



Draft Genome Sequences of *Micrococcus luteus* MFP06 and MFP07, Isolated from the Skin of Healthy Volunteers

Djouhar Souak,^a  Amine M. Boukerb,^a Magalie Barreau,^a Cecile Duclairoir-Poc,^a  Marc G. J. Feuilloley^a

^aLaboratoire de Microbiologie, Signaux et Microenvironnement (LMSM) EA412, Université de Rouen Normandie, Evreux, France

ABSTRACT We report the draft genome sequences of two *Micrococcus luteus* strains, MFP06 and MFP07, isolated from human skin. The genome assemblies consist of 2,480 and 2,417 kbp with 2,337 and 2,240 coding sequences, respectively. The genomes contain genes potentially involved in osmotic stress tolerance, DNA repair, monoacylglycerol hydrolysis, and beta-lactone synthesis.

Micrococcus luteus is a high-GC-content, Gram-positive, strictly aerobic coccus typically occurring in tetrads and phylogenetically affiliated with the family *Micrococcaceae* in the phylum *Actinobacteria*. This bacterium is found in environments such as soil (1), air (2), and human skin (3, 4). *M. luteus* is also known as an opportunistic pathogen involved in severe infections such as meningitis and septic shock in immunocompromised patients with reported antibioresistance (5). Despite this opportunistic behavior, little is known about its role within the skin microbiome.

M. luteus strains MFP06 and MFP07 were collected under the control of the Bio-EC CRO (Longjumeau, France) and according to the French and European ethical directives (ARS Biomedical Research Agreement 2012-12-010, Bioethics Agreement DC-2008-542) (6). These strains were isolated by swabbing the right antecubital fossa of an adult woman (50 to 65 years old) and the right scapula of an adult man (50 to 65 years old), respectively. Bacterial colonies cultured on tryptic soy agar (TSA) at 37°C were identified as *M. luteus* by analysis of their total proteome using an Autoflex III matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometer coupled to the MALDI-Biotyper 3.0 algorithmic system (Bruker, Marcy-l'Étoile, France) (6). For genome sequencing, DNA was extracted from an overnight culture in tryptic soy broth (TSB) at 37°C with a genomic extraction kit (GeneJET genomic DNA purification kit, catalog number K0721; Thermo Scientific) following the supplied procedure, with pretreatment using a lysis solution (20 mM Tris-HCl [pH 8.0], 2 mM EDTA, 1.2% Triton X-100, and 20 mg/ml lysozyme). Library preparation and sequencing were conducted at the LMSM genomics platform (LMSM Evreux, University of Rouen Normandy). Briefly, libraries were prepared with the Nextera XT DNA sample preparation kit (Illumina, USA) and sequenced on an Illumina MiSeq system (MiSeq reagent kit v.3, 600 cycles), generating 1,604,848 and 2,745,926 high-quality raw paired-end (PE) 250-bp reads, respectively.

All bioinformatic tools were used with default parameters unless otherwise stated. Reads were quality screened and trimmed with FastQC v.0.11.8 (7) and Trim Galore v.0.6.2 (8), respectively. Genome assembly was conducted using Unicycler v.0.4.7 (9), and sequences were assessed for contamination with CheckM v.1.1.2 (10). Automated gene predictions and functional annotations were performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (11). Coding genes for proteins possibly involved in secondary metabolite production were identified with antiSMASH v.5.1.2 (12). The draft genomes of MFP06 and MFP07 consisted of 2,480,672 and 2,417,497 bp in 221 and 165 contigs (N_{50} , 19,847 and 30,452) with 147.6 and 219.4× mean coverage, respectively. The GC content for both draft genomes was 72.96%. There are 2,337 and

Citation Souak D, Boukerb AM, Barreau M, Duclairoir-Poc C, Feuilloley MJG. 2020. Draft genome sequences of *Micrococcus luteus* MFP06 and MFP07, isolated from the skin of healthy volunteers. *Microbiol Resour Announc* 9:e00545-20. <https://doi.org/10.1128/MRA.00545-20>.

Editor Irene L. G. Newton, Indiana University, Bloomington

Copyright © 2020 Souak et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Amine M. Boukerb, amine.boukerb@univ-rouen.fr, or Marc G. J. Feuilloley, marc.feuilloy@univ-rouen.fr.

Received 12 May 2020

Accepted 24 May 2020

Published 18 June 2020

2,240 protein-coding genes, 51 and 52 tRNA genes, 1 copy each of 5S, 16S, and 23S rRNA genes, and 36 and 34 insertion sequence (IS) elements, respectively. Genes coding for monoacylglycerol lipases involved in lipid metabolism (13), L-ectoine synthase, which helps resist shifts in salt concentration (14), and a biosynthetic gene cluster (BGC) coding for a beta-lactone (15) were also detected. Future detailed analysis of these loci and the genomic characterization of these strains will provide further information about adaptation and success of *M. luteus* on human skin.

Data availability. Sample information, genomic assembly and annotation, and raw sequences are accessible under the NCBI BioProject number [PRJNA626598](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA626598). The whole-genome shotgun (WGS) projects are available under the GenBank accession numbers [JABBWS000000000](https://www.ncbi.nlm.nih.gov/genbank/JABBWS000000000) and [JABBWT000000000](https://www.ncbi.nlm.nih.gov/genbank/JABBWT000000000), and the SRA accession numbers are [SRR11574543](https://www.ncbi.nlm.nih.gov/sra/SRR11574543) and [SRR11574542](https://www.ncbi.nlm.nih.gov/sra/SRR11574542).

ACKNOWLEDGMENTS

We acknowledge the contribution of the Normandy Network of Microbial Metagenomics (RNAMB), supported by Région Normandie and the European Union (FEDER). This work was supported by grants from Evreux Portes de Normandie and the FUI SKIN O FLOR research program. D.S. is the recipient of a doctoral grant financed by the ANRT.

The LMSM is a member of the industrial cluster Cosmetic Valley and of CNRS GDR 3711 Cosm'Actifs.

REFERENCES

1. Sims GK, Sommers LE, Konopka A. 1986. Degradation of pyridine by *Micrococcus luteus* isolated from soil. *Appl Environ Microbiol* 51:963–968. <https://doi.org/10.1128/AEM.51.5.963-968.1986>.
2. Kutmutia SK, Drautz-Moses DI, Uchida A, Purbojati RW, Wong A, Kushwaha KK, Putra A, Premkrishnan BNV, Heinle CE, Vettath VK, Junqueira ACM, Schuster SC. 2019. Complete genome sequence of *Micrococcus luteus* strain SGAir0127, isolated from indoor air samples from Singapore. *Microbiol Resour Announc* 8:381e00646-19. <https://doi.org/10.1128/MRA.00646-19>.
3. Kloos WE, Musselwhite MS. 1975. Distribution and persistence of *Staphylococcus* and *Micrococcus* species and other aerobic bacteria on human skin. *Appl Microbiol* 30:381–385. <https://doi.org/10.1128/AEM.30.3.381-395.1975>.
4. Kloos WE, Tornabene TG, Schleifer KH. 1974. Isolation and characterization of *Micrococci* from human skin, including two new species: *Micrococcus lylae* and *Micrococcus kristinae*. *Int J Syst Bacteriol* 24:79–101. <https://doi.org/10.1099/00207713-24-1-79>.
5. Eady EA, Coates P, Ross JI, Ratyal AH, Cove JH. 2000. Antibiotic resistance patterns of aerobic coryneforms and furazolidone-resistant Gram-positive cocci from the skin surface of the human axilla and fourth toe cleft. *J Antimicrob Chemother* 46:205–213. <https://doi.org/10.1093/jac/46.2.205>.
6. Hillion M, Mijouin L, Jaouen T, Barreau M, Meunier P, Lefeuvre L, Lati E, Chevalier S, Feuilloley MG. 2013. Comparative study of normal and sensitive skin aerobic bacterial populations. *Microbiologyopen* 2:953–961. <https://doi.org/10.1002/mbo3.138>.
7. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>.
8. Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet J* 17:10–12. <https://doi.org/10.14806/ej.17.1.200>.
9. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
10. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
11. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
12. Blin K, Shaw S, Steinke K, Villebro R, Ziemert N, Lee SY, Medema MH, Weber T. 2019. antiSMASH 5.0: updates to the secondary metabolite genome mining pipeline. *Nucleic Acids Res* 47:W81–W87. <https://doi.org/10.1093/nar/gkz310>.
13. Vromman F, Subtil A. 2014. Exploitation of host lipids by bacteria. *Curr Opin Microbiol* 17:38–45. <https://doi.org/10.1016/j.mib.2013.11.003>.
14. Young M, Artsatbanov V, Beller HR, Chandra G, Chater KF, Dover LG, Goh EB, Kahan T, Kaprelyants AS, Kyrpides N, Lapidus A, Lowry SR, Lykidis A, Mahillon J, Markowitz V, Mavromatis K, Mukamolova GV, Oren A, Rokem JS, Smith MC, Young DI, Greenblatt CL. 2010. Genome sequence of the Fleming strain of *Micrococcus luteus*, a simple free-living actinobacterium. *J Bacteriol* 192:841–860. <https://doi.org/10.1128/JB.01254-09>.
15. Robinson SL, Christenson JK, Wackett LP. 2019. Biosynthesis and chemical diversity of beta-lactone natural products. *Nat Prod Rep* 36:458–475. <https://doi.org/10.1039/c8np00052b>.