilograms of protein and fat fed per day in the liquid diet.

Additives in the liquid diet were categorized as antibiotics, direct-fed microbials, fly control, acids/preservatives, coccidiostats, vitamins and minerals, or other additives. Information regarding calf starter included the percent protein and the age starter was first offered, but starter intake was not recorded.

## **Biological Sampling**

Before enrollment in the study, calves were screened for persistent infection with bovine viral diarrhea virus (**BVDV**). V-Cut ear notchers (Nasco, Fort Atkinson, WI) were used to collect ear notch samples from all calves, which were tested on-farm for BVDV using the Idexx SNAP BVDV Antigen Test (Idexx, Westbrook, ME). Calves that tested positive for BVDV were excluded from the study. Colostrum samples (40–50 mL) from the first feeding of colostrum administered to each calf were collected in conical screw-top tubes by farm personnel and frozen until shipping. Blood samples (5 mL) from calves between 1 and 7 d of age were collected in serum separator tubes by veterinary medical officers or animal health technicians, and samples were centrifuged if possible before shipping. Colostrum and blood samples were shipped together on ice to USDA's National Veterinary Services Laboratories (Ames, IA). Blood samples were centrifuged at the National Veterinary Services Laboratories and serum was separated. Serum and colostrum samples were then accumulated and shipped in bulk to the Saskatoon Colostrum Company (Saskatoon, SK, Canada) for testing. Colostrum samples were tested for IgG concentration using radial immunodiffusion and Brix score using a digital Brix refractometer. Serum samples were tested for IgG concentration using radial immunodiffusion and for total protein and Brix score using a digital Brix refractometer. Blood samples collected within 24 h of birth or after 7 d of age were excluded from this analysis. A complete description of the methods used for Brix scoring and IgG determination can be found in Shivley et al. (2018).

For evaluation of enteric parasites, veterinary medical officers or animal health technicians collected approximately 50 g of feces directly from the rectum of calves between 2 and 4 wk of age. Fecal samples were placed in cylindrical screw-top containers and shipped on ice to the USDA Agricultural Research Service's Environmental Microbial Food Safety Laboratory (Beltsville, MD), as quickly as possible after sample collection. Fecal samples were tested by immunofluorescence microscopy for Cryptosporidium parvum and Giardia duodenalis. A complete description of the methods used for fecal parasite testing can be found in Urie et al. (2018b).

# Growth Measurements

Calves were measured approximately every 2 wk during the preweaning period to measure growth. Height/ weight tapes from Coburn (Nasco, Fort Atkinson, WI) were provided for measuring calves. For consistency, it was recommended that 1 trained veterinary medical officer or animal health technician complete the measurements on all calves enrolled on an operation. Birth weights were estimated using a scale, hoof circumference, or heart girth circumference and reported in pounds or kilograms. The method operations used to estimate birth weight were not captured. Hip height and heart girth circumference were measured approximately every 2 wk and recorded in centimeters. Heart girth circumference in centimeters was converted to kilograms using the following equation (Heinrichs et al., 1992):

BW (kg) = 
$$[(0.02655 \times \text{cm}^2) + (-2.876 \times \text{cm}) + 102.71].$$

Preweaning weight gain was calculated by subtracting the birth weight from the final weight. Average daily gain was calculated by taking the weight gain during the preweaning period divided by the number of days between birth weight and final weight (approximately the preweaning period). Final weights did not always occur at the same time as weaning. However, only calves with final weight measurements within 14 d of weaning were included in the analysis.

#### Environmental Factors

Temperature-humidity index (THI) data were obtained monthly by county from the National Oceanic and Atmospheric Administration (NOAA, 2016). The THI accounts for the effects of temperature and relative humidity, and the equation uses dry bulb temperature (T, °F) and relative humidity (RH). The equation used for this analysis was THI = T -  $[0.55 - (0.55 \times \text{RH}/100)] \times (\text{T} - 58)$ . Each calf was assigned a THI for each month during the preweaning period and averaged over the months of their preweaning period (**pTHI**).

# Statistical Analysis

When each calf was weaned, the Calf Card was mailed to USDA NAHMS (Fort Collins, CO). Initial ongoing validation was performed on every calf card, followed by data entry into SAS (version 9.4; SAS Institute Inc., Cary, NC). Once all Calf Cards were entered, the data were validated again by NAHMS staff and merged with the results from the colostrum, serum, and fecal testing obtained from the laboratories.

After data validation was complete, primary morbidity categories, monthly calf enrollment, and incidence rates were calculated. Primary morbidity categories for digestive, respiratory, concurrent digestive and respiratory, and other clinical signs were created to allow for single classification of each case (i.e., each case was only listed under the primary reported clinical sign). These categories excluded nondescript signs, such as dull and dehydrated. For example, if a calf was listed as having diarrhea and being dull and dehydrated, it was classified under the primary morbidity category of "digestive." If a calf was listed as being only dull or only dehydrated, it was classified under "other" clinical signs. Although each case was assigned a primary morbidity category, an individual calf could have been reported as displaying digestive signs, respiratory signs, concurrent digestive and respiratory signs, and other signs at different times throughout the preweaning period. If multiple cases for a single calf were recorded, digestive events and other events were considered independent cases if a minimum of 3 d had elapsed between each episode. Respiratory events and concurrent digestive and respiratory events were considered independent cases if at least 7 d had elapsed between each event.

Study morbidity incidence rates for any cases and independent classification of cases were calculated using calf-days at risk. Calf-days at risk were calculated by multiplying the total calves enrolled in the study by the average age at weaning. Monthly calf enrollment was calculated for each month of the study to estimate monthly morbidity incidence rates. A calf was included in every month it was enrolled in the study from birth until weaning or death, regardless of the exact number of days the calf may have spent in the month. Monthly morbidity incidence rates were calculated as a ratio of the number of cases observed in each month divided by the number of calves at risk in that month. Weekly morbidity rates were calculated as a ratio of the number of cases observed in each week divided by the number of calves at risk in that week of age. Write-in data for cause of death were reviewed for all calves and compared with disease data reported at the time of death. If a calf had concurrent respiratory and digestive signs, but the cause of death was only reported as one or the other, the cause of death was reclassified into concurrent respiratory and digestive.

Descriptive data were analyzed using the FRE-QUENCY and MEANS procedures in SAS for categorical and continuous variables, respectively. To evaluate the time in days to morbidity and mortality events for preweaned heifer calves, Kaplan-Meier survival graphs were constructed using the LIFETEST procedure in SAS. The STRATA statement was used to visualize differences based on serum IgG level, serum total protein level, and serum Brix percent. PROC GLM was used to model serum total protein level and serum Brix score by serum IgG level to develop comparable categories. The log-rank test was used to evaluate differences of the survival curves throughout the preweaning period (Hosmer and Lemeshow, 1999) for each serum measurement. Censored calves were calves that were weaned before 60 d of age.

Two separate logistic regression models were constructed to determine the factors associated with morbidity and mortality. Operation was included in the model as a random variable to account for clustering of calves within operations. Predictor variables were considered for each model based on biological plausibility. To be included in the analysis, calves had to have data for all variables initially included in each model. Univariate screening was used to determine which variables to include in each of the initial models and variables with P < 0.20 were selected for inclusion in the multivariate models. Stepwise backward elimination model selection in PROC GENMOD in SAS was used to construct each final model, with P < 0.05considered significant and thus included in the model. All first-order interactions of the final model variables were evaluated for significance. Model fit was evaluated using the Akaike information criteria.

Figures to demonstrate the relationship of continuous variables in the morbidity and mortality model were created using PROC GPLOT. The log means of morbidity and mortality were taken and graphed against the model variable. The points were combined into 12 different groups and a line of best fit was constructed based on the 12 groups. These graphs were created for all final continuous variables in the morbidity and mortality models.

**Table 1.** Reported clinical signs by number and percent of heifer calves and number and percent of cases (n = 2.545 calves; n = 859 ill calves; n = 1.103 cases)

Reported clinical sign	No. of calves	Percent of all calves	Percent of reported ill calves	No. of cases	Percent of reported cases
Digestive	483	18.9	56.0	533	48.3
Dull	330	13.0	38.4	383	34.7
Respiratory	287	11.3	33.4	349	31.6
Dehydration	108	4.2	12.6	112	10.2
Lameness	28	1.1	3.3	29	2.6
Neurological	8	0.3	0.9	8	0.7
$Other^1$	183	7.2	21.3	267	24.2
Any	859	33.8		1,103	

<sup>1</sup>Other included calves reported with infections or injuries.

#### RESULTS

#### Descriptive Morbidity

Overall, 33.8% of enrolled calves had at least one morbidity case, with 6% of calves experiencing more than one case. Therefore, 859 of the 2,545 enrolled calves were reported ill, resulting in 1,103 reported cases (Table 1). The majority of calves reported ill had digestive signs (56.0%), appeared dull (38.4%), or displayed respiratory signs (33.4%). Primary categories of "digestive," "respiratory," "concurrent digestive and respiratory," and "other" clinical signs were developed to eliminate nondescript signs of illness, such as dull and dehydrated, and to allow for classification of each individual case. Classification of these clinical categories is described under Statistical Analysis. Once primary clinical categories were developed, digestive signs remained the most prominent, with 17.2% of all calves reported having digestive signs, representing 50.9% of all ill calves and 43.6% of all cases (Table 2). Respiratory, concurrent digestive and respiratory, and other signs were lower, with 9.5, 1.8, and 8.0% of all calves, respectively, representing 27.0, 4.6, and 24.8% of all cases, respectively. Of the 24.8% "other" cases, 75.5%were reported as only dull, dehydrated, or febrile, and 24.5% were reported as having infections, injuries, lameness, or neurological signs.

The overall morbidity incidence rate for the study was 0.450 cases per 65.7 calf-days at risk. Incidence rates for digestive cases, respiratory cases, concurrent digestive and respiratory cases, and other clinical cases were 0.196, 0.122, 0.021, and 0.111 cases/65.7 calf-days at risk for the study (Supplemental Table S1; https:// doi.org/10.3168/jds.2017-14019).

The number of calves enrolled each month was relatively evenly distributed, with the highest enrollment in August (689 calves) and the lowest in February (556 calves). Digestive morbidity incidence rates were highest during December (0.086 cases/calf-month), February (0.076 cases/calf-month), and April (0.081 cases)calf-month). Respiratory morbidity incidence rates were elevated during the months of January (0.045 cas)es/calf-month), March (0.062 cases/calf-month), and April (0.044 cases/calf-month). Concurrent digestive and respiratory signs were consistently low throughout the months, with a slight increase in morbidity incidence rate during the month of October (0.012 cases)calf-month). "Other" clinical signs were highest during January, February, and April with morbidity rates of 0.053, 0.051, and 0.051 cases/calf-month, respectively. Ultimately, any morbidity was highest during the spring, with total morbidity incidence rates of 0.163 cases/calf-month in February, 0.174 cases/calf-month in April, and 0.184 cases/calf-month in May (Figure

Table 2. Reported primary clinical signs by number and percent of heifer calves and number and percent of independent cases (n = 2,545 calves; n = 859 ill calves; n = 1,103 cases)

Reported primary clinical sign	No. of calves	Percent of all calves	Percent of reported ill calves	No. of cases	Percent of cases
Digestive	437	17.2	50.9	481	43.6
Respiratory	241	9.5	28.1	298	27.0
Concurrent digestive and respiratory	46	1.8	5.4	51	4.6
Other <sup>1</sup>	203	8.0	23.6	273	24.8

<sup>1</sup>"Other" included calves reported as only dull or dehydrated or febrile and calves with reported infections, injuries, lameness, or neurological signs.

1, Supplemental Table S2; https://doi.org/10.3168/jds .2017-14019).

Weekly incidence rates were calculated for each primary clinical case by calf-weeks and graphed by week of age (Figure 2, Supplemental Table S3; https://doi.org/ 10.3168/jds.2017-14019). The weekly incidence rates showed a peak in digestive disease at approximately 2 wk of age (0.0727 cases/calf-week). The incidence rate of digestive disease quickly declined by wk 4 to 0.0090 cases/calf-week and continued to decline throughout the rest of the preweaning period. "Other" disease followed a similar trend as digestive disease with a maximum incidence rate of 0.0306 cases/calf-week at 1 wk of age. Respiratory disease cases had a maximum incidence rate of 0.0169 cases/calf-week at wk 5 of age. Concurrent respiratory and digestive disease had low incidence rates throughout the preweaning period, with a maximum incidence rate of 0.0039 during wk 2 of age.

Most ill calves (90.2%) received treatment. The majority (73.8%) of ill calves received antibiotics, 40.5% received fluids (oral or injectable), 27.6% were given a nonsteroidal anti-inflammatory (NSAID), and 8.0% of calves received a decreased liquid diet (Table 3).

Antibiotics were administered to over one-fourth (26.8%) of all enrolled calves (for treatment and prevention) and 73.8% of all ill calves. The most prominent antibiotic classes administered to calves were fluoroquinolones (29.1%), sulfonamides (23.5%), and penicillins (26.8%). Sulfonamides were the most common antibiotics used for cases with digestive signs (30.2%), whereas fluoroquinolones were administered to 17.6% of digestive cases. The primary antibiotics used for respiratory

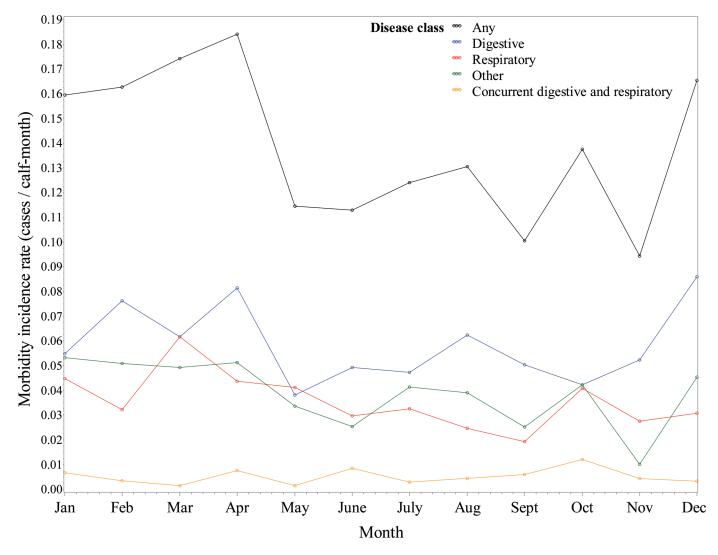


Figure 1. Calculated morbidity incidence rates for preweaned heifer calves by reported exclusive clinical cases and month from March 2014 through September 2015. Color version available online.

cases were macrolides (32.3%) and florfenicols (24.1%; Table 4). Unlike digestive cases, for which only 68.8% received antibiotics, 88.3% of respiratory cases received antibiotics, and 90.2% of concurrent respiratory and digestive cases were treated with antibiotics. Additionally, 50 calves received antibiotics as a preventative treatment, with the most common antibiotic classes being penicillin (64.0%) and fluoroquinolone (32.0%).

# **Descriptive Mortality**

The overall study mortality was 5.0%; 128 of the 2,545 calves died during the preweaning period (Table 5). Recorded causes of death were digestive (32.0% of deaths), respiratory (14.1% of deaths), digestive and respiratory

(7.0% of deaths), and other (13.3% of deaths), which included injuries, infections, and calves that were sold for unreported reasons. One-fourth of deceased calves (25.0%) had an unknown cause of death and 8.6% had no reported cause of death. The mean age at death was 24.4 d. The mean age for digestive causes of death was 18.3 d, the mean age for respiratory causes of death was 37.5 d, and the mean age for concurrent respiratory and digestive causes of death was 18.6 d. Unknown causes of death had a mean age of 25.7 d. The case-fatality rates were 8.5% of digestive cases, 6.0% of respiratory cases, 17.7% of concurrent digestive and respiratory cases, and 6.2% of other cases. However, 2.5% of calves that were never reported ill died during the preweaning period (Table 6).

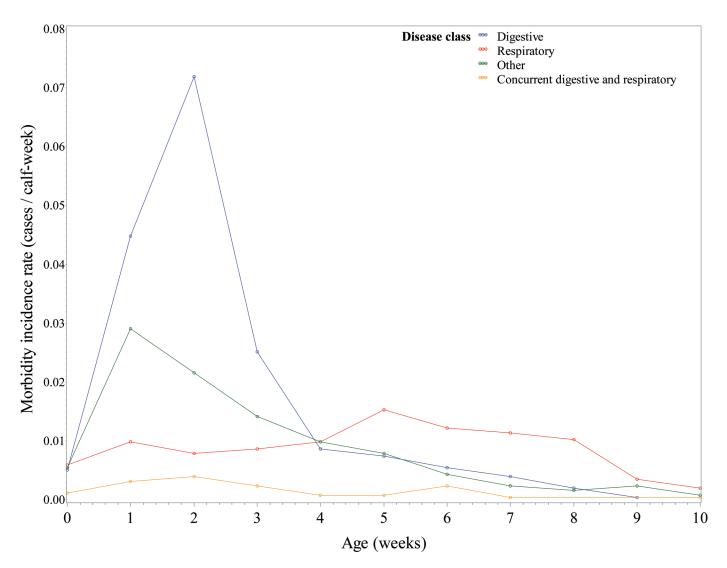


Figure 2. Calculated morbidity incidence rates for preweaned heifer calves by reported exclusive clinical cases and age in weeks. Color version available online.

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Table 3. Number and percentage of heifer calves by treatment (n = 2,545 calves; n = 859 ill calves; n = 1,103 cases; n = 826 treated calves)

Treatment	No. of calves	Percent of all calves	Percent of ill calves	Percent of cases	Percent of treated calves
Antibiotic	634	24.9	73.8	57.5	76.8
Oral electrolytes	317	12.5	36.9	28.7	38.4
Nonsteroidal anti-inflammatory drugs	237	9.3	27.6	21.5	28.7
Decreased liquid diet	69	2.7	8.0	6.3	8.4
Gut soothers	38	1.5	4.4	3.4	4.6
Vitamins	34	1.3	4.0	3.1	4.1
Injectable fluids	31	1.2	3.6	2.8	3.8
Steroids	27	1.1	3.1	2.4	3.3
Coccidiostats	19	0.7	2.2	1.7	2.3
Minerals	17	0.7	2.0	1.5	2.1
Changed liquid diet	10	0.4	1.2	0.9	1.2
Clostridium antitoxin	8	0.3	0.9	0.7	1.0
Antihistamines	4	0.2	0.5	0.4	0.5
Immune boosters	2	0.1	0.2	0.2	0.2
Analgesics	1	0.0	0.1	0.1	0.1
Preventative treatment <sup>1</sup>	51	2.0			6.2
Any treatment	826	30.7	90.2	74.9	

<sup>1</sup>Fifty calves received preventive antibiotics; 1 received a preventative coccidiostat.

# Survival Analysis

*Morbidity.* Kaplan-Meier survival curves through 60 d of age were created for overall morbidity stratified by serum IgG concentration with corresponding serum total protein levels and serum Brix scores listed (Figure 3). The morbidity survival curve illustrates that the majority of reported clinical signs occurred in the first 20 d of life and then slowly tapered off throughout the rest of the preweaning period. Serum total protein level and serum Brix scores were modeled with serum IgG to develop comparable categories ( $R^2 = 0.803$  and  $R^2$ ) = 0.797, respectively). The morbidity survival curve stratified by serum IgG level with corresponding serum total protein levels and serum Brix scores had a significant log-rank (P < 0.0001, P < 0.0001) when comparing serum IgG  $\geq 15.0$  g/L to < 15.0 g/L and  $\geq 10.0$  g/L to <10.0 g/L, respectively. These suggest that the survival curve for each serum test differed at the beginning and toward the end of the 60-d period and that there was a dose-dependent response regarding serum IgG, serum total protein, and serum Brix, and morbidity.

**Mortality.** Kaplan-Meier survival curves through 60 d of age were created for overall mortality stratified by serum IgG concentration with corresponding serum total protein levels and serum Brix scores listed (Figure 4). The mortality survival curve is similar to the morbidity curve, in that most mortality events occurred in the first 20 d of life. The mortality survival curve stratified by serum IgG level with corresponding serum total protein levels and serum Brix scores had a significant log-rank when comparing serum IgG levels of  $\geq 15.0$  g/L to <15.0 g/L and  $\geq 10.0$  g/L to <10 g/L (P = 0.0019 and P < 0.0001, respectively). Once again, these results illustrate a dose-dependent response with

serum IgG level, serum total protein, and serum Brix and mortality, suggesting that calves should have a serum IgG concentration of at least 15.0 g/L, a serum total protein  $\geq 5.4$  g/L, or a serum Brix score  $\geq 8.6\%$  to help prevent both morbidity and mortality.

## Morbidity Model

Of the 2,545 calves in the study, 2,374 calves had complete data and were included in the morbidity model for analysis. Initial variables included in the multivariable model after univariate screening for morbidity included sex of the primary caretaker during the preweaning period, primary housing ventilation type, a categorized housing number based on group size, vaccination status during the preweaning period, any preventative antibiotic treatments given during the preweaning period, liquid diet type, direct-fed microbial additives in the liquid diet, birth weight (kg), serum IgG concentration (g/L), amount of protein in the liquid diet per day (kg), and pTHI (Supplemental Tables S4 and S5; https://doi .org/10.3168/jds.2017-14019). The final multivariable model for morbidity in calves included birth weight (P= 0.004), serum IgG (P = 0.001), ventilation type (P =0.015), and pTHI (P = 0.008) (Table 7).

After controlling for other independent variables in the model, calves born at higher birth weights had a lower risk of morbidity than calves with lower birth weights [odds ratio (**OR**) = 0.962]. Selecting a low birth weight of 35.0 kg for this continuous variable resulted in a predicted morbidity risk of 40.0%, whereas a birth weight of 45.0 kg had a predicted morbidity risk of 31.2% (Supplemental Figure S1a; https://doi.org/ 10.3168/jds.2017-14019). Similarly, an increase in se-

	, ,	:	Percent	Percent of	Digesti	Digestive signs	Respiratory signs	ory signs	Concurrent digestive and respiratory signs	urrent stive biratory ns	Other signs	signs	Preve	Preventative treatment
Antibiotic	No. of calves	No. of Percent of all calves calves	of treated calves	calves given antibiotic	No.	8	No.	%	No.	%	No.	%	No.	%
Fluoroquinolone	197	7.7	23.8	29.1	81	17.6	36	11.9	6	17.6	4	1.4	16	32.0
Sulfonamide	159	6.2	19.2	23.5	139	30.2	14	4.6	10	19.6	19	6.7	0	0.0
Penicillin	182	7.2	22.0	26.8	68	14.8	22	7.3	12	23.5	85	29.9	32	64.0
Macrolide	138	5.4	16.7	20.4	18	3.9	98	32.3	7	13.7	9	2.1	0	0.0
Florfenicol	120	4.7	14.5	17.7	10	2.2	73	24.1	11	21.6	75	26.4	1	2.0
Cephalosporin	96	3.8	11.6	14.2	43	9.3	33	10.9	c,	5.9	26	9.2	0	0.0
Tetracycline	63	2.5	7.6	9.3	24	5.2	21	6.9	2	3.9	21	7.4	1	2.0
Aminoglycoside	11	0.4	1.3	1.6	ഹ	1.1	2	0.7	1	2.0	27	9.5	0	0.0
Any antibiotic <sup>1</sup>	684	26.8	76.8	100.0	331	68.8	263	88.3	46	90.2	237	86.8	50	98.0

= 826 treated calves; n = 481 digestive cases;

= 2,545 calves; n

Table 4. Reported antibiotic administered by number and percent of heifer calves, and exclusive clinical signs (n

rum IgG concentration decreased the risk of morbidity (OR = 0.977). A serum IgG of 8.0 g/L (consistent with failure of passive transfer) resulted in a predicted morbidity risk of 40.3%, whereas a serum IgG of 30.0 g/L (consistent with excellent passive transfer) had a predictive morbidity risk of 29.3% (Supplemental Figure S1b). Ventilation type was correlated with morbidity. Calves not housed in natural ventilation systems had 2.218 times higher odds of developing disease compared with calves housed in natural ventilation systems. Finally, pTHI was inversely correlated with morbidity (OR = 0.991). A pTHI of 20, below the reported calf thermoneutral zone, had a predicted morbidity risk of 39.5%, whereas a pTHI of 70 had a predictive morbidity risk of 29.1% (Supplemental Figure S1c).

#### Mortality Model

Of the 2,545 calves in the study, 2,272 calves had complete data and were included in the mortality model for analysis. Initial variables included in the multivariable model after univariate screening for mortality included vaccination status during the preweaning period, morbidity during the preweaning period, amount of fat in the liquid diet per day, direct-fed microbial additives in the liquid diet, any additives in the liquid diet, antibiotic additives in the liquid diet, pasteurization of milk, birth weight, serum IgG concentration, amount of protein in the liquid diet per day, and pTHI (Supplemental Tables S6 and S7; https://doi.org/10.3168/jds .2017-14019).

The final multivariable model for mortality in calves included birth weight (P = 0.011), serum IgG (P = 0.004), amount (kg) of fat/d in the liquid diet (P = 0.011), and morbidity during the preweaning period (P < 0.001) (Table 8).

After controlling for other independent variables in the model, calves born at a higher birth weight had a lower risk of mortality than calves with a lower birth weight (OR = 0.946). Selecting 2 arbitrary birth weights resulted in a relative mortality risk of 4.7%at a birth weight of 35.0 kg and a predicted mortality risk of 2.3% at a birth weight of 45.0 kg (Supplemental Figure S2a; https://doi.org/10.3168/jds.2017-14019). Similarly, an increase in serum IgG decreased the risk of mortality (0.963). A serum IgG of 8.0 g/L (consistent with failure of passive transfer) resulted in a predicted mortality risk of 5.2%, whereas a serum IgG of 30.0 g/L(consistent with excellent passive transfer) had a predicted mortality risk of 2.0%. Fat per day in the liquid diet was also correlated with mortality (Supplemental Figure S2b). The odds of mortality was 3.010 times higher for calves fed <0.15 kg of fat/d compared with calves fed >0.22 kg of fat/d. Mortality did not differ

Cause of death	No. of calves	Percent of all calves	Percent of deaths	Mean age (d)	SE	Minimum	Maximum
Digestive	41	1.6	32.0	18.3	2.3	6.0	78.0
Respiratory	18	0.7	14.1	37.5	4.6	3.0	70.0
Concurrent digestive and respiratory	9	0.4	7.0	18.6	5.03	5.0	44.0
Unknown	32	1.3	25.0	25.7	3.6	2.0	71.0
$Other^1$	17	0.7	13.3	27.0	4.5	8.0	75.0
Not reported	11	0.4	8.6	23.0	6.5	5.0	62.0
Any	128	5.0	100.0	24.4	1.64	2.0	78.0

**Table 5.** Reported cause of death by number and percent of heifer calves (n = 2,545 calves; n = 128 deaths)

<sup>1</sup>Other included infection, injury, or calves that were sold without a recorded reason.

significantly between calves fed 0.16 to 0.21 kg/d of fat in the liquid diet and those fed  $\geq 0.22$  kg/d of fat (P = 0.768). Calves that experienced any disease throughout the preweaning period had a 4.567 times higher odds of mortality than calves that did not experience any disease.

#### DISCUSSION

The reported morbidity rate of this study was 33.8% compared with a morbidity rate of 36.1% reported in the 1992 National Dairy Heifer Evaluation project (USDA, 1994). The Dairy Calf and Heifer Association report that the target morbidity rate for calves 24 to 60 d of age is less than 25%. Thus, there is still room to improve overall morbidity in preweaned dairy heifer calves (Dairy Calf and Heifer Association, 2010). Similar to the 1992 and 2007 USDA studies, digestive and respiratory signs were the most commonly identified problems, with digestive signs peaking at 2 wk of age and respiratory signs being more prevalent later in the preweaning period (USDA, 1994, 2010).

Cases of disease, including digestive and respiratory signs, were highest during the winter and spring months. This is likely due to changing temperatures and possible inclement weather, which results in a calf spending more time outside its thermoneutral zone. Wiseman et al. (1976) showed that the incidence of pneumonia increased during inclement weather events. Additionally, harsh weather events that include rain, snow, and high levels of moisture have been shown to increase the susceptibility of calves to diarrhea (Perez et al., 1990; Larson and Tyler, 2005; Carroll and Forsberg, 2007). Calculated incidence rates demonstrated that most calves developed digestive signs at approximately 2 wk of age. Diarrhea at 2 wk of age is often seen in at-risk calves, given that the infectious causes of diarrhea, such as bovine rotavirus and coronavirus, have a short incubation period and tend to cause diarrhea in calves to peak at 1 to 2 wk of age (Steele et al., 2004; Dhama et al., 2009). Additionally, calves developed respiratory signs that peaked at approximately 5 wk of age. Respiratory disease seen at this age is common in at-risk calves, because viral and bacterial agents, such as respiratory syncytial virus and parainfluenza type 3, associated with pneumonia in calves are normally found in calves over 4 wk of age (Lorenz et al., 2011).

Almost all (90.6%) ill calves reported in this study received treatment. The most common treatments were antibiotics and oral electrolytes. The use of antibiotics for clinical signs has stayed relatively constant since 1992, with 24.5 to 29.8% of all calves receiving antibiotics compared with 26.6% of all calves in the current study (USDA, 1994). The most commonly used antibiotic class to treat calves was fluoroquinolone, accounting for 29.1% of all calves administered an antibiotic, 17.6% of digestive cases, 11.9% of respiratory cases, 17.6% of concurrent digestive and respiratory cases, and 31% of preventative antibiotic administration. However, the use of fluoroquinolones is only approved for the treatment of respiratory disease in food-producing animals in the United States (Constable, 2004).

Decreasing milk or milk replacer during a diarrheic event is a practice that is no longer recommended

Table 6. Calculated case fatali	y rates by number $(\%)$ of calv	ves and by exclusive clinical signs
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Item	Digestive signs	Respiratory signs	Concurrent digestive and respiratory signs	$\frac{\text{Other}}{\text{signs}^1}$	Unknown/ not reported
Morbidity (% of cases) Mortality (% of dead calves) Case fatality rate (%; mortality/morbidity cases)	$\begin{array}{c} 481 \ (43.6) \\ 41 \ (32.0) \\ 8.5 \end{array}$	$\begin{array}{c} 298 \ (27.0) \\ 18 \ (14.1) \\ 6.0 \end{array}$	51 (4.6) 9 (7.0) 17.7	$273 (24.8) \\ 17 (0.7) \\ 6.2$	$\begin{array}{r} 1,693 \ (\mathrm{NA}^2) \\ 43 \ (33.6) \\ 2.5 \end{array}$

 $^1\mathrm{Other}$  included infection, injury, or calves that were sold without a recorded reason.  $^2\mathrm{Not}$  available.

(Garthwaite et al., 1994). Decreasing the amount of liquid diet fed as treatment during a disease event has diminished over the years; currently 2.7% of all calves received this treatment method compared with 15.9% of all calves in 1992 (USDA, 1994, 2010). It is recommended that calves be fed through the course of the diarrheic event to help maintain caloric intake and to add fluid volume and electrolytes to supplemental fluid administration (McGuirk, 2008).

The mortality rate of 5.0% was lower than mortality rates reported in previous US studies that ranged from 8 to 11% (USDA, 1994, 2002, 2010). The target mortality rate for calves 24 to 60 d of age is less than 5% (Dairy Calf and Heifer Association, 2010). The reported cause of death in this study was consistent with previous studies, highlighting digestive and respiratory disease as the most common contributors to calf death (USDA, 1994, 2010; Sivula et al., 1996; Tyler et al., 1999; Svensson et al., 2003). Consistent with previously reported data, the mean age of death due to digestive signs was 18.3 d, and the mean age of death due to respiratory signs was 37.5 d of age (USDA, 1994, 2010). Diarrhea is the most common cause of death in calves less than 30 d old, and pneumonia is the most important disease in calves greater than 30 d of age (USDA, 1994, 2010; Agerholm et al., 1993; Virtala et al., 1996; Svensson et al., 2003). However, approximately one-fourth of calves died from unknown causes. This highlights the importance of working with calf caretakers to identify and classify the cause of death for individual calves through necropsy and diagnostics. Case-fatality rates for digestive (8.5%)and respiratory (6.0%) cases decreased compared with those in the Sivula et al. (1996) study, in which the case-fatality rate for enteritis was 17.9% and that for

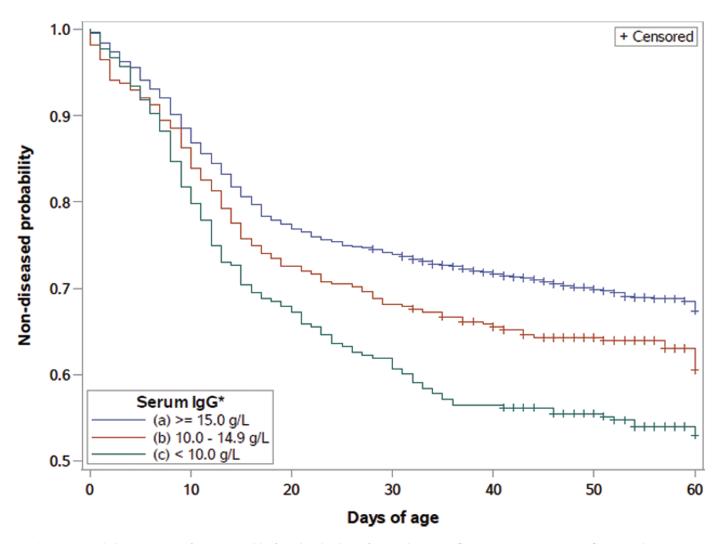


Figure 3. Morbidity percentage for preveaned heifer calves by days of age and serum IgG concentration categories. Corresponding to serum IgG levels of  $\geq$ 15.0 mg/dL, 10 to 14.9 mg/dL, and <10.0 mg/dL were serum total protein categories of  $\geq$ 5.4 g/dL, 5.1–5.3 g/dL, and <5.1 g/dL, and Brix score categories of  $\geq$ 8.6%, 8.1–8.5%, and <8.1%, respectively. Color version available online.

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**Table 7.** Results of multivariable modeling of factors associated with morbidity in heifer calves (n = 2,374)

		Model results			Example model outcomes		
Final model variable	Variable level	β-Estimate (SE)	Odds ratio (SE)	Factor P-value	Example variable level	Predicted morbidity risk (SE)	
Birth weight (kg)		-0.038(0.012)	0.962(0.011)	0.004	35.0	0.400(0.148)	
					45.0	0.312(0.132)	
Serum IgG $(g/L)$		$-0.023 \ (0.006)$	$0.977 \ (0.003)$	0.001	8.0	0.403(0.148)	
					30.0	0.293(0.141)	
Housing ventilation	Natural ventilation	Referent	Referent	0.015	Natural ventilation	0.296(0.242)	
	Other ventilation	$0.311 \ (0.294)$	2.218(0.646)		Other ventilation	$0.482 \ (0.355)$	
Average pTHI <sup>1</sup>		-0.009(0.003)	0.991 (0.003)	0.008	70.0	0.395(0.149)	
0.		()	()		20.0	0.291(0.154)	

<sup>1</sup>Temperature-humidity index (THI) accounts for the effects of temperature and relative humidity, and the equation uses the dry bulb temperature (T, °F) and the relative humidity (RH). The equation used for this analysis was THI =  $T - [0.55 - (0.55 \times \text{RH}/100)] \times (T - 58)$ .

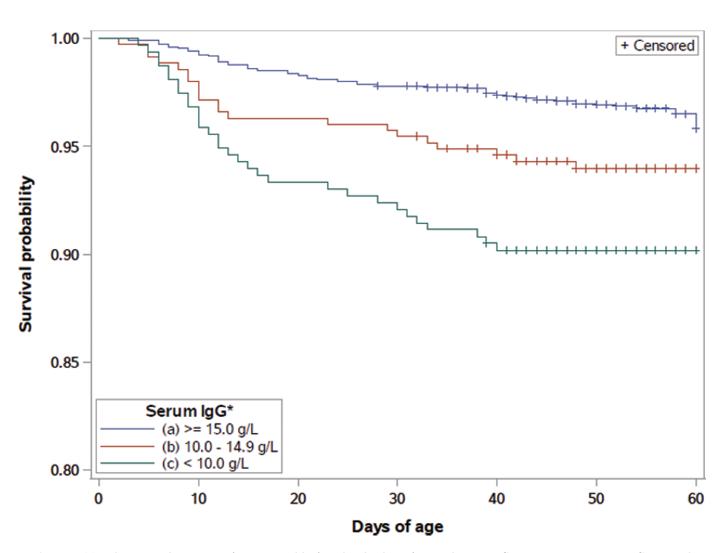


Figure 4. Mortality survival percentage for preveated heifer calves by days of age and serum IgG concentration categories. Corresponding to serum IgG levels of  $\geq$ 15.0 mg/dL, 10 to 14.9 mg/dL, and <10.0 mg/dL were serum total protein categories of  $\geq$ 5.4 g/dL, 5.1–5.3 g/dL, and <5.1 g/dL, and Brix score categories of  $\geq$ 8.6%, 8.1–8.5%, and <8.1%, respectively. Color version available online.

#### PREWEANED HEIFER MORBIDITY AND MORTALITY

	Mo	del results			Example model outcomes		
Final model variable	Variable level	β-Estimate (SE)	Odds ratio (SE)	Factor P-value	Example variable level	Predicted mortality risk (SE)	
Birth weight (kg)		-0.0553 (0.021)	$0.946\ (0.0198)$	0.011	$35.0 \\ 45.0$	$0.047 (0.202) \\ 0.023 (0.1714)$	
Serum IgG $(g/L)$		$-0.037 \ (0.013)$	$0.963\ (0.013)$	0.004	49.0 8.0 30.0	0.023 (0.1714) 0.052 (0.248) 0.020 (0.190)	
Liquid diet fat per day (kg)	$\leq 0.15$ 0.16-0.21	$\begin{array}{c} \text{Referent} \\ 0.049 \ (0.341) \end{array}$	$\begin{array}{c} \text{Referent} \\ 1.05 \ (0.341) \end{array}$	0.011	$\leq 0.15$ 0.16-0.21	$\begin{array}{c} 0.099 & (0.064) \\ 0.037 & (0.223) \end{array}$	
Morbidity	$\geq 0.22$ Any None	$\begin{array}{c} 1.103 \ (0.340) \\ 1.519 \ (0.259) \\ \text{Referent} \end{array}$	$\begin{array}{c} 3.01 \ (0.340) \\ 4.567 \ (0.219) \\ \text{Referent} \end{array}$	< 0.001	$\geq 0.22$ Any None	$\begin{array}{c} 0.035 \ (0.023) \\ 0.079 \ (0.058) \\ 0.018 \ (0.012) \end{array}$	

Table 8. Results of multivariable modeling of factors associated with mortality in heifer calves (n = 2,272)

pneumonia was 9.4%. A decrease in case-fatality rates is most likely due to routine management changes, the development of better treatment protocols, and the implementation of vaccinations for specific digestive and respiratory diseases (McGuirk, 2008).

Multivariable evaluation of management and health factors showed that low birth weight and serum IgG concentration were associated with disease and death. Additionally, housing ventilation and average THI during the preweaning period were associated with increased morbidity. Moreover, disease and the amount (kg) of fat per day in liquid were associated with increased mortality.

Calves born with higher birth weights were less likely to have clinical signs and less likely to die during the preveating period than calves with lower birth weights. Previous studies reported that calves with a birth weight that deviated from the mean (i.e., both heavy and light calves) were more likely to have digestive disease (Paré et al., 1993; Fallon et al., 1987). In the current study, calves were required to live beyond 24 h to be enrolled in the study; therefore, it is possible that calves with severe dystocia due to heavy birth weights that could be at a higher risk for morbidity and mortality were excluded from the study. Nonetheless, for low-birthweight calves, the majority of fetal growth is determined by the maternal uterine environment (Penrose, 1952). Thus, low birth weight could be due to a restricted supply of nutrients available to the developing fetus, which may occur due to maternal age and body condition at the time of conception and maternal nutrient supply during the gestation period (McCrabb et al., 1991; Bauer et al., 1998; McMillen et al., 2001; Osgerby et al., 2003a,b). Wolfenson et al. (1988) demonstrated that heat stress in dams resulted in calves with lower birth weight. Additionally, the size of the placenta may inhibit fetal growth, especially in late gestation, when maximal fetal growth occurs (McMillen et al., 2001; Mellor and Stafford, 2004). Low birth weight due to

intrauterine restrictions of nutrients and space may be a potentiating factor to increase morbidity and mortality in smaller calves. A post-uterine factor associated with low-birth-weight calves that may increase the likelihood of morbidity and mortality is a greater body surface-area-to-mass ratio. This greater ratio allows increased heat dissipation and increases energy requirements directly after birth, resulting in a compromised ability to acquire passive transfer (Paré et al., 1993; Mellor and Stafford, 2004; Monteiro et al., 2014). However, management interventions, such as drying off all calves at birth, providing dry housing, and immediately providing high-quality colostrum, may mitigate most environmental risk factors associated with morbidity and mortality in low-birth-weight calves. Nonetheless, ensuring that cows are healthy throughout the gestation period and providing high-quality nutrients will promote healthy newborns with decreased morbidity and mortality and increased lifelong production measures, such as fertility and milk production (Linden et al., 2009).

Although birth weight is a difficult variable to control on the farm, serum IgG, which was directly associated with morbidity and mortality, can be controlled through the implementation of a quality colostrum feeding program (DeNise et al., 1989; Donovan et al., 1998; Furman-Fratczak et al., 2011). Because calves are born agammaglobulinemic, they rely on consumption and absorption of maternal immunoglobulins via colostrum to fight infections, especially during the first few weeks of life (Gulliksen et al., 2008). Absorption of colostral immunoglobulins can be measured via serum IgG concentration. In this study, as serum IgG concentration increased, the likelihood of disease and death decreased. Several other studies support this result, demonstrating that calves with a serum IgG concentration >10g/L had decreased morbidity and mortality during the preweaning period (Arthington et al., 2000; Gulliksen et al., 2008; Godden et al., 2009). Calves with a serum IgG concentration <10 g/L are considered to have failure of passive transfer and are at a higher risk of disease and death. However, survival analyses of morbidity and mortality by serum IgG concentration in this study demonstrated that serum IgG concentration has a dose-dependent effect. Therefore, increasing the minimum serum IgG concentration target to 15 g/L may reduce morbidity and mortality, compared with the current 10 g/L industry standard. Additionally, serum total protein level and serum Brix score, which act as on-farm proxies for serum IgG concentration, also demonstrated a dose-dependent effect in the survival analyses for morbidity and mortality. Thus, if the industry standard for serum IgG concentration is increased, it is recommended to increase the target of serum total protein to 5.4 g/dL and serum Brix score to 8.6%.

Housing ventilation was another significant predictor of morbidity. Calves housed in ventilation systems other than natural ventilation were more likely to develop disease. Previous research associated enzootic pneumonia with poorly ventilated housing conditions, because poor ventilation can increase pathogen density (Nardell et al., 1991; Callan and Garry, 2002). Additionally, Lago et al. (2006) demonstrated that factors related to ventilation, including air changes per hour, were associated with the prevalence of calf respiratory disease, and decreased ventilation was associated with increased alleyway bacterial counts. Ultimately, increasing natural ventilation rates and providing a hygienic environment with decreased pathogen loads, regardless of the provided ventilation type, will help reduce morbidity, especially in regard to respiratory disease.

An increased average THI during the preweaning period was associated with decreased morbidity. The thermoneutral zone for a calf less than 1 mo of age is 10.0° to 26.6°C (Scibilia et al., 1987). Averaging THI throughout the calf's preweaning period allowed us to better account for seasonal variability across the United States. Because the THI was an average per month throughout each calf's preweaning period, extremely high or low temperatures outside the thermoneutral zone (and likely resulting in heat or cold stress) were not observed. However, past research demonstrated that both cold stress and heat stress affect the immune and stress responses of the body, specifically elevating pro-inflammatory cytokines, tumor necrosis factor- $\alpha$ , ACTH, and cortisol (Frank et al., 2003; Hangalapura et al., 2006; Dahl et al., 2016). Additionally, Godden et al. (2005) demonstrated that calves were more likely to receive treatment for clinical signs in the winter and summer months. Nonnecke et al. (2009) reported that respiratory disease and antibiotic costs were higher for

calves in cold environments. Therefore, minimizing cold stress and, to a degree, heat stress is extremely important in raising healthy calves throughout the preweaning period.

In this study, an increase in the amount (kg) of fat per day in the liquid diet resulted in a decrease in calf mortality. Research has demonstrated the importance of adipose tissue in mammalian survival—it provides energy, a source of heat and water, and thermal insulation (Young, 1976). Furthermore, early research regarding milk replacers in dairy calves demonstrated that higher fat replacers produced a lower incidence of scours and improved rates of gain (Lister, 1971; Leaver and Yarrow, 1972; Appleman and Owen, 1975). Thus, it would be expected that calves with limited fat intake would have lower energy levels available to fight infections, such as scours, and be at greater risk for cold and heat stress, and more likely to die during the preweaning period.

Last, morbidity status was highly correlated with mortality. Keeping calves healthy throughout the preweaning period is essential to decreasing overall mortality rates. However, it is important to recall that morbidity is affected by many other factors, such as birth weight, passive transfer, housing ventilation, and THI. Therefore, management techniques to decrease morbidity rates will ultimately help decrease mortality rates.

Although calving ease did not remain in the final morbidity and mortality models, dystocia is known to have a negative influence on calf health. Because calves were required to live beyond 24 h to be enrolled in the study, it is probable that calves born with any dystocia were less likely to be enrolled due to decreased thrift and risk of dying before 24 h of age. Additionally, most calves were tested for *Cryptosporidium* and *Giardia*; however, there was no association between the enteric protozoa and morbidity. For more information regarding *Cryptosporidium* and *Giardia* in preweaned dairy heifers, please see Urie et al. (2018b).

# CONCLUSIONS

Various management factors can help minimize morbidity and mortality on US dairy operations. The morbidity rate of 33.9% and mortality rate of 5.0% in this study suggest that US dairy operations have improved in terms of overall calf care. However, areas for improvement remain, including properly treating calves with antibiotics, feeding calves through scours events, and classifying causes of death. To continue reducing morbidity and mortality, the minimum serum IgG concentration target should be increased to 15 g/L compared with the current industry standard of 10 g/L. Additionally, more research should be conducted on factors that were shown to affect morbidity and mortality in this study, especially birth weight, housing ventilation, THI, and amount of fat fed per day in the liquid diet.

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## REFERENCES

- Agerholm, J. S., A. Basse, H. V. Krogh, K. Christensen, and L. Rønsholt. 1993. Abortion and calf mortality in Danish cattle herds. Acta Vet. Scand. 34:371–377.
- Appleman, R. D., and F. G. Owen. 1975. Breeding, housing, and feeding management. J. Dairy Sci. 58:447–464. https://doi.org/10 .3168/jds.S0022-0302(75)84588-7.
- Arthington, J. D., M. B. Cattell, and J. D. Quigley. 2000. Effect of dietary IgG source (colostrum, serum, or milk-derived supplement) on the efficiency of Ig absorption in newborn Holstein calves. J. Dairy Sci. 83:1463–1467. https://doi.org/10.3168/jds.S0022 -0302(00)75018-1.
- Bauer, R., B. Walter, A. Hoppe, E. Gaser, V. Lampe, E. Kauf, and U. Zwiener. 1998. Body weight distribution and organ size in newborn swine (Sus scrofa domestica)—A study describing an animal model for asymmetrical intrauterine growth retardation. Exp. Toxicol. Pathol. 50:59–65. https://doi.org/10.1016/S0940-2993(98)80071-7.
- Callan, R. J., and F. B. Garry. 2002. Biosecurity and bovine respiratory disease. Vet. Clin. North Am. Food Anim. Pract. 18:57–77.
- Carroll, J. A., and N. E. Forsberg. 2007. Influence of stress and nutrition on cattle immunity. Vet. Clin. North Am. Food Anim. Pract. 23:105–149.
- Chester-Jones, H., and P. C. Hoffman. 2003. Calf Nutrition: Raising Dairy Replacements. P. C. Hoffman, and R. Plourd, ed. Midwest Plan Service, Ames, IA.
- Constable, P. D. 2004. Antimicrobial use in the treatment of calf diarrhea. J. Vet. Intern. Med. 18:8–17.
- Dahl, G. E., S. Tao, and A. P. Monteiro. 2016. Effects of late-gestation heat stress on immunity and performance of calves. J. Dairy Sci. 99:3193–3198.
- Dairy Calf and Heifer Association. 2010. Gold Standards. Accessed Jul. 13, 2016. http://calfandheifer.org/gold\_standards/index.php.

- DeNise, S. K., J. D. Robison, G. H. Stott, and D. V. Armstrong. 1989. Effects of passive immunity on subsequent production in dairy heifers. J. Dairy Sci. 72:552–554.
- Dhama, K., R. S. Chauhan, M. Mahendran, and S. V. Malik. 2009. Rotavirus diarrhea in bovines and other domestic animals. Vet. Res. Commun. 33:1–23.
- Donovan, G. A., I. R. Dohoo, D. M. Montgomery, and F. L. Bennett. 1998. Associations between passive immunity and morbidity and mortality in dairy heifers in Florida, USA. Prev. Vet. Med. 34:31–46.
- Fallon, R. J., F. J. Harte, and D. Harrington. 1987. The effect of calf purchase weight, serum IG level and feeding systems on the incidence of diarrhoea, respiratory disease and mortality. Bov. Pract. 22:104–106.
- Frank, J. W., J. A. Carroll, G. L. Allee, and M. E. Zannelli. 2003. The effects of thermal environment and spray-dried plasma on the acute-phase response of pigs challenged with lipopolysaccharide. J. Anim. Sci. 81:1166–1176.
- Furman-Fratczak, K., A. Rzasa, and T. Stefaniak. 2011. The influence of colostral immunoglobulin concentration in heifer calves serum on their health and growth. J. Dairy Sci. 94:5536–5543.
- Garthwaite, B. D., J. K. Drackley, G. C. McCoy, and E. H. Jaster. 1994. Whole milk and oral rehydration solution for calves with diarrhea of spontaneous origin. J. Dairy Sci. 77:835–843.
- Godden, S. M., J. P. Fetrow, J. M. Feirtag, L. R. Green, and S. J. Wells. 2005. Economic analysis of feeding pasteurized nonsaleable milk versus conventional milk replacer to dairy calves. J. Am. Vet. Med. Assoc. 226:1547–1554.
- Godden, S. M., D. M. Haines, K. Konkol, and J. Peterson. 2009. Improving passive transfer of immunoglobulins in calves. II: Interactions between feeding method and volume of colostrum fed. J. Dairy Sci. 92:1758–1764. https://doi.org/10.3168/jds.2008-1847.
- Gulliksen, S. M., K. I. Lie, L. Sølverød, and O. Østerås. 2008. Risk factors associated with colostrum quality in Norwegian dairy cows. J. Dairy Sci. 91:704–712.
- Hangalapura, B. N., M. G. Kaiser, J. J. Poel, H. K. Parmentier, and S. J. Lamont. 2006. Cold stress equally enhances in vivo proinflammatory cytokine gene expression in chicken lines divergently selected for antibody responses. Dev. Comp. Immunol. 30:503–511.
- Heinrichs, A. J., G. W. Rogers, and J. B. Cooper. 1992. Predicting body weight and wither height in Holstein heifers using body measurements. J. Dairy Sci. 75:3576–3581.
- Hosmer, D. W., and S. Lemeshow. 1999. Applied Survival Analysis: Regression Modeling of Time to Event Data. John Wiley & Son Inc., New York, NY.
- Lago, A., S. M. McGuirk, T. B. Bennett, N. B. Cook, and K. V. Nordlund. 2006. Calf respiratory disease and pen microenvironments in naturally ventilated calf barns in winter. J. Dairy Sci. 89:4014– 4025. https://doi.org/10.3168/jds.S0022-0302(06)72445-6.
- Larson, R. L., and J. W. Tyler. 2005. Reducing calf losses in beef herds. Vet. Clin. North Am. Food Anim. Pract. 21:569–584.
- Leaver, J. D., and N. H. Yarrow. 1972. Rearing of dairy cattle: 3. A note on the use of whole milk for calves fed once daily. Ann. Sci. 15:315–318. https://doi.org/10.1017/S0003356100011594.
- Linden, T. C., R. C. Bicalho, and D. V. Nydam. 2009. Calf birth weight and its association with calf and cow survivability, disease incidence, reproductive performance, and milk production. J. Dairy Sci. 92:2580–2588. https://doi.org/10.3168/jds.2008-1603.
- Lister, E. E. 1971. Effects of heat treatment of skim-milk powder and levels of fat and protein in milk replacer diets on the growth of calves. Can. J. Anim. Sci. 51:735–742.
- Lorenz, I., B. Earley, R. Fallon, J. Gilmore, I. Hogan, E. Kennedy, and S. J. More. 2011. Calf health from birth to weaning. III. Disease prevention and management with particular reference to calf pneumonia. Ir. Vet. J. 64:9–10.
- McCrabb, G. J., A. R. Egan, and B. J. Hosking. 1991. Maternal undernutrition during mid-pregnancy in sheep. Placental size and its relationship to calcium transfer during late pregnancy. Br. J. Nutr. 65:157–168.

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- McGuirk, S. M. 2008. Disease management of dairy calves and heifers. Vet. Clin. North Am. Food Anim. Pract. 24:139–153.
- McMillen, I. C., M. B. Adams, J. T. Ross, C. L. Coulter, G. Simonetta, J. A. Owens, J. S. Robinson, and L. J. Edwards. 2001. Fetal growth restriction: Adaptations and consequences. Reproduction 122:195–204.
- Mellor, D. J., and K. J. Stafford. 2004. Animal welfare implications of neonatal mortality and morbidity in farm animals. Vet. J. 168:118–133.
- Monteiro, A. P., S. Tao, I. M. Thompson, and G. E. Dahl. 2014. Effect of heat stress during late gestation on immune function and growth performance of calves: isolation of altered colostral and calf factors. J. Dairy Sci. 97:6426–6439.
- Nardell, E. A., J. Keegan, S. A. Cheney, and S. C. Etkind. 1991. Airborne infection: Theoretical limits of protection achievable by building ventilation. Am. Rev. Respir. Dis. 144:302–306. https:// doi.org/10.1164/ajrccm/144.2.302.
- NOAA. 2016. Quality controlled local climatological data (QCLCD). Accessed Mar. 1, 2016. https://www.ncdc.noaa.gov/data-access/ land-based-station-data/land-based-datasets/quality-controlled -local-climatological-data-qclcd.
- Nonnecke, B. J., M. R. Foote, B. L. Miller, M. Fowler, T. E. Johnson, and R. L. Horst. 2009. Effects of chronic environmental cold on growth, health, and select metabolic and immunologic responses of preruminant calves1. J. Dairy Sci. 92:6134–6143.
- Osgerby, J. C., T. S. Gadd, and D. C. Wathes. 2003a. Effect of maternal body condition on placental and fetal growth and the insulinlike growth factor axis in Dorset ewes. Reproduction 125:717–731.
- Osgerby, J. C., T. S. Gadd, and D. C. Wathes. 2003b. The effects of maternal nutrition and body condition on placental and foetal growth in the ewe. Placenta 24:236–247.
- Paré, J., M. C. Thurmond, I. A. Gardner, and J. P. Picanso. 1993. Effect of birthweight, total protein, serum IgG and packed cell volume on risk of neonatal diarrhea in calves on two California dairies. Can. J. Vet. Res. 57:241–246.
- Penrose, L. S. 1952. Data on the genetics of birth weight. Ann. Eugen. 16:378–381.
- Perez, E., J. P. T. M. Noordhuizen, L. A. van Wuijkhuise, and E. N. Stassen. 1990. Management factors related to calf morbidity and mortality rates. Livest. Prod. Sci. 25:79–93.
- Scibilia, L. S., L. D. Muller, R. S. Kensinger, T. F. Sweeney, and P. R. Shellenberger. 1987. Effect of environmental temperature and dietary fat on growth and physiological responses of newborn calves. J. Dairy Sci. 70:1426–1433.
- Shivley, C. B., J. E. Lombard, N. J. Urie, D. M. Haines, R. Sargent, C. A. Kopral, T. J. Earleywine, J. D. Olson, and F. B. Garry. 2018. Preweaned heifer management on US dairy operations: Part II. Factors associated with colostrum quality and passive transfer status of dairy heifer calves. J. Dairy Sci. 101:9185–9198. https:// doi.org/10.3168/jds.2017-14008.
- Sivula, N. J., T. R. Ames, W. E. Marsh, and R. E. Werdin. 1996. Descriptive epidemiology of morbidity and mortality in Minnesota dairy heifer calves. Prev. Vet. Med. 27:155–171.
- Stanton, A. L., D. F. Kelton, S. J. LeBlanc, J. Wormuth, and K. E. Leslie. 2012. The effect of respiratory disease and a preventative antibiotic treatment on growth, survival, age at first calving, and milk production of dairy heifers. J. Dairy Sci. 95:4950–4960.
- Steele, A. D., A. Geyer, and G. H. Gerdes. 2004. Rotavirus infections. Pages 1256–1264 in Infectious Diseases of Livestock. 2nd ed. Oxford University Press, Oxford, UK.
- Svensson, C., K. Lundborg, U. Emanuelson, and S.-O. Olsson. 2003. Morbidity in Swedish dairy calves from birth to 90 days of age and

individual calf-level risk factors for infectious diseases. Prev. Vet. Med.  $58{:}179{-}197.$ 

- Swali, A., and D. C. Wathes. 2006. Influence of the dam and sire on size at birth and subsequent growth, milk production and fertility in dairy heifers. Theriogenology 66:1173–1184.
- Tyler, J. W., D. D. Hancock, J. G. Thorne, C. C. Gay, and J. M. Gay. 1999. Partitioning the mortality risk associated with inadequate passive transfer of colostral immunoglobulins in dairy calves. J. Vet. Intern. Med. 13:335–337.
- Urie, N. J., J. E. Lombard, C. B. Shivley, A. E. Adams, C. A. Kopral, and M. Santin. 2018b. Preweaned heifer management on US dairy operations: Part III. Factors associated with *Cryptosporidium* and *Giardia* in preweaned dairy heifer calves. J. Dairy Sci. 101:9199– 9213. https://doi.org/10.3168/jds.2017-14060.
- Urie, N. J., J. E. Lombard, C. B. Shivley, C. A. Kopral, A. E. Adams, T. J. Earleywine, J. D. Olson, and F. B. Garry. 2018a. Preweaned heifer management on US dairy operations: Part I. Descriptive characteristics of preweaned heifer raising practices. J. Dairy Sci. 101:9168–9184. https://doi.org/10.3168/jds.2017-14010.
- USDA. 1994. National dairy health evaluation project. Dairy heifer morbidity, mortality, and health management focusing on preweaned heifers. USDA National Animal Health Monitoring System (NAHMS). Accessed Jul. 21, 2016. https://www.aphis.usda .gov/animal\_health/nahms/dairy/downloads/ndhep/NDHEP\_dr \_Report.pdf.
- USDA. 2002. Dairy 2002. Part 1: Reference of dairy health and management in the United States. USDA National Animal Health Monitoring System (NAHMS). Accessed Nov. 29, 2016. https:// www.aphis.usda.gov/animal\_health/nahms/dairy/downloads/ dairy02/Dairy02\_dr\_PartI.pdf.
- USDA. 2010. Heifer calf health and management practices on U.S. Dairy Operations, 2007. USDA–APHIS-VS-CEAH-NAHMS. Fort Collins, CO. Accessed Jun. 15, 2016. https://www.aphis.usda .gov/animal\_health/nahms/dairy/downloads/dairy07/Dairy07\_ir \_CalfHealth.pdf.
- USDA. 2016. Dairy 2014: Dairy Cattle Management Practices in the United States, 2014. USDA–APHIS-VS-CEAH-NAHMS. Fort Collins, CO. Accessed Jul. 21, 2016. https://www.aphis.usda.gov/ animal\_health/nahms/dairy/downloads/dairy14/Dairy14\_dr \_PartI.pdf.
- Virtala, A. M., G. D. Mechor, Y. T. Gröhn, and H. N. Erb. 1996. Morbidity from nonrespiratory diseases and mortality in dairy heifers during the first three months of life. J. Am. Vet. Med. Assoc. 208:2043–2046.
- Waltner-Toews, D., S. W. Martin, and A. H. Meek. 1986. Dairy calf management, morbidity and mortality in Ontario Holstein herds. II. Age and seasonal patterns. Prev. Vet. Med. 4:125–135.
- Wells, S. J., D. A. Dargatz, and S. L. Ott. 1996. Factors associated with mortality to 21 days of life in dairy heifers in the United States. Prev. Vet. Med. 29:9–19.
- Wells, S. J., L. P. Garber, and G. W. Hill. 1997. Health status of preweaned dairy heifers in the United States. Prev. Vet. Med. 29:185–199.
- Wiseman, A., I. Selman, H. Pirie, and I. Harvey. 1976. Pneumonia in single-suckled calves. Vet. Rec. 98:409–410.
- Wolfenson, D., I. Flamenbaum, and A. Berman. 1988. Dry period heat stress relief effects on prepartum progesterone, calf birth weight, and milk production. J. Dairy Sci. 71:809–818.
- Young, R. A. 1976. Fat, energy and mammalian survival. Am. Zool. 16:699–710. https://doi.org/10.1093/icb/16.4.699.