CASE REPORT

http://dx.doi.org/10.5653/cerm.2011.38.4.238 pISSN 2233-8233 • eISSN 2233-8241 Clin Exp Reprod Med 2011;38(4):238-241



Duplication of intrachromosomal insertion segments 4q32→q35 confirmed by comparative genomic hybridization and fluorescent *in situ* hybridization

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A 35-year-old man with infertility was referred for chromosomal analysis. In routine cytogenetic analysis, the patient was seen to have additional material of unknown origin on the terminal region of the short arm of chromosome 4. To determine the origin of the unknown material, we carried out high-resolution banding, comparative genomic hybridization (CGH), and FISH. CGH showed a gain of signal on the region of $4q32 \rightarrow q35$. FISH using whole chromosome painting and subtelomeric region probes for chromosome 4 confirmed the aberrant chromosome as an intrachromosomal insertion duplication of $4q32 \rightarrow q35$. Duplication often leads to some phenotypic abnormalities; however, our patient showed an almost normal phenotype except for congenital dysfunction in spermatogenesis.

Keywords: Chromosome 4; Insertion 4p; Duplication 4q; Comparative genomic hybridization; Fluorescent in situ hybridization; Human

Introduction

Insertions occur when a single chromosome breaks at 2 places and a piece of the chromosome is reinserted elsewhere. The piece of chromosome may be reinserted into a different location in the original chromosome (intrachromosomal insertion) or it may be reinserted into a different chromosome (interchromosomal insertion). Intrachromosomal insertion can occur on the same side of the centromere (paracentric) or on the opposite side of the centromere (pericentric).

Intrachromosomal insertions are very rare forms of chromosomal rearrangements; about 40 cases have been published [1-4]. Balanced carriers of intrachromosomal insertions are phenotypically normal, except a case report of cri-du-chat syndrome [5]. On the other hand,

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they are at risk for unbalanced meiotic segregation due to crossing over, and the frequency of a child having an unbalanced karyotype is about 15-50% [2].

Patients with trisomy 4q syndrome have variable clinical features, often including growth and developmental delays, mental retardation, and a dysmorphic appearance [6]. Pure partial trisomy of 4q may contribute significantly to the clarification of region specific phenotypes, but very few cases have been reported. In this study, we present an intrachromosomal insertion duplication of chromosome 4q that was detected in an infertile man with non-obstructive azoospermia.

Case report

A 35-year-old man with infertility was referred for chromosomal analysis. His height was 175 cm and weight was 63 kg. The results of semen analysis according to the guidelines of the World Health Organization [7] showed azoospermia. The histological examination of the testicular tissue revealed spermatogenic arrest in meiosis, confirming the diagnosis of non-obstructive azoospermia. He was diag-



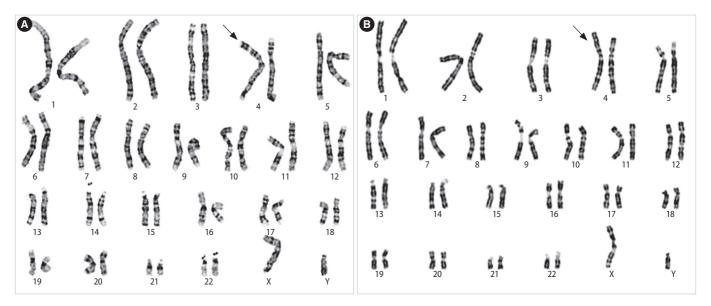


Figure 1. Karyotype of the patient. (A) Giemsa (GTG) and (B) reverse (RBG) banding. The arrow is an abnormal chromosome 4, which has an additional material of unknown origin on the terminal region of the short arm (4p).

nosed with sertoli-cell-only syndrome because all the seminiferous tubules contained only Sertoli cells and did not contain germ cells. He was psychologically normal. Chromosome analysis was performed using Giemsa banding according to usual procedures. The patient was seen to have additional material of unknown origin on the terminal region of the short arm of chromosome 4 (4p). The additional material was relatively large in size with two dark bands.

To confirm the origin of the unknown material, we carried out high-resolution banding, comparative genomic hybridization (CGH) and FISH. High-resolution GTG and RBG banding analysis showed an unbalanced karyotype with additional material attached to the terminal band of 4p16 (Figure 1). However, we could not estimate the origin of the extra segment or the type of rearrangement. DNA was extracted from the peripheral blood of the patient and used in a CGH method. CGH revealed a gain of material on the region of 4q32-35 (Figure 2). FISH was carried out with multiple probes for chromosome 4, including the whole chromosome painting (wcp) probe, telomeric probes for 4p (D4S3359)/4q (D4S2930), and the Wolf-Hirschhorn region (D4S96)/centromeric (D4Z1) probe (Vysis, Downers Grove, IL, USA).

For each analysis, a minimum of ten metaphase and 100 interphase cells were scored. The whole chromosome 4 painting probe hybridized on the entire chromosome of the abnormal chromosome 4 including the additional material (Figure 3A). Using telomeric probes, the presence of 4p and 4q telomeric regions was found on derivative chromosome 4 (Figure 3B). The Wolf Hirschhorn region (4p16.3) and centromeric probe revealed the presence of two signals on derivative chromosome 4 (Figure 3C). We concluded that the additional material of the aberrant chromosome was an intrachromosomal insertion

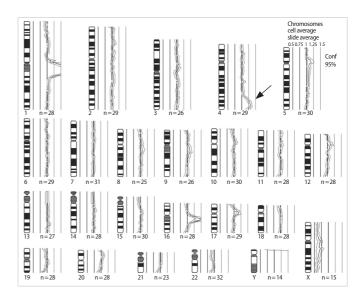


Figure 2. Characterization of the additional material using comparative genomic hybridization. The arrow denotes the gain of chromosome region $4q32 \rightarrow q35$.

duplication of 4q32-35; 46,XY,add(4)(p16.3).ish der(4)dup(4)(q32q35) ins(4)(p16.3q32q35)(wcp+,D4S3359+,D4S2930+,D4S96+,D4Z1+).

Discussion

Trisomy 4q syndrome was a rare condition, first described in 1971 [8]. The clinical findings are frequently presented including mental retardation, developmental delays, and multiple abnormalities such as microcephaly, acrocephaly, as well as malformed ears, a high/bro-ad/depressed nasal bridge, and teeth and thumb anomalies. Geno-

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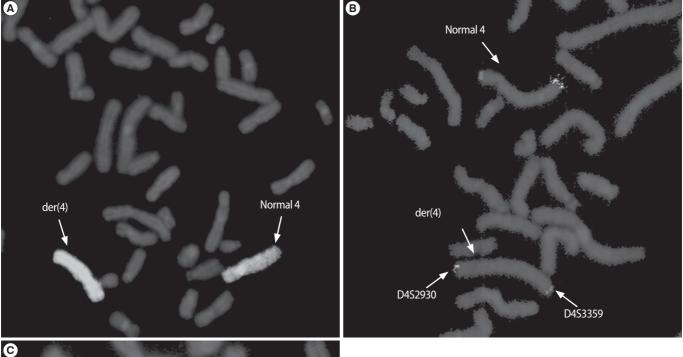


Figure 3. Results of FISH. (A) FISH with the whole chromosome painting probe for chromosome 4 showed additional material attached on the entire body of the derivative chromosome 4, (B) FISH using telomeric 4p (D4S3359) and 4q (D4S2930) probes confirmed two signals on both ends of derivative chromosome 4, and (C) FISH with chromosome 4p16.3 (D4S96) and 4 centromeric (D4Z1) probes showed two signals on derivative chromosome 4.

type-phenotype correlation was difficult because of partial chromosome monosomy associated with 4q trisomy.

One boy with a duplication of 4q12-q13 was reported to have relatively few features, merely including microcephaly and mental retardation [9]. Likewise, the additional duplicated material from 4q25-q28 seems to produce no major clinical effects. The segment 4q27-4q31 seemed to be preferentially engaged in the trisomy 4q syndrome with severe clinical effects including growth retardation, mental retardation, microcephaly, facial asymmetry, thumb anomalies, hearing impairment, epilepsy, and a congenital heart defect [10]. Two fa-

milial cases of a small region duplication of 4q31.1-q32.3 and 4q31.3-q33 showed minimal clinical features [11,12]. Additional duplicated 4q34 material seems to be without major clinical effects, but telomeric band 4q35 may be involved in the development of microcephaly and severe mental and growth retardation [13]. Our patient with pure trisomy 4q32-35 was phenotypically normal except for azoospermia.

We wanted to confirm whether the meiotic segregation error appears in carriers with intrachromosomal insertion of chromosome 4 or de novo. Unfortunately, the chromosomal analysis of the parents was not available. However, we predicted that one of the parents



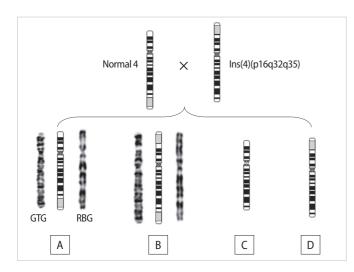


Figure 4. Diagrammatic representation for gamete production following a recombination between the normal 4 and the abnormal 4 with ins(4)(p16q32q35). There are four types of gametes: (A) normal 4, (B) the abnormal 4 of the present patient, der(4)dup(4q)ins(4)(p16q32q35), (C) del(4)(q32q35), and (D) ins(4)(p16q32q35).

might have a ins(4)(p16.3q32q35) because the der(4)dup(4q)ins(4) (p16.3q32q35) was a type of possible recombinant from pairing and crossing over during meiosis in carriers with the pericentric intrachromosomal insertion of chromosome 4 (Figure 4). Also, the risk of having a child with an unbalanced chromosome aberration from a carrier depends on both the type and size of the insertion. Individuals with a pericentric insertion are at a greater risk of having unbalanced offspring, but those with a paracentric insertion are at a relatively low risk.

To investigate rapidly whether the karyotype is balanced or unbalanced, CGH and FISH analyses can be a useful diagnostic tool. These techniques have been particularly useful in achieving an even higher resolution, such that small alterations (< 3 Mb) in DNA copy number or location can be detected. Recently, array CGH analysis has been applied to the clinical diagnosis of chromosomal abnormalities and genomic aberrations [14].

In conclusion, we reported a case with an unbalanced karyotype of pure trisomy 4q32-35 which showed a normal phenotype except for spermatogenesis, though duplication often leads to some phenotypic abnormalities. Identification of the insertion is difficult. In this case, molecular cytogenetic analysis helped us in the precise detection. The precise characterization of an abnormal chromosome is clinically important for genetic counseling.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

References

- Farrell SA, Chow G. Intrachromosomal insertion of chromosome 7. Clin Genet 1992;41:299-302.
- 2. Madan K, Menko FH. Intrachromosomal insertions: a case report and a review. Hum Genet 1992;89:1-9.
- Ardalan A, Prieur M, Choiset A, Turleau C, Goutieres F, Girard-Orgeolet S. Intrachromosomal insertion mimicking a pericentric inversion: molecular cytogenetic characterization of a three break rearrangement of chromosome 20. Am J Med Genet A 2005; 138A:288-93.
- 4. Lybaek H, Orstavik KH, Prescott T, Hovland R, Breilid H, Stansberg C, et al. An 8.9 Mb 19p13 duplication associated with precocious puberty and a sporadic 3.9 Mb 2q23.3q24.1 deletion containing NR4A2 in mentally retarded members of a family with an intrachromosomal 19p-into-19q between-arm insertion. Eur J Hum Genet 2009;17:904-10.
- South ST, Swensen JJ, Maxwell T, Rope A, Brothman AR, Chen Z. A new genomic mechanism leading to cri-du-chat syndrome. Am J Med Genet A 2006;140:2714-20.
- 6. Cui YX, Wang YH, Hao LJ, Hou L, Li W, Huang YF. Partial trisomy 4q: a case report. Chin Med J (Engl) 2006;119:1136-9.
- World Health Organization. WHO laboratory manual for the examination of human semen and sperm–cervical mucus interactions. 4th ed. Cambridge (UK): Cambridge University Press; 1999.
- 8. Hoehn H, Sander C, Sander LZ. Aneusomie de recombinaison: rearrangement between paternal chromosomes 4 and 18 yielding offspring with features of the 18q-syndrome. Ann Genet 1971; 14:187-92.
- 9. Shashi V, Berry MN, Santos C, Pettenati MJ. Partial duplication of 4q 12q13 leads to a mild phenotype. Am J Med Genet 1999;86: 51-3.
- Lundin C, Zech L, Sjörs K, Wadelius C, Annerén G. Trisomy 4q syndrome: presentation of a new case and review of the literature. Ann Genet 2002;45:53-7.
- 11. Goodman BK, Capone GT, Hennessey J, Thomas GH. Familial tandem duplication of bands q31.1 to q32.3 on chromosome 4 with mild phenotypic effect. Am J Med Genet 1997;73:119-24.
- 12. Maltby EL, Barnes IC, Bennett CP. Duplication involving band 4q32 with minimal clinical effect. Am J Med Genet 1999;83:431.
- 13. Elghezal H, Sendi HS, Monastiri K, Lapierre JM, Romdhane SI, Mougou S, et al. Large duplication 4q25-q34 with mild clinical effect. Ann Genet 2004;47:419-22.
- Xiang B, Li A, Valentin D, Nowak NJ, Zhao H, Li P. Analytical and clinical validity of whole-genome oligonucleotide array comparative genomic hybridization for pediatric patients with mental retardation and developmental delay. Am J Med Genet A 2008; 146A:1942-54.

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